

Review

Status of β_1 -Adrenoceptor Signal Transduction System in Cardiac Hypertrophy and Heart Failure

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Academic Editors: Zoltán Papp and Maurizio Pieroni

Submitted: 16 March 2023 Revised: 30 May 2023 Accepted: 15 June 2023 Published: 21 September 2023

Abstract

Although β_1 -adrenoceptor (β_1 -AR) signal transduction, which maintains cardiac function, is downregulated in failing hearts, the mechanisms for such a defect in heart failure are not fully understood. Since cardiac hypertrophy is invariably associated with heart failure, it is possible that the loss of β_1 -AR mechanisms in failing heart occurs due to hypertrophy at 4 and 24 weeks after inducing pressure overload as well as adaptive cardiac hypertrophy and heart failure at 4 and 24 weeks after inducing volume overload, respectively. Varying degrees of alterations in β_1 -AR density as well as isoproterenol-induced increases in cardiac function, intracellular Ca²⁺-concentration in cardiomyocytes and adenylyl cyclase activity in crude membranes have been reported under these hypertrophic conditions. Adaptive hypertrophy at 4 weeks of pressure or volume overload showed unaltered or augmented increases in the activities of different components of β_1 -AR signaling. On the other hand, maladaptive hypertrophy due to pressure overload and heart failure due to volume overload at 24 weeks revealed depressions in the activities of β_1 -AR signal transduction pathway. These observations provide evidence that β_1 -AR signal system is either unaltered or upregulated in adaptive cardiac hypertrophy and downregulated in maladaptive cardiac hypertrophy or heart failure. Furthermore, the information presented in this article supports the concept that downregulation of β_1 -AR mechanisms in heart failure or maladaptive cardiac hypertrophy is not due to hypertrophy and downregulated in adaptive cardiac hypertrophy is not due to hypertrophy and downregulated in maladaptive cardiac hypertrophy is not due to hypertrophy and downregulation of β_1 -AR mechanisms in heart failure or maladaptive cardiac hypertrophy is not due to hypertrophy and downregulation of β_1 -AR mechanisms in heart failure or maladaptive cardiac hypertrophy is not due to hypertrophy and downregulation in complex mechanisms in heart failure or mal

Keywords: adaptive cardiac hypertrophy; maladaptive cardiac hypertrophy; heart failure; β_1 -adrenoceptors; intracellular Ca²⁺; adenylyl cyclase; cardiac function

1. Introduction

In Canada, more than 100,000 patients with heart failure are diagnosed annually and about 2.6 million adults aged 20 and over are living with this heart disease. Since heart failure is one of the top reasons for hospitalization, the associated healthcare costs have been estimated to reach \$2.8 billion by 2030 in this country [1-4]. However, it should be pointed out that significant advances have been made for the development of medical therapies, which are used for the treatment of this disease. Several interventions have reduced morbidity, mortality, and economic burden of this devastating disorder, and in fact a great deal of effort is being made to further improve its pharmacotherapy [5-11]. Although extensive research is also being done to understand the pathogenesis of heart failure, the exact mechanism for its progression remains unclear at present [12–18]. Nonetheless, it is evident that heart failure is a complex problem, which is associated with different disorders such as cardiac dysfunction, cardiac arrhythmias, loss of adrenergic support, exercise intolerance and fluid retention. Since a number of vasoactive hormones are elevated in heart failure, several hormone receptor antagonists are now available for its therapy. In this regard, guanine nucleotide protein coupled receptors (GPCRs) have been identified as the most promising targets for drug discovery and a few of their blockers have been shown to exert beneficial effects in heart failure [19–24].

It is noteworthy that β -adrenergic receptors (β -AR) are the most prominent class of GPCRs, which along with their modulators, are shown to play a critical role in cardiac health and disease [25–38]. Since alterations in β -AR mechanisms are reported in heart failure, these targets have been manipulated to achieve clinically relevant therapies [39–42]. Furthermore, attenuated responses of the heart to sympathetic stimulation have been observed at different stages of heart failure [28,43–45]. The activities of various



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components of β_1 -AR system are unaltered, upregulated, or downregulated in different types of heart failure [42,46]. Since cardiac hypertrophy is generally associated with development of heart failure [23,47-52], it is not clear whether upregulation or downregulation of β_1 -AR mechanisms are involved in adaptive or maladaptive cardiac hypertrophy [53-58]. In this article, we have briefly reviewed the role of β_1 -AR signaling activation in the regulation of cardiac function upon stimulation of the sympathetic nervous system (SNS). Furthermore, the status of this system in the development of cardiac hypertrophy and heart failure is discussed. We have also reviewed the evidence regarding β_1 -AR signal alterations in adaptive and maladaptive cardiac hypertrophy due to pressure overload. In addition, some observations regarding changes in β_1 -AR mechanisms in adaptive hypertrophy and heart failure due to volume overload are described to evaluate the role of hypertrophic process in heart failure.

2. Role of β -AR Signal Transduction in Cardiac Function

It is now well known that stimulation of β_1 -AR signal transduction by activation of the SNS or exogenous catecholamines for a short duration augments cardiac function and produces cardiac hypertrophy whereas its stimulation for a prolonged period results in heart failure. Furthermore, several β_1 -AR blockers have been reported to exert cardiodepressant action under physiological conditions but improve cardiac function in heart failure [27-31,34-41,59–63]. The activation of β_1 -AR stimulates adenylyl cyclase activity to form 3'-5'-cyclic adenosine monophosphate (cyclic AMP) in the myocardium. The elevated level of cyclic AMP promotes protein kinase A (PKA)-mediated phosphorylations of different Ca²⁺-handling proteins in the sarcolemma and sarcoplasmic reticulum for increasing the intracellular concentration of Ca^{2+} ($[Ca^{2+}]_i$) and producing positive inotropic effect in the heart [39,64-69]. The increased activation of β_1 -AR signal transduction is considered to provide circulatory support during early stages of heart failure [70-73] but prolonged stimulation triggers β_1 -AR desensitization in the failing heart [42,55,69,74–80]. Such changes due to elevated levels of circulatory catecholamines or prolonged stimulation of β_1 -AR system are associated with worsening cardiac outcome, cardiac dysfunction and sudden cardiac death [41,63,81-86].

It is pointed out that the β -AR family in healthy human heart comprises subtypes that include 80% β_1 -AR, 20% β_2 -AR and about 3% β_3 -AR [22,87–89]. β_1 -AR subtype displays localization in the sarcolemma in the heart whereas β_2 -AR and β_3 -AR subtypes are mainly confined to the Ttubular network [90,91]. The density of β_1 -ARs is reduced by about 50% depending upon the severity of heart failure, whereas the β_2 -AR density remains unchanged. A substantial reduction in β_1 -AR receptor density in heart failure has been shown to be due to downregulation of these receptors [44,71,72,92,93]. It should be mentioned that the activation of both β_1 -AR and β_2 -AR subtypes occurs with different potencies by catecholamines (norepinephrine and epinephrine) in general. β_1 -ARs are coupled to $G\alpha_s$ proteins and β_2 -ARs are coupled to both $G\alpha_s$ -and $G\alpha_i$ proteins. The acute activation of β_1 -AR through G α_s proteins produces positive chronotropic and inotropic responses as well as cardiac hypertrophy whereas the chronic stimulation of β_1 -AR is associated with heart failure. The effects of both acute and chronic stimulation of the SNS are illustrated in Fig. 1. It needs to be emphasized that acute stimulation of β_1 -AR system results in adaptive hypertrophy whereas prolonged β_1 -AR signaling accounts for the development of maladaptive hypertrophy and subsequent heart failure [53,94-97]. Furthermore, overexpression of β_1 -AR in transgenic mice has also been reported to exhibit depressed cardiac function, progressive hypertrophy, and myocardial fibrosis [54,98]. On the other hand, $G\alpha_i$ -protein mediated signaling via β_2 -AR is generally believed to be cardioprotective due to its anti-apoptotic and anti-fibrotic effects [99,100].

In certain types of heart failure such as that due to aortic stenosis, it has been reported that β_2 -AR signaling may change to β_1 -AR-like signaling, become more susceptible to ischemic injury and contribute to the development of heart failure [101,102]. It has been suggested that such pathological manifestations of β_2 -AR overexpression are mediated primarily by $G\alpha_s$ - proteins rather than $G\alpha_i$ proteins [102]. Thus, it has been indicated that β_2 -AR signaling may be either protective or deleterious in the heart depending on transducer coupling with G-proteins [103-108]. It should also be noted that both β_1 -AR and β_2 -AR subtypes are coupled to β -arrestins, which may induce cardioprotective signaling cascades in the heart. Although the role of β_3 -AR in cardiac pathology is unclear, some studies have suggested that β_3 -AR may be involved in the development of heart disease [89,109–112]. The β_3 -AR expression in the myocardium has been shown to be upregulated in heart failure [67,113,114]. In addition, β_3 -AR has been reported to signal through endothelial nitric oxide synthase/nitric oxide/cyclic guanosine monophosphate (eNOS/NO/cGMP) pathway for the attenuation of cardiac contractility [90]. While extensive work needs to be carried out for establishing the exact role of both β_2 -AR and β_3 -AR signaling systems in cardiac hypertrophy and heart failure, there is overwhelming evidence that β_1 -AR signal transduction is activated. In this regard, it is noteworthy that blocking β_1 -AR signaling by several antagonists such as carvedilol, metoprolol, atenolol, and bisoprolol has been shown cardioprotection and other beneficial effects in heart failure [73,108,115–129].



Fig. 1. Acute and chronic effects of the sympathetic nervous system on β -adrenoceptor-mediated signal transduction components. NE, norepinephrine; EPI, epinephrine; Gs-Proteins, stimulatory guanine nucleotide proteins; \uparrow , increased.

3. Role of β_1 -AR Signal Transduction in Cardiac Hypertrophy and Heart Failure

Several studies have indicated that a wide variety of both extrinsic and intrinsic stimuli induce activation of different signal transduction pathways to increase the muscle mass for the occurrence of cardiac hypertrophy. This process is initiated by mechanical stress as well as different hormones, cytokines and growth factors that are sensed by different receptors in the cell membrane of cardiomyocytes. It is evident that cardiac hypertrophy at initial stages is an adaptive process in which the heart does not show any structural abnormalities and cardiac function is usually unaltered or augmented [25,56,130–134]. However, if the stimulus is not removed within a certain time period, there occurs a transition of adaptive hypertrophy to maladaptive hypertrophy, which exhibit a set of complexities, including cardiac remodeling, cardiac dysfunction, metabolic alterations, electrophysiological defects and increased ventricular wall stress. Progressive metabolic alterations in maladaptive hypertrophy are considered to result in the progression of subcellular abnormalities for Ca²⁺-handling, cardiac dysfunction and heart failure [51,57,58]. The loss of inotropic mechanism in the hypertrophied heart has been reported to occur due to changes in membrane receptors, protein kinase activities, and associated signal transduction system as well as defects in subcellular organelles during the progression of heart failure [23,34,49,50,52,69,135–143].

Involvement of β_1 -AR signaling in both adaptive and maladaptive hypertrophy as well as in heart failure is now well established [30,37,42,54,140] and the SNS is considered to regulate the status of β_1 -AR signal pathway during occurrence of these phenotypes. At early stages, activation of the SNS and subsequent elevation in the levels of plasma norepinephrine and epinephrine stimulate β_1 -AR and increase cardiac contractile force. However, prolonged hyperactivity of the SNS and elevated plasma catecholamines result in the derangement of one or more components of the β_1 -AR signaling transduction system, including β_1 -AR, Gs-proteins, adenylyl cyclase, β_1 -AR-Gs-protein coupling, and Gs-protein-adenylyl cyclase interactions. It is pointed out that an increase in Gs-protein or content results in augmenting cardiac function by increasing the adenylyl cyclase activity whereas an increase in G_i-protein activity or content is known to depress cardiac function by decreasing the adenylyl cyclase activity. Furthermore, exposure of cardiomyocytes to high amount of norepinephrine has been shown to cause a reduction in β_1 -AR expression, adenylyl cyclase activity, and contractile activity. Thus, excessive circulating levels of catecholamines can be seen to induce abnormalities in the β_1 -AR signal transduction pathway and result in cardiac dysfunction [133,135,144–148].

Depressed sensitivity of β_1 -AR to catecholamines as well as reduction in β_1 -AR number are reported to occur in heart failure [149]. Furthermore, overexpression of β_1 -AR in the heart in transgenic mice was found to develop hypertrophy at young age followed by progressive heart failure in later life [54,98,150–152]. Chronic stimulation of β_1 -AR by agonists such as isoproterenol has also been observed to induce cardiac hypertrophy [53] due to activation of PKA by elevated levels of cyclic AMP. Another study has indicated that β_1 -AR signaling stimulates hypertrophy in a PKA-independent manner via the activation of cyclic AMP binding protein, Epac [153]. However, other investigators have shown that mice overexpressing PKA are protected against isoproterenol-induced cardiac hypertrophy [154]. It is also pointed that the level of Gi-proteins is elevated in heart failure and this reduces cyclic AMP content for overall depression in β_1 -AR-mediated signaling [68]. Since PKA signaling microdomains regulate Ca²⁺-handling, it has been suggested that some PKA catalytic subunit may cause maladaptive hypertrophy and result in heart failure [48]. It should also be mentioned that PKA may directly enhance the stimulation of calcium-calmodulin kinase (CaMKII) or calcineurin/nuclear factor of activated T cells (NFAT) signaling [155]. Furthermore, the activation of PKA has also been suggested to inhibit cardiac hypertrophy via some signaling protein changes such as histone deacetylases (HDAC)5 phosphorylation or HDAC4 proteolysis [156]. While most of these observations support the view that β_1 -AR stimulation results in cardiac hypertrophy and progression to heart failure [53,93,94,118,125,157], the specific mechanisms remain unclear because of the complex nature of β_1 -AR signaling transduction pathway. It is also likely that changes in β_1 -AR signaling may depend on the stage and type of hypertrophy and heart failure.

4. Dependence of Changes in β_1 -AR Signal Transduction on Type and Stage of Pathological Stimulus

Since hypertrophy and heart failure are known to occur in response to several pathological stimuli, it was considered of great interest to determine if alterations in β_1 -AR signal pathway occur in different types of cardiac diseases. It may be noted that pressure overload in cardiovascular diseases such as hypertension, aortic stenosis, and aortic valve stenosis is associated with an increase in the ventricular wall thickness (concentric cardiac hypertrophy). On the other hand, volume overload in pathological conditions such as anemia, heart block, regurgitant mitral or aortic valves, as well as atrial or ventricular septal defects, and different congenital diseases, is associated with dilatation of the left ventricle chamber (eccentric cardiac hypertrophy) [61,158,159]. Varying degrees of changes in β_1 -AR signaling system due to both pressure overload [160–164] and volume overload [165–169] have been observed at the end-stage heart failure. Alterations in β_1 -AR signal transduction have also been reported to occur in other types of heart diseases [170–172] and heart failure due to chronic myocardial infarction [173–175].

Downregulation of β_1 -AR has been shown to occur in patients with left heart valvular disease as well as chronic mitral regurgitation [166,176]. Depressions in myocardial β_1 -AR density, adenylyl cyclase activity, and response to isoproterenol were observed after inducing volume overload [177]. A reduction in the adenylyl cyclase response to norepinephrine has been reported due to volume overload [167]. Furthermore, upregulation of β_1 -AR mechanisms was seen in the hypertrophic stage whereas these changes were depressed in heart failure [178]. Alterations in β_1 -AR signaling system, sensitivity of the myocardium to β_1 -AR stimulation, as well as changes in the subcellular distribution of regulatory proteins namely G-proteincoupled receptor kinase (GRK) isoforms and β -arrestins were observed at different stages of heart failure due to volume overload [165,168]. Other studies have also shown increased β_1 -AR expression and GRK activity as well as depressed activities of different components of β_1 -AR signaling pathways in heart failure [169,179-181]. Such variable alterations in β_1 -AR signal transduction system in the hypertrophied and failing hearts due to volume overload appear to be related to the stage of heart disease.

Varying degrees of changes in β_1 -AR, adenylyl cyclase and Gs-protein have also been identified in cardiac hypertrophy under several conditions associated with pressure overload [160]. Modification of cardiac adenylyl cyclase activities and changes in Gs-protein function have been observed in hypertension [172,182]. Pressure overload induced heart failure in guinea pigs was accompanied by an increase in β_1 -AR density [183] whereas depressions in the density of β_1 -AR as well as isoproterenol-induced increase in cardiac contraction and stimulation of adenylyl cyclase activity were observed in dogs with heart failure due to pressure overload [161,184]. Overexpression of cyclic AMP-hydrolyzing protein phosphodiesterase 4B (PDE4B), a key negative regulator of cardiac β_1 -AR stimulation, was shown to blunt the β_1 -AR signaling whereas its deficiency resulted in abnormal Ca²⁺-handling in pressure overload induced cardiac hypertrophy [185]. Furthermore, overexpression of a dominant negative mutant of Gs α -proteins decreased β_1 -AR responsiveness and protected against isoproterenol-induced cardiac hypertrophy in transgenic Gs α -DN-mice [186]. These observations showing variable changes in β_1 -AR signaling transduction system due to pressure overload also support the view that alterations in β_1 -AR signaling are dependent upon the stage of cardiac hypertrophy and heart failure.

5. Experimental Evidence for Alterations in β_1 -AR Mechanisms in Cardiac Hypertrophy

Since heart failure is commonly associated with cardiac hypertrophy, we have evaluated the existing information to determine if alterations in β_1 -AR mediated activities in the failing hearts are a consequence of the hypertrophic process. In this regard, we monitored changes in β_1 -AR signal transduction in pressure overload induced cardiac hypertrophy which was induced upon occluding the abdominal aorta in rats for 4 and 24 weeks [34,42,172,187,188]. The results in Fig. 2 (Ref. [42]) indicate that increased heart weight/body weight ratio (an index of cardiac hypertrophy) at 4 weeks of pressure overload was accompanied by increased left ventricle developed pressure (LVDP), left ventricle end-diastolic pressure (LVEDP) as well as rates of both rise and decline of ventricular pressures ($\pm dP/dt$) without any changes in the lung or liver weight to body weight ratios. On the other hand, hypertrophy induced by pressure overload for 24 weeks was associated with increased LVEDP and depressions in both LVDP and $\pm dP/dt$ parameters without any changes in lung or liver weight to body weight ratios (Fig. 2). These observations suggest that pressure overload for 4 weeks induces adaptive hypertrophy whereas that for 24 weeks induces maladaptive hypertrophy without any changes in lung or liver congestion (wellknown indices of heart failure).

Fig. 3 (Ref. [42]) shows that increased cardiac function (as reflected by increase in LVDP) and intracellular Ca^{2+} -concentration ($[Ca^{2+}]_i$) in cardiomyocytes by isoproterenol were not affected in adaptive hypertrophy due to pressure overload at 4 weeks. In contrast, both isoproterenol-induced increase in LVDP in the heart and $[Ca^{2+}]_i$ in cardiomyocytes were depressed in maladaptive hypertrophy due to pressure overload at 24 weeks. Furthermore, the results in Fig. 4 (Ref. [42]) show that pressure overload induced adaptive hypertrophy for 4 weeks did not show any changes in β_1 -AR density (B_{max} value); without any changes in dissociation constant (K_d value) or isoproterenol-induced increase in adenylyl cyclase activity. In contrast, pressure overload reduced maladaptive hypertrophy for 24 weeks showed depressions in β_1 -AR density and isoproterenol-induced increase the adenylyl cyclase activity (without any changes in K_d value) (Fig. 4). These data have been interpreted to reflect that adaptive cardiac hypertrophy due to pressure overload did not show any changes in β_1 -AR signal transduction mechanisms whereas maladaptive cardiac hypertrophy due to pressure overload was associated with some defects in the β_1 -AR signaling.



Fig. 2. General characteristics and ventricular function in rats at 4 and 24 weeks due to pressure overload (PO) after occluding the abdominal aorta. Data are based on the results described in our paper —Journal of Applied Physiology. 2007; 102: 978– 984 [42]. LVDP, left ventricle developed pressure; LVEDP, left ventricle end diastolic pressure; \pm dP/dt, rates of rise and decline of ventricle pressures. *p < 0.05 versus respective sham.

6. Experimental Evidence for Alterations in β_1 -AR Mechanisms in Heart Failure

In order to show if changes in β_1 -AR signal transduction system in heart failure are similar to those seen in adaptive cardiac hypertrophy, the data from studies in which volume overload was induced by aorto-venous (AV) shunt in rats at 4 and 24 weeks was evaluated [42,84,165,168,169, 189,190]. The results in Fig. 5 (Ref. [42]) show that increased heart weight to body weight ratio was accompanied by increased LVEDP and lung weight to body weight ratio without any changes in LVDP, $\pm dP/dt$ and liver weight to body weight ratios upon inducing AV-shunt for a 4-week period. It is pointed out that since no changes in cardiac function (as represented by LVDP and $\pm dP/dt$ parameters) were evident upon inducing volume overload for 4 weeks, we believe that cardiac hypertrophy at this stage is of adaptive type. Since lung weight to body weight ratios was significantly increased at 4 weeks of inducing volume overload, it can be argued that it may represent an early stage



Fig. 3. Effects of isoproterenol (ISO) on ventricular developed pressure and $[Ca^{2+}]_i$ in cardiomyocytes at 4 and 24 weeks due to pressure overload (PO) in rats. Data are based on the results described in our paper —Journal of Applied Physiology. 2007; 102: 978–984 [42]. Con, control; LVDP, left ventricle developed pressure. *p < 0.05 versus respective sham.

of heart failure. However, this may not be the case as no changes in cardiac function were observed at this stage. On the other hand, increases in heart weight to body weight ratio and LVEDP upon inducing volume overload for 24 weeks were associated with depressions of both LVDP and \pm dP/dt as well as increases in both lung or liver weight to body weight ratios, indicating the occurrence of heart failure. These data are consistent with the view that adaptive cardiac hypertrophy and heart failure due to volume overload become evident at 4 weeks and 24 weeks after inducing AV-shunt, respectively.



Fig. 4. Ventricular B_{max} (maximal number of binding) and K_d (dissociation constant) values for β_1 -adrenoceptors and effect of isoproterenol (ISO) on adenylyl cyclase activity at 4 and 24 weeks due to pressure overload (PO) in rats. Data are based on the results described in our paper — Journal of Applied Physiology. 2007; 102: 978–984 [42]. *p < 0.05 versus respective sham.

The results described in Fig. 6 (Ref. [42]) indicate that isoproterenol-induced increases in LVDP in the heart and $[Ca^{2+}]_i$ in cardiomyocytes were augmented by volume overload at 4 weeks of inducing AV-shunt whereas these responses of the heart to isoproterenol showed marked depressions at 24 weeks AV-shunt. Furthermore, β_1 -AR density as well as activation of adenylyl cyclase by isoproterenol were markedly augmented by volume overload at 4 weeks after inducing AV-shunt whereas both β_1 -AR density and isoproterenol-induced activation of adenylyl cyclase were attenuated at 24 weeks after inducing AV-shunt. No changes in K_d values for β_1 -AR were observed either at 4 weeks or 24 weeks after inducing AV-shunt (Fig. 7, Ref. [42]). These data indicate that alterations in β_1 -AR signal transduction pathways in the failing heart are not similar to those in adaptive cardiac hypertrophy due to volume overload.



Fig. 5. General characteristics and ventricular function in rats at 4 and 24 weeks due to volume overload (VO) after the aortocaval shunt. Data are based on the results described in our paper — Journal of Applied Physiology. 2007; 102: 978–984 [42]. LVDP, left ventricle developed pressure; LVEDP, left ventricle end diastolic pressure; \pm dP/dt, rates of rise and decline of ventricle pressure. *p < 0.05 versus respective sham.

7. Conclusions and Perspectives

Although heart failure is associated with cardiac dysfunction, there also occurs a loss of adrenergic support, which is considered to maintain cardiac performance in this syndrome. The depression of inotropic responses to stimulation of the SNS or exogenously administrated catecholamines is considered to be a consequence of a defect in the β_1 -AR signal transduction in heart failure. However, the exact mechanisms for such an alteration are not fully understood. Since the β_1 -AR signaling system is known to include β_1 -AR, Gs-and Gi-proteins and adenylyl cyclase, it has been observed that alterations in anyone of these components may result in reduced formation of cyclic AMP and subsequent impaired PKA-mediated phosphorylation of subcellular proteins in the failing heart. In view of the importance of β_1 -AR signaling and PKA-induced phosphorylation of Ca²⁺- pump and Ca²⁺- release proteins in the sarcoplasmic reticulum as well as troponin and other regulatory proteins in myofilaments for regulating cardiac function, it is likely that augmentation and depression of isoproterenol - induced responses of cardiac function in adap-



Fig. 6. Effects of isoproterenol (ISO) on left ventricular developed pressure (LVDP) in rats and $[Ca^{2+}]_i$ in cardiomyocytes at 4 and 24 weeks due to volume overload in rats. Data are based on the results described in our paper —Journal of Applied Physiology. 2007; 102: 978–984 [42]. LVDP, left ventricle developed pressure; Con, control; VO, volume overload. *p < 0.05versus respective sham.

tive cardiac hypertrophy and failing hearts are due to corresponding alterations in PKA associated phosphorylations [13,14,32,34,63,65], respectively. In fact, various studies in heart failure have shown that the depressed β_1 -AR signaling in failing hearts is due to desensitization of β_1 -AR [67,74,85] but these changes are considered to be dependent on the stage of heart failure. Since catecholamines for a short period increase cardiac contractile force whereas these responses are attenuated over a prolonged period, it



Fig. 7. Ventricular Bmax (maximal number of binding) and Kd (dissociation constant) values for β_1 -adrenoceptors and effect of isoproterenol (ISO) on adenylyl cyclase activity at 4 and 24 weeks due to volume overload (VO) in rats. Data are based on the results described in our paper —Journal of Applied Physiology. 2007; 102:978–984 [42]. *p < 0.05 versus respective sham.

appears that downregulation of β_1 -AR signal transduction in heart failure may be due to elevated levels of plasma catecholamines for a prolonged period. It is also pointed out that oxidative stress plays an important role in the pathogenesis of heart failure and it is likely that defects in β_1 -AR signaling at the advanced stage of heart failure may be due to the development of oxidative stress as a consequence of circulating catecholamines and other vasoactive hormones such as angiotensin II [34,80,191]. Accordingly, it is suggested that therapy of heart failure with some antioxidants may prove useful in preventing downregulation of β_1 -AR mechanisms in the failing heart.

From the foregoing discussion, it is evident that not only changes in β_1 -AR signal transduction are dependent upon the stage of heart failure, marked differences in β_1 -AR signaling have also been observed in adaptive and maladaptive cardiac hypertrophy. Particularly, it is noteworthy that adaptive hypertrophy induced by pressure overload or volume overload for a 4-week period was found to exhibit either unaltered or augmented responses of heart function,

 $[Ca^{2+}]_i$ in cardiomyocytes and adenylyl cyclase activity to isoproterenol as well as unaltered or increased β_1 -AR density. On the other hand, all these responses or parameters for β_1 -AR signal transduction mechanisms were depressed in maladaptive hypertrophy at 24 weeks of inducing pressure overload as well as in heart failure at 24 weeks of inducing volume overload. Such differences in β_1 -AR signaling in adaptive and maladaptive cardiac hypertrophy as well as heart failure can be explained on the basis of differences in the development of progressive levels of oxidative stress as a consequence of circulating catecholamines and other vasoactive hormones for a prolonged duration [143,191]. Furthermore, it is pointed out that, unlike the adaptive cardiac hypertrophy, both maladaptive cardiac hypertrophy at 24 weeks due to pressure overload and heart failure due to volume overload for 24 weeks were found to exhibit a similar pattern of depressions in all parameters of β_1 -AR signal transduction system. Thus, it appears that downregulation of the β_1 -AR signaling in heart failure or maladaptive cardiac hypertrophy may not be associated with the hypertrophic process per se. Although occurrence of oxidative stress has been suggested to be involved in transition of adaptive hypertrophy to maladaptive hypertrophy as well as progression to heart failure [80,143,191], extensive research work needs to be carried out with respect to establishing any relationship between oxidative stress and changes in β_1 -AR signal transduction pathway during the development of heart failure to make any meaningful conclusion.

Several investigators have reported a wide variety of changes in β_1 -AR signal transduction in cardiac hypertrophy and heart failure [25,34,35,46,54,73,149]; however, the exact mechanisms for such variable alterations in this pathway have not been identified. It needs to be emphasized that adaptive cardiac hypertrophy has been suggested to be a consequence of changes in the redox status of myocardium due to formation of a small amount of oxyradicals [137,191]. On the other hand, excessive formation of oxyradicals for the occurrence of oxidative stress is considered to be involved in the development of maladaptive cardiac hypertrophy and subsequent heart failure [137,191]. However, the participation of other mechanisms such as alterations in the levels of proinflammatory cytokines and intracellular Ca²⁺ - overload as well as metabolic abnormalities [51,60,61,136,138,155,192] cannot be ruled out for explaining the difference in the status of β_1 -AR signaling in non failing and failing hypertrophied hearts. Since the activation of baroreceptors in the heart is known to play a critical role in the regulation of cardiac function and β_1 -AR mechanism [193], alterations in the baroreflex mechanisms during the development of hypertension and heart failure have been implicated in changing the intensity of adrenergic stimuli and β_1 -AR signal transduction pathway [194,195]. This view is also supported by the observations that there occurs an increase in the sympathetic activity and a decrease

in the parasympathetic activity in patients with heart failure [196]. In addition, newer approaches for activating the baroreflex system or vagal stimulation have been shown to exert promising effects in correcting the autonomic imbalance for improving cardiac performance in heart failure [197,198]. Accordingly, progressive changes in the baroreflex system due to both pressure and volume overload can also be seen to induce upregulation and downregulation of β_1 -AR signaling during the development of cardiac hypertrophy and heart failure. Thus, it appears that the pathophysiological and molecular mechanisms in changing the status of β_1 -AR signal transduction pathway in cardiac hypertrophy and heart failure are of complex nature and require further studies for establishing the exact relationship among diverse pathogenic factors for the induction of alterations in β_1 -AR signaling.

Author Contributions

NSD developed the concept and outline for this project whereas SKB searched the literature, prepared figures and wrote the first draft of this manuscript. AA, KOM and CMLdeV participated in analysis and interpretation of data as well as in editing and revising the manuscript. All authors have contributed sufficiently in preparing, editorial changes and completing this manuscript and have approved its submission for publication. All authors have read and approved the final manuscript and have agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

We thank the St. Boniface Hospital Albrechtsen Research Centre for infrastructural support. Thanks are also due to Ms. Khushman Kaur for her help in editing this paper.

Funding

This research received no external funding.

Conflict of Interest

The authors declare no conflict of interest. Although the data presented in this paper are based on earlier work from our laboratory, none of the figures in this article show any similarity with those in our previous paper. Naranjan S. Dhalla is serving as one of the Editorial Board members of this journal. We declare that Naranjan S. Dhalla had no involvement in the peer review of this article and has no access to information regarding its peer review. Full responsibility for the editorial process for this article was delegated to Zoltán Papp and Maurizio Pieroni.

References

- Hamm NC, Robitaille C, Ellison J, O'Donnell S, McRae L, Hutchings K, *et al.* Population coverage of the Canadian Chronic Disease Surveillance System: a survey of the contents of health insurance registries across Canada. Health Promotion and Chronic Disease Prevention in Canada: Research, Policy and Practice. 2021; 40: 230–241.
- [2] Tran DT, Ohinmaa A, Thanh NX, Howlett JG, Ezekowitz JA, McAlister FA, *et al.* The current and future financial burden of hospital admissions for heart failure in Canada: a cost analysis. CMAJ Open. 2016; 4: E365–E370.
- [3] Canadian Institute for Health Information. Hospital Stays in Canada. Available at: https://www.cihi.ca/en/hospital-stays-i n-canada (Accessed: 23 February 2023).
- [4] Poon S, Leis B, Lambert L, MacFarlane K, Anderson K, Blais C, et al. The State of Heart Failure Care in Canada: Minimal Improvement in Readmissions Over Time Despite an Increased Number of Evidence-Based Therapies. CJC Open. 2022; 4: 667–675.
- [5] McDonald MA, Ashley EA, Fedak PWM, Hawkins N, Januzzi JL, McMurray JJV, *et al.* Mind the Gap: Current Challenges and Future State of Heart Failure Care. The Canadian Journal of Cardiology. 2017; 33: 1434–1449.
- [6] Virani SA, Bains M, Code J, Ducharme A, Harkness K, Howlett JG, et al. The Need for Heart Failure Advocacy in Canada. The Canadian Journal of Cardiology. 2017; 33: 1450–1454.
- [7] Ponikowski P, Voors AA, Anker SD, Bueno H, Cleland JGF, Coats AJS, *et al.* 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure. Polish Heart Journal. 2016; 74: 1037–1147.
- [8] Yancy CW, Jessup M, Bozkurt B, Butler J, Casey DE, Jr, Colvin MM, et al. 2017 ACC/AHA/HFSA Focused Update of the 2013 ACCF/AHA Guideline for the Management of Heart Failure: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines and the Heart Failure Society of America. Circulation. 2017; 136: e137– e161.
- [9] Ezekowitz JA, O'Meara E, McDonald MA, Abrams H, Chan M, Ducharme A, *et al.* 2017 Comprehensive Update of the Canadian Cardiovascular Society Guidelines for the Management of Heart Failure. The Canadian Journal of Cardiology. 2017; 33: 1342– 1433.
- [10] Greene SJ, Fonarow GC, DeVore AD, Sharma PP, Vaduganathan M, Albert NM, *et al.* Titration of Medical Therapy for Heart Failure with Reduced Ejection Fraction. Journal of the American College of Cardiology. 2019; 73: 2365–2383.
- [11] DeFilippis EM, Butler J, Vaduganathan M. Waiting Period Before Implantable Cardioverter-Defibrillator Implantation in Newly Diagnosed Heart Failure with Reduced Ejection Fraction: A Window of Opportunity. Circulation: Heart Failure. 2017; 10: e004478.
- [12] Dhalla NS, Afzal N, Beamish RE, Naimark B, Takeda N, Nagano M. Pathophysiology of cardiac dysfunction in congestive heart failure. The Canadian Journal of Cardiology. 1993; 9: 873–887.
- [13] Dhalla NS, Dent MR, Tappia PS, Sethi R, Barta J, Goyal RK. Subcellular remodeling as a viable target for the treatment of congestive heart failure. Journal of Cardiovascular Pharmacology and Therapeutics. 2006; 11: 31–45.
- [14] Machackova J, Barta J, Dhalla NS. Myofibrillar remodeling in cardiac hypertrophy, heart failure and cardiomyopathies. The Canadian Journal of Cardiology. 2006; 22: 953–968.
- [15] Mudd JO, Kass DA. Tackling heart failure in the twenty-first century. Nature. 2008; 451: 919–928.

- [16] Dhalla NS, Rangi S, Babick AP, Zieroth S, Elimban V. Cardiac remodeling and subcellular defects in heart failure due to myocardial infarction and aging. Heart Failure Reviews. 2012; 17: 671–681.
- [17] Sriram K, Insel PA. G Protein-Coupled Receptors as Targets for Approved Drugs: How Many Targets and How Many Drugs? Molecular Pharmacology. 2018; 93: 251–258.
- [18] Grogan A, Lucero EY, Jiang H, Rockman HA. Pathophysiology and pharmacology of G protein-coupled receptors in the heart. Cardiovascular Research. 2023; 119: 1117–1129.
- [19] Allen JA, Roth BL. Strategies to discover unexpected targets for drugs active at G protein-coupled receptors. Annual Review of Pharmacology and Toxicology. 2011; 51: 117–144.
- [20] Rask-Andersen M, Masuram S, Schiöth HB. The druggable genome: Evaluation of drug targets in clinical trials suggests major shifts in molecular class and indication. Annual Review of Pharmacology and Toxicology. 2014; 54: 9–26.
- [21] Santos R, Ursu O, Gaulton A, Bento AP, Donadi RS, Bologa CG, et al. A comprehensive map of molecular drug targets. Nature Reviews Drug Discovery. 2017; 16: 19–34.
- [22] Rockman HA, Koch WJ, Lefkowitz RJ. Seven-transmembranespanning receptors and heart function. Nature. 2002; 415: 206– 212.
- [23] Brower GL, Janicki JS. Contribution of ventricular remodeling to pathogenesis of heart failure in rats. American Journal of Physiology. Heart and Circulatory Physiology. 2001; 280: H674–H683.
- [24] Wang J, Gareri C, Rockman HA. G-Protein-Coupled Receptors in Heart Disease. Circulation Research. 2018; 123: 716–735.
- [25] Bristow MR, Hershberger RE, Port JD, Gilbert EM, Sandoval A, Rasmussen R, *et al.* Beta-adrenergic pathways in nonfailing and failing human ventricular myocardium. Circulation. 1990; 82: 112–125.
- [26] Homey CJ, Vatner SF, Vatner DE. Beta-adrenergic receptor regulation in the heart in pathophysiologic states: abnormal adrenergic responsiveness in cardiac disease. Annual Review of Physiology. 1991; 53: 137–159.
- [27] Campbell AS, Johnstone SR, Baillie GS, Smith G. β-Adrenergic modulation of myocardial conduction velocity: Connexins vs. sodium current. Journal of Molecular and Cellular Cardiology. 2014; 77: 147–154.
- [28] Machackova J, Sanganalmath SK, Barta J, Dhalla KS, Dhalla NS. Amelioration of cardiac remodeling in congestive heart failure by beta-adrenoceptor blockade is associated with depression in sympathetic activity. Cardiovascular Toxicology. 2010; 10: 9–16.
- [29] Ahles A, Engelhardt S. Polymorphic variants of adrenoceptors: pharmacology, physiology, and role in disease. Pharmacological Reviews. 2014; 66: 598–637.
- [30] Meyer EE, Clancy CE, Lewis TJ. Dynamics of adrenergic signaling in cardiac myocytes and implications for pharmacological treatment. Journal of Theoretical Biology. 2021; 519: 110619.
- [31] Jiang H, Galtes D, Wang J, Rockman HA. G protein-coupled receptor signaling: transducers and effectors. American Journal of Physiology Cell Physiology. 2022; 323: C731–C748.
- [32] Bers DM. Calcium cycling and signaling in cardiac myocytes. Annual Review of Physiology. 2008; 70: 23–49.
- [33] Dhalla NS, Müller AL. Protein Kinases as Drug Development Targets for Heart Disease Therapy. Pharmaceuticals (Basel, Switzerland). 2010; 3: 2111–2145.
- [34] Dhalla NS, Wang X, Sethi R, Das PK, Beamish RE. β-adrenergic linked signal transduction mechanisms in failing hearts. Heart Failure Reviews. 1997; 2: 55–65.
- [35] Chakraborti S, Chakraborti T, Shaw G. beta-adrenergic mechanisms in cardiac diseases: a perspective. Cellular Signalling. 2000; 12: 499–513.

- [36] Lohse MJ, Engelhardt S, Eschenhagen T. What is the role of beta-adrenergic signaling in heart failure? Circulation Research. 2003; 93: 896–906.
- [37] Feldman DS, Carnes CA, Abraham WT, Bristow MR. Mechanisms of disease: beta-adrenergic receptors-alterations in signal transduction and pharmacogenomics in heart failure. Nature Clinical Practice. Cardiovascular Medicine. 2005; 2: 475–483.
- [38] Katz AM. The "modern" view of heart failure: how did we get here? Circulation: Heart Failure. 2008; 1: 63–71.
- [39] Al-Gobari M, El Khatib C, Pillon F, Gueyffier F. β-Blockers for the prevention of sudden cardiac death in heart failure patients: a meta-analysis of randomized controlled trials. BMC Cardiovascular Disorders. 2013; 13: 52.
- [40] Wachter SB, Gilbert EM. Beta-adrenergic receptors, from their discovery and characterization through their manipulation to beneficial clinical application. Cardiology. 2012; 122: 104–112.
- [41] Lymperopoulos A. Physiology and pharmacology of the cardiovascular adrenergic system. Frontiers in Physiology. 2013; 4: 240.
- [42] Sethi R, Saini HK, Guo X, Wang X, Elimban V, Dhalla NS. Dependence of changes in beta-adrenoceptor signal transduction on type and stage of cardiac hypertrophy. Journal of Applied Physiology (Bethesda, Md.: 1985). 2007; 102: 978–984.
- [43] Ganguly PK, Lee SL, Beamish RE, Dhalla NS. Altered sympathetic system and adrenoceptors during the development of cardiac hypertrophy. American Heart Journal. 1989; 118: 520–525.
- [44] Yagishita D, Chui RW, Yamakawa K, Rajendran PS, Ajijola OA, Nakamura K, et al. Sympathetic nerve stimulation, not circulating norepinephrine, modulates T-peak to T-end interval by increasing global dispersion of repolarization. Circulation: Arrhythmia and Electrophysiology. 2015; 8: 174–185.
- [45] Zekios KC, Mouchtouri ET, Lekkas P, Nikas DN, Kolettis TM. Sympathetic Activation and Arrhythmogenesis after Myocardial Infarction: Where Do We Stand? Journal of Cardiovascular Development and Disease. 2021; 8: 57.
- [46] Lamba S, Abraham WT. Alterations in adrenergic receptor signaling in heart failure. Heart Failure Reviews. 2000; 5: 7–16.
- [47] Okwuosa TM, Soliman EZ, Lopez F, Williams KA, Alonso A, Ferdinand KC. Left ventricular hypertrophy and cardiovascular disease risk prediction and reclassification in blacks and whites: the Atherosclerosis Risk in Communities Study. American Heart Journal. 2015; 169: 155–161.e5.
- [48] Yang JH, Polanowska-Grabowska RK, Smith JS, Shields CW, 4th, Saucerman JJ. PKA catalytic subunit compartmentation regulates contractile and hypertrophic responses to β-adrenergic signaling. Journal of Molecular and Cellular Cardiology. 2014; 66: 83–93.
- [49] Frey N, Olson EN. Cardiac hypertrophy: the good, the bad, and the ugly. Annual Review of Physiology. 2003; 65: 45–79.
- [50] Hannan RD, Jenkins A, Jenkins AK, Brandenburger Y. Cardiac hypertrophy: a matter of translation. Clinical and Experimental Pharmacology & Physiology. 2003; 30: 517–527.
- [51] Wikman-Coffelt J, Parmley WW, Mason DT. The cardiac hypertrophy process. Analyses of factors determining pathological vs. physiological development. Circulation Research. 1979; 45: 697–707.
- [52] Mann DL, Spann JF, Cooper G. Basic mechanisms and models in cardiac hypertrophy: part 1. Pathophysiological models. Modern Concepts of Cardiovascular Disease. 1988; 57: 7–11.
- [53] Morisco C, Zebrowski DC, Vatner DE, Vatner SF, Sadoshima J. Beta-adrenergic cardiac hypertrophy is mediated primarily by the beta(1)-subtype in the rat heart. Journal of Molecular and Cellular Cardiology. 2001; 33: 561–573.
- [54] Engelhardt S, Hein L, Wiesmann F, Lohse MJ. Progressive hypertrophy and heart failure in beta1-adrenergic receptor transgenic mice. Proceedings of the National Academy of Sciences

of the United States of America. 1999; 96: 7059-7064.

- [55] Khalilimeybodi A, Daneshmehr A, Sharif-Kashani B. Investigating β-adrenergic-induced cardiac hypertrophy through computational approach: classical and non-classical pathways. The Journal of Physiological Sciences: JPS. 2018; 68: 503–520.
- [56] Weber KT, Clark WA, Janicki JS, Shroff SG. Physiologic versus pathologic hypertrophy and the pressure-overloaded myocardium. Journal of Cardiovascular Pharmacology. 1987; 10 Suppl 6: S37–S50.
- [57] Dhalla NS, Heyliger CE, Beamish RE, Innes IR. Pathophysiological aspects of myocardial hypertrophy. The Canadian Journal of Cardiology. 1987; 3: 183–196.
- [58] Lyon RC, Zanella F, Omens JH, Sheikh F. Mechanotransduction in cardiac hypertrophy and failure. Circulation Research. 2015; 116: 1462–1476.
- [59] Post SR, Hammond HK, Insel PA. Beta-adrenergic receptors and receptor signaling in heart failure. Annual Review of Pharmacology and Toxicology. 1999; 39: 343–360.
- [60] Stiles GL, Caron MG, Lefkowitz RJ. Beta-adrenergic receptors: biochemical mechanisms of physiological regulation. Physiological Reviews. 1984; 64: 661–743.
- [61] Mann DL, Bristow MR. Mechanisms and models in heart failure: the biomechanical model and beyond. Circulation. 2005; 111: 2837–2849.
- [62] Dent MR, Singal T, Tappia PS, Sethi R, Dhalla NS. β adrenoceptor-linked signal transduction mechanisms in congestive heart failure. Advances in Biochemistry in Health and Disease. 2008; 3: 27–49.
- [63] Madamanchi A. Beta-adrenergic receptor signaling in cardiac function and heart failure. McGill Journal of Medicine. 2007; 10: 99–104.
- [64] Bedioune I, Lefebvre F, Lechêne P, Varin A, Domergue V, Kapiloff MS, *et al.* PDE4 and mAKAPβ are nodal organizers of β2-ARs nuclear PKA signalling in cardiac myocytes. Cardiovascular Research. 2018; 114: 1499–1511.
- [65] Liu Y, Chen J, Fontes SK, Bautista EN, Cheng Z. Physiological and pathological roles of protein kinase A in the heart. Cardiovascular Research. 2022; 118: 386–398.
- [66] Kamide T, Okumura S, Ghosh S, Shinoda Y, Mototani Y, Ohnuki Y, *et al*. Oscillation of cAMP and Ca(2+) in cardiac myocytes: a systems biology approach. The Journal of Physiological Sciences: JPS. 2015; 65: 195–200.
- [67] Violin JD, DiPilato LM, Yildirim N, Elston TC, Zhang J, Lefkowitz RJ. beta2-adrenergic receptor signaling and desensitization elucidated by quantitative modeling of real time cAMP dynamics. The Journal of Biological Chemistry. 2008; 283: 2949–2961.
- [68] Nikolaev VO, Moshkov A, Lyon AR, Miragoli M, Novak P, Paur H, et al. Beta2-adrenergic receptor redistribution in heart failure changes cAMP compartmentation. Science (New York, N.Y.). 2010; 327: 1653–1657.
- [69] Kirchhefer U, Schmitz W, Scholz H, Neumann J. Activity of cAMP-dependent protein kinase and Ca2+/calmodulindependent protein kinase in failing and nonfailing human hearts. Cardiovascular Research. 1999; 42: 254–261.
- [70] Lymperopoulos A, Rengo G, Koch WJ. Adrenal adrenoceptors in heart failure: fine-tuning cardiac stimulation. Trends in Molecular Medicine. 2007; 13: 503–511.
- [71] Armour JA. Cardiac neuronal hierarchy in health and disease. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology. 2004; 287: R262–R271.
- [72] Gordan R, Gwathmey JK, Xie LH. Autonomic and endocrine control of cardiovascular function. World Journal of Cardiology. 2015; 7: 204–214.
- [73] Clark AL, Cleland JG. The control of adrenergic function in heart failure: therapeutic intervention. Heart Failure Reviews.

2000; 5: 101–114.

- [74] Fu LX, Waagstein F, Hjalmarson A. Beta-adrenoceptor–Gprotein–adenylyl cyclase system in cardiac disease: a new insight into desensitization. Clinical Physiology (Oxford, England). 1991; 11: 1–7.
- [75] Iwai-Kanai E, Hasegawa K, Araki M, Kakita T, Morimoto T, Sasayama S. alpha- and beta-adrenergic pathways differentially regulate cell type-specific apoptosis in rat cardiac myocytes. Circulation. 1999; 100: 305–311.
- [76] Mangmool S, Shukla AK, Rockman HA. beta-Arrestindependent activation of Ca(2+)/calmodulin kinase II after beta(1)-adrenergic receptor stimulation. The Journal of Cell Biology. 2010; 189: 573–587.
- [77] Yoo B, Lemaire A, Mangmool S, Wolf MJ, Curcio A, Mao L, et al. Beta1-adrenergic receptors stimulate cardiac contractility and CaMKII activation *in vivo* and enhance cardiac dysfunction following myocardial infarction. American Journal of Physiology-Heart and Circulatory Physiology. 2009; 297: H1377–H1386.
- [78] Communal C, Singh K, Sawyer DB, Colucci WS. Opposing effects of beta(1)- and beta(2)-adrenergic receptors on cardiac myocyte apoptosis: role of a pertussis toxin-sensitive G protein. Circulation. 1999; 100: 2210–2212.
- [79] Burchfield JS, Xie M, Hill JA. Pathological ventricular remodeling: mechanisms: part 1 of 2. Circulation. 2013; 128: 388–400.
- [80] Dhalla NS, Elimban V, Shah AK, Mohamad N. Mechanisms of cardiac dysfunction in heart failure due to myocardial infarction. Journal of Integrative Cardiology. 2019; 2: 1–7.
- [81] Dorian P. Antiarrhythmic action of beta-blockers: potential mechanisms. Journal of Cardiovascular Pharmacology and Therapeutics. 2005; 10 Suppl 1: S15–S22.
- [82] Randhawa AS, Dhadial RS, Adameova A, Ashgar E, Dhalla NS. The role of the sympathetic nervous system in sudden cardiac death. Current Research: Cardiology. 2016; 3: 83–88.
- [83] Fu Y, Westenbroek RE, Scheuer T, Catterall WA. Basal and βadrenergic regulation of the cardiac calcium channel CaV1.2 requires phosphorylation of serine 1700. Proceedings of the National Academy of Sciences of the United States of America. 2014; 111: 16598–16603.
- [84] Wang X, Sentex E, Chapman D, Dhalla NS. Alterations of adenylyl cyclase and G proteins in aortocaval shunt-induced heart failure. American Journal of Physiology. Heart and Circulatory Physiology. 2004; 287: H118–H125.
- [85] Choi DJ, Koch WJ, Hunter JJ, Rockman HA. Mechanism of beta-adrenergic receptor desensitization in cardiac hypertrophy is increased beta-adrenergic receptor kinase. The Journal of Biological Chemistry. 1997; 272: 17223–17229.
- [86] Cellini A, Höfler D, Arias-Loza PA, Bandleon S, Langsenlehner T, Kohlhaas M, *et al.* The α2-isoform of the Na⁺/K⁺-ATPase protects against pathological remodeling and β-adrenergic desensitization after myocardial infarction. American Journal of Physiology. Heart and Circulatory Physiology. 2021; 321: H650–H662.
- [87] Woo AYH, Xiao RP. β-Adrenergic receptor subtype signaling in heart: from bench to bedside. Acta Pharmacologica Sinica. 2012; 33: 335–341.
- [88] Myagmar BE, Flynn JM, Cowley PM, Swigart PM, Montgomery MD, Thai K, *et al.* Adrenergic Receptors in Individual Ventricular Myocytes: The Beta-1 and Alpha-1B Are in All Cells, the Alpha-1A Is in a Subpopulation, and the Beta-2 and Beta-3 Are Mostly Absent. Circulation Research. 2017; 120: 1103–1115.
- [89] Michel LYM, Farah C, Balligand JL. The Beta3 Adrenergic Receptor in Healthy and Pathological Cardiovascular Tissues. Cells. 2020; 9: 2584.
- [90] Schobesberger S, Wright PT, Poulet C, Sanchez Alonso Mardones JL, Mansfield C, Friebe A, et al. β_3-Adrenoceptor re-

distribution impairs NO/cGMP/PDE2 signalling in failing cardiomyocytes. ELife. 2020; 9: e52221.

- [91] Bathe-Peters M, Gmach P, Boltz HH, Einsiedel J, Gotthardt M, Hübner H, *et al.* Visualization of β-adrenergic receptor dynamics and differential localization in cardiomyocytes. Proceedings of the National Academy of Sciences of the United States of America. 2021; 118: e2101119118.
- [92] Saucerman JJ, McCulloch AD. Cardiac beta-adrenergic signaling: from subcellular microdomains to heart failure. Annals of the New York Academy of Sciences. 2006; 1080: 348–361.
- [93] Bristow MR, Ginsburg R, Umans V, Fowler M, Minobe W, Rasmussen R, et al. Beta 1- and beta 2-adrenergic-receptor subpopulations in nonfailing and failing human ventricular myocardium: coupling of both receptor subtypes to muscle contraction and selective beta 1-receptor down-regulation in heart failure. Circulation Research. 1986; 59: 297–309.
- [94] Schäfer M, Frischkopf K, Taimor G, Piper HM, Schlüter KD. Hypertrophic effect of selective beta(1)-adrenoceptor stimulation on ventricular cardiomyocytes from adult rat. American Journal of Physiology. Cell Physiology. 2000; 279: C495–C503.
- [95] Brodde OE. Beta 1- and beta 2-adrenoceptors in the human heart: properties, function, and alterations in chronic heart failure. Pharmacological Reviews. 1991; 43: 203–242.
- [96] Steinberg SF. The molecular basis for distinct beta-adrenergic receptor subtype actions in cardiomyocytes. Circulation Research. 1999; 85: 1101–1111.
- [97] Xiao RP. Beta-adrenergic signaling in the heart: dual coupling of the beta2-adrenergic receptor to G(s) and G(i) proteins. Science's STKE: Signal Transduction Knowledge Environment. 2001; 2001: re15.
- [98] Bisognano JD, Weinberger HD, Bohlmeyer TJ, Pende A, Raynolds MV, Sastravaha A, *et al*. Myocardial-directed overexpression of the human beta(1)-adrenergic receptor in transgenic mice. Journal of Molecular and Cellular Cardiology. 2000; 32: 817–830.
- [99] Chesley A, Lundberg MS, Asai T, Xiao RP, Ohtani S, Lakatta EG, *et al.* The beta(2)-adrenergic receptor delivers an antiapoptotic signal to cardiac myocytes through G(i)-dependent coupling to phosphatidylinositol 3'-kinase. Circulation Research. 2000; 87: 1172–1179.
- [100] Zhu WZ, Zheng M, Koch WJ, Lefkowitz RJ, Kobilka BK, Xiao RP. Dual modulation of cell survival and cell death by beta(2)adrenergic signaling in adult mouse cardiac myocytes. Proceedings of the National Academy of Sciences of the United States of America. 2001; 98: 1607–1612.
- [101] Du XJ, Autelitano DJ, Dilley RJ, Wang B, Dart AM, Woodcock EA. beta(2)-adrenergic receptor overexpression exacerbates development of heart failure after aortic stenosis. Circulation. 2000; 101: 71–77.
- [102] Cross HR, Steenbergen C, Lefkowitz RJ, Koch WJ, Murphy E. Overexpression of the cardiac beta(2)-adrenergic receptor and expression of a beta-adrenergic receptor kinase-1 (betaARK1) inhibitor both increase myocardial contractility but have differential effects on susceptibility to ischemic injury. Circulation Research. 1999; 85: 1077–1084.
- [103] Woo AYH, Song Y, Xiao RP, Zhu W. Biased β 2-adrenoceptor signalling in heart failure: pathophysiology and drug discovery. British Journal of Pharmacology. 2015; 172: 5444–5456.
- [104] Zhu W, Petrashevskaya N, Ren S, Zhao A, Chakir K, Gao E, et al. Gi-biased β2AR signaling links GRK2 upregulation to heart failure. Circulation Research. 2012; 110: 265–274.
- [105] Lang D, Holzem K, Kang C, Xiao M, Hwang HJ, Ewald GA, et al. Arrhythmogenic remodeling of $\beta 2$ versus $\beta 1$ adrenergic signaling in the human failing heart. Circulation: Arrhythmia and Electrophysiology. 2015; 8: 409–419.
- [106] Wang Y, Yuan J, Qian Z, Zhang X, Chen Y, Hou X, et al.

 β 2 adrenergic receptor activation governs cardiac repolarization and arrhythmogenesis in a guinea pig model of heart failure. Scientific Reports. 2015; 5: 7681.

- [107] Zhang X, Szeto C, Gao E, Tang M, Jin J, Fu Q, *et al.* Cardiotoxic and cardioprotective features of chronic β-adrenergic signaling. Circulation Research. 2013; 112: 498–509.
- [108] Wang J, Pani B, Gokhan I, Xiong X, Kahsai AW, Jiang H, et al. β-Arrestin-Biased Allosteric Modulator Potentiates Carvedilol-Stimulated β Adrenergic Receptor Cardioprotection. Molecular Pharmacology. 2021; 100: 568–579.
- [109] Trappanese DM, Liu Y, McCormick RC, Cannavo A, Nanayakkara G, Baskharoun MM, *et al.* Chronic β 1-adrenergic blockade enhances myocardial β 3-adrenergic coupling with nitric oxide-cGMP signaling in a canine model of chronic volume overload: new insight into mechanisms of cardiac benefit with selective β 1-blocker therapy. Basic Research in Cardiology. 2015; 110: 456.
- [110] Arioglu-Inan E, Kayki-Mutlu G, Michel MC. Cardiac β_3 adrenoceptors-A role in human pathophysiology? British Journal of Pharmacology. 2019; 176: 2482–2495.
- [111] Belge C, Hammond J, Dubois-Deruy E, Manoury B, Hamelet J, Beauloye C, *et al.* Enhanced expression of β 3-adrenoceptors in cardiac myocytes attenuates neurohormone-induced hypertrophic remodeling through nitric oxide synthase. Circulation. 2014; 129: 451–462.
- [112] Skeberdis VA, Gendviliene V, Zablockaite D, Treinys R, Macianskiene R, Bogdelis A, *et al.* beta3-adrenergic receptor activation increases human atrial tissue contractility and stimulates the L-type Ca2+ current. The Journal of Clinical Investigation. 2008; 118: 3219–3227.
- [113] Gauthier C, Sèze-Goismier C, Rozec B. Beta 3-adrenoceptors in the cardiovascular system. Clinical Hemorheology and Microcirculation. 2007; 37: 193–204.
- [114] Dessy C, Balligand JL. Beta3-adrenergic receptors in cardiac and vascular tissues emerging concepts and therapeutic perspectives. Advances in Pharmacology (San Diego, Calif.). 2010; 59: 135–163.
- [115] Brodde OE, Zerkowski HR, Borst HG, Maier W, Michel MC. Drug- and disease-induced changes of human cardiac beta 1- and beta 2-adrenoceptors. European Heart Journal. 1989; 10 Suppl B: 38–44.
- [116] Soppa GKR, Lee J, Stagg MA, Felkin LE, Barton PJR, Siedlecka U, *et al.* Role and possible mechanisms of clenbuterol in enhancing reverse remodelling during mechanical unloading in murine heart failure. Cardiovascular Research. 2008; 77: 695–706.
- [117] Lowes BD, Gill EA, Abraham WT, Larrain JR, Robertson AD, Bristow MR, *et al.* Effects of carvedilol on left ventricular mass, chamber geometry, and mitral regurgitation in chronic heart failure. The American Journal of Cardiology. 1999; 83: 1201–1205.
- [118] Lowes BD, Gilbert EM, Abraham WT, Minobe WA, Larrabee P, Ferguson D, *et al.* Myocardial gene expression in dilated cardiomyopathy treated with beta-blocking agents. The New England Journal of Medicine. 2002; 346: 1357–1365.
- [119] Papa A, Kushner J, Hennessey JA, Katchman AN, Zakharov SI, Chen BX, *et al.* Adrenergic Ca_V1.2 Activation via Rad Phosphorylation Converges at $\alpha_1 C$ I-II Loop. Circulation Research. 2021; 128: 76–88.
- [120] Cannavo A, Koch WJ. Targeting β 3-Adrenergic Receptors in the Heart: Selective Agonism and β -Blockade. Journal of Cardiovascular Pharmacology. 2017; 69: 71–78.
- [121] Rozec B, Erfanian M, Laurent K, Trochu JN, Gauthier C. Nebivolol, a vasodilating selective beta(1)-blocker, is a beta(3)adrenoceptor agonist in the nonfailing transplanted human heart. Journal of the American College of Cardiology. 2009; 53: 1532– 1538.

- [122] Kotecha D, Flather MD, Altman DG, Holmes J, Rosano G, Wikstrand J, *et al.* Heart Rate and Rhythm and the Benefit of Beta-Blockers in Patients With Heart Failure. Journal of the American College of Cardiology. 2017; 69: 2885–2896.
- [123] Poole-Wilson PA, Swedberg K, Cleland JGF, Di Lenarda A, Hanrath P, Komajda M, *et al.* Comparison of carvedilol and metoprolol on clinical outcomes in patients with chronic heart failure in the Carvedilol or Metoprolol European Trial (COMET): Randomised controlled trial. The Lancet. 2003; 362: 7–13.
- [124] Sigmund M, Jakob H, Becker H, Hanrath P, Schumacher C, Eschenhagen T, *et al.* Effects of metoprolol on myocardial betaadrenoceptors and Gi alpha-proteins in patients with congestive heart failure. European Journal of Clinical Pharmacology. 1996; 51: 127–132.
- [125] Zhuo J, Geng H, Wu X, Fan M, Sheng H, Yao J. AKT-mTOR signaling-mediated rescue of *PRKAG2* R302Q mutant-induced familial hypertrophic cardiomyopathy by treatment with β -adrenergic receptor (β -AR) blocker metoprolol. Cardiovascular Diagnosis and Therapy. 2022; 12: 360–369.
- [126] Ferguson SSG, Feldman RD. β-adrenoceptors as molecular targets in the treatment of hypertension. The Canadian Journal of Cardiology. 2014; 30: S3–S8.
- [127] Böhm M, Ungerer M, Erdmann E. Beta adrenoceptors and mcholinoceptors in myocardium of hearts with coronary artery disease or idiopathic dilated cardiomyopathy removed at cardiac transplantation. The American Journal of Cardiology. 1990; 66: 880–882.
- [128] Velmurugan BK, Baskaran R, Huang CY. Detailed insight on β -adrenoceptors as therapeutic targets. Biomedicine & Pharma-cotherapy. 2019; 117: 109039.
- [129] Frigerio M, Roubina E. Drugs for left ventricular remodeling in heart failure. The American Journal of Cardiology. 2005; 96: 10L–18L.
- [130] Zak R. Cell proliferation during cardiac growth. The American Journal of Cardiology. 1973; 31: 211–219.
- [131] Weber KT. Cardiac interstitium in health and disease: the fibrillar collagen network. Journal of the American College of Cardiology. 1989; 13: 1637–1652.
- [132] Weber KT, Brilla CG. Pathological hypertrophy and cardiac interstitium. Fibrosis and renin-angiotensin-aldosterone system. Circulation. 1991; 83: 1849–1865.
- [133] Briest W, Hölzl A, Rassler B, Deten A, Baba HA, Zimmer HG. Significance of matrix metalloproteinases in norepinephrineinduced remodelling of rat hearts. Cardiovascular Research. 2003; 57: 379–387.
- [134] Feldman AM. Modulation of adrenergic receptors and Gtransduction proteins in failing human ventricular myocardium. Circulation. 1993; 87: IV27–IV34.
- [135] Vatner DE, Asai K, Iwase M, Ishikawa Y, Shannon RP, Homcy CJ, et al. Beta-adrenergic receptor-G protein-adenylyl cyclase signal transduction in the failing heart. The American Journal of Cardiology. 1999; 83: 80H–85H.
- [136] Heger J, Schulz R, Euler G. Molecular switches under TGF β signalling during progression from cardiac hypertrophy to heart failure. British Journal of Pharmacology. 2016; 173: 3–14.
- [137] Schirone L, Forte M, Palmerio S, Yee D, Nocella C, Angelini F, et al. A Review of the Molecular Mechanisms Underlying the Development and Progression of Cardiac Remodeling. Oxidative Medicine and Cellular Longevity. 2017; 2017: 3920195.
- [138] Hu ST, Shen YF, Liu GS, Lei CH, Tang Y, Wang JF, et al. Altered intracellular Ca2+ regulation in chronic rat heart failure. The Journal of Physiological Sciences: JPS. 2010; 60: 85–94.
- [139] Chang CWJ, Lee L, Yu D, Dao K, Bossuyt J, Bers DM. Acute β -adrenergic activation triggers nuclear import of histone deacetylase 5 and delays G(q)-induced transcriptional activa-

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tion. The Journal of Biological Chemistry. 2013; 288: 192-204.

- [140] Heineke J, Molkentin JD. Regulation of cardiac hypertrophy by intracellular signalling pathways. Nature Reviews Molecular Cell Biology. 2006; 7: 589–600.
- [141] Sygitowicz G, Maciejak-Jastrzębska A, Sitkiewicz D. MicroR-NAs in the development of left ventricular remodeling and postmyocardial infarction heart failure. Polish Archives of Internal Medicine. 2020; 130: 59–65.
- [142] Bhullar SK, Dhalla, NS. Angiotensin II-induced signal transduction mechanisms for cardiac hypertrophy. Cells. 2022; 11: 3336.
- [143] Oldfield CJ, Duhamel TA, Dhalla NS. Mechanisms for the transition from physiological to pathological cardiac hypertrophy. Canadian Journal of Physiology and Pharmacology. 2020; 98: 74–84.
- [144] Brodde OE. Beta-adrenoceptors in cardiac disease. Pharmacology & Therapeutics. 1993; 60: 405–430.
- [145] Cotecchia S, Stanasila L, Diviani D. Protein-protein interactions at the adrenergic receptors. Current Drug Targets. 2012; 13: 15–27.
- [146] Molkentin JD, Dorn GW, 2nd. Cytoplasmic signaling pathways that regulate cardiac hypertrophy. Annual Review of Physiology. 2001; 63: 391–426.
- [147] Sethi R, Elimban V, Chapman D, Dixon IM, Dhalla NS. Differential alterations in left and right ventricular G-proteins in congestive heart failure due to myocardial infarction. Journal of Molecular and Cellular Cardiology. 1998; 30: 2153–2163.
- [148] Mishra S, Ling H, Grimm M, Zhang T, Bers DM, Brown JH. Cardiac hypertrophy and heart failure development through Gq and CaM kinase II signaling. Journal of Cardiovascular Pharmacology. 2010; 56: 598–603.
- [149] Bristow MR, Ginsburg R, Minobe W, Cubicciotti RS, Sageman WS, Lurie K, *et al.* Decreased catecholamine sensitivity and beta-adrenergic-receptor density in failing human hearts. The New England Journal of Medicine. 1982; 307: 205–211.
- [150] Mishra S, Kass DA. Cellular and molecular pathobiology of heart failure with preserved ejection fraction. Nature Reviews. Cardiology. 2021; 18: 400–423.
- [151] Böhm M. Alterations of beta-adrenoceptor-G-proteinregulated adenylyl cyclase in heart failure. Molecular and Cellular Biochemistry. 1995; 147: 147–160.
- [152] Wang X, Dhalla NS. Modification of beta-adrenoceptor signal transduction pathway by genetic manipulation and heart failure. Molecular and Cellular Biochemistry. 2000; 214: 131–155.
- [153] Morel E, Marcantoni A, Gastineau M, Birkedal R, Rochais F, Garnier A, *et al.* cAMP-binding protein Epac induces cardiomyocyte hypertrophy. Circulation Research. 2005; 97: 1296–1304.
- [154] Yin Q, Yang C, Wu J, Lu H, Zheng X, Zhang Y, et al. Downregulation of β-Adrenoceptors in Isoproterenol-Induced Cardiac Remodeling through HuR. PLoS ONE. 2016; 11: e0152005.
- [155] Houser SR, Molkentin JD. Does contractile Ca2+ control calcineurin-NFAT signaling and pathological hypertrophy in cardiac myocytes? Science Signaling. 2008; 1: pe31.
- [156] Backs J, Worst BC, Lehmann LH, Patrick DM, Jebessa Z, Kreusser MM, et al. Selective repression of MEF2 activity by PKA-dependent proteolysis of HDAC4. The Journal of Cell Biology. 2011; 195: 403–415.
- [157] Freedman NJ, Lefkowitz RJ. Anti-beta(1)-adrenergic receptor antibodies and heart failure: causation, not just correlation. The Journal of Clinical Investigation. 2004; 113: 1379–1382.
- [158] Carabello BA. Concentric versus eccentric remodeling. Journal of Cardiac Failure. 2002; 8: S258–S263.
- [159] Carabello BA. Models of volume overload hypertrophy. Journal of Cardiac Failure. 1996; 2: 55–64.
- [160] Galinier M, Sénard JM, Valet P, Arias A, Daviaud D, Glock Y, *et al.* Cardiac beta-adrenoceptors and adenylyl cyclase activity

in human left ventricular hypertrophy due to pressure overload. Fundamental & Clinical Pharmacology. 1994; 8: 90–99.

- [161] Vatner DE, Homcy CJ, Sit SP, Manders WT, Vatner SF. Effects of pressure overload, left ventricular hypertrophy on betaadrenergic receptors, and responsiveness to catecholamines. The Journal of Clinical Investigation. 1984; 73: 1473–1482.
- [162] Akazawa Y, Taneike M, Ueda H, Kitazume-Taneike R, Murakawa T, Sugihara R, *et al*. Rubicon-regulated beta-1 adrenergic receptor recycling protects the heart from pressure overload. Scientific Reports. 2022; 12: 41.
- [163] Brancaccio M, Fratta L, Notte A, Hirsch E, Poulet R, Guazzone S, et al. Melusin, a muscle-specific integrin beta1-interacting protein, is required to prevent cardiac failure in response to chronic pressure overload. Nature Medicine. 2003; 9: 68–75.
- [164] Okumura S, Takagi G, Kawabe JI, Yang G, Lee MC, Hong C, et al. Disruption of type 5 adenylyl cyclase gene preserves cardiac function against pressure overload. Proceedings of the National Academy of Sciences of the United States of America. 2003; 100: 9986–9990.
- [165] Wang X, Sentex E, Saini HK, Chapman D, Dhalla NS. Upregulation of beta-adrenergic receptors in heart failure due to volume overload. American Journal of Physiology. Heart and Circulatory Physiology. 2005; 289: H151–H159.
- [166] Sakagoshi N, Nakano S, Taniguchi K, Hirata N, Matsuda H. Relation between myocardial beta-adrenergic receptor and left ventricular function in patients with left ventricular volume overload due to chronic mitral regurgitation with or without aortic regurgitation. The American Journal of Cardiology. 1991; 68: 81–84.
- [167] Sullebarger J, D'Ambra P, Clark L, Thanikarry L, Fontanet H. Effect of Digoxin on Ventricular Remodeling and Responsiveness of beta-Adrenoceptors in Chronic Volume Overload. Journal of Cardiovascular Pharmacology and Therapeutics. 1998; 3: 281–290.
- [168] Wang X, Ren B, Liu S, Sentex E, Tappia PS, Dhalla NS. Characterization of cardiac hypertrophy and heart failure due to volume overload in the rat. Journal of Applied Physiology (Bethesda, Md.: 1985). 2003; 94: 752–763.
- [169] Di Fusco F, Hashim S, Anand-Srivastava MB. Volume overload cardiac hypertrophy exhibits decreased expression of g(s)alpha and not of g(i)alpha in heart. American Journal of Physiology. Cell Physiology. 2000; 279: C990–C998.
- [170] Anderson KM, Eckhart AD, Willette RN, Koch WJ. The myocardial beta-adrenergic system in spontaneously hypertensive heart failure (SHHF) rats. Hypertension (Dallas, Tex.: 1979). 1999; 33: 402–407.
- [171] Sun F, Lu Z, Zhang Y, Geng S, Xu M, Xu L, *et al.* Stage dependent changes of $\beta 2$ adrenergic receptor signaling in right ventricular remodeling in monocrotaline induced pulmonary arterial hypertension. International Journal of Molecular Medicine. 2018; 41: 2493–2504.
- [172] Böhm M, Gierschik P, Knorr A, Schmidt U, Weismann K, Erdmann E. Cardiac adenylyl cyclase, beta-adrenergic receptors, and G proteins in salt-sensitive hypertension. Hypertension (Dallas, Tex.: 1979). 1993; 22: 715–727.
- [173] Sethi R, Dhalla KS, Beamish RE, Dhalla NS. Differential changes in left and right ventricular adenylyl cyclase activities in congestive heart failure. The American Journal of Physiology. 1997; 272: H884–H893.
- [174] Ishigai Y, Mori T, Moriyama S, Shibano T. Induction of cardiac beta-adrenergic receptor kinase 1 in rat heart failure caused by coronary ligation. Journal of Molecular and Cellular Cardiology. 1999; 31: 1261–1268.
- [175] Vinge LE, Øie E, Andersson Y, Grøgaard HK, Andersen G, Attramadal H. Myocardial distribution and regulation of GRK and beta-arrestin isoforms in congestive heart failure in rats. Amer-

ican Journal of Physiology. Heart and Circulatory Physiology. 2001; 281: H2490-H2499.

- [176] Dzimiri N, Moorji A. Relationship between alterations in lymphocyte and myocardial beta-adrenoceptor density in patients with left heart valvular disease. Clinical and Experimental Pharmacology & Physiology. 1996; 23: 498–502.
- [177] Hammond HK, Roth DA, Insel PA, Ford CE, White FC, Maisel AS, *et al.* Myocardial beta-adrenergic receptor expression and signal transduction after chronic volume-overload hypertrophy and circulatory congestion. Circulation. 1992; 85: 269–280.
- [178] Cartagena G, Sapag-Hagar M, Jalil J, Tapia V, Guarda E, Foncea R, *et al.* Changes in beta-adrenergic receptors of rat heart and adipocytes during volume-overload induced cardiac hypertrophy. International Journal of Clinical Pharmacology, Therapy, and Toxicology. 1993; 31: 198–203.
- [179] Daaka Y, Luttrell LM, Lefkowitz RJ. Switching of the coupling of the beta2-adrenergic receptor to different G proteins by protein kinase A. Nature. 1997; 390: 88–91.
- [180] Rengo G, Lymperopoulos A, Leosco D, Koch WJ. GRK2 as a novel gene therapy target in heart failure. Journal of Molecular and Cellular Cardiology. 2011; 50: 785–792.
- [181] Salazar NC, Vallejos X, Siryk A, Rengo G, Cannavo A, Liccardo D, *et al.* GRK2 blockade with β ARKct is essential for cardiac β 2-adrenergic receptor signaling towards increased contractility. Cell Communication and Signaling: CCS. 2013; 11: 64.
- [182] Iaccarino G, Dolber PC, Lefkowitz RJ, Koch WJ. Bbetaadrenergic receptor kinase-1 levels in catecholamine-induced myocardial hypertrophy: regulation by beta- but not alphaladrenergic stimulation. Hypertension (Dallas, Tex.: 1979). 1999; 33: 396–401.
- [183] Karliner JS, Barnes P, Brown M, Dollery C. Chronic heart failure in the guinea pig increases cardiac alpha 1- and betaadrenoceptors. European Journal of Pharmacology. 1980; 67: 115–118.
- [184] Fan TH, Liang CS, Kawashima S, Banerjee SP. Alterations in cardiac beta-adrenoceptor responsiveness and adenylate cyclase system by congestive heart failure in dogs. European Journal of Pharmacology. 1987; 140: 123–132.
- [185] Karam S, Margaria JP, Bourcier A, Mika D, Varin A, Bedioune I, *et al.* Cardiac Overexpression of PDE4B Blunts β-Adrenergic Response and Maladaptive Remodeling in Heart Failure. Circulation. 2020; 142: 161–174.
- [186] Streit MR, Weiss CS, Meyer S, Ochs MM, Hagenmueller M, Riffel JH, *et al.* Cardiac Effects of Attenuating $Gs\alpha$ - Dependent Signaling. PLoS ONE. 2016; 11: e0146988.
- [187] Cantor EJF, Babick AP, Vasanji Z, Dhalla NS, Netticadan T. A comparative serial echocardiographic analysis of cardiac structure and function in rats subjected to pressure or volume overload. Journal of Molecular and Cellular Cardiology. 2005; 38: 777–786.
- [188] Carabello BA, Zile MR, Tanaka R, Cooper G, 4th. Left ventricular hypertrophy due to volume overload versus pressure overload. The American Journal of Physiology. 1992; 263: H1137– H1144.
- [189] Modesti PA, Vanni S, Bertolozzi I, Cecioni I, Polidori G, Paniccia R, *et al.* Early sequence of cardiac adaptations and growth factor formation in pressure- and volume-overload hypertrophy. American Journal of Physiology. Heart and Circulatory Physiology. 2000; 279: H976–H985.
- [190] Plehn JF, Foster E, Grice WN, Huntington-Coats M, Apstein CS. Echocardiographic assessment of LV mass in rabbits: models of pressure and volume overload hypertrophy. The American Journal of Physiology. 1993; 265: H2066–H2072.
- [191] Shah AK, Bhullar SK, Elimban V, Dhalla NS. Oxidative Stress as A Mechanism for Functional Alterations in Cardiac Hypertro-

phy and Heart Failure. Antioxidants (Basel, Switzerland). 2021; 10: 931.

- [192] Bartekova M, Radosinska J, Jelemensky M, Dhalla NS. Role of cytokines and inflammation in heart function during health and disease. Heart Failure Reviews. 2018; 23: 733–758.
- [193] Gronda E, Lovett EG, Tarascio M, Georgakopoulos D, Grassi G, Vanoli E. The baroreceptor as a therapeutic target for heart failure. Journal of Cardiovascular Translational Research. 2014; 7: 301–309.
- [194] Piani F, Landolfo M, Fiorini G, D'Addato S, Mancia G, Borghi C. Severe impaired blood pressure control caused by baroreflex failure as a late sequela of neck irradiation. Journal of Hypertension. 2020; 38: 553–556.
- [195] Zile MR, Lindenfeld J, Weaver FA, Zannad F, Galle E, Rogers T, *et al.* Baroreflex Activation Therapy in Patients with Heart

Failure with Reduced Ejection Fraction. Journal of the American College of Cardiology. 2020; 76: 1–13.

- [196] Rundqvist B, Eisenhofer G, Elam M, Friberg P. Attenuated cardiac sympathetic responsiveness during dynamic exercise in patients with heart failure. Circulation. 1997; 95: 940–945.
- [197] Zipes DP, Neuzil P, Theres H, Caraway D, Mann DL, Mannheimer C, *et al.* Ventricular Functional response to spinal cord stimulation for advanced heart failure: primary results of the randomized DEFEAT-HF Trial. Circulation. 2014; 130: 2105–2126.
- [198] Zannad F, De Ferrari GM, Tuinenburg AE, Wright D, Brugada J, Butter C, *et al.* Chronic vagal stimulation for the treatment of low ejection fraction heart failure: results of the NEural Cardiac TherApy foR Heart Failure (NECTAR-HF) randomized controlled trial. European Heart Journal. 2015; 36: 425–433.

