

Original Research

Two Adverse Early Life Events Induce Differential Changes in Brain CRH and Serotonin Systems in Rats along with Hyperphagia and Depression

Viridiana Alcántara-Alonso¹, Cinthia García-Luna¹, Paulina Soberanes-Chávez¹, Erika Estrada-Camarena², Patricia de Gortari^{1,*}¹Lab. de Neurofisiología Molecular, Dirección de Neurociencias, Instituto Nacional de Psiquiatría Ramón de la Fuente Muñiz, 14370 Mexico City, Mexico²Lab. de Neuropsicofarmacología, Dirección de Neurociencias, Instituto Nacional de Psiquiatría Ramón de la Fuente Muñiz, 14370 Mexico City, Mexico*Correspondence: gortari@imp.edu.mx (Patricia de Gortari)

Academic Editor: Yoshihiro Noda

Submitted: 28 July 2023 Revised: 14 November 2023 Accepted: 16 November 2023 Published: 20 February 2024

Abstract

Background: Different types of stress inflicted in early stages of life elevate the risk, among adult animals and humans, to develop disturbed emotional-associated behaviors, such as hyperphagia or depression. Early-life stressed (ELS) adults present hyperactivity of the hypothalamus-pituitary-adrenal (HPA) axis, which is a risk factor associated with mood disorders. However, the prevalence of hyperphagia (17%) and depression (50%) is variable among adults that experienced ELS, suggesting that the nature, intensity, and chronicity of the stress determines the specific behavioral alteration that those individuals develop. **Methods:** We analyzed corticosterone serum levels, *Crh*, *GR*, *Crhr1* genes expression in the hypothalamic paraventricular nucleus, amygdala, and hippocampus due to their regulatory role on HPA axis in adult rats that experienced maternal separation (MS) or limited nesting material (LNM) stress; as well as the serotonergic system activity in the same regions given its association with the corticotropin-releasing hormone (CRH) pathway functioning and with the hyperphagia and depression development. **Results:** Alterations in dams' maternal care provoked an unresponsive or hyper-responsive HPA axis function to an acute stress in MS and LNM adults, respectively. The differential changes in amygdala and hippocampal CRH system seemed compensating alterations to the hypothalamic desensitized glucocorticoids receptor (GR) in MS or hypersensitive in LNM. However, both adult animals developed hyperphagia and depression-like behavior when subjected to the forced-swimming test, which helps to understand that both hypo and hypercortisolemic patients present those disorders. **Conclusion:** Different ELS types induce neuroendocrine, brain CRH and 5-hydroxytryptamine (5-HT) systems' alterations that may interact converging to develop similar maladaptive behaviors.

Keywords: early life stress; abandonment; neglect maternal care; hyperphagia; depression; CRH; serotonin

1. Introduction

Different psychiatric disorders such as depression, anxiety and hyperphagia-induced obesity [1–5] develop during adulthood due to alterations in the hypothalamic-pituitary-adrenal (HPA) axis function [6–9] resulting after being exposed to a variety of stressful events during early life periods.

Threatening experiences in early childhood, such as abandonment, neglect, physical or sexual abuse, parental illnesses or poverty, among other adversities, induce the disruption of the HPA axis functioning, given that it is a particularly susceptible period to be impaired by stressful events [8].

Chronic exposure to early-life stress (ELS) is specifically associated with hyperactivity of the HPA axis characterized by elevated glucocorticoids serum levels (cortisol in humans, corticosterone (CORT) in rodents), along with

an impaired negative feedback regulation of the axis, recognized by high hypothalamic corticotropin-releasing hormone (CRH) mRNA and protein content in the paraventricular nucleus of the hypothalamus (PVN), which contains the CRH neurons involved in directing the functioning of the neuroendocrine axis [10]. The high serum glucocorticoids levels in early life periods, when sustained during long terms, are also able to disrupt different brain neuropeptides and neurotransmitters' systems, such as those of CRH and serotonin (5-hydroxytryptamine, 5-HT) particularly in the amygdala (Amy) and hippocampus (Hipp) [11,12].

Amygdalar and hippocampal CRHergic neurons project their axons to the hypothalamic PVN and are known to activate and inhibit, respectively, the functioning of the HPA axis. Thus, alterations in the expression of their components, such as that of *Crh*, type 1 corticotropin-releasing hormone receptor (*Crhr1*), and glucocorticoids receptor (*GR*) of those regions, may account for the impairments found in neuroendocrine and neurobiological sys-



tems in early-life stressed offspring that persist until adulthood [3,9,13,14] and that underly the development of depression or hyperphagia-induced obesity [4,15].

Besides that of CRH, alterations in the serotonin system are also involved in the development of depressive-like behaviors and in stress-induced hyperphagia [11,12]. In fact, CRH itself, as well as corticosterone, are able to modify 5-HT neurotransmission, supporting the assumption that early-life stress-induced impairments in CRH and in 5-HT systems are linked to specific maladaptive behavioral outcomes in adults.

However, although the participation of altered amygdalar, hippocampal and hypothalamic CRH and 5-HT pathways in the development of psychiatric disorders is well accepted, there is no description of specific changes to their components that may account for various behavioral maladaptations that lead to depression or to hyperphagia-induced obesity among adults that suffered from early-life stress events.

Therefore, we subjected rats to two different models of early life stress that mimic childhood abandonment (maternal separation, MS) or neglect (limited nesting material, LNM) and evaluated their food intake and body weight since weaning and up to adulthood, as well as their depression-like behavior in adulthood by subjecting them to the forced swim test (FST). FST is a paradigm to evaluate neurobiological changes related to coping strategies for stressors [16]. It is reported that if a stressor persists, it may increase the vulnerability to develop depression-like behaviors [17]. From this point of view, forced swim could be adequate to detect behavioral changes induced by early-stress interventions, such as the increase of immobility behavior [18]. MS consists in separating the offspring from their mothers 3 h daily during post-natal days 2 to 14 [19], whereas LNM refers to an erratic behavior of dams resulting from having limited amount of nest material during lactation [20]. Those paradigms induce fragmented maternal care in the weaning dams that increases their corticosterone serum levels and those of the offspring, leading to HPA axis alterations known to underly depression-like behavior, as well as increased food intake and body weight, in ten-week old adult rats [15]. Groups with or with no performance on the FST were sacrificed at the same time, and their trunk blood collected for evaluating corticosterone serum levels as evidence of stress and of the response to an acute challenge, the FST. Also, we extracted the brains and analyzed the genes' expression of *Crh*, *Crhr1*, *GR*, as well as the activity of 5-HT pathway in PVN, amygdala, and hippocampus of adult rats, and compared those parameters between groups from the two different early-life stress paradigms and between animals with or with no performance of the FST.

We hypothesized that both groups of rats will show differential behavioral alterations linked to specific modifications in HPA axis-functioning parameters, as well as

in the expression of brain CRHergic system components or in serotonin pathway activity that explain the diversity of mood disorders developed in adults exposed to various early-life adversities.

2. Materials and Methods

2.1 Animals

Twelve pregnant Wistar rats in the third week of gestation were obtained from the animal house of the Instituto Nacional de Psiquiatría Ramón de la Fuente Muñiz (IN-PRFM), and maintained in a 12 h light-dark cycle (lights on 07:00–19:00 h) under controlled temperature (22 ± 1 °C) and ad libitum access to water and food (Lab rodent diet # 5001, PMI Nutrition International; Brentwood, MO, USA) during gestation and lactation periods. All procedures were conducted with the approval of the local Ethics Committee for Animal Experimentation of the INPRFM (CEI/C/024/2014) following the guidelines outlined by the Mexican Official Standard NOM-0620ZOO-1999. Animals were checked daily for delivery, considering it as day 0. On postnatal day (PND) 2, rats (dams and pups) were randomly separated in 3 experimental groups (Fig. 1):

(1) Control group (C): Dams and pups ($n = 4$) were left undisturbed and, after weaning (PND 21), dams were euthanized by decapitation, and pups were individually housed and left undisturbed until PND 71.

(2) Maternal separation group (MS): From PND 2 to 14, dams ($n = 4$) were withdrawn from their home cages to an adjacent room, while pups were left in their home cages for 180 min/day during the light period (10:00 to 13:00 h). At the end of the separation period, dams were returned to the nest cage. On PND 15, dams and pups were left undisturbed and housed in cages like C group. After weaning (PND 21), dams were euthanized, and pups were individually housed and left undisturbed until PND 71.

(3) Limited nesting material group (LNM): From PND 2 to PND 9, dams ($n = 4$) and their pups were placed on a bored metal surface 2.5 cm above the cage floor that contained sawdust to pick up droppings throughout the day. The only material offered to the dams to build the nest was one paper towel, according to the model described by Baram's group [21]. On PND 10, the metal surface was removed, and dams and pups were housed in cages like C group. After weaning (PND 21), dams were decapitated, and pups were individually housed and left undisturbed until PND 71.

At PND 21, all dams were sacrificed to measure corticosterone serum levels and adrenal glands' weight. Male offspring ($n = 12$ /group) were weaned, single housed, and maintained under controlled conditions with ad libitum access to food and water up to adulthood (PND 70). Since single housing is considered social isolation and a stressful condition, all our animals in fact were exposed to two different stressors at different periods of life [22]. Offspring food intake and body weight were registered on a weekly

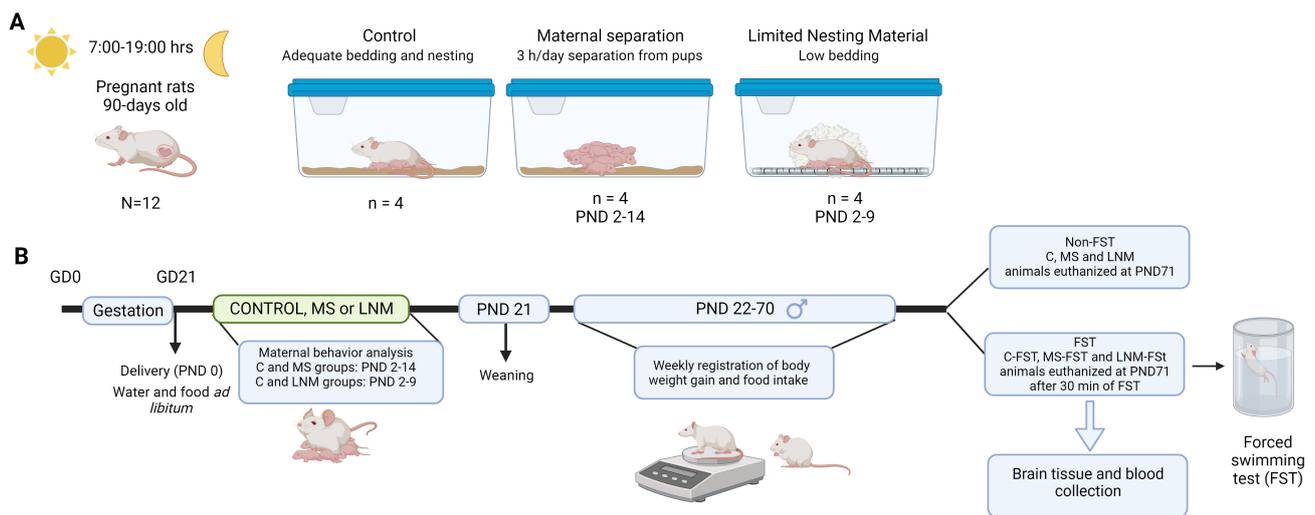


Fig. 1. Experimental design. (A) Housing conditions of lactating dams and offspring. Pregnant rats were checked for delivery that was considered postnatal day (PND) 0. After delivery, dams and offspring were divided in control group (C: adequate bedding material and all the time together in the same cage), maternal separation group (MS: dams were withdrawn from home cages 3 h/day from PND 2 to 14), and limited nesting material group (LNM: from PND 2 to 9, dams and pups were together but the material provided for build the nest was not enough). (B) Timeline of the experimental design. After delivery (PND 0), rats were subjected to C, MS or LNM conditions; after weaning, dams were euthanized and pups were singly housed and left undisturbed until PND 71 when a subset of C, MS and LNM rats were euthanized, and another subset of rats were subjected to the forced swimming test (FST; C-FST, MS-FST, LNM-FST), and after 30 min of FST, they were euthanized. Brains and blood were collected. Created with Biorender. GD, gestation day; MS, maternal separation; PND, postnatal day; LNM, limited nesting material; FST, forced swim test.

basis. On PND 71, a set of animals from each group (4 rats/group) were euthanized by decapitation (C, MS, LNM) and another set of rats (8 animals/group) were subjected to the FST and euthanized after 30 min (C-FST, MS-FST, LNM-FST). Trunk blood was collected, and the serum was obtained by centrifugation at 3000 rpm and used to analyze corticosterone levels. Brains were excised and stored at -70°C until analyzed for *Crh*, *Crhr1* and *GR* mRNA expression ($n = 4/\text{group}$), or for 5-HT and 5-hydroxyindole acetic acid (5-HIAA) content ($n = 4/\text{group}$). PVN, Amy and Hipp were hand dissected from coronal slices of frozen brain: for PVN and Amy -1.08 to -3.24 mm from bregma, for Hipp -3.72 to -5.16 mm from bregma [23]. The right hemisphere was used for mRNA content determinations, and the left hemisphere was used for 5-HT and 5-HIAA measurements.

2.2 Maternal Care

Maternal care behaviors were analyzed in C, MS, and LNM dams. All animals were videotaped for a 30-minute period between 8 AM and 2 PM and different behaviors were analyzed. For MS group, the recordings were made after returning dams to home cages and compared to a control dams' group who were separated from their pups for 5 minutes from PND 2 to 14. For LNM rats, we recorded the animals' behavior in their home cages from PND 2 to 9. We assessed the spent time of mothers engaged in different behaviors, such as being near the litter, outside the

nest, grooming the pups and themselves, passive nursing, and arched-nursing posture. We also counted the number of pups that the dams left out of the care area while nursing [24].

2.3 Forced Swim Test (FST)

On PND 70, a subset of animals ($n = 8/\text{group}$) were allowed to swim for 15 min (pre-test) in a modified version of the Porsolt's FST (C-FST, MS-FST, LNM-FST groups) [25]. Briefly, each rat was placed in a glass cylinder (46 cm tall \times 20 cm in diameter) containing water (30 cm depth) at $24 \pm 1^{\circ}\text{C}$. On PND 71, the same animals were subjected to a 5 min videotaped swimming test session. At the end of each session, rats were removed from the container and dried before returning to their home cages. Animals were sacrificed by decapitation 30 min after the FST, blood was extracted to evaluate their corticosterone serum levels, and brains were excised and stored at -70°C until used. We analyzed every 5 s of FST videos to identify the display of the following behaviors: (1) immobility: floating, making only movements needed to keep the head above the water; (2) swimming: active movements, diving and moving around the cylinder; (3) climbing: directing and moving forepaws against the wall of the container [25–27].

Table 1. RT-PCR primers for gene expression semi-quantification.

Gene name	Protein	Sense primer (5'-3')	Antisense primer (5'-3')	Product length (bp)	NCBI reference sequence
<i>Nr3c1</i>	Glucocorticoids receptor	AAAAAGCACATCACA CATAAATCTG	TAAATAAGAGGGGAGC AAACTACTGG	688	NM_012576.3
<i>Crh</i>	Corticotropin releasing hormone	AGAAGAGAGCGCCCC TAAAC	ATCAGAATCGGCTGA GGTTG	190	NM_031019.2
<i>Crhr1</i>	Corticotropin releasing hormone receptor 1	TCCACTACATCTGAGA CCATTCAGTACA	TCCTGCCACCGGCGC CACCTCTTCCGGA	248	NM_030999.5
<i>Ppia</i>	Cyclophilin	GGGGAGAAAGGATTT GGCTA	ACATGCTTGCCATCC AGCC	257	NM_017101.1

RT-PCR, reverse transcriptase polymerase chain reaction; NCBI, National Center for Biotechnology Information.

2.4 Determination of Serum Corticosterone Levels

We used 50 μ L of each rat' serum to determine corticosterone content using a radioimmunoassay (RIA) kit (Coat-a-Count, Siemens, Los Angeles, CA, USA): limit of detection 5.7 ng/mL, inter and intra-assay variation 15 and 13%, respectively.

2.5 mRNA Semi-quantification by RT-PCR

For total RNA extraction, frozen PVN, Amy and Hipp were homogenized in 4 M guanidine thiocyanate (ICN Biomedical Research Products, Aurora, OH, USA) and treated as described [28]. RNA quality was confirmed by 260/280 nm and 260/230 nm ratios of O.D. absorbance, in addition to an agarose gel electrophoresis to analyze RNA integrity by quantifying 28S/18S ratio, considering an optimal extraction when both values were >1.8 . Levels of *Crh*, *Crhr1* and *GR* mRNAs were semi-quantified by reverse transcriptase polymerase chain reaction (RT-PCR); cyclophilin (CYC) transcript was used as control [29]. cDNA was obtained using 1.5 μ g of RNA, Moloney-Murine Leukemia Virus (M-MLV) reverse transcriptase (Invitrogen, Carlsbad, CA, USA) and oligo-dT, followed by PCR reaction: 6 μ L of cDNA, specific oligonucleotides (25 pmol for each problem primer or 50 pmol for CYC primer) and 0.5 μ L Taq DNA polymerase (5 U/ μ L) (Biotecnologías Universitarias, Universidad Nacional Autónoma de México, Mexico), as previously described [29]. Specific primers and product sizes are listed in Table 1.

The number of cycles was optimized for each primer in a particular region. In the Amy, 29 cycles were used for pro-CRH, 31 cycles for CRH-R1, 30 cycles for GR, and 21 cycles for CYC. In the Hipp, 30 cycles were performed for pro-CRH, 32 cycles for CRH-R1, 29 cycles for GR, and 21 cycles for CYC. In the PVN, 28 cycles were used for pro-CRH, 30 cycles for CRH-R1, 27 cycles for GR, and 21 cycles for CYC. Each cycle consisted of 1 min at 94 $^{\circ}$ C, followed by 1 min at 64 $^{\circ}$ C for GR or CYC, 55 $^{\circ}$ C for CRH-R1 or 63 $^{\circ}$ C for pro-CRH, and a final minute at 72 $^{\circ}$ C. All cDNAs had a final extension of 10 min at 72 $^{\circ}$ C. PCR products (10 μ L of each problem gene, and 5 μ L of CYC)

were separated by 2% agarose gel electrophoresis (Ultra-pure Bio-Rad, Hercules, CA, USA), stained with ethidium bromide (1 mg/L; Sigma-Aldrich, St. Louis, MO, USA), and revealed under a UV light. The optical density (O.D.) was measured with the Advanced American Biotech Imaging software (American-Applied Biotechnology, Fullerton, CA, USA), and the relative expression of mRNA was calculated as the ratio of each OD problem gene over that of CYC.

2.6 Determination of Serotonin (5-HT) and 5-hydroxyindoleacetic Acid (5-HIAA) Content by High Performance Liquid Chromatography (HPLC)

C, MS, and LNM adult offspring samples were homogenized in ice-cold 0.1 M perchloric acid (Sigma-Aldrich, St. Louis, MO, USA) (PVN in 400 μ L, Amy in 500 μ L and Hipp in 600 μ L), and a 30 μ L aliquot was used to determine protein content by the Folin-Ciocalteu method [30]. The remaining homogenate was filtered to dispose of impurities (0.25 μ m pore, Whatman, Piscataway, NJ, USA).

High-performance liquid chromatography (HPLC) system consisted of an auto-sampler (Waters Alliance 2695, Milford, MA, USA) equipped with a Phenomenex® pre-column to filter impurities, an analytical Phenomenex C-18 column (150 \times 1 mm with 5- μ m silica particles), and electrochemical detector (Waters 2465, Milford, MA, USA). The composition of the mobile phase was similar to previous reports [31]. A stock buffer was prepared as follows: 25 mM sodium dihydrogen orthophosphate, 27 μ M ethylene-diamine tetra-acetic acid (EDTA), 50 mM sodium citrate, 10 mM diethylamine, 10 mM sodium chloride, and 2 mM sodium decanesulfonate dissolved in 1 L of deionized distilled water, pH 3.1. The mobile phase flux was 0.1 mL/min. The working electrode potential was set at +750 mV versus an Ag/AgCl reference electrode. Detector sensitivity was maintained at 20 nA full-scale detection, with injection volume being 5 μ L at 25 $^{\circ}$ C.

Standard solutions were prepared from a 1 mg/mL stock standard of each analyte and dissolved in ice-cold 0.1 M perchloric acid. Retention time was 7.2 minutes for 5-HT

Table 2. Corticosterone serum content, adrenal glands size and maternal behavior of control (C), maternal separation (MS) and limited-nest material (LNM) dams.

	Group		
	Control	MS	LNM
Cort (ng/mL)	178.9 ± 51.6	591 ± 71.8**	399.6 ± 64.2*
Adrenal glands (g/kg b.w.)	0.3 ± 0.009	0.31 ± 0.025*	0.31 ± 0.007*
Maternal care (min)			
Outside the nest	12.6 ± 2.0	5.9 ± 0.1.0**	12.3 ± 1.3 [#]
Passive nursing	6.3 ± 1.2	11.6 ± 1.4**	9.8 ± 1.4
Arched nursing posture	1.2 ± 0.31	0.9 ± 0.4	2.3 ± 0.4 [#]
Grooming the pups	7.4 ± 0.9	11 ± 1.1*	4.0 ± 0.5*,###
Self-grooming	2.7 ± 0.5	0.7 ± 0.22***	1.6 ± 0.3
Pups outside the nest	1.3 ± 0.4	1.2 ± 0.5	3.3 ± 0.4**, [#]
Total number of male pups PND 1	16	16	12
Total number of female pups PND 1	15	16	12

Data are presented as mean ± standard error of mean (SEM). n = 4 rats/group * $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$ vs C; [#] $p < 0.05$, and ### $p < 0.001$ vs MS. PND, postnatal day.

standards and 2.9 min for 5-HIAA standards. Data was acquired using the Waters Empower software for HPLC (Milford, MA, USA).

2.7 Statistical Analysis

Differences between groups of dams in corticosterone serum content and pups' caring behavior were analyzed by one-way analysis of variance (ANOVA). Comparisons of body weight and food intake from weeks 3 to 10 after weaning were analyzed by two-way repeated measures ANOVA. The scores of behavioral parameters (FST) were analyzed by one-way ANOVA, while differences between pups' groups in adulthood for gene expression, serotonin metabolism, and corticosterone serum levels were analyzed by two-way ANOVA (groups with no FST and those subjected to FST). When $p < 0.05$, analyses were followed by Fisher's post-hoc test. All analyses were performed using the StatView software (Abacus Concepts, Piscataway, NJ, USA).

3. Results

3.1 ELS on HPA Axis and Maternal Behavior of Dams

3.1.1 HPA Response of Dams Subjected to Stress in Lactation

Serum CORT levels of dams subjected to MS and LNM were elevated to 330 and 223%, respectively, compared to C dams (100%; 178.9 ± 52 ng/mL). One-way ANOVA showed differences between groups ($F_{(2,22)} = 8.976$, $p < 0.05$; Table 2).

Accordingly, the weight of adrenal glands of the same dams increased to 125 and 128% in MS and LNM dams, respectively, compared to that of C dams (100%; 0.25 ± 0.01 g/kg B.W.). One-way ANOVA showed differences between groups ($F_{(2,13)} = 4.845$, $p < 0.05$; Table 2). Both CORT and adrenal glands' weight showed that MS and LNM dams were under chronic stress.

3.1.2 Maternal Care of Dams Subjected to Stress during Lactation

We analyzed the stress effect on maternal behaviors. We found that the time MS dams spent outside the nest was 47% less than C (100%; 12.6 ± 2 min) and LNM groups; as well as the time spent of MS dams grooming themselves vs C (100%; 2.7 ± 0.5 min; Table 2). One-way ANOVA exhibited differences between groups (time outside the nest: $F_{(2,49)} = 7.628$, $p < 0.005$; self-grooming time: $F_{(2,49)} = 6.689$, $p < 0.005$). On the contrary, the time MS dams spent nursing and grooming the pups increased to 184% and 149%, respectively, when compared to C dams (100%; passive nursing: 6.3 ± 1.2 min; grooming the pups: 7.4 ± 0.9 min). The LNM group did not show differences in passive nursing behavior; in contrast, LNM dams decreased the time spent grooming the pups to 53% vs C rats (Table 2). One way ANOVA showed differences for passive nursing ($F_{(2,49)} = 6.822$, $p < 0.01$) and pup grooming ($F_{(2,49)} = 15.75$, $p < 0.001$) behaviors. We also observed that the number of pups left outside the nest was 260% higher in the LNM group than in C and MS rats (C = 100%; 1.3 ± 0.4 pups; Table 2). One-way ANOVA showed difference between groups ($F_{(2,24)} = 6.689$, $p < 0.05$). These behavioral changes could be due to an overcompensation of the time spent away from the pups by MS rats, and to a fragmented maternal care of the pups by LNM dams.

3.1.3 Body Weight and Food Intake of Adult Male Rats Exposed to Early Postnatal Stress

Both MS and LNM offspring displayed more body weight than C group, reaching 12 and 24% at adulthood (week 10; C = 100%, 202 ± 4.6 g). Moreover, LNM animals exhibited 11% more body weight than MS at the end of the experiment (LNM: 251 ± 7.1 g; Fig. 2A). Two-way repeated measures ANOVA showed an effect of groups

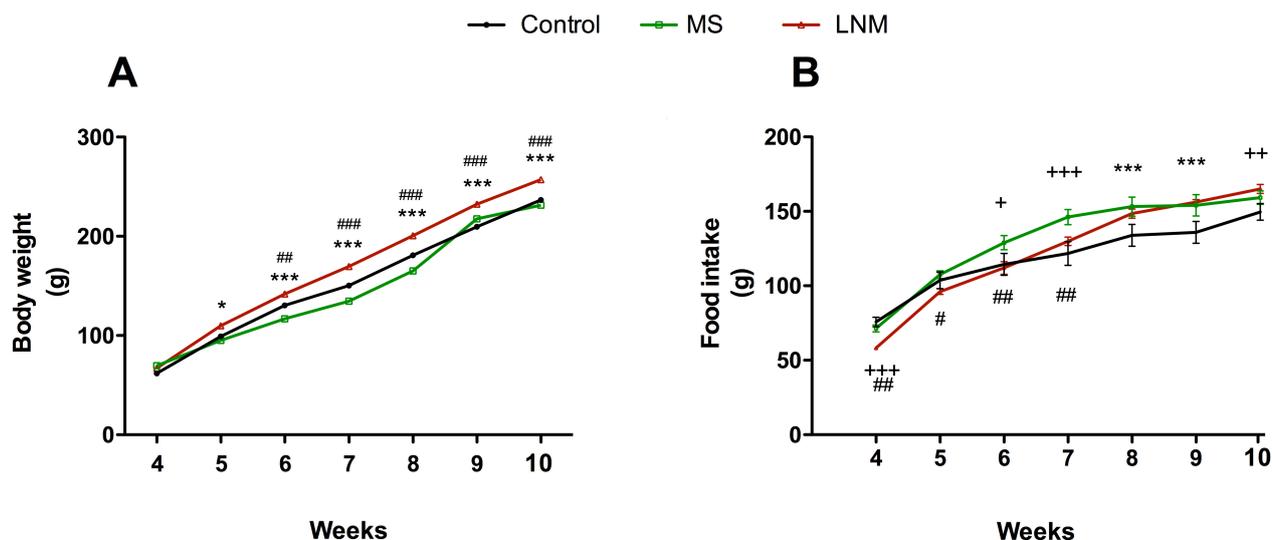


Fig. 2. Body weight and food intake of control (C), maternal separation (MS), and limited-nest material (LNM) offspring from weeks 4 to 10 after delivery. (A) Body weight from 3 to 10 weeks. (B) Weekly food intake from 4 to 10 weeks. Data are expressed in grams (g) and presented as the mean \pm standard error of mean (SEM); $n = 10$ rats/group. * $p < 0.05$, *** $p < 0.001$ of MS and LNM vs C, # $p < 0.05$, ## $p < 0.01$ and ### $p < 0.001$ vs MS; + $p < 0.05$, ++ $p < 0.01$, +++ $p < 0.001$ in week 5 LNM vs C for body weight, and in weeks 6 and 7 MS vs C and in weeks 4 and 10 LNM vs C for food intake.

($F_{(2,150)} = 27.89, p < 0.001$), time ($F_{(6,150)} = 91.30, p < 0.001$), but not in the variables interaction.

Regarding food intake, MS rats ingested 13% more food than C group only between weeks 6 and 9 (C= 100%; week 6: 114 ± 7.3 g; week 9: 139 ± 7.3 g). The LNM group increased it 11% from week 8 to 10 compared to C (C=100%; week 10: 150 ± 5.6 g), but results were similar to MS rats (Fig. 2B). Two-way repeated measures ANOVA showed an effect of time ($F_{(2,264)} = 29.55, p < 0.001$), in the interaction between variables ($F_{(12,264)} = 5.97, p < 0.001$), but not in groups.

3.1.4 Depression-like Behavior of Adult Male Rats Exposed to Early Postnatal Stress

Swimming and climbing behaviors were not different between groups subjected to FST (Table 3). However, both MS-FST and LNM-FST rats exhibited a 54% higher immobility behavior when compared to C rats (100%, 13 ± 0.8 immobility episodes; Table 3) suggesting that both early postnatal stress paradigms induced depression in adult rats. One-way ANOVA showed differences between groups ($F_{(2,55)} = 8.74, p < 0.001$).

3.1.5 Corticosterone Serum Levels and *Crh*, *GR* mRNA Expression in the PVN, and *Crh*, *Crhr1*, and *GR* in Amygdala and Hippocampus of FST-performing Adult Male Rats Exposed to Early Postnatal Stress

The MS group showed an increase of 197% in CORT serum content when compared to those of C animals (100%; 236 ± 111 ng/mL) and of 182% vs that of LNM rats (Fig. 3A). C-FST and LNM-FST groups increased their

Table 3. Depression-like behavior in the forced swimming test (FST) of control (C), maternal separation (MS), and limited-nest material (LNM) adult offspring.

Frequency	Group		
	Control	MS	LNM
Immobility	13.0 ± 0.9	$20.1 \pm 1.8^{***}$	$20.2 \pm 1.7^{**}$
Swimming	34.9 ± 1.5	30.3 ± 2.1	30 ± 1.9
Climbing	12.1 ± 1.3	9.5 ± 1.5	10.7 ± 1.2

Frequency is referred as the total 5 sec counts of each behavior in the FST over a 5 min period. $n = 8$ rats/group. Data represent the mean \pm SEM. ** $p < 0.01$ and *** $p < 0.001$ vs C.

CORT serum concentration by 317% and 341%, respectively, when compared to their respective group with no FST. Interestingly, MS-FST rats did not increase CORT serum levels, which remained similar to those of MS (Fig. 3A). Two-way ANOVA showed differences in the behavioral test exposure ($F_{(1,22)} = 58.1, p < 0.001$) and in the interaction of groups and behavioral test ($F_{(2,22)} = 18.18, p < 0.001$).

MS and LNM groups increased *Crh* mRNA content in the PVN to 241 and 150% respectively, compared to that of C animals (100%, 1.3 ± 0.07 a.u.). The C-FST group showed an increase in *Crh* expression to 206% when compared to C rats. Interestingly, MS-FST showed a decrease in *Crh* expression to 26% when compared to its non-FST group (MS), but LNM-FST rats showed a similar increase as that of the LNM group. However, both MS-FST and LNM-FST groups exhibited 76 and 32% more *Crh* mRNA content than C rats, respectively (Fig. 3B). Two-

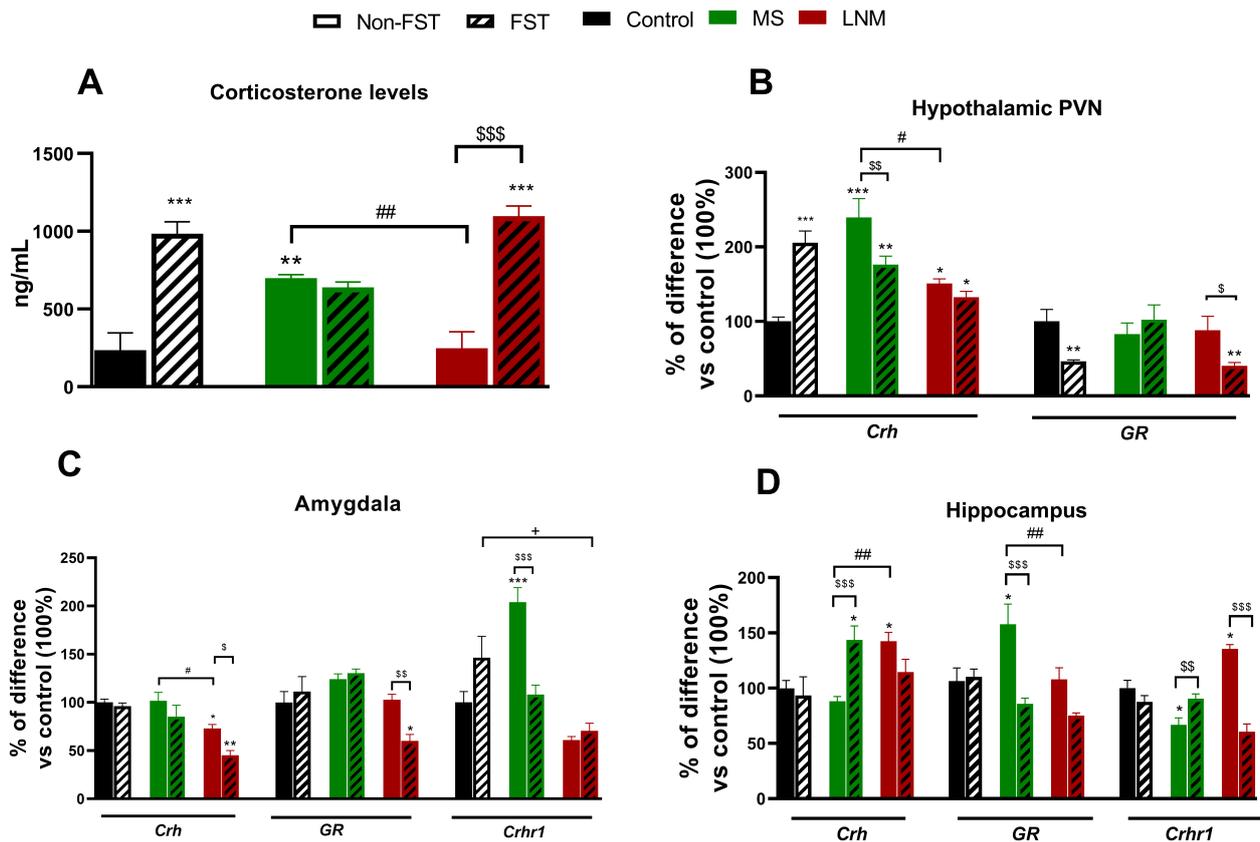


Fig. 3. Corticosterone serum content and central gene expression of *Crh*, *GR* and *Crhr1* in control (C), maternal separation (MS), and limited nest material (LNM) adult offspring that were or not subjected to the forced swimming test (FST). (A) Serum corticosterone levels (ng/mL); mRNA expression of *Crh*, *GR* and *Crhr1* expressed in percentage of difference vs C in (B) hypothalamic paraventricular nucleus (PVN), (C) amygdala and (D) hippocampus of non-FST rats (C, MS and LNM) (open bars) and rats subjected to the forced swimming test (FST; C-FST, MS-FST, LNM-FST) (lines in bars), $n = 4-6$ rats/group. Data represent mean \pm S.E.M. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs non-FST C group; # $p < 0.05$, ## $p < 0.01$ vs non-FST MS group; \$ $p < 0.05$, \$\$ $p < 0.01$, \$\$\$ $p < 0.001$ vs its respective-FST group; + $p < 0.05$ C-FST vs LNM-FST. *Crh*, corticotropin-releasing hormone; *GR*, glucocorticoid receptor; *Crhr1*, type 1 corticotropin-releasing hormone receptor.

way ANOVA showed differences between groups ($F_{(2,57)} = 7.58, p < 0.001$) and in the interaction between variables ($F_{(2,57)} = 14.89, p < 0.001$).

Regarding *GR* mRNA levels, MS and LNM rats that were not subjected to the FST had not changed. In contrast, C-FST and LNM-FST decreased *GR* expression in the PVN to 46% and 40% to that of C group (100%, 3.13 ± 0.5 a.u.), as well as LNM-FST decreased to 46% when compared to the LNM group with no FST. MS-FST rats did not exhibit *GR* expression changes when compared to C or MS (Fig. 3B). Two-way ANOVA showed differences between groups ($F_{(2,14)} = 3.53, p < 0.05$), behavioral test ($F_{(1,14)} = 5.29, p < 0.05$), and the interaction between variables ($F_{(2,14)} = 3.70, p = 0.05$).

In the Amy of LNM rats, we found a reduction of 27% in *Crh* mRNA expression compared to C group (100%; 1.07 ± 0.05 a.u.). Furthermore, *Crh* mRNA content in the LNM-FST group exhibited a more pronounced decrease reaching 62% of LNM values and 45% of that of C rats.

MS and MS-FST groups did not show differences vs C rats (Fig. 3C). Two-way ANOVA showed differences between groups ($F_{(2,20)} = 18.79, p < 0.001$) and behavioral test ($F_{(1,20)} = 7.23, p < 0.01$). *GR* mRNA expression in the Amy only decreased in LNM-FST animals to 60% of C group (100%; 1.90 ± 0.26 a.u.) (Fig. 3C). Two-way ANOVA showed differences between groups ($F_{(2,19)} = 13.77, p < 0.001$) and in the interaction between variables ($F_{(2,19)} = 9.3, p < 0.01$). The expression of *Crhr1* mRNA in the Amy was significantly higher in the MS group, increasing 204% when compared to C group (C=100%; 1.22 ± 0.22 a.u.; Fig. 3A). In contrast, MS-FST exhibited a similar *Crhr1* expression vs C, but a 47% reduction when compared to MS rats. LNM-FST showed a significant decrease in comparison to C-FST (Fig. 3A). Two-way ANOVA showed significant differences between groups ($F_{(2,31)} = 44.66, p < 0.001$) and in the interaction between variables ($F_{(2,31)} = 14.69, p < 0.001$).

In the Hipp, we observed that LNM rats increased *Crh* expression to 142% when compared to that of C group (100%; 0.87 ± 0.1 a.u.). The behavioral test induced in MS-FST rats showed an increase to 139% of C and MS values (Fig. 3D). Two-way ANOVA showed differences between groups ($F_{(2,19)} = 3.5, p < 0.05$) and in the interaction between variables ($F_{(2,19)} = 4.74, p < 0.05$). *GR* expression only increased in MS rats reaching 158% vs that of C group (100%; 4.8 ± 1.24 a.u.) and LNM rats, and was 72% higher than MS-FST (Fig. 3D). Two-way ANOVA showed differences between groups ($F_{(2,17)} = 3.72, p < 0.05$), behavioral test ($F_{(1,17)} = 11.37, p < 0.01$), and in the interaction between variables ($F_{(2,17)} = 6.3, p < 0.01$). *Crhr1* mRNA expression in MS rats decreased 43%, while the LNM group increased to 135% when compared to C group (C=100%; 0.89 ± 0.37 a.u.). There was a 35% increase in MS-FST compared to MS rats, and LNM-FST showed a 60% reduction when compared to LNM rats (Fig. 3D). Two-way ANOVA showed significant differences between groups ($F_{(2,31)} = 6.39; p < 0.01$), behavioral test ($F_{(1,31)} = 16.51; p < 0.001$), and in the interaction between variables ($F_{(2,31)} = 33.34; p < 0.001$).

3.2 Serotonergic System in the PVN, Amygdala, and Hippocampus of FST-performing Adult Male Rats Exposed to Early Postnatal Stress

The levels of 5-HT in the PVN of rats exposed to LNM were 345% higher than those of the control group (C = 100%; 4.73 ± 1.07 pg/ μ g protein) and 245% higher than the MS group (Fig. 4A). Two-way ANOVA revealed significant differences between groups ($F_{(2,24)} = 8.49; p < 0.001$) and in the interaction between variables ($F_{(2,24)} = 12.22; p < 0.001$). The ratio of 5-HIAA/5-HT decreased by 42% for MS and 28% for LNM rats, respectively, when compared to the control group (C = 100%, 8.02 ± 1.83 ratio; Fig. 4C). Similarly, FST reduced in MS-FST the 5-HIAA/5-HT ratio to 36% when compared to C group (Fig. 4C). There were no differences in 5-HIAA levels between any group (Fig. 4B). Two-way ANOVA showed differences for groups ($F_{(2,24)} = 3.96; p < 0.01$) and in the interaction between variables ($F_{(2,24)} = 5.99; p < 0.01$).

In the Amy, there were no changes in 5-HT content (Fig. 4D). In contrast, the metabolite of serotonin 5-HIAA increased to 150% in MS group when compared to C rats (100%, 6.37 ± 1.11 pg/ μ g protein). This effect was absent in animals exposed to FST, MS-FST (Fig. 4E). Two-way ANOVA revealed significant differences between groups ($F_{(2,24)} = 3.45; p < 0.05$), behavioral test ($F_{(2,24)} = 5.58; p < 0.05$), and in the interaction ($F_{(2,24)} = 8.56; p < 0.001$). The ratio of 5-HIAA/5-HT only diminished in MS-FST rats to 15% vs C animals (C=100%, 4.26 ± 1.06 ratio; Fig. 4F). Two-way ANOVA revealed significant differences between groups ($F_{(2,24)} = 5.07; p < 0.05$) and the behavioral test ($F_{(2,24)} = 5.77; p < 0.05$).

Hippocampal 5-HT levels increased to 200% in MS group vs C animals (100%; 2.49 ± 0.67 pg/ μ g protein; Fig. 4G). Also, C-FST animals increased 239% when compared to C rats (Fig. 4G). Two-way ANOVA revealed significant differences between groups ($F_{(2,23)} = 12.01; p < 0.001$), the behavioral test ($F_{(2,23)} = 5.61; p < 0.05$), and in the interaction between variables ($F_{(2,23)} = 11.02; p < 0.001$). On the other hand, there was a reduction of 5-HIAA levels in the Hipp of LNM group to 47% vs C animals (100%, 20.96 ± 2.96 pg/ μ g protein; Fig. 4H). Two-way ANOVA showed significant differences between groups ($F_{(2,24)} = 7.16; p < 0.01$). The ratio of 5-HIAA/5-HT decreased to 39% in MS and to 31% in LNM groups, respectively, in comparison to C group (C=100%, 11.19 ± 2.87 ratio; Fig. 4I). In rats subjected to the behavioral test, the ratio of C-FST and MS-FST decreased to 35% when compared to C group (Fig. 4I). Two-way ANOVA showed significant differences between groups ($F_{(2,23)} = 5.12; p < 0.01$) and in the interaction between variables ($F_{(2,23)} = 7.14; p < 0.01$).

4. Discussion

Although different early-life stress paradigms induced different HPA axis maladaptations (Fig. 5), the behavioral outcomes are similar, which show that mental illness, such as depression, may arise from different stressful events suffered during the early postnatal period.

The elevated CORT serum levels observed in dams from MS and LNM models positively correlated with their enlarged adrenal glands, supporting that they were chronically stressed during the weaning period. As a consequence, they showed alterations in their maternal care, which were different between MS and LNM dams: MS dams spent less time on the nest, but seemed to compensate it with higher offspring care, whereas LNM dams showed a more erratic behavior, since they left a greater number of their offspring outside of the caring place and groomed them more sparingly than MS dams, which had previously been described [32,33]. Increased CORT serum levels in the dams not only impact their maternal care but it can also be transferred to their pups via milk during lactation [34,35], and the chronicity of the exposure to corticosterone through milk could also influence CORT serum levels in pups after weaning. Interestingly, we observed that only MS offspring showed high CORT concentration compared to controls: a fact associated to the longer period that MS dams were stressed when compared to the LNM group. High CORT content in mother's milk may program the development of phenotypes involving impaired emotional processing, such as depression-like behavior and greater infant weight gain across time [36,37].

The hyperphagia and higher body weight of MS and LNM adults are outcomes already associated with a variety of adverse early-life conditions in humans [38] that, even if they differ in intensity, chronicity, or co-occurrence, consistently alter the development of neuroendocrine systems,

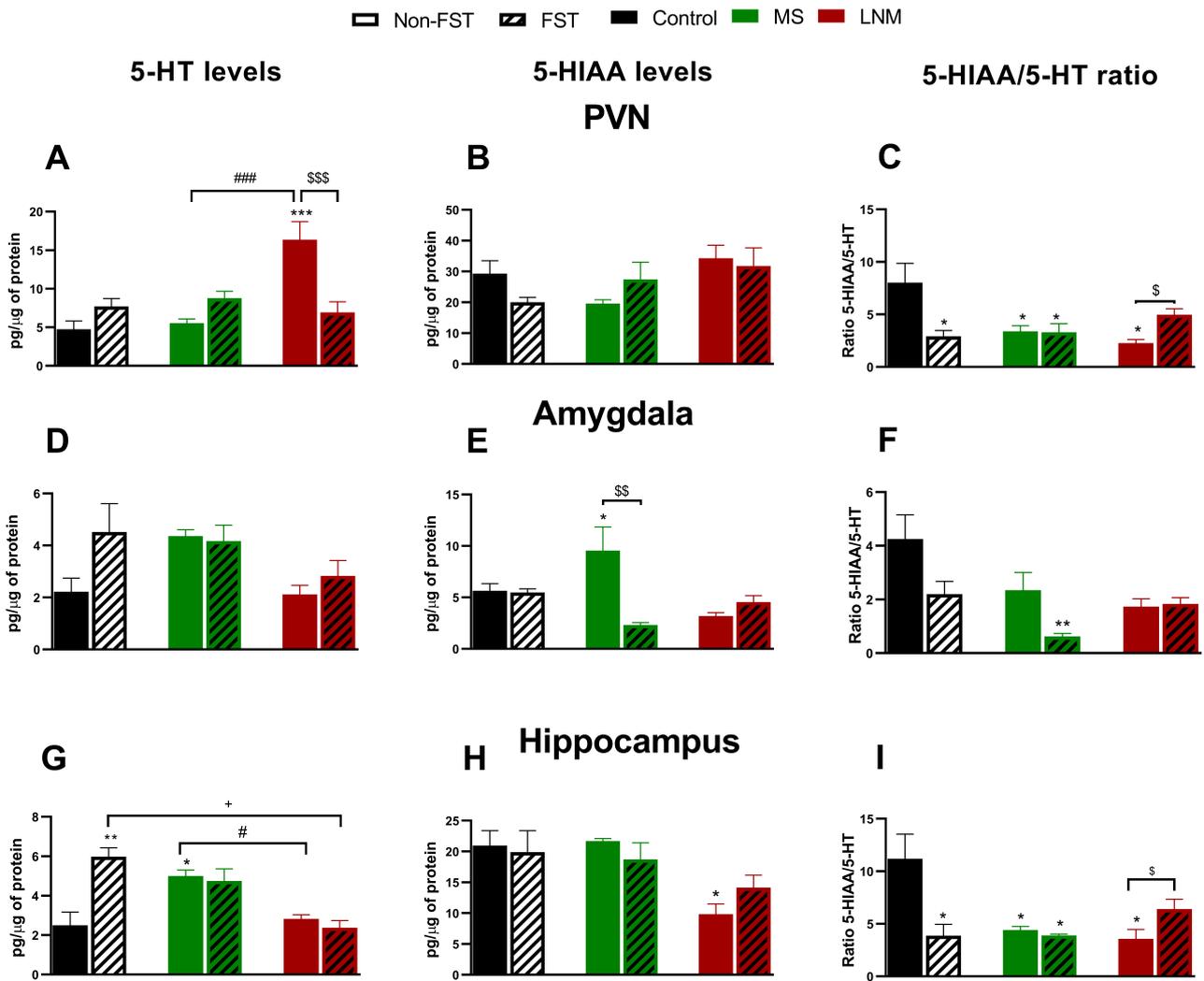


Fig. 4. Central serotonergic system in control (C), maternal separation (MS), and limited nest material (LNM) adult offspring that were or not subjected to the forced swimming test (FST). Serotonin (5-HT) and 5-hydroxyindoleindole acetic acid (5-HIAA) levels (pg/μg protein), and 5-HT/5-HIAA ratio in (A,B,C) hypothalamic paraventricular nucleus (PVN), (D,E,F) amygdala and (G,H,I) hippocampus of non-FST rats (C, MS and LNM) (open bars) and rats subjected to the forced swimming test (FST; C-FST, MS-FST, LNM-FST) (lines in bars). $n = 4$ rats/group. Data represent the mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs non-FST C group; # $p < 0.05$, ### $p < 0.001$ vs non-FST MS group; \$ $p < 0.05$, \$\$ $p < 0.01$, \$\$\$ $p < 0.001$ vs its respective FST group; + $p < 0.05$ C-FST vs LNM-FST.

and impact their lipid accumulation and appetite. Our results agree with previous studies showing that only animals with MS along with social isolation exhibit higher food intake and body weight [22]; however, regarding LNM animals we were not able to distinguish the effect of each stressor in this outcome.

Our results corroborated that the HPA axis was modified in early life-stressed adult rats, although that of MS changed more drastically than that of LNM. Both stressed groups showed increased PVN *Crh* expression, higher in the MS than in the LNM group, which had a congruent impact on CORT serum levels that only increased in MS rats. Furthermore, only adult LNM rats subjected to the FST (LNM-FST) showed GR downregulated, which is the

expected result due to the high CORT levels also observed in this group. This change contrasted with the blunted levels of CORT in MS-FST, supporting a hypersensitive or desensitized HPA axis in LNM and MS animals, respectively, most likely due to ligand-induced GR sensitivity alterations. This probably avoided a greater increase in *Crh* mRNA levels and higher HPA axis activity of LNM than in MS.

However, HPA axis was activated in both early-life stressed adults, favoring their greater appetite and food intake most likely due to increases in neuropeptide Y (NPY) and agouti related protein (AgRP) protein expression in the arcuate nucleus due to the high CORT levels [39,40]. Also, since CORT participates in lipid accumulation [41], it might

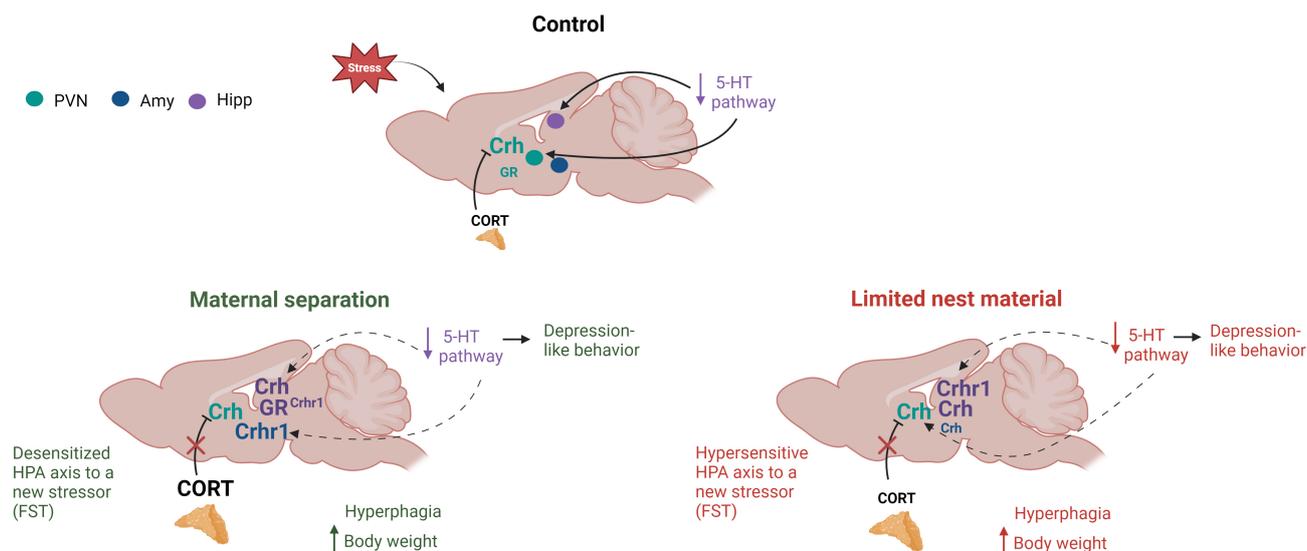


Fig. 5. Summary of the effects of two different early-life stress (ELS) paradigms on the hypothalamic-pituitary-adrenal (HPA) axis and serotonergic pathway activity. The image shows a sagittal section of a rat brain, highlighting three cerebral regions: the hypothalamic paraventricular nucleus (PVN), amygdala (Amy), and hippocampus (Hipp). When control animals were subjected to an acute stressor (forced swimming test, FST), the HPA axis was activated, resulting in higher levels of CRH and lower levels of serotonin in the PVN. Maternal separation induced a desensitization of the HPA axis to a new stressor (FST), which caused depression-like behavior, hyperphagia, and an increase in body weight associated with a low serotonergic activation. On the other hand, limited nesting material promoted a hypersensitized HPA axis to a new stressor (FST), leading to hyperphagia, increased body weight, depression-like behavior, and a decrease in the serotonergic pathway. Dashed lines depict a decrease in serotonergic pathway, and bigger letters show an enhanced synthesis or content of CRH system's components or corticosterone (CORT) serum content, while smaller letters represent a decrease in those parameters. Corticotropin releasing hormone (Crh), glucocorticoid receptor (GR), corticotropin releasing hormone receptor 1 (Crhr1), corticosterone (CORT), serotonin (5-HT).

facilitate the elevated body weight of adults of the two post-natal stress models. The association between an impaired development of neuroendocrine axis and obesity has also been observed in humans [38,42–47], and explains the 17% prevalence of obesity among adults experiencing early-life stress. Our results are helpful to understand that a specific outcome, such as obesity and hyperphagia, may result from various postnatal adversities that differ in type or intensity of the inflicted stress during early periods of life.

Besides hyperphagia, MS and LNM adults also displayed a high-immobility behavior when performing the FST, which represents an impaired coping strategy to stressful situations. This high immobility was observed even when their dams' care, HPA axis activity, and stress duration was different between the two models, which agrees with the 50% incidence of depression among humans that experienced a variety of ELS [38]. In contrast to hyperphagia and higher body weight that are induced by the combination of ELS and isolation, high immobility behavior in FST may be observed in animals subjected to MS or to isolation; interestingly, the combined stressors do not induce an additive immobility score [22,48]. In this study, we are observing that the behavioral outcome in the FST resulted from both stressors, but we were not able to discriminate their individual effect in the experimental groups.

In MS animals, the observed depression-like behavior was associated to their unresponsive HPA axis, since neither their *Crh* expression nor CORT serum levels further arose in rats performing the FST. This result supported the inability of these animals to face a new energy-demanding challenge, due to impaired CORT-induced glucose availability. In contrast, LNM rats did respond to the new stressor (FST) with increased CORT serum levels even when their *Crh* mRNA levels remained elevated as compared to animals not subjected to the test. This result suggested a greater sensitivity of GR to CORT-induced regulation than in MS rats, and supports the assumption that ELS induces epigenetic alterations in adults' *GR* gene that could alter the receptor responsiveness and HPA regulation [49].

Another explanation for the differential responsiveness of the HPA axis between MS and LNM adults to FST might be that only the amygdalar CRH pathway changed in the LNM group. LNM rats showed low *Crh* expression in that brain area, which could maintain CORT serum levels under basal values until adulthood, thus avoiding an exacerbated activity of the HPA axis during their development and allowing it to respond to the FST. This could be due to the amygdalar efferent CRH connections to the PVN that stimulate the HPA axis, given that the overexpression of CRH in the central nucleus of Amy is associated with im-

paired CORT-induced negative feedback of the HPA axis, thus inducing its hyperactivity, as well as different behavioral alterations (Gillespie *et al.*, 2009) [50]. Since specific changes in maternal care affect the interactions between Amy and the HPA axis, those displayed by LNM dams were likely milder than those of MS. Moreover, even when LNM pups presented depression-like behavior and hyperphagia, they showed an advantage when compared to MS, which is the normal responsiveness of their HPA axis to other stressful stimuli. Also, LNM rats subjected to FST increased CORT levels that might reach the Amy and activate GR evidenced by its low expression, thus being able to downregulate the CRH pathway and its impact on HPA axis. Amygdalar *Crhr1* expression also changed differentially between groups, increasing in MS and decreasing in LNM adult rats, which seemed to correlate with their HPA axis desensitization or hypersensitivity, respectively. *Crhr1* upregulation in MS rats could result as a compensating change to the blocked negative feedback of the axis that maintains elevated CRH and CORT levels, which may impact the amygdalar CRH stress-responsive system. This interpretation is plausible mainly because the LNM group displayed a hypersensitive HPA axis and presented an inverse change in *Crhr1* in Amy.

Crhr1 mRNA downregulation is also observed in other ELS-exposed adults, i.e. among restrain-stressed rats during early postnatal days [15,51,52], but not among those stressed during adulthood [51]. This supports that the time-limited CORT increase in LNM offspring, as well as the intensity and type of stress, might induce changes in *Crhr1* expression, which may be representative of the brain-adaptive alterations to different types of ELS and might be associated to behavioral changes in adulthood.

In contrast to Amy, the hippocampal CRHergic system seemed deregulated only in the MS group, since its *Crh* expression increased when subjected to FST, avoiding a further activation of their HPA axis to the new stress. This is supported by the resulting decreased CORT serum levels after hippocampus electrical stimulation, as well as its increase by hippocampotomy or hippocampal lesions [53–57]. Present data align with previous studies showing that MS animals are unresponsive to acute stress such as the FST [58]. An elevated expression of *GR* was also specific for the MS group, which might favor their higher body weight and depression-like behavior, since other chronic stressful paradigms also developed similar symptoms in ELS adults [59]. Upregulated *GR* expression is interpreted as resistance mainly induced by the high CORT levels these animals had since early-life periods [59]. Furthermore, the sudden decrease in *GR* expression among MS-FST rats supported a deregulation of this hippocampal pathway by CORT that might upregulate *Crh* mRNA levels, and a specific MS-induced change in Hipp that favored the lack of response of their HPA axis to the FST, as it is thoroughly described [52,56]. The repercussion of this hippocampal

deregulated system only in MS animals resulted evident on the HPA axis by its unchanged hypothalamic PVN *GR* expression after exposure to FST, supporting MS rats' inability to respond to the new challenge, reinforcing the hippocampal regulation of HPA axis sensitivity to corticosteroid feedback [56].

Hippocampal *Crhr1* expression also resulted differentially altered between groups, decreasing in MS and increasing in LNM in basal conditions. This change in the MS group, associated with helplessness behavior of stressed rats [60] and a low 5-HT system functioning, is supported by the reversion induced by a fluoxetine treatment of the low *Crhr1* expression and with reduced depression-like behavior and hyperphagia [61]. The decreased hippocampal 5-HT system activity in MS animals was inferred by their higher intracellular content (accumulation), along with a low metabolism of the neurotransmitter (decreased 5-HIAA/5-HT ratio) observed in the non-FST group, which were similar to the changes observed by C-FST. In contrast, the LNM group showed higher *Crhr1* expression already associated to cognitive alterations [62] that are avoided with CRHR1 antagonists injections during ELS [63].

By analyzing the brain 5-HT system activity, we found that the sole FST performance was enough to induce 5-HT accumulation in Hipp and decrease serotonin pathway activity in C-FST, which resulted from 5-HT release induced by the FST in midbrain-projecting sites, such as Amy, Hipp, and PVN [64]. This could activate pre-synaptic 5-HT_{1A} receptors in those regions favoring 5-HT accumulation. Hippocampus of the MS group also had this accumulation in rats either performing or not the FST. Decelerating 5-HT pathway in LNM rats' Hipp was not as clear as in MS animals, but their low metabolite concentration and 5-HIAA/5-HT ratio in limbic regions supported a similar change. Specific 5-HT system alterations were found in PVN and limbic regions; but, in general, pointed out to a decreased functioning, which agreed with previous studies and supported its association with hyperphagia and depression development [65]. We did not directly prove the association between low activity of the 5-HT pathway and alterations in CRH system in the PVN and limbic regions. However, substantial evidence for this fact has been previously described [64].

5. Conclusion

We concluded that blood corticosterone elevation of stressed dams from the two paradigms differentially disturbed the HPA axis of their offspring, these results being more intense in MS than in LNM groups and seemed to be also associated to the duration of the increase. As a consequence, the HPA axis of MS adults resulted desensitized, whereas that of LNM was hypersensitized to the negative feedback effects of corticosterone. Specific differential changes in the CRH system in amygdala and hippocampus between MS and LNM adult animals might explain the variations in responsiveness of their HPA axis.

A slow activity of serotonergic neurons in the amygdala and hippocampus might contribute to the similar behavioral maladaptive outcomes that both groups of early-life stress showed in adulthood: depression-like behavior and overweight-induced hyperphagia. Moreover, early-life stressed adults' exposure to the FST was needed for unraveling the greater vulnerability of MS animals than that of LNM when facing a new acute stressful stimulus, as the HPA axis of MS was unable to increase corticosterone levels and, in turn, generate available energy for animals to escape or to face the threat.

Availability of Data and Materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Author Contributions

VAA and PdG design the research study. VAA performed the research. VAA, CGL, PSC, EEC analyzed the data. PdG interpretation of data. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

All procedures were conducted with the approval of the local Ethics Committee for Animal Experimentation of the Instituto Nacional de Psiquiatría Ramón de la Fuente Muñiz (INPRFM) (CEI/C/024/2014) following the guidelines outlined by the Mexican Official Standard NOM-0620ZOO-1999.

Acknowledgment

We thank Orlando Jaimes for his technical assistance with HPLC determinations.

Funding

This research received no external funding.

Conflict of Interest

The authors declare no conflict of interest.

References

[1] Carr CP, Martins CMS, Stingel AM, Lemgruber VB, Juruena MF. The role of early life stress in adult psychiatric disorders: a systematic review according to childhood trauma subtypes. *The Journal of Nervous and Mental Disease*. 2013; 201: 1007–1020.

[2] Ventriglio A, Gentile A, Baldessarini RJ, Bellomo A. Early-life stress and psychiatric disorders: epidemiology, neurobiology and innovative pharmacological targets. *Current Pharmaceutical Design*. 2015; 21: 1379–1387.

[3] Gill H, El-Halabi S, Majeed A, Gill B, Lui LMW, Mansur RB, *et al*. The Association Between Adverse Childhood Experiences and Inflammation in Patients with Major Depressive Disorder:

A Systematic Review. *Journal of Affective Disorders*. 2020; 272: 1–7.

[4] Gardner R, Feely A, Layte R, Williams J, McGavock J. Adverse childhood experiences are associated with an increased risk of obesity in early adolescence: a population-based prospective cohort study. *Pediatric Research*. 2019; 86: 522–528.

[5] Antoniou G, Lambourg E, Steele JD, Colvin LA. The effect of adverse childhood experiences on chronic pain and major depression in adulthood: a systematic review and meta-analysis. *British Journal of Anaesthesia*. 2023; 130: 729–746.

[6] Turecki G, Ota VK, Belangero SI, Jackowski A, Kaufman J. Early life adversity, genomic plasticity, and psychopathology. *The Lancet. Psychiatry*. 2014; 1: 461–466.

[7] Dube SR, Felitti VJ, Dong M, Giles WH, Anda RF. The impact of adverse childhood experiences on health problems: evidence from four birth cohorts dating back to 1900. *Preventive Medicine*. 2003; 37: 268–277.

[8] van Bodegom M, Homberg JR, Henckens MJAG. Modulation of the Hypothalamic-Pituitary-Adrenal Axis by Early Life Stress Exposure. *Frontiers in Cellular Neuroscience*. 2017; 11: 87.

[9] Mathur A, Graham-Engeland JE, Slavish DC, Smyth JM, Lipton RB, Katz MJ, *et al*. Recalled early life adversity and pain: the role of mood, sleep, optimism, and control. *Journal of Behavioral Medicine*. 2018; 41: 504–515.

[10] Stratakis CA, Chrousos GP. Neuroendocrinology and pathophysiology of the stress system. *Annals of the New York Academy of Sciences*. 1995; 771: 1–18.

[11] Lee JH, Kim HJ, Kim JG, Ryu V, Kim BT, Kang DW, *et al*. Depressive behaviors and decreased expression of serotonin reuptake transporter in rats that experienced neonatal maternal separation. *Neuroscience Research*. 2007; 58: 32–39.

[12] Malave L, van Dijk MT, Anacker C. Early life adversity shapes neural circuit function during sensitive postnatal developmental periods. *Translational Psychiatry*. 2022; 12: 306.

[13] Elovainio M, Pulkki-Råback L, Jokela M, Kivimäki M, Hintsanen M, Hintsanen T, *et al*. Socioeconomic status and the development of depressive symptoms from childhood to adulthood: a longitudinal analysis across 27 years of follow-up in the Young Finns study. *Social Science & Medicine* (1982). 2012; 74: 923–929.

[14] Ehlert U. Enduring psychobiological effects of childhood adversity. *Psychoneuroendocrinology*. 2013; 38: 1850–1857.

[15] Alcántara-Alonso V, Amaya MI, Matamoros-Trejo G, de Gortari P. Altered functionality of the corticotrophin-releasing hormone receptor-2 in the hypothalamic paraventricular nucleus of hyperphagic maternally separated rats. *Neuropeptides*. 2017; 63: 75–82.

[16] Commons KG, Cholanians AB, Babb JA, Ehlinger DG. The Rodent Forced Swim Test Measures Stress-Coping Strategy, Not Depression-like Behavior. *ACS Chemical Neuroscience*. 2017; 8: 955–960.

[17] Molendijk ML, de Kloet ER. Coping with the forced swim stressor: Current state-of-the-art. *Behavioural Brain Research*. 2019; 364: 1–10.

[18] Vega-Rivera NM, Fernández-Guasti A, Ramírez-Rodríguez G, Estrada-Camarena E. Acute stress further decreases the effect of ovariectomy on immobility behavior and hippocampal cell survival in rats. *Psychoneuroendocrinology*. 2013; 38: 1407–1417.

[19] Plotsky PM, Thiruvikraman KV, Nemeroff CB, Caldji C, Sharma S, Meaney MJ. Long-term consequences of neonatal rearing on central corticotropin-releasing factor systems in adult male rat offspring. *Neuropsychopharmacology*. 2005; 30: 2192–2204.

[20] Ivy AS, Brunson KL, Sandman C, Baram TZ. Dysfunctional nurturing behavior in rat dams with limited access to nesting material: a clinically relevant model for early-life stress. *Neuroscience*. 2008; 154: 1132–1142.

- [21] Gilles EE, Schultz L, Baram TZ. Abnormal corticosterone regulation in an immature rat model of continuous chronic stress. *Pediatric Neurology*. 1996; 15: 114–119.
- [22] Ryu V, Yoo SB, Kang DW, Lee JH, Jahng JW. Post-weaning isolation promotes food intake and body weight gain in rats that experienced neonatal maternal separation. *Brain Research*. 2009; 1295: 127–134.
- [23] Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates*. Fifth (edn.). Academic Press Elsevier Inc.: San Diego. 2005.
- [24] Myers MM, Brunelli SA, Shair HN, Squire JM, Hofer MA. Relationships between maternal behavior of SHR and WKY dams and adult blood pressures of cross-fostered F1 pups. *Developmental Psychobiology*. 1989; 22: 55–67.
- [25] Martínez-Mota L, Ulloa RE, Herrera-Pérez J, Chavira R, Fernández-Guasti A. Sex and age differences in the impact of the forced swimming test on the levels of steroid hormones. *Physiology & Behavior*. 2011; 104: 900–905.
- [26] Porsolt RD, Le Pichon M, Jalfre M. Depression: a new animal model sensitive to antidepressant treatments. *Nature*. 1977; 266: 730–732.
- [27] Detke MJ, Rickels M, Lucki I. Active behaviors in the rat forced swimming test differentially produced by serotonergic and noradrenergic antidepressants. *Psychopharmacology*. 1995; 121: 66–72.
- [28] Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Analytical Biochemistry*. 1987; 162: 156–159.
- [29] Jaimes-Hoy L, Joseph-Bravo P, de Gortari P. Differential response of TRHergic neurons of the hypothalamic paraventricular nucleus (PVN) in female animals submitted to food-restriction or dehydration-induced anorexia and cold exposure. *Hormones and Behavior*. 2008; 53: 366–377.
- [30] Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *The Journal of Biological Chemistry*. 1951; 193: 265–275.
- [31] Patel BA, Arundell M, Parker KH, Yeoman MS, O'Hare D. Simple and rapid determination of serotonin and catecholamines in biological tissue using high-performance liquid chromatography with electrochemical detection. *Journal of Chromatography. B, Analytical Technologies in the Biomedical and Life Sciences*. 2005; 818: 269–276.
- [32] Rincón-Cortés M, Grace AA. Postpartum scarcity-adversity disrupts maternal behavior and induces a hypodopaminergic state in the rat dam and adult female offspring. *Neuropsychopharmacology*. 2022; 47: 488–496.
- [33] Walker CD, Bath KG, Joels M, Korosi A, Larauche M, Lucassen PJ, *et al.* Chronic early life stress induced by limited bedding and nesting (LBN) material in rodents: critical considerations of methodology, outcomes and translational potential. *Stress (Amsterdam, Netherlands)*. 2017; 20: 421–448.
- [34] Angelucci L, Patacchioli FR, Chierichetti C, Laureti S. Perinatal mother-offspring pituitary-adrenal interrelationship in rats: corticosterone in milk may affect adult life. *Endocrinologia Experimentalis*. 1983; 17: 191–205.
- [35] Brummelte S, Schmidt KL, Taves MD, Soma KK, Galea LAM. Elevated corticosterone levels in stomach milk, serum, and brain of male and female offspring after maternal corticosterone treatment in the rat. *Developmental Neurobiology*. 2010; 70: 714–725.
- [36] Hinde K, Skibieli AL, Foster AB, Del Rosso L, Mendoza SP, Capitano JP. Cortisol in mother's milk across lactation reflects maternal life history and predicts infant temperament. *Behavioral Ecology*. 2015; 26: 269–281.
- [37] Martins IP, Vargas R, Saavedra LPJ, Rickli S, Matusso CCI, Pavanello A, *et al.* Protein-caloric restriction induced HPA axis activation and altered the milk composition imprint metabolism of weaned rat offspring. *Nutrition*. 2023; 108: 111945.
- [38] Schroeder K, Schuler BR, Kobulsky JM, Sarwer DB. The association between adverse childhood experiences and childhood obesity: A systematic review. *Obesity Reviews: an Official Journal of the International Association for the Study of Obesity*. 2021; 22: e13204.
- [39] Chagra SL, Zavala JK, Hall MV, Gosselink KL. Acute and repeated restraint differentially activate orexigenic pathways in the rat hypothalamus. *Regulatory Peptides*. 2011; 167: 70–78.
- [40] Hagimoto S, Arima H, Adachi K, Ito Y, Suga H, Sugimura Y, *et al.* Expression of neuropeptide Y and agouti-related protein mRNA stimulated by glucocorticoids is attenuated via NF- κ B p65 under ER stress in mouse hypothalamic cultures. *Neuroscience Letters*. 2013; 553: 165–169.
- [41] García-Eguren G, Sala-Vila A, Giró O, Vega-Beyhart A, Hanzu FA. Long-term hypercortisolism induces lipogenesis promoting palmitic acid accumulation and inflammation in visceral adipose tissue compared with HFD-induced obesity. *American Journal of Physiology. Endocrinology and Metabolism*. 2020; 318: E995–E1003.
- [42] Noll JG, Zeller MH, Trickett PK, Putnam FW. Obesity risk for female victims of childhood sexual abuse: a prospective study. *Pediatrics*. 2007; 120: e61–7.
- [43] Mason SM, Bryn Austin S, Bakalar JL, Boynton-Jarrett R, Field AE, Gooding HC, *et al.* Child Maltreatment's Heavy Toll: The Need for Trauma-Informed Obesity Prevention. *American Journal of Preventive Medicine*. 2016; 50: 646–649.
- [44] Zeller MH, Noll JG, Sarwer DB, Reiter-Purtill J, Rofey DL, Baughcum AE, *et al.* Child Maltreatment and the Adolescent Patient With Severe Obesity: Implications for Clinical Care. *Journal of Pediatric Psychology*. 2015; 40: 640–648.
- [45] Hemmingsson E, Johansson K, Reynisdottir S. Effects of childhood abuse on adult obesity: a systematic review and meta-analysis. *Obesity Reviews: an Official Journal of the International Association for the Study of Obesity*. 2014; 15: 882–893.
- [46] Ahn S, Zhang H, Berlin KS, Levy M, Kabra R. Adverse Childhood Experiences and Childhood Obesity: A Path Analysis Approach. *Children's Health Care*. 2019; 49: 247–266.
- [47] Burke NJ, Hellman JL, Scott BG, Weems CF, Carrion VG. The impact of adverse childhood experiences on an urban pediatric population. *Child Abuse & Neglect*. 2011; 35: 408–413.
- [48] Vargas J, Junco M, Gomez C, Lajud N. Early Life Stress Increases Metabolic Risk, HPA Axis Reactivity, and Depressive-Like Behavior When Combined with Postweaning Social Isolation in Rats. *PloS One*. 2016; 11: e0162665.
- [49] McGowan PO, Sasaki A, D'Alessio AC, Dymov S, Labonté B, Szyf M, *et al.* Epigenetic regulation of the glucocorticoid receptor in human brain associates with childhood abuse. *Nature Neuroscience*. 2009; 12: 342–348.
- [50] Gillespie CF, Phifer J, Bradley B, Ressler KJ. Risk and resilience: genetic and environmental influences on development of the stress response. *Depress Anxiety*. 2009; 26: 984–992.
- [51] Vazquez DM, Bailey C, Dent GW, Okimoto DK, Steffek A, López JF, *et al.* Brain corticotropin-releasing hormone (CRH) circuits in the developing rat: effect of maternal deprivation. *Brain Research*. 2006; 1121: 83–94.
- [52] Alcántara-Alonso V, Panetta P, de Gortari P, Grammatopoulos DK. Corticotropin-Releasing Hormone As the Homeostatic Rheostat of Feto-Maternal Symbiosis and Developmental Programming *In Utero* and Neonatal Life. *Frontiers in Endocrinology*. 2017; 8: 161.
- [53] Momose KJ, Kjellberg RN, Kliman B. High incidence of cortical atrophy of the cerebral and cerebellar hemispheres in Cushing's disease. *Radiology*. 1971; 99: 341–348.
- [54] Rabins PV, Pearson GD, Aylward E, Kumar AJ, Dowell K. Cortical magnetic resonance imaging changes in elderly inpa-

- tients with major depression. *The American Journal of Psychiatry*. 1991; 148: 617–620.
- [55] Rothschild AJ, Benes F, Hebben N, Woods B, Luciana M, Bakanas E, *et al*. Relationships between brain CT scan findings and cortisol in psychotic and nonpsychotic depressed patients. *Biological Psychiatry*. 1989; 26: 565–575.
- [56] Jacobson L, Sapolsky R. The role of the hippocampus in feedback regulation of the hypothalamic-pituitary-adrenocortical axis. *Endocrine Reviews*. 1991; 12: 118–134.
- [57] McEwen BS, Sapolsky RM. Stress and cognitive function. *Current Opinion in Neurobiology*. 1995; 5: 205–216.
- [58] Alonso SJ, Arevalo R, Afonso D, Rodríguez M. Effects of maternal stress during pregnancy on forced swimming test behavior of the offspring. *Physiology & Behavior*. 1991; 50: 511–517.
- [59] Aleksic M, Brkic Z, Petrovic Z, Francija E, Lukic I, Adzic M. Sex-specific contribution of glucocorticoid receptor alpha isoforms to anxiety and depressive-like behavior in mice. *Journal of Neuroscience Research*. 2022; 100: 1239–1253.
- [60] Bravo JA, Dinan TG, Cryan JF. Alterations in the central CRF system of two different rat models of comorbid depression and functional gastrointestinal disorders. *The International Journal of Neuropsychopharmacology*. 2011; 14: 666–683.
- [61] Fernández Macedo GV, Cladouchos ML, Sifonios L, Cassanelli PM, Wikinski S. Effects of fluoxetine on CRF and CRF1 expression in rats exposed to the learned helplessness paradigm. *Psychopharmacology*. 2013; 225: 647–659.
- [62] Ivy AS, Rex CS, Chen Y, Dubé C, Maras PM, Grigoriadis DE, *et al*. Hippocampal dysfunction and cognitive impairments provoked by chronic early-life stress involve excessive activation of CRH receptors. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*. 2010; 30: 13005–13015.
- [63] Brunson KL, Kramár E, Lin B, Chen Y, Colgin LL, Yanagihara TK, *et al*. Mechanisms of late-onset cognitive decline after early-life stress. *The Journal of Neuroscience*. 2005; 25: 9328–9338.
- [64] Valentino RJ, Lucki I, Van Bockstaele E. Corticotropin-releasing factor in the dorsal raphe nucleus: Linking stress coping and addiction. *Brain Research*. 2010; 1314: 29–37.
- [65] Zangen A, Overstreet DH, Yadid G. High serotonin and 5-hydroxyindoleacetic acid levels in limbic brain regions in a rat model of depression: normalization by chronic antidepressant treatment. *Journal of Neurochemistry*. 1997; 69: 2477–2483.