

Systematic Review

Neurofilament Light Chain as a Potential Biomarker in Plasma for Alzheimer's Disease and Mild Cognitive Impairment: A Systematic Review and a Meta-Analysis

Zhenyi Fan¹, Xiaoxia Liu^{1,†}, Jie Liu^{2,†}, Caijing Chen², Min Zhou^{1,*}

¹Department of Neurology, Ningbo NO.2 Hospital, 315000 Ningbo, Zhejiang, China

²Department of Geriatrics, Ningbo NO.2 Hospital, 315000 Ningbo, Zhejiang, China

*Correspondence: zmn1989@163.com (Min Zhou)

†These authors contributed equally.

Academic Editor: Gernot Riedel

Submitted: 8 December 2022 Revised: 1 January 2023 Accepted: 11 January 2023 Published: 30 June 2023

Abstract

Background: Plasma neurofilament light (NfL) is an intermediate filamentous protein involved in stabilizing axonal structure and promoting axon growth. Recent clinical studies have reported increased NfL levels in the plasma of Alzheimer's disease (AD) patients and patients with mild cognitive impairment (MCI). This study used meta-analysis to evaluate the potential of plasma NfL as a biomarker for patients with AD and MCI. **Methods:** PubMed, Embase, and Web of Science databases were systematically searched for studies of plasma NfL levels in AD and MCI, and a meta-analysis was employed to identify whether it was suited as a reliable biomarker and discrimination of healthy controls. **Results:** A total of 24 published articles that included 2397 AD and 3242 MCI patients were analysed. The level of plasma NfL was significantly increased in patients with AD and MCI when compared with healthy control subjects (standard mean difference [SMD]: 14.33 [12.42–16.24], $z = 14.71$, $p < 0.00001$; SMD: 4.95 [3.82–6.80], $z = 8.59$, $p < 0.00001$) and higher in AD patients than MCI patients (SMD: 9.32 [8.07–10.57], $z = 14.62$, $p < 0.00001$). Meta-regression analysis showed a negative relationship between Mini-Mental State Examination (MMSE) scores and plasma NfL levels in MCI patients (slope = -0.399 [95% confidence interval (CI): -0.518 to -0.281], $p < 0.05$). **Conclusions:** The meta-analysis suggested that NfL levels increased in the plasma of patients with AD and MCI and were associated with cognitive decline. Results provide the clinical evidence to support plasma NfL as a cognitive biomarker for AD and MCI.

Keywords: Alzheimer's disease; mild cognitive impairment; neurofilament light chain; meta-analysis

1. Introduction

Alzheimer's disease (AD), a severe neurodegenerative disease of the central nervous system that usually occurs in senile and pre-senile patients, is characterized by progressive loss of thinking, memory, language, and impairment of cognitive ability. It is the most common form of dementia in old age, with about 60% to 80% of the dementia diagnosed in people over 65 years old being attributed to it [1]. New cases of AD are projected to increase to more than 1 million by 2050, which will place an enormous financial burden on families and society [2]. Mild cognitive impairment (MCI) is an intermediate stage in which a person has problems with memory, language, or other cognitive functions that can be detected by others or by testing but is not serious enough to affect the activities of daily living [3]. If it is not prevented from progressing in a timely manner, such subjects are at a significantly increased risk of evolving toward AD at the rate of 15–25% over two years [4,5]. However, there are neither disease-modifying therapies nor successful late-stage clinical trials currently available [6]. It is believed that continuous pathophysiological changes begin many years prior to clinical symptom onset, thus further investigations is required with the aim of discovering a

practical biomarker for the early diagnosis and detection of AD.

Neurofilaments (Nfs) are intermediate filamentous proteins, expressed in neurons and particularly abundant in axons, responsible for the structural stability of axon morphology, forming the neuronal cytoskeleton, and maintaining cytoarchitecture and transport functions [7]. Nfs are divided by molecular weight into three subunits, the neurofilament heavy, medium, and light chains [8]. Neurofilament light chain (NfL) has the lowest molecular weight, and is thought to be the leading Nfs for stabilization of axonal structure and the promotion of axon growth [9,10]. There are many studies demonstrating that NfL levels are higher in AD patients than controls, associated with poorer cognitive performance, and short survival time in demented patients [11]. NfL was used as a specifically diagnostic biomarker, meanwhile, it is potentially a valuable tool for the detection of the initial pathological changes associated with AD, even at the MCI stage, while in the differential diagnosis, monitoring, and prognosis of AD. Recently, there has been great interest in the utility of NfL in plasma as a biomarker for AD. It avoids the invasiveness of cerebrospinal fluid (CSF) sampling, which is restricted in its clinical application [12].



This meta-analysis aimed to investigate whether NfL in peripheral blood is accompanied by improved levels of AD or MCI and to evaluate NfL as a cognitive biomarker for the diagnosis of AD and MCI. The findings reported here may be used for risk assessment and assessment of disease progression, even for clinically aided diagnosis of MCI or AD. Plasma NfL levels were measured in three groups of patients, AD, MCI, and healthy subjects, obtained from cross-sectional and longitudinal studies.

2. Materials and Methods

2.1 Search Strategy

The meta-analysis was conducted according to the guidelines of the Preferred Reporting Items for Systematic Review and Meta-analysis (PRISMA) [13]. PRISMA checklist is shown in **Supplementary Material**. Two independent investigators (MZ and XL) performed a systematic literature research in English through June 2022 in the following three electronic databases: PubMed, Embase, and Web of Science. The search strategy was by Mesh phrases and keywords included (neurofilament light chain OR NfL) AND (plasma) AND (Alzheimer's disease OR Mild cognitive impairment). All articles were imported into the management software. Two investigators independently screened the titles, abstracts, and full texts of the most eligible publications. Any conflicts were resolved through discussion or, if necessary, adjudicated by a third investigator (ZF).

2.2 Inclusion and Exclusion Criteria

All of the included publications compared peripheral NfL data in AD or MCI patients with controls. The qualifying studies satisfied the following criteria, including: (a) Study designs must be either cross-sectional or longitudinal; (b) Studies must include AD and/or MCI patients and healthy controls; (c) AD and MCI patients must fulfil the diagnosis criteria of National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association (NINCDS-ADRDA) Criteria [14], or National Institute on Aging and Alzheimer's Association (NIA-AA) Criteria [15] or Petersen (Mayo Clinic) diagnostic criteria [16]; (d) The NfL concentration of plasma must be one of the main interests in patients with AD or MCI and in the healthy control group; (e) The article must report NfL levels in plasma measured with an in-house assay on the single molecule array platform (Simoa); (f) The study should be available from the original paper rather than case-reports, protocols, conference abstracts, reviews, or meta-analyses. Studies were excluded according to the following criteria: (a) Non-human studies; (b) The number of subjects was less than 20; (c) Studies without healthy controls or without disease controls; (d) Articles that were unrelated to NfL, AD, or MCI; (e) Studies with missing data such as details about demographic information and the method employed to measure NfL.

2.3 Data Extraction

One investigator independently extracted the following data from every study for the purposes of this meta-analysis, and another investigator independently checked them to ensure accuracy. The relevant data extracted included the name of the first author, year of publication, study design, number of the AD or MCI patients and healthy control groups, the average age, percentage of females, diagnosis criteria of AD and MCI, the plasma concentration of NfL, the measurement methods of NfL, and a Mini-Mental State Examination (MMSE). This information was entered into a standardized Excel spreadsheet with any disagreement resolved by discussion and agreement.

2.4 Quality Assessment

The Newcastle-Ottawa scale (NOS) was employed to assess the quality of the available studies [17]. The assessment process was individually performed by two investigators, and any discrepancy was solved by discussion or by a third investigator. NOS scores range from zero to nine, with a higher score indicating better quality. A study was given a maximum score of one star for each numbered item within three domains: The Selection (0–4 scores), Comparability (0–2 scores), and Exposure categories (0–3 scores). A score greater than five was considered to imply that the study was of high quality. The more stars allocated, the better the quality.

2.5 Statistical Analysis

Review Manager (version 5.3, Cochrane Collaboration, Oxford, United Kingdom) and Stata statistical software (version 14.0, Stata Corporation, College Station, TX, USA) were used to pool all statistical analyses. The statistical analysis was performed using a standardized mean difference methodology. When only median and interquartile ranges were available from the included articles, means and standard deviations were estimated following Wan *et al.* [18] and Luo *et al.* [19]. The standardized mean difference (SMD) and corresponding 95% confidence interval (CI) were calculated, and forest plots were generated to compare the mean plasma NfL levels between AD or MCI patients with healthy controls (HC). Either a random-effects or a fixed-effects model was based on the heterogeneity of the articles for each comparison. In the following statistical analysis, an overall meta-analysis was performed for AD vs. HC, MCI vs. HC, and AD vs. MCI. Heterogeneity between the studies was assessed by the I^2 test. When $I^2 > 50\%$, indicating heterogeneity was significant, a random-effects model was employed for calculations; otherwise, the fixed-effects model was applied to the data. Secondly, one article was removed from each group for a sensitivity analysis to evaluate the influence of an individual study on the stability of the obtained estimate. Thirdly, meta-regression analysis and subgroup analyses were conducted to test whether there was a significant difference in sample size, age, sex

ratio, and MMSE scores. A RevMan funnel plot was employed to identify any potential publication bias for each meta-analysis. All tests were two-sided, and except where noted, all statistical significances were set at $p < 0.05$.

3. Results

3.1 Study Characteristics

According to the search strategy, a total of 1316 articles were obtained from the three databases. Among them, 520 duplicate articles were removed. By screening the titles and abstract reviews, 680 articles were excluded for the following reasons: 40 were irrelevant topics, 119 were reviews, 43 used non-human subjects, 123 were conferences, editorials or meta-analysis, 262 were non-primary dementia studies, and 93 were not related to either plasma NfL or analytical methods for NfL. 116 potentially relevant publications were subjected to full-text reviews. Ninety-two publications were excluded for the following reasons: 43 were not controlled studies, 30 had incomplete or unavailable data, 9 lacked necessary plasma NfL data, 8 were non-dementia studies, and 2 had a total sample size less than 20. The Flow Diagram showing the detailed process of selection is given in Fig. 1. Table 1 (Ref. [20–43]) presents a summary of the main characteristics of the included studies.

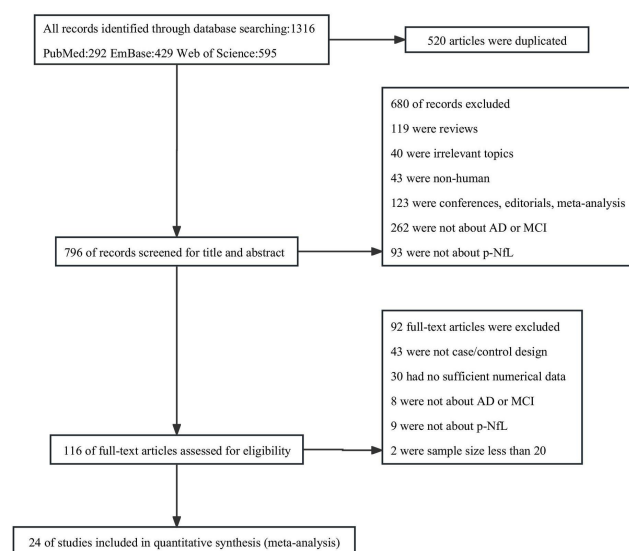


Fig. 1. Flow diagram of studies selected for meta-analysis.

3.2 Quality Assessment

The NOS assessment tool was used to independently evaluate the quality of the articles by two authors (MZ and XL), as shown in Table 2 (Ref. [20–43]). A study was awarded a maximum score of one star for each numbered item within the Selection (0–4 points) and Exposure (0–3 points) categories. A maximum score of up to two stars was given to a study for its Comparability (0–2 points). Gener-

ally, the quality of the studies was moderate and high, with the more stars allocated to a study, the better quality the methodology employed. All the publications in this review scored greater than or equal to six stars, indicating good quality.

3.3 Statistical Results

3.3.1 Association between Plasma Neurofilament Light Chain Levels in AD and HC

Plasma neurofilament light chain levels in AD patients were compared with those in HC subjects [20–22,24,25,27–31,33,34,36–40,42] with extracted data from 19 studies comprising a sample of 2397 AD subjects and 3219 HC subjects. The heterogeneity of these studies was high ($\text{Tau}^2 = 11.21$; $\chi^2 = 58.66$, $\text{df} = 18$, $p < 0.00001$; $I^2 = 69\%$), therefore a random effect model was performed, and results showed that AD subjects had significantly higher levels of plasma NfL when compared with HC subjects (SMD: 14.33 [12.42–16.24], $z = 14.71$, $p < 0.00001$, Fig. 2) and sensitivity analysis indicated that none of the studies changed the nature of the effect value.

3.3.2 Association between Plasma Neurofilament Light Chain Levels in MCI and HC

The NfL levels between MCI subjects and healthy controls were then compared; the total number of MCI and HC subjects were 3242 and 3801, respectively, from 19 studies [20,23,24,26–29,31–40,42,43]. Results showed a trend of high plasma NfL levels in MCI patients when compared with healthy control subjects (SMD: 4.95 [3.82–6.08], $z = 8.59$, $p < 0.00001$, Fig. 3). There was significant heterogeneity among the studies ($\text{Tau}^2 = 3.79$; $\chi^2 = 92.01$, $\text{df} = 18$, $p < 0.00001$; $I^2 = 80\%$), while sensitivity analysis showed the conclusions were robust.

3.3.3 Association between Plasma Neurofilament Light Chain Levels in AD and MCI

Fourteen studies including 1716 AD and 2707 MCI subjects tested NfL levels in the plasma [20,24,27–29,31,33,34,36–40,42]. Results demonstrated a significantly higher mean level of plasma NfL in the AD subjects when compared with MCI (SMD: 9.32 [8.07–10.57], $z = 14.62$, $p < 0.00001$, Fig. 4). The heterogeneity was low [$\chi^2 = 16.63$, $\text{df} = 13$, $p = 0.22$, $I^2 = 22\%$] between the studies and was not significantly affected by the specific study.

3.4 Investigation of Heterogeneity

Meta-regression analysis was conducted, and the results showed that age, gender (male ratio), and MMSE scores could not be regarded as possible sources of the heterogeneity observed between AD subjects and HC subjects (see Table 3). Subgroup analyses were performed to identify the cause of high heterogeneity in the comparison of plasma NfL levels between AD and healthy controls. It revealed that the different sex ratio and $\text{MMSE} \geq 20$ and

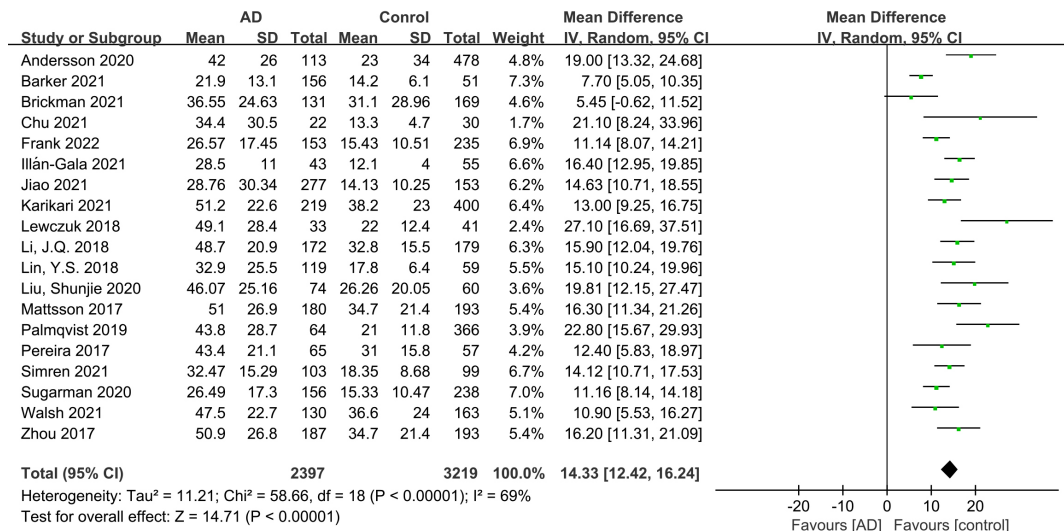


Fig. 2. Forest plot of random effects meta-analysis of plasma NfL levels in AD patients and HC subjects.

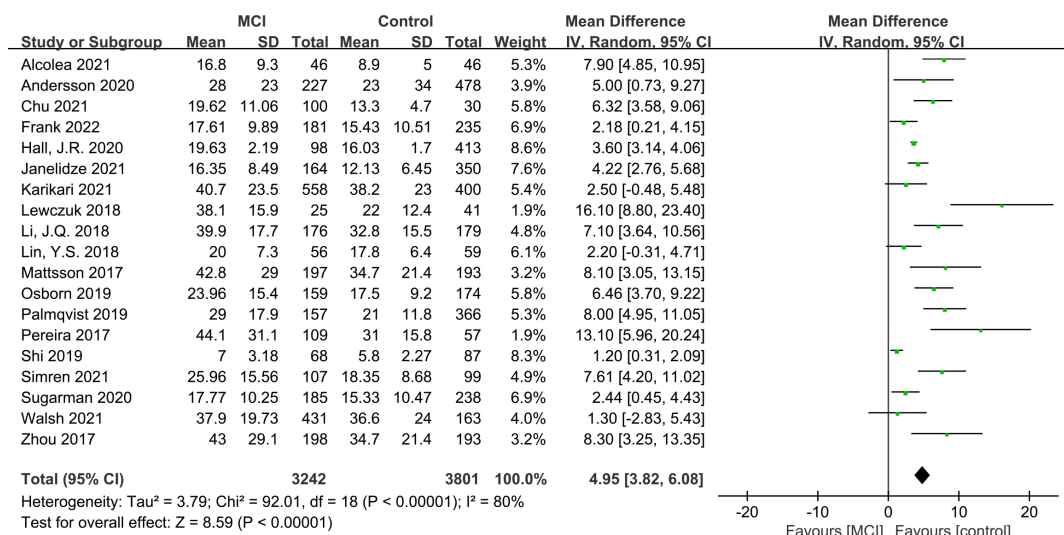


Fig. 3. Forest plot of random effects meta-analysis of plasma NfL levels in MCI patients and HC subjects.

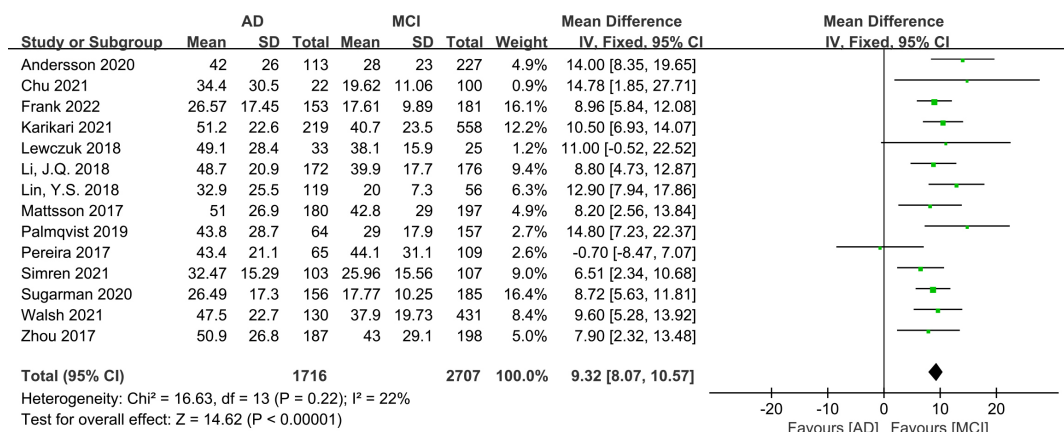


Fig. 4. Forest plot of fixed effects meta-analysis of plasma NfL levels in AD and MCI patients.

Table 1. Characteristics of included studies.

Author, Year	Country	Study group	N	Sex (F/M)	Age	MMSE	p-NfL level	Type of study	Analytical method
Andersson <i>et al.</i> , 2020 [20]	Sweden (BioFINDER)	AD	113	72/41	75 (7.2)	21.7 (3.7)	42 (26)	cross-sectional	Simoa
		MCI	227	92/135	70.6 (5.4)	27.1 (1.8)	28 (23)		
		HC	478	278/200	72.1 (5.5)	28.9 (1.2)	23 (34)		
Barker <i>et al.</i> , 2021 [21]	America (ADRC)	AD	156	88/68	74.8 (8.2)	25.3 (4.3)	21.9 (13.1)	cross-sectional	Simoa
		HC	51	40/11	70.8 (5.9)	29.4 (0.8)	14.2 (6.1)		
Brickman <i>et al.</i> , 2021 [22]	Spain, America	AD	131	69/62	82.99 (6.49)	NR	36.55 (24.63)	cross-sectional	Simoa
		HC	169	64/105	81.01 (6.31)	NR	31.10 (28.96)		
Janelidze <i>et al.</i> , 2021 [23]	Sweden (BioFINDER)	MCI	164	79/85	71 (7.48)	27 (2.99)	16.35 (8.49)	cross-sectional	Simoa
		HC	350	183/167	64 (16.38)	29 (1.49)	12.13 (6.45)		
Karikari <i>et al.</i> , 2021 [24]	America, Canada (ADNI)	AD	219	89/130	75.9 (7.9)	22.7 (3.0)	51.2 (22.6)	cross-sectional	Simoa
		MCI	558	237/321	72.9 (7.9)	28.0 (1.8)	40.7 (23.5)		
		HC	400	213/187	74.8 (6.6)	29.0 (1.3)	38.2 (23.0)		
Illán-Gala <i>et al.</i> , 2021 [25]	America	AD	43	27/16	65.2 (10)	21.5 (6)	28.5 (11)	cross-sectional	Simoa
		HC	55	30/25	52.2 (13)	28.9 (1)	12.1 (4)		
Hall, J.R. <i>et al.</i> , 2020 [26]	America	MCI	98	63/35	65.6 (8.48)	23.9 (3.65)	19.63 (2.19)	cross-sectional	Simoa
		HC	413	326/87	59.2 (6.97)	27.0 (2.60)	16.03 (1.7)		
Lewczuk <i>et al.</i> , 2018 [27]	Germany	AD	33	20/13	70.8 (7.6)	21.2 (3.4)	49.1 (28.4)	cross-sectional	Simoa
		MCI	25	15/10	71.3 (8.4)	26.7 (2.1)	38.1 (15.9)		
		HC	41	19/22	52.5 (13.1)	29.3 (0.9)	22.0 (12.4)		
Li, J.Q. <i>et al.</i> , 2018 [28]	America, Canada (ADNI-1)	AD	172	82/90	76 (7)	NR	48.7 (20.9)	cross-sectional	Simoa
		MCI	176	59/117	75 (8)	NR	39.9 (17.7)		
		HC	179	76/103	76 (5)	NR	32.8 (15.5)		
Lin, Y.S. <i>et al.</i> , 2018 [29]	China	AD	119	63/56	77.3 (5.1)	18.6 (6.2)	32.9 (25.5)	cross-sectional	Simoa
		MCI	56	29/27	76.0 (5.6)	26.4 (2.3)	20.0 (7.3)		
		HC	59	28/31	77.0 (6.2)	27.8 (2.1)	17.8 (6.4)		
Liu, Shunjue <i>et al.</i> , 2020 [30]	China	AD	74	42/32	73.2 (5.46)	21.12 (1.99)	46.07 (25.16)	cross-sectional	Simoa
		HC	60	34/36	71.95 (4.74)	28.53 (1.62)	26.26 (20.05)		
Mattsson <i>et al.</i> , 2017 [31]	America, Canada (ADNI)	AD	180	86/94	75.3 (7.3)	23.2 (2.1)	51.0 (26.9)	Longitudinal	Simoa
		MCI	197	65/132	74.7 (7.5)	26.9 (1.8)	42.8 (29.0)		
		HC	193	87/106	75.9 (4.9)	29.1 (1.0)	34.7 (21.4)		
Osborn <i>et al.</i> , 2019 [32]	Sweden	MCI	159	65/94	73 (7.7)	NR	23.96 (15.4)	cross-sectional	Simoa
		HC	174	71/103	72 (7.0)	NR	17.50 (9.2)		
Palmqvist <i>et al.</i> , 2019 [33]	Sweden (BioFINDER)	AD	64	39/25	76 (5)	21.8 (3.7)	43.8 (28.7)	Longitudinal	Simoa
		MCI	157	78/79	72 (5)	26.7 (1.8)	29.0 (17.9)		
		HC	366	214/152	72 (5)	28.9 (1.1)	21.0 (11.8)		

Table 1. Continued.

Author, Year	Country	Study group	N	Sex (F/M)	Age	MMSE	p-NfL level	Type of study	Analytical method
Pereira <i>et al.</i> , 2017 [34]	America, Canada (ANDI)	AD	65	31/34	73.7 (7.6)	23.5 (1.8)	43.4 (21.1)	cross-sectional	Simoa
		MCI	109	42/67	74.2 (6.9)	26.7 (1.8)	44.1 (31.1)		
		HC	57	30/27	74.8 (5.2)	29 (1)	31 (15.8)		
Shi. <i>et al.</i> , 2019 [35]	China	MCI	68	39/29	64.53 (7.68)	27.26 (1.67)	7.0 (3.18)	cross-sectional	Simoa
		HC	87	51/36	64.77 (7.40)	28.55 (1.16)	5.8 (2.27)		
Simrén <i>et al.</i> , 2021 [36]	Europe (AddNeuroMed)	AD	103	63/40	76.35 (5.76)	21.07 (4.42)	32.47 (15.29)	cross-sectional	Simoa
		MCI	107	56/51	74.47 (5.89)	27.21 (1.82)	25.96 (15.56)		
		HC	99	53/46	73 (6.14)	29.07 (1.26)	18.35 (8.68)		
Sugarman <i>et al.</i> , 2020 [37]	America (BU ADRC)	AD	156	69/87	76.74 (8.12)	21.11 (6.17)	26.49 (17.30)	cross-sectional	Simoa
		MCI	185	108/77	74.99 (7.24)	28.20 (1.67)	17.77 (10.25)		
		HC	238	149/89	72.38 (7.69)	29.39 (0.91)	15.33 (10.47)		
Walsh <i>et al.</i> , 2021 [38]	America, Canada (ANDI)	AD	130	57/73	74.2 (8.0)	23.1 (2.1)	47.5 (22.7)	cross-sectional	Simoa
		MCI	431	196/235	71.5 (7.5)	28.0 (1.7)	37.9 (19.7)		
		HC	163	86/77	73.6 (6.2)	29.0 (1.3)	36.6 (24.0)		
Zhou <i>et al.</i> , 2017 [39]	America, Canada (ANDI)	AD	187	90/97	75.5 (7.4)	23.3 (2.1)	50.9 (26.8)	cross-sectional	Simoa
		MCI	198	65/133	74.5 (7.4)	26.9 (1.8)	43.0 (29.1)		
		HC	193	87/106	75.7 (4.9)	29.1 (0.99)	34.7 (21.4)		
Chu <i>et al.</i> , 2021 [40]	America (ADRC)	AD	22	10/12	71.5 (9.2)	19.1 (7.8)	34.4 (30.5)	cross-sectional	Simoa
		MCI	100	51/49	73.4 (7.9)	27.6 (2.7)	19.6 (11.1)		
		HC	30	21/9	70.5 (6.7)	29.3 (1.0)	13.3 (4.7)		
Jiao <i>et al.</i> , 2021 [41]	China	AD	277	172/105	65.11 (10.57)	12 (6.44)	28.76 (30.34)	cross-sectional	Simoa
		HC	153	99/54	64.5 (8.2)	27.7 (2.3)	14.13 (10.25)		
Frank <i>et al.</i> , 2022 [42]	America (BU ADRC)	AD	153	67/86	76.82 (8.13)	21.12 (6.21)	26.57 (17.45)	cross-sectional	Simoa
		MCI	181	105/76	74.96 (7.25)	28.20 (1.68)	17.61 (9.89)		
		HC	235	148/87	72.38 (7.69)	29.39 (0.91)	15.43 (10.51)		
Alcolea <i>et al.</i> , 2021 [43]	Spain	MCI	46	28/18	72.6 (6.5)	25.8 (2.7)	16.8 (9.3)	cross-sectional	Simoa
		HC	46	24/22	54.8 (12.3)	29.1 (1.1)	8.9 (5)		

Note: AD, Alzheimer's disease; MCI, mild cognitive impairment; HC, healthy controls; BioFINDER, Biomarkers For Identifying Neurodegenerative Disorders Early and Reliably; ADNI, Alzheimer's Disease Neuroimaging Initiative; BU, Boston University; ADRC, Wisconsin Alzheimer's Disease Research Center; MMSE, Mini-Mental State Examination; Simoa, Single-molecule Array; NR, not reported.

MMSE <20, but not sample size, the age-matched and age-mismatched were possible sources of the heterogeneity observed. It was found that the studies with a large proportion of men and lower MMSE scores had lower heterogeneity ($I^2 = 35.6\%$, $p = 0.133$; $I^2 = 15.3\%$, $p = 0.307$) (see Table 4).

It was also found that for the comparison of plasma NfL levels between MCI and controls, the meta-regression analysis result showed a negative relationship (slope = -0.400 [95% CI: -0.519 to -0.281], $p < 0.05$) between the MMSE scores and effect size in MCI patients, indicating that the lower the MMSE scores, the SMD increased implying larger plasma NfL levels compared with healthy controls (see Table 3). In the subgroup analysis of age matching and sex ratio, the heterogeneity was unaffected, but it was found that the large sample size had no heterogeneity ($I^2 = 41.1\%$, $p = 0.104$) and that the small sample size had higher heterogeneity ($I^2 = 91.7\%$, $p < 0.05$) (see Table 4). Therefore, for the different sample sizes, the proportion of men and MMSE scores were both important sources of heterogeneity regarding plasma NfL levels in patients with AD and MCI.

3.5 Publication Bias

In the present study, publication bias was evaluated by visual inspection of a funnel plot and then confirmed by Egger's test. There was significant publication bias for plasma NfL level comparisons between AD and HC ($t = 2.69$, $p = 0.016$) and MCI and HC ($t = 2.38$, $p = 0.03$), while the results of Egger's test confirmed no significant publication bias among patients with AD and MCI. When six and eight virtual studies were separately added using the trim-and-fill-method the publication bias remained significant (all $p < 0.05$, see Fig. 5).

4. Discussion

Alzheimer's patients have a long, mild pre-clinical phase of cognitive impairment before showing the clinical symptoms typical of dementia. A clinical performance on memory impairment characterized as amnesic MCI (aMCI), a subtype of MCI, has increasingly been accepted as a high-risk condition for conversion to AD. Unfortunately, currently there are no effective treatments available to halt, slow, or reverse the progression of AD. Consequently, there is a significant clinical need for rapid and non-invasive diagnostic biomarkers to identify AD or MCI patients. Low levels of NfL can be detected in the blood and CSF of normal persons, with an increased concentration of NfL correlated with age. Recent research suggests that NfL is abnormally released into the CSF and blood with damage and degeneration following damage to various central and peripheral neurons [10]. The exact mechanism is not completely understood, although it most probably involves the destruction of cell membrane integrity. Recently, studies have suggested that NfL levels in the CSF and blood play an important role in patients with AD or MCI. In this

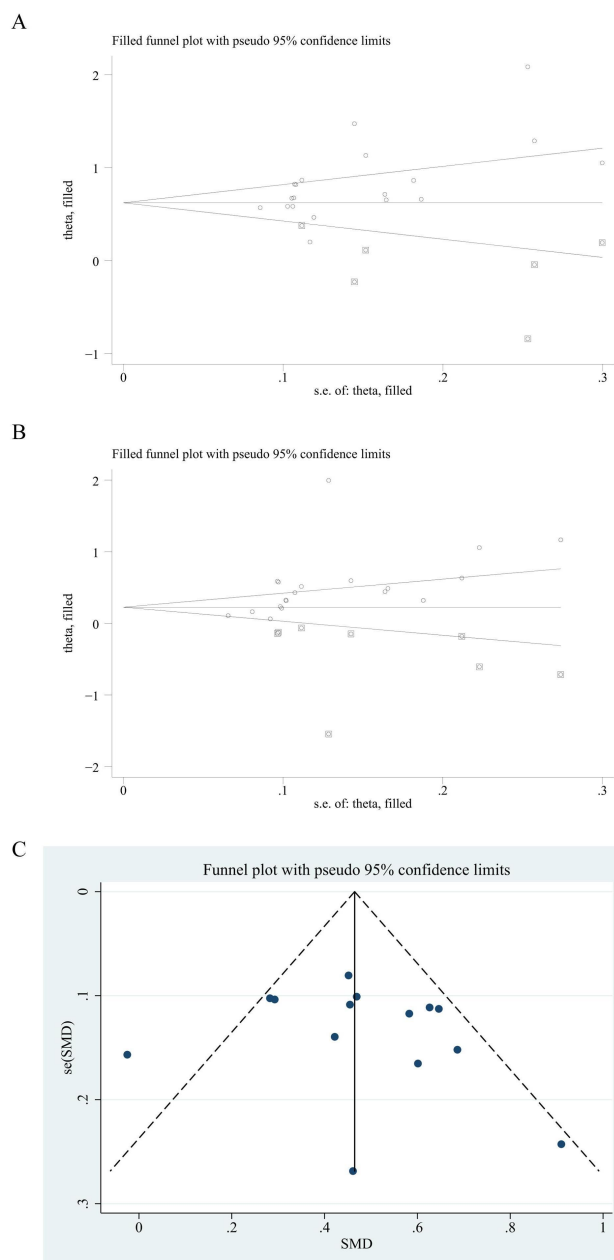


Fig. 5. Funnel plot of plasma NfL levels. (A) AD patients and HC subjects. (B) MCI patients and HC subjects. (C) AD and MCI patients.

study, due to the fact that it is inexpensive and simple to acquire, meta-analysis was used to explore the peripheral blood, for the development of diagnostic biomarkers of patients with AD or MCI. Plasma NfL levels may serve as one of the most promising fluid biomarkers for the validation of AD or MCI diagnosis [44]. Further, some animal studies have also reported that NfL is a potentially reliable biomarker for the severity of neuronal apoptosis [20,45]. Although there is a potential rationale that NfL can be regarded as a measure of the intensity of ongoing AD or MCI, the data remain controversial. Other work corroborates that there is no significant association between plasma NfL and

Table 2. The Newcastle-Ottawa Scale (NOS) for the quality assessment of studies.

Case-control studies				
Author, year	Selection	Comparability	Exposure	Score
Andersson <i>et al.</i> , 2020 [20]	☆☆☆☆	☆☆	☆☆	8
Barker <i>et al.</i> , 2021 [21]	☆☆☆	☆☆	☆☆☆	8
Brickman <i>et al.</i> , 2021 [22]	☆☆☆☆	☆☆	☆	7
Janelidze <i>et al.</i> , 2021 [23]	☆☆☆☆	☆☆	☆	7
Karikari <i>et al.</i> , 2021 [24]	☆☆☆	☆☆	☆☆	7
Illán-Gala <i>et al.</i> , 2021 [25]	☆☆☆	☆☆	☆☆	7
Hall, J.R. <i>et al.</i> , 2020 [26]	☆☆☆	☆☆	☆	6
Lewczuk <i>et al.</i> , 2018 [27]	☆☆	☆☆	☆☆	6
Li, J.Q. <i>et al.</i> , 2018 [28]	☆☆☆	☆☆	☆☆	7
Lin, Y.S. <i>et al.</i> , 2018 [29]	☆☆	☆☆	☆☆	6
Liu, Shunjie <i>et al.</i> , 2020 [30]	☆☆☆☆	☆☆	☆☆	8
Mattsson <i>et al.</i> , 2017 [31]	☆☆☆	☆☆	☆☆	7
Osborn <i>et al.</i> , 2019 [32]	☆☆☆	☆☆	☆	6
Palmqvist <i>et al.</i> , 2019 [33]	☆☆☆	☆☆	☆☆	7
Pereira <i>et al.</i> , 2017 [34]	☆☆☆	☆☆	☆	6
Shi <i>et al.</i> , 2019 [35]	☆☆☆	☆☆	☆☆	7
Simren <i>et al.</i> , 2021 [36]	☆☆☆	☆☆	☆	6
Sugarman <i>et al.</i> , 2020 [37]	☆☆☆	☆☆	☆	6
Walsh <i>et al.</i> , 2021 [38]	☆☆☆	☆☆	☆	6
Zhou <i>et al.</i> , 2017 [39]	☆☆☆	☆☆	☆	6
Chu <i>et al.</i> , 2021 [40]	☆☆☆	☆☆	☆☆	7
Jiao <i>et al.</i> , 2021 [41]	☆☆☆	☆☆	☆☆	7
Frank <i>et al.</i> , 2022 [42]	☆☆☆	☆☆	☆☆	7
Alcolea <i>et al.</i> , 2021 [43]	☆☆☆	☆☆	☆☆☆	8

Table 3. Meta-regression analysis of plasma NfL levels in patients with AD and MCI and HC subjects.

Group	Moderators	Coef.	95% CI		<i>p</i>
AD vs. HC	Age	−0.035	−0.083	0.013	0.144
	Gender	−0.014	−0.039	0.011	0.260
	MMSE	−0.002	−0.073	0.070	0.960
MCI vs. HC	Age	−0.052	−0.120	0.017	0.129
	Gender	−0.018	−0.038	0.003	0.091
	MMSE	−0.400	−0.519	−0.281	0.000

Note: AD, Alzheimer's disease; MCI, mild cognitive impairment; HC, healthy controls; NfL, neurofilament light chain; MMSE, Mini-Mental State Examination; CI, confidence interval.

cognitive decline [46]. Here, a total of 24 studies were extracted from the literature for this meta-analysis, and the following findings were obtained: Firstly, the level of NfL in plasma of patients with AD and MCI increased, simultaneously, and it was higher in patients with AD than in MCI subjects. Secondly, the concentration of NfL in plasma increased with cognitive decrease and was negatively correlated with MMSE scores. These findings provide clinical evidence that peripheral NfL levels can potentially be used as a biomarker for AD and MCI.

In the last few years, new biomarkers have emerged for the early diagnosis of AD, among which plasma NfL has recently been considered as a diagnostic and prognos-

tic biomarker for the preclinical stages of AD [47]. NfL levels in the plasma are closely related to several traditional biomarkers of AD, including amyloid beta ($A\beta$)₄₂, $A\beta$ _{42/40}, and $A\beta$ ₄₂/t-tau (total tau) [48]. NfL is a strongly proposed marker for the detection of neuronal injury or loss before the onset of the clinical symptoms of AD and cognitive dysfunction, brain atrophy, and disease progression monitored by the increase of the plasma NfL level [49], unlike the pathological mechanism of $A\beta$ and Tau. Neuronal damage and neuronal death are an important characteristic of AD pathology from the beginning of the presymptomatic stage of AD, and cognitive functions are associated with dendritic and axonal integrity [50]. Axonal integrity and

Table 4. Subgroup meta-analysis of plasma NfL levels in patients with AD and MCI and HC subjects.

Group	Assign criteria	No. of studies	SMD	95% CI		Heterogeneity		
						Q	p	I ²
AD vs. HC Subgroup								
Sample size	small	9	1.071	0.774	1.367	53.11	0.000	84.9%
	large	10	0.633	0.514	0.752	27.10	0.001	66.8%
Age (years)	matched	7	0.955	0.610	1.300	42.08	0.000	85.7%
	mismatched	12	0.755	0.596	0.914	62.96	0.000	82.5%
Proportion of men	≤50%	10	0.927	0.633	1.220	91.32	0.000	90.1%
	>50%	9	0.702	0.605	0.799	12.43	0.133	35.6%
MMSE	<20	3	0.668	0.477	0.858	2.36	0.307	15.3%
	≥20	14	0.867	0.697	1.038	78.64	0.000	83.5%
MCI vs. HC Subgroup								
Sample size	small	11	0.757	0.458	1.055	121.01	0.000	91.7%
	large	8	0.216	0.134	0.299	11.89	0.104	41.1%
Age (years)	matched	7	0.701	0.242	1.160	182.81	0.000	96.7%
	mismatched	12	0.389	0.269	0.509	34.45	0.000	68.1%
Proportion of men	≤50%	9	0.734	0.300	1.167	158.18	0.000	94.9%
	>50%	10	0.346	0.216	0.476	41.75	0.000	78.4%

Note: AD, Alzheimer's disease; MCI, mild cognitive impairment; HC, healthy controls; MMSE, Mini-Mental State Examination; SMD, standard mean difference; CI, confidence interval.

transport are directly associated with the degree of cognitive decline and neurodegeneration [51]. Therefore, NfL as a structural component of axons could be a promising tool for early diagnosis of AD. There is a growing body of evidence that axonal degeneration is an indicator of AD progression, affecting both brain structure and cortical metabolism, thus influencing AD's cognition [52]. Additionally, recent studies have demonstrated that the expression of NfL in plasma is elevated in patients with AD and is significantly associated with the degree of A β and tau in the CSF and positron emission tomography [31]. Here, the level of NfL was analysed in plasma of patients with AD and MCI and was found to be significantly higher than in HC. Additionally, a meta-analysis also found that blood NfL levels are higher in AD when compared to HC [53]. However, another study has shown that there was no association between plasma NfL levels in MCI and HC subjects [54]. They included small sample size to clarify whether blood NfL was a reliable biomarker, whereas here 24 articles were included, giving a larger patient number for analysis. A series of regression and subgroup analyses were also employed to confirm the conclusion reported here. Consequently, plasma NfL may have the potential to reflect axonal degeneration and be employed as a biomarker for AD and MCI.

In summary, these findings further validated that NfL in plasma can be used as a significant cognitive biomarker that distinguishes patients with AD and MCI from HC, but that the different ranges of NfL are unable to identify the different stages of the disease; therefore, in the future, other studies are needed to define the optimal range of plasma NfL values for prediction at the different stages of the disease.

There were several limitations to the meta-analysis reported here. Firstly, despite an exhaustive literature search, it is possible that some studies may have been missed and some publications had to be excluded due to small sample size and low quality. Secondly, the trim-and-fill-method which detects and adjusts for publication bias may affect the robustness of the results reported here. Thirdly, a high heterogeneity was found in this meta-analysis where comparison of AD with HCs and MCI with HCs may have reduced its statistical power.

5. Conclusions

The results of this meta-analysis suggest that there was a significant difference of plasma NfL between AD, MCI and the healthy controls and that there was a correlation between plasma NfL and cognitive dysfunction levels. From these results, it can be concluded that plasma NfL can serve as a biomarker for AD or MCI, but it cannot discriminate AD from other dementias or neurodegenerative diseases. Given the limited data, more cohort studies are required to confirm the results reported here.

Abbreviations

AD, Alzheimer's disease; MCI, mild cognitive impairment; Nfs, neurofilaments; NfL, neurofilament light; CSF, cerebrospinal fluid; PRISMA, Preferred Reporting Items for Systematic Review and Meta-analysis; NINCDS-ADRDA, National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association; NIA-AA, National Institute on Aging and Alzheimer's Association; Simoa,

single molecule array; MMSE, Mini-Mental State Examination; NOS, Newcastle-Ottawa scale; SMD, standardized mean difference; CI, confidence interval; A β , amyloid beta; HC, healthy controls.

Availability of Data and Materials

The datasets analyzed in this article are available upon request to: zmn1989@163.com.

Author Contributions

MZ and ZF designed the review. XL and JL collected and analyzed the data. CC was responsible for the result analysis and make figures. ZF and CC supervised the procedures. MZ wrote the manuscript. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Not applicable.

Funding

This research was funded by Zhejiang Medicine and Health Science and Technology Project, grant number 2020KY848; the Project of NINGBO Leading Medical & Health Discipline, grant number 2022-B12.

Conflict of Interest

The authors declare no conflict of interest.

Supplementary Material

Supplementary material associated with this article can be found, in the online version, at <https://doi.org/10.31083/j.jin2204085>.

References

- [1] Alzheimer's Association. 2022 Alzheimer's disease facts and figures. *Alzheimer's & Dementia*. 2022; 18: 700–789.
- [2] Querfurth HW, LaFerla FM. Alzheimer's disease. *The New England Journal of Medicine*. 2010; 362: 329–344.
- [3] Roberts R, Knopman DS. Classification and epidemiology of MCI. *Clinics in Geriatric Medicine*. 2013; 29: 753–772.
- [4] Petersen RC, Stevens JC, Ganguli M, Tangalos EG, Cummings JL, DeKosky ST. Practice parameter: early detection of dementia: mild cognitive impairment (an evidence-based review). Report of the Quality Standards Subcommittee of the American Academy of Neurology. *Neurology*. 2001; 56: 1133–1142.
- [5] Scheltens P, Blennow K, Breteler MMB, de Strooper B, Frisoni GB, Salloway S, *et al.* Alzheimer's disease. *Lancet*. 2016; 388: 505–517.
- [6] Vaz M, Silvestre S. Alzheimer's disease: Recent treatment strategies. *European Journal of Pharmacology*. 2020; 887: 173554.
- [7] Gaetani L, Blennow K, Calabresi P, Di Filippo M, Parnetti L, Zetterberg H. Neurofilament light chain as a biomarker in neurological disorders. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2019; 90: 870–881.
- [8] Puentes F, Lombardi V, Lu C, Yildiz O, Fratta P, Isaacs A, *et al.* Humoral response to neurofilaments and dipeptide repeats in ALS progression. *Annals of Clinical and Translational Neurology*. 2021; 8: 1831–1844.
- [9] Yuan A, Rao MV, Veeranna, Nixon RA. Neurofilaments and Neurofilament Proteins in Health and Disease. *Cold Spring Harbor Perspectives in Biology*. 2017; 9: a018309.
- [10] van Lieverloo GGA, Wieske L, Verhamme C, Vrancken AFJ, van Doorn PA, Michalak Z, *et al.* Serum neurofilament light chain in chronic inflammatory demyelinating polyneuropathy. *Journal of the Peripheral Nervous System*. 2019; 24: 187–194.
- [11] Molinuevo JL, Ayton S, Batrla R, Bednar MM, Bittner T, Cummings J, *et al.* Current state of Alzheimer's fluid biomarkers. *Acta Neuropathologica*. 2018; 136: 821–853.
- [12] Barro C, Chitnis T, Weiner HL. Blood neurofilament light: a critical review of its application to neurologic disease. *Annals of Clinical and Translational Neurology*. 2020; 7: 2508–2523.
- [13] Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *Annals of Internal Medicine*. 2009; 151: 264–269.
- [14] Dubois B, Feldman HH, Jacova C, Dekosky ST, Barberger-Gateau P, Cummings J, *et al.* Research criteria for the diagnosis of Alzheimer's disease: revising the NINCDS-ADRDA criteria. *The Lancet Neurology*. 2007; 6: 734–746.
- [15] Albert MS, DeKosky ST, Dickson D, Dubois B, Feldman HH, Fox NC, *et al.* The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's & Dementia*. 2011; 7: 270–279.
- [16] Petersen RC, Smith GE, Waring SC, Ivnik RJ, Tangalos EG, Kokmen E. Mild cognitive impairment: clinical characterization and outcome. *Archives of Neurology*. 1999; 56: 303–308.
- [17] Lo CK, Mertz D, Loeb M. Newcastle-Ottawa Scale: comparing reviewers' to authors' assessments. *BMC Medical Research Methodology*. 2014; 14: 45.
- [18] Wan X, Wang W, Liu J, Tong T. Estimating the sample mean and standard deviation from the sample size, median, range and/or interquartile range. *BMC Medical Research Methodology*. 2014; 14: 135.
- [19] Luo D, Wan X, Liu J, Tong T. Optimally estimating the sample mean from the sample size, median, mid-range, and/or mid-quartile range. *Statistical Methods in Medical Research*. 2018; 27: 1785–1805.
- [20] Andersson E, Janelidze S, Lampinen B, Nilsson M, Leuzy A, Stomrud E, *et al.* Blood and cerebrospinal fluid neurofilament light differentially detect neurodegeneration in early Alzheimer's disease. *Neurobiology of Aging*. 2020; 95: 143–153.
- [21] Barker W, Quinonez C, Greig MT, Behar R, Chirinos C, Rodriguez RA, *et al.* Utility of Plasma Neurofilament Light in the 1Florida Alzheimer's Disease Research Center (ADRC). *Journal of Alzheimer's Disease*. 2021; 79: 59–70.
- [22] Brickman AM, Manly JJ, Honig LS, Sanchez D, Reyes-Dumeyer D, Lantigua RA, *et al.* Plasma p-tau181, p-tau217, and other blood-based Alzheimer's disease biomarkers in a multi-ethnic, community study. *Alzheimer's & Dementia*. 2021; 17: 1353–1364.
- [23] Janelidze S, Palmqvist S, Leuzy A, Stomrud E, Verberk IMW, Zetterberg H, *et al.* Detecting amyloid positivity in early Alzheimer's disease using combinations of plasma A β 42/A β 40 and p-tau. *Alzheimer's & Dementia*. 2022; 18: 283–293.
- [24] Karikari TK, Benedet AL, Ashton NJ, Lantero Rodriguez J, Snellman A, Suárez-Calvet M, *et al.* Diagnostic performance

and prediction of clinical progression of plasma phospho-tau181 in the Alzheimer's Disease Neuroimaging Initiative. *Molecular Psychiatry*. 2021; 26: 429–442.

- [25] Illán-Gala I, Lleó A, Karydas A, Staffaroni AM, Zetterberg H, Sivasankaran R, *et al.* Plasma Tau and Neurofilament Light in Frontotemporal Lobar Degeneration and Alzheimer Disease. *Neurology*. 2021; 96: e671–e683.
- [26] Hall JR, Johnson LA, Peterson M, Julovich D, Como T, O'Bryant SE. Relationship of Neurofilament Light (NfL) and Cognitive Performance in a Sample of Mexican Americans with Normal Cognition, Mild Cognitive Impairment and Dementia. *Current Alzheimer Research*. 2020; 17: 1214–1220.
- [27] Lewczuk P, Ermann N, Andreasson U, Schultheis C, Podhorna J, Spitzer P, *et al.* Plasma neurofilament light as a potential biomarker of neurodegeneration in Alzheimer's disease. *Alzheimer's Research & Therapy*. 2018; 10: 71.
- [28] Li J, Yuan X, Li H, Cao X, Yu J, Tan L, *et al.* Genome-wide association study identifies two loci influencing plasma neurofilament light levels. *BMC Medical Genomics*. 2018; 11: 47.
- [29] Lin Y, Lee W, Wang S, Fuh J. Levels of plasma neurofilament light chain and cognitive function in patients with Alzheimer or Parkinson disease. *Scientific Reports*. 2018; 8: 17368.
- [30] Liu S, Huang Z, Zhang L, Pan J, Lei Q, Meng Y, *et al.* Plasma Neurofilament Light Chain May Be a Biomarker for the Inverse Association Between Cancers and Neurodegenerative Diseases. *Frontiers in Aging Neuroscience*. 2020; 12: 10.
- [31] Mattsson N, Andreasson U, Zetterberg H, Blennow K, Alzheimer's Disease Neuroimaging Initiative. Association of Plasma Neurofilament Light With Neurodegeneration in Patients With Alzheimer Disease. *JAMA Neurology*. 2017; 74: 557–566.
- [32] Osborn KE, Khan OA, Kresge HA, Bown CW, Liu D, Moore EE, *et al.* Cerebrospinal fluid and plasma neurofilament light relate to abnormal cognition. *Alzheimer's & Dementia*. 2019; 11: 700–709.
- [33] Palmqvist S, Janelidze S, Stomrud E, Zetterberg H, Karl J, Zink K, *et al.* Performance of Fully Automated Plasma Assays as Screening Tests for Alzheimer Disease-Related β -Amyloid Status. *JAMA Neurology*. 2019; 76: 1060–1069.
- [34] Pereira JB, Westman E, Hansson O, Alzheimer's Disease Neuroimaging Initiative. Association between cerebrospinal fluid and plasma neurodegeneration biomarkers with brain atrophy in Alzheimer's disease. *Neurobiology of Aging*. 2017; 58: 14–29.
- [35] Shi Y, Lu X, Zhang L, Shu H, Gu L, Wang Z, *et al.* Potential Value of Plasma Amyloid- β , Total Tau, and Neurofilament Light for Identification of Early Alzheimer's Disease. *ACS Chemical Neuroscience*. 2019; 10: 3479–3485.
- [36] Simrén J, Leuzy A, Karikari TK, Hye A, Benedet AL, Lantero-Rodriguez J, *et al.* The diagnostic and prognostic capabilities of plasma biomarkers in Alzheimer's disease. *Alzheimer's & Dementia*. 2021; 17: 1145–1156.
- [37] Sugarman MA, Zetterberg H, Blennow K, Tripodis Y, McKee AC, Stein TD, *et al.* A longitudinal examination of plasma neurofilament light and total tau for the clinical detection and monitoring of Alzheimer's disease. *Neurobiology of Aging*. 2020; 94: 60–70.
- [38] Walsh P, Sudre CH, Fiford CM, Ryan NS, Lashley T, Frost C, *et al.* The age-dependent associations of white matter hyperintensities and neurofilament light in early- and late-stage Alzheimer's disease. *Neurobiology of Aging*. 2021; 97: 10–17.
- [39] Zhou W, Zhang J, Ye F, Xu G, Su H, Su Y, *et al.* Plasma neurofilament light chain levels in Alzheimer's disease. *Neuroscience Letters*. 2017; 650: 60–64.
- [40] Chu WT, Wang W, Zaborszky L, Golde TE, DeKosky S, Duara R, *et al.* Association of Cognitive Impairment With Free Water in the Nucleus Basalis of Meynert and Locus Coeruleus to Transentorhinal Cortex Tract. *Neurology*. 2022; 98: e700–e710.
- [41] Jiao B, Liu H, Guo L, Liao X, Zhou Y, Weng L, *et al.* Performance of Plasma Amyloid β , Total Tau, and Neurofilament Light Chain in the Identification of Probable Alzheimer's Disease in South China. *Frontiers in Aging Neuroscience*. 2021; 13: 749649.
- [42] Frank B, Ally M, Brekke B, Zetterberg H, Blennow K, Sugarman MA, *et al.* Plasma p-tau₁₈₁ shows stronger network association to Alzheimer's disease dementia than neurofilament light and total tau. *Alzheimer's & Dementia*. 2022; 18: 1523–1536.
- [43] Alcolea D, Delaby C, Muñoz L, Torres S, Estellés T, Zhu N, *et al.* Use of plasma biomarkers for AT(N) classification of neurodegenerative dementias. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2021; 92: 1206–1214.
- [44] Leuzy A, Mattsson-Carlgren N, Palmqvist S, Janelidze S, Dage JL, Hansson O. Blood-based biomarkers for Alzheimer's disease. *EMBO Molecular Medicine*. 2022; 14: e14408.
- [45] Liu S, Zhang Z, Shi S, Meng Y, Zhang X, Lei Q, *et al.* NREM sleep loss increases neurofilament light chain levels in APP/PS1 and C57BL/6 J mice. *Sleep & Breathing*. 2022. (online ahead of print)
- [46] Moscoso A, Grothe MJ, Ashton NJ, Karikari TK, Lantero-Rodriguez J, Snellman A, *et al.* Longitudinal Associations of Blood Phosphorylated Tau181 and Neurofilament Light Chain With Neurodegeneration in Alzheimer Disease. *JAMA Neurology*. 2021; 78: 396–406.
- [47] Chatterjee P, Pedrini S, Ashton NJ, Tegg M, Goozee K, Singh AK, *et al.* Diagnostic and prognostic plasma biomarkers for pre-clinical Alzheimer's disease. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2022; 18: 1141–1154.
- [48] Ditttrich A, Ashton NJ, Zetterberg H, Blennow K, Simrén J, Geiger F, *et al.* Plasma and CSF NfL are differentially associated with biomarker evidence of neurodegeneration in a community-based sample of 70-year-olds. *Alzheimer's & Dementia*. 2022; 14: e12295.
- [49] Parvizi T, König T, Wurm R, Silvaieh S, Altmann P, Klotz S, *et al.* Real-world applicability of glial fibrillary acidic protein and neurofilament light chain in Alzheimer's disease. *Frontiers in Aging Neuroscience*. 2022; 14: 887498.
- [50] Teipel SJ, Stahl R, Dietrich O, Schoenberg SO, Perneczky R, Bokde ALW, *et al.* Multivariate network analysis of fiber tract integrity in Alzheimer's disease. *NeuroImage*. 2007; 34: 985–995.
- [51] Zetterberg H, Skillbäck T, Mattsson N, Trojanowski JQ, Portelius E, Shaw LM, *et al.* Association of Cerebrospinal Fluid Neurofilament Light Concentration With Alzheimer Disease Progression. *JAMA Neurology*. 2016; 73: 60–67.
- [52] Chen Y, Theriault J, Luo J, Ba M, Zhang H, Initiative ADN. Neurofilament light as a biomarker of axonal degeneration in patients with mild cognitive impairment and Alzheimer's disease. *Journal of Integrative Neuroscience*. 2021; 20: 861–870.
- [53] Koychev I, Jansen K, Dette A, Shi L, Holling H. Blood-Based ATN Biomarkers of Alzheimer's Disease: A Meta-Analysis. *Journal of Alzheimer's Disease*. 2021; 79: 177–195.
- [54] Zhang J, Cheng H, Liu W, Li H, Song Y, Jia L. Neurofilament light chain in cerebrospinal fluid or blood as a biomarker for mild cognitive impairment: A systematic review and meta-analysis. *Medicine*. 2022; 101: e28932.