Mitochondrial dysfunction and prostate cancer racial disparities among American men

Ajay K. Chaudhary<sup>1</sup>, Jordan O'Malley<sup>1</sup>, Sandeep Kumar<sup>1</sup>, Joseph R. Inigo<sup>1</sup>, Rahul Kumar<sup>1</sup>, Neelu Yadav<sup>1</sup>, Dhyan Chandra<sup>1</sup>

<sup>1</sup>Department of Pharmacology and Therapeutics, Roswell Park Cancer Institute, Buffalo, New York, USA

## TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. Reduced level of mtDNA associates with OXPHOS defects and predisposition to cancer in African American men
- 4. P53 regulation of mtDNA depletion and prostate cancer racial disparity
- 5. OXPHOS defects inhibit apoptosis leading to resistance to therapy in prostate cancer
- 6. OXPHOS defects contribute to aggressive nature of prostate cancer in African American men
- 7. Differential effect of cancer stem-like cells (CSCs) killing in prostate cancer
- 8. Conclusions and future perspectives
- 9. Acknowledgements
- 10. References

#### **1. ABSTRACT**

The gap between prostate cancer disparities among American men is narrowing, which is mostly due to increased screening of African American (AA) men. However, the biological reasons for prostate cancer disparities among American men still remain undefined. Mitochondrion, an organelle within cells, regulates both cell survival and cell death mechanisms. These cellular signaling pathways require various proteins localized to mitochondria, which are encoded by both nuclear DNA (nDNA) and mitochondrial DNA (mtDNA). Interestingly, prostate tissues from AA men harbor reduced mtDNA content compared to Caucasian American (CA) men. Therefore, changes in mitochondrial genes may have detrimental consequences in terms of cellular signaling regulated by mitochondria in AA men. This review describes the plausible underlying mechanism of mtDNA depletion and its impact in driving resistance to therapy leading to faster progression and poor prognosis in African American men with prostate cancer. Since defective cellular signaling is critical for prostate cancer cell survival, restoring mitochondrial function may provide strategies to alleviate prostate cancer disparities among American men.

#### 2. INTRODUCTION

Prostate cancer is one of the most commonly diagnosed cancers among American men. It is now well established that higher incidence and mortality rate associate with African American (AA) men compared to Caucasian American (CA) men (1-3). This gap between prostate cancer disparities among American men is narrowing, which is mostly due to increased screening of AA men (4, 5). Higher incidence and mortality rates in AA men with prostate cancer may occur due to various reasons, which may involve differences in molecular or genetics changes between AA and CA men with prostate cancer. Indeed, noteworthy molecular differences occur between AA and CA men with prostate cancer (6). Our findings suggest that mitochondrial DNA (mtDNA) depletion may play an important role in prostate cancer disparities among American men (7). MtDNA encodes 13 essential proteins required for proper functioning of the oxidative phosphorylation (OXPHOS) system. Therefore, reduced levels of mtDNA will contribute to reduced synthesis of mitochondrial-encoded OXPHOS proteins, causing a stoichiometric imbalance between nuclear- and mitochondrial-encoded subunits. The "mito-nuclear imbalance" causes defective OXPHOS complexes and leads to the development of an aggressive phenotype (8). Indeed, reduction of mtDNA content associates with acquisition of an androgen-independent phenotype and prostate cancer progression (9-11). In addition, reduced levels of mtDNA and mutations in mtDNA have been reported in multiple cancer types including prostate (12, 13) breast (14), renal (15), and liver (16). In contrast, increased variance in mtDNA copy number with paralleled increase in mtDNA mutations have also been reported in other types of cancer including prostate cancer (17-19). Mutations in mtDNA render respective protein products less active or dysfunctional and thus mimics the phenotype produced by mtDNA depletion. Thus, any imbalance of mtDNA may have deleterious consequences for prostate cancer patients. For example, depletion or reduction of mtDNA inhibits apoptosis, thereby contributing to resistance to

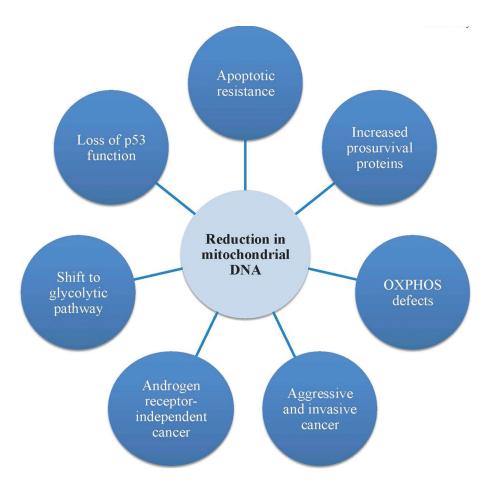


Figure 1. Impact of mitochondrial DNA depletion on various cellular functions in prostate cancer.

current available therapeutics (20-22), such as cisplatin and adriamycin (23). The present review highlights the importance of mtDNA reduction and mitochondrial dysfunction between AA in CA men with prostate cancer. We comprehensively highlight the significant impact of mtDNA depletion on many cellular functions in prostate cancer (Figure 1). Future studies may provide new avenues on how an imbalance of mtDNA and altered cellular signaling in cancer cells can predict therapy outcome and may allow for better personalized therapy for American men with prostate cancer.

## 3. REDUCED LEVEL OF mtDNA ASSOCIATES WITH OXPHOS DEFECTS AND PREDISPOSITION TO CANCER IN AFRICAN AMERICAN MEN

Although various factors contribute to tumorigenesis, a defective OXPHOS system is one of the hallmarks of cancer (24). The OXPHOS system consists of five multisubunit complexes including Complex I, II, III, IV and V. These complexes play critical roles during electron transfer and energy production in the form of ATP. The protein subunits of OXPHOS complexes are encoded by both nuclear DNA (nDNA) and mtDNA. MtDNA encodes 7 subunits of complex I, 1 subunit of complex III, 3 subunits of complex IV, and 2 subunits of complex V (19, 25, 26). We observed reduced level of mtDNA in prostate epithelial cells of AA men (7). Thus it could be concluded that except for Complex II, reduced level of mtDNA may cause imbalance between protein synthesis encoded by mtDNA and nDNA, leading to defective OXPHOS complexes. These assumptions are supported by the notion that mutation, reduction or deletion of mtDNA lead to defective OXPHOS, enhanced reactive oxygen species (ROS) production, shift to the glycolytic pathway, as well as increased expression of prosurvival proteins (26, 27). These activities may ultimately predispose normal cells with reduced mtDNA to cancerous phenotypes leading to increased cancer cell proliferation and tumorigenicity (22, 28-30) in AA men compared to CA men.

Prostate cancer incidence in AA men is higher than in CA men and AA men with prostate cancer present with a more aggressive phenotype (1-3). Reduced level of mtDNA in prostate cancer cells was observed in AA men (7). Thus reduction in mtDNA may be considered one

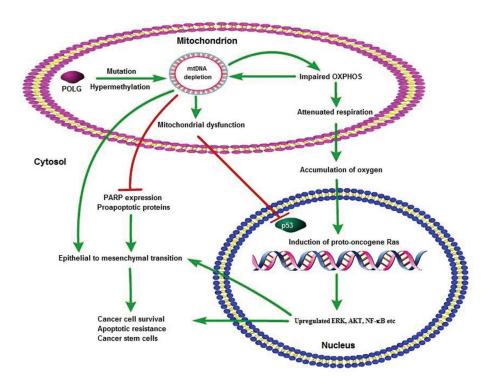


Figure 2. Mitochondrial DNA depletion modulates multiple cellular signaling leading to prostate cancer racial disparities among American men.

of the contributing factors for prostate cancer disparities among American men. As mentioned above, human mtDNA encodes 13 protein subunits of the mitochondrial respiratory chain, 22 transfer RNAs, and 2 ribosomal RNAs (25, 26). Thus, reduced mtDNA content may cause an imbalance in the OXPHOS complex assembly leading to the attenuation of mitochondrial respiration (25, 31), which will ultimately slow down oxygen consumption. This persistent reduction in mtDNA contributes to accumulation of oxygen, which induces a signaling cascade that leads to increased expression and activation of proto-oncogenic Ras in prostate epithelial cells (32). Ras activation constitutively triggers various signaling events including ERK, AKT, and NF-kB pathways. The combined effects of these survival signaling over a long period of time may contribute to other oncogenic changes and ultimately lead to initiation and progression of prostate cancer at greater extent in AA men compared to CA men (Figure 2). Together, the available evidence suggests a correlation of reduced mtDNA content with higher incidence of prostate cancer in AA men compared to CA men (1, 2). Since abrogation of OXPHOS function favors faster growth and invasiveness (33), reduced mtDNA level in already transformed prostate epithelial cells will promote more aggressive and invasive prostate cancer in AA men. Indeed, our recent published findings show that cancer cells derived from AA men with prostate cancer are highly invasive in nature compared to cancer cells derived from CA men (34). Animal models of mtDNA mutations further support our conclusion that mtDNA

mutations and reduced mtDNA content results in tumor development in mice (35).

## 4. *P53* REGULATION OF mtDNA DEPLETION AND PROSTATE CANCER RACIAL DISPARITY

We have demonstrated earlier that prostate tumors as well as normal tissues from AA men harbor reduced mtDNA levels compared to CA men (7). Although causes and consequences of these observations require further investigation, decreased level of mtDNA in AA men with prostate cancer suggests a strong correlation of prostate cancer incidence and aggressive prostate cancer in AA men compared to CA men. Thus the identification of underlying mechanisms of mtDNA reduction may have future significance in understanding the disease process in African American men with prostate cancer. It has been shown previously that mutations in the D-loop region of mtDNA may contribute to depletion/reduction of mtDNA, as the D-loop is the critical site responsible for replication of the mitochondrial genome because it contains the leading-strand of the origin of replication and the major promoter elements required for transcription (26, 27). MtDNA replication and repair are maintained by polymerase y (POLG), the only DNA polymerase in mitochondria (36). POLG is encoded by nDNA and is mutated in breast cancer and other mitochondria-associated diseases (37-39). It is interesting to note that POLG genes are also hypermethylated in

cancer stem-like cells and hypermethylation leads to silencing of POLG protein leading to the reduction of mtDNA (40). Thus it is possible that POLG mutations result in loss of POLG, which may lead to reduction in mtDNA causing mitochondrial dysfunction. Mitochondrial dysfunction could then trigger p53 mutations, which are rare in prostate adenocarcinomas. More likely, continued dysfunction of mitochondria may lead to loss of p53 function, which is a hallmark of transdifferentiated neuroendocrine prostate cancer, and is highly resistant to radiotherapy and androgen deprivation therapy (ADT) (41-43). Because mtDNA integrity is maintained by p53 protein (44) through its interaction with POLG (45), it is also possible that POLG mutations are due to nonfunctional p53 or due to p53 mutations in AA men with prostate cancer. These findings may have significance because higher tumor stage and grade, androgenindependence, and metastatic prostate cancer are associated with high levels of p53 mutations (46-48). Mitochondrial dysfunction causes the suppression of p53 activity (49) and p53 positively regulates mitochondrial respiration (50, 51). Therefore, reduced mtDNA levels in normal prostate epithelial and cancer cells from AA men may be associated with defects in POLG and p53 functions (Figure 2).

## 5. OXPHOS DEFECTS INHIBIT APOPTOSIS LEADING TO RESISTANCE TO THERAPY IN PROSTATE CANCER

Reduced mtDNA content in prostate cancer cells may suppress apoptosis either by directly targeting mitochondria and/or by depleting p53 protein in cancer cells derived from AA men. Apoptosis is a programmed cell death that requires the permeabilization of mitochondrial membrane leading to the release of apoptogenic proteins such as cytochrome c, Smac, and AIF (52-54). The permeabilization of mitochondrial membrane is regulated by Bcl-2 family proteins, which are divided into two broad groups: proapoptotic and antiapoptotic Bcl-2 proteins based on their function in the apoptosis program (55). Antiapoptotic proteins Bcl2, Bcl-xL and Mcl-1 inhibit channel-forming proteins Bax and Bak. Proapoptotic BH3-only proteins such as Bim/ Bid inhibit antiapoptotic proteins while activating Bax and Bak. The activation of BH3-only proteins triggers the permeabilization of the outer mitochondrial membrane leading to the release of apoptogenic proteins such as cytochrome c, which interacts with an adaptor protein Apaf-1 to form the apoptosome (56, 57). In the cytosol, the apoptosome initiates the caspase cascade leading to the execution of apoptosis. P53 is also known to activate mitochondrial apoptosis both by active transcription of proapoptotic proteins including Apaf-1, Bak and by direct activation of Bax on the mitochondrial membrane (58).

Reduced mtDNA level in both normal prostate epithelial and prostate cancer cells of AA origin may

inhibit apoptosis both by regulating mitochondrial channel activity and by deactivating/suppressing *p53* protein. It is interesting to note that depletion of mtDNA leads to the increased expression of antiapoptotic Bcl-2 protein (59). Reduction of mtDNA or mitochondria function also suppresses *p53* protein in mammalian cells (49). As mentioned above, *p53* regulates apoptosis by targeting mitochondria or transcriptional activation of proapoptotic proteins Apaf-1 and Bak. In addition, *p53* also positively regulates mitochondrial respiration (60).

Mitochondrial respiration is required for apoptosis execution (61-63). Reduction or depletion of mtDNA in cancer cells further abrogates mitochondrial respiration leading to the inhibition of apoptosis induced by multiple anticancer agents (21, 63-66). Thus reduced mtDNA will make prostate epithelial and tumor cells refractory to anticancer agents at greater level in AA men with prostate cancer (7). Indeed, reduction of mtDNA content associates with acquisition of an androgen-independent phenotype and prostate cancer progression (9-11). Although higher mtDNA level and resistance to anticancer agent docetaxel have been reported (67), which may be due increased mtDNA replication in response to oxidative stress. The lack of mtDNA in cancer cells of AA origin creates mitochondrial dysfunction leading to development of therapy resistance. The relatively higher level of mtDNA in CA men with prostate cancer will maintain relatively better functional mitochondria and thus respiration, which predicts better response to therapy compared to AA men with prostate cancer. Additionally, increased level of mtDNA in tumors from CA prostate cancer patients may lead to mitochondrial biogenesis and relatively functional mitochondria, which promote apoptosis (68) in response to chemotherapy.

Our recent findings demonstrate that prostate cancer cells from CA origin are sensitive to cell death in response to taxol upon restoration of mitochondria function by dichloroacetate (DCA) (34), a small molecule that restores mitochondria function. It is interesting to note that anticancer agent-induced cell death does not require caspase activation, which suggests that DCA restores mitochondria function and suppresses survival pathways leading to increased cell death without an increase in caspase activation. These findings also suggest the involvement of either caspase-independent cell death or non-apoptotic cell death in CA prostate cancer cells (69). Forcing cancer cells to switch from a glycolytic state (procancerous) to an OXPHOS (anti-cancerous) state is a new approach in cancer management/treatment (70) and may provide a therapeutic advantage to sensitize prostate cancer cells in both populations. However, such an effect was not observed in prostate cancer cells of AA origin, which may be due to various reasons. For example, AA cells possess very low basal levels of ROS and have decreased mitochondrial membrane potential (mtMP) compared to CA cells, thus DCA-induced increase in

mitochondrial ROS and mtMP in AA cells may not be sufficient to induce apoptosis. Thus severe mitochondria dysfunction is an important factor contributing to the development of resistance to conventional chemotherapies. Indeed, due to the presence of severe mitochondrial dysfunction characterized by highly reduced ROS levels, reduced mitochondrial biomass, and reduced mtMP, AA subjects respond poorly to first line chemotherapy (20, 71-74). Anticancer agents that restore mitochondria function will have therapeutic advantages for AA men with prostate cancer. We observed that doxorubicin increases mitochondrial biomass and function, with a subsequent increase in apoptotic cell death (75). Importantly, doxorubicin shows higher toxicity to prostate cancer cells of AA origin in the presence of DCA. Thus doxorubicin-induced cell death in prostate cancer of AA origin may be associated with restoration of some mitochondrial function to show higher toxic effects in AA cancer cells. However, due to the presence of severe mitochondria dysfunction in prostate cancer cells of AA men, doxorubicin alone and/or in combination with DCA is unable to restore mitochondria function above the threshold level to induce efficient cell death in prostate cancer cells of AA origin.

## 6. OXPHOS DEFECTS CONTRIBUTE TO AGGRESSIVE NATURE OF PROSTATE CANCER IN AFRICAN AMERICAN MEN

Depletion of mtDNA induces prostate cancer progression (9, 30, 76), prevents apoptosis, and promotes cell motility via upregulation of phosphatidylinositol 3-kinase (PI3K)/Akt2 signaling (77). Depletion of mtDNA also induces epithelial-mesenchymal transition (EMT) (10) and reduces PARP-1 levels (76), which indirectly inhibits apoptosis while promoting a more aggressive phenotype of the disease. For example, the androgen-independent derivative of LNCaP, LNCaP C42, expresses less mtDNA and consumes less oxygen than the parental LNCaP line (9). Strikingly, mtDNA-depleted LNCaP (LNp0) cells do not respond to androgens and grow significantly faster than the parental LNCaP cell line, which directly showcases the transformative property of mtDNA on prostate cancer cells (9). Furthermore, LNp0 cells are more motile and have increased expression of mesenchymal markers than the parental line, which demonstrates how mtDNA depletion may induce metastasis (10). Therefore, mtDNA reduction in prostate cancer cells is likely to inhibit apoptosis and promote invasiveness to a greater extent in AA men with prostate cancer. Based on these discussions, we suggest that reduced mtDNA content in AA men may be associated with more aggressive prostate cancers.

Mutated mtDNA also induces defective OXPHOS systems. Specific mutations in mtDNA have been shown to cause tumorigenesis and increased growth of the primary tumor. For example, a specific mutation in the gene ATP6 causes an increase in ROS formation and promotes growth of PC3 prostate cancer cells in vivo (28). Furthermore, PC3 cybrids containing the same ATP6 mutation grow more readily in a bone environment than wild-type PC3 cells, which indicates that mtDNA mutations augment bone metastatic events (11). Additionally, germline mutations in the cytochrome c oxidase subunit 1 gene (COI) are associated with prostate cancer in CA men (28). However, of the 15 COI SNPs identified, 9 were specific to AA men (78). Interestingly, SNP T7389C is significantly associated with AA men with prostate cancer (78). However, SNP T7389C is part of the African-specific mitochondrial haplogroup (L), which is used as a marker for African ancestry (79). Thus, more studies are required in order to ascertain the importance of this SNP in prostate cancer progression in AA men.

Mutated mtDNA has also been shown to directly affect metastatic potential. Alterations in the ND6 gene cause respiratory dysfunction of Complex I, induce ROS formation, and increase metastatic potential (29). Intriguingly, transferring the mutated MT-ND6 (mitochondrially encoded NADH: ubiquinone oxidoreductase core subunit 6) gene from a highly metastatic tumor cell line into a line with poor metastatic potential greatly heightened metastatic activity of the recipient cell line, thus illustrating the importance of mtDNA in inducing malignant phenotypes in cancer. Prostate cancer metastasis to the bone is a hallmark characteristic of advanced prostate cancer and is typically lethal. Indeed, the frequency of mtDNA mutations correlates to aggressiveness of the disease. For example, mtDNA mutations were detected in primary prostate tumors from a cohort of patients, but at a higher frequency in their visceral metastases and a higher frequency still in their bone metastases (80). Interestingly, a single missense mutation within the ND3 gene of complex I was detected in 77% of all bone metastases (80).

## 7. DIFFERENTIAL EFFECT OF CANCER STEM-LIKE CELLS (CSCs) KILLING IN PROSTATE CANCER

CSCs represent a small subpopulation of cells are responsible for tumor progression, metastasis, drug resistance, tumor relapse, and are associated with poor response to conventional interventions (81). CSCs isolated from prostate cancer are critical for prostate cancer development and metastatic progression (81). Our recent findings demonstrate that DCA treatment induces higher enrichment of CD44<sup>+</sup> and CD44<sup>+</sup>CD133<sup>+</sup> cell populations in AA cells compared to CA cells, which may be associated with higher metastatic potential (34). Mitochondrial dysfunction in CSCs may confer higher metastatic features in AA cells compared to CA prostate cancer cells. Indeed, prostate cancer cells that undergo mtDNA depletion also become more stem-like. PC3 and DU145 cells significantly overexpressed stemness markers, such as Oct3/4, Nanog, and CD44 upon treatment with ethidium bromide (82). Similar observations were made in ovarian cancer cells, whereby depleting mtDNA induced expression of stem cell markers (83). Conversely, mammospheres were found to have greater than 60 mitochondrial-related genes upregulated compared to cells cultured as monolayer, which illustrates a requirement for mitochondrial biogenesis in cancer stem cells (84). Indeed, PC3 prostate cancer spheres were greatly reduced in number when treated with various classes of antibiotics that inhibit mitochondrial biogenesis or OXPHOS (85). However, most studies reviewed here utilized AR-independent cell lines, which limit observation-based conclusions to address endstage, ADT-resistant prostate cancer. Therefore it is still uncertain how CSC-derived mitochondria may play a role in tumorigenesis.

## 8. CONCLUSIONS AND FUTURE PERSPECTIVES

Based on available evidence, we conclude that defective mitochondria are a leading cause of racial disparities among American men with prostate cancer. Mitochondrial dysfunction is caused by mtDNA depletion, which induces a mito-nuclear imbalance and subsequent defective assembly of OXPHOS complexes (8). The impaired OXPHOS complex function increases ROS production, which further mutates mtDNA, and is detrimental to anti-cancer proteins, such as p53. Dysfunctional mitochondria increase metastatic potential and stemness of prostate cancer cells and are associated with radioresistant disease (29, 80, 82, 83, 86). The insufficiency of mtDNA in AA men may render them more susceptible to developing prostate cancer, and helps to explain the higher incidence rate of prostate cancer within this group as well as why AA men typically have a more aggressive phenotype than CA men (Figure 1 and 2). Future studies focusing on the role of mtDNA copy number and/or mtDNA mutations and mitochondria-based cellular signaling may provide new strategies to differentially target metastatic diversity among American men for prostate cancer therapy and define novel biomarkers for prostate cancer aggressiveness.

# 9. ACKNOWLEDGEMENTS

Ajay K. Chaudhary, Jordan O'Malley contributed equally to this work. This work was supported in part by the National Cancer Institute of the National Institutes of Health under Award Number R01CA160685, and the American Cancer Society Research Scholar Grant RSG-12-214-01 – CCG to DC; and the National Cancer Institute Center Support Grant P30 CA016056 to the Roswell Park Cancer Institute. We apologize to those colleagues whose publications inadvertently could not be cited.

# **10. REFERENCES**

- A. Jemal, R. Siegel, E. Ward, Y. Hao, J. Xu, T. Murray and M. J. Thun: Cancer statistics, 2008. *CA Cancer J Clin*, 58(2), 71-96 (2008) DOI: 10.3322/CA.2007.0010
- G. Chornokur, K. Dalton, M. E. Borysova and N. B. Kumar: Disparities at presentation, diagnosis, treatment, and survival in African American men, affected by prostate cancer. *Prostate*, 71(9), 985-97 (2011) DOI: 10.1002/pros.21314
- P. A. Godley, A. P. Schenck, M. A. Amamoo, V. J. Schoenbach, S. Peacock, M. Manning, M. Symons and J. A. Talcott: Racial differences in mortality among Medicare recipients after treatment for localized prostate cancer. *J Natl Cancer Inst*, 95(22), 1702-10 (2003) DOI: 10.1093/jnci/djg094
- I. J. Powell, F. D. Vigneau, C. H. Bock, J. Ruterbusch and L. K. Heilbrun: Reducing prostate cancer racial disparity: evidence for aggressive early prostate cancer PSA testing of African American men. *Cancer Epidemiol Biomarkers Prev*, 23(8), 1505-11 (2014) DOI: 10.1158/1055-9965.EPI-13-1328
- C. E. DeSantis, R. L. Siegel, A. G. Sauer, K. D. Miller, S. A. Fedewa, K. I. Alcaraz and A. Jemal: Cancer statistics for African Americans, 2016: Progress and opportunities in reducing racial disparities. *CA Cancer J Clin* (2016) DOI: 10.3322/caac.21340
- F. Khani, J. M. Mosquera, K. Park, M. Blattner, C. O'Reilly, T. Y. MacDonald, Z. Chen, A. Srivastava, A. K. Tewari, C. E. Barbieri, M. A. Rubin and B. D. Robinson: Evidence for molecular differences in prostate cancer between African American and Caucasian men. *Clin Cancer Res*, 20(18), 4925-34 (2014) DOI: 10.1158/1078-0432.CCR-13-2265
- S. Koochekpour, T. Marlowe, K. K. Singh, K. Attwood and D. Chandra: Reduced mitochondrial DNA content associates with poor prognosis of prostate cancer in African American men. *PLoS One*, 8(9), e74688 (2013) DOI: 10.1371/journal.pone.0074688
- 8. R. H. Houtkooper, L. Mouchiroud, D. Ryu, N. Moullan, E. Katsyuba, G. Knott, R. W. Williams and J. Auwerx: Mitonuclear protein imbalance as a conserved longevity mechanism. *Nature*,

497(7450), 451-7 (2013) DOI: 10.1038/nature12188

- M. Higuchi, T. Kudo, S. Suzuki, T. T. Evans, R. Sasaki, Y. Wada, T. Shirakawa, J. R. Sawyer and A. Gotoh: Mitochondrial DNA determines androgen dependence in prostate cancer cell lines. *Oncogene*, 25(10), 1437-45 (2006) DOI: 10.1038/sj.onc.1209190
- A. Naito, C. C. Cook, T. Mizumachi, M. Wang, C. H. Xie, T. T. Evans, T. Kelly and M. Higuchi: Progressive tumor features accompany epithelial-mesenchymal transition induced in mitochondrial DNA-depleted cells. *Cancer Sci*, 99(8), 1584-8 (2008) DOI: 10.1111/j.1349-7006.2008.00879.x
- R. S. Arnold, C. Q. Sun, J. C. Richards, G. Grigoriev, I. M. Coleman, P. S. Nelson, C. L. Hsieh, J. K. Lee, Z. Xu, A. Rogatko, A. O. Osunkoya, M. Zayzafoon, L. Chung and J. A. Petros: Mitochondrial DNA mutation stimulates prostate cancer growth in bone stromal environment. *Prostate*, 69(1), 1-11 (2009)

DOI: 10.1002/pros.20854

- C. Jeronimo, S. Nomoto, O. L. Caballero, H. Usadel, R. Henrique, G. Varzim, J. Oliveira, C. Lopes, M. S. Fliss and D. Sidransky: Mitochondrial mutations in early stage prostate cancer and bodily fluids. *Oncogene*, 20(37), 5195-8 (2001) DOI: 10.1038/sj.onc.1204646
- J. Z. Chen, N. Gokden, G. F. Greene, P. Mukunyadzi and F. F. Kadlubar: Extensive somatic mitochondrial mutations in primary prostate cancer using laser capture microdissection. *Cancer Res*, 62(22), 6470-4 (2002)
- E. Mambo, A. Chatterjee, M. Xing, G. Tallini, B. R. Haugen, S. C. Yeung, S. Sukumar and D. Sidransky: Tumor-specific changes in mtDNA content in human cancer. *Int J Cancer*, 116(6), 920-4 (2005) DOI: 10.1002/ijc.21110
- A. Heddi, H. Faure-Vigny, D. C. Wallace and G. Stepien: Coordinate expression of nuclear and mitochondrial genes involved in energy production in carcinoma and oncocytoma. *Biochim Biophys Acta*, 1316(3), 203-9 (1996) DOI: 10.1016/0925-4439(96)00026-9
- 16. K. Luciakova and S. Kuzela: Increased

steady-state levels of several mitochondrial and nuclear gene transcripts in rat hepatoma with a low content of mitochondria. *Eur J Biochem*, 205(3), 1187-93 (1992) DOI: 10.1111/j.1432-1033.1992.tb16889.x

- M. S. Fliss, H. Usadel, O. L. Caballero, L. Wu, M. R. Buta, S. M. Eleff, J. Jen and D. Sidransky: Facile detection of mitochondrial DNA mutations in tumors and bodily fluids. *Science*, 287(5460), 2017-9 (2000) DOI: 10.1126/science.287.5460.2017
- J. B. Jones, J. J. Song, P. M. Hempen, G. Parmigiani, R. H. Hruban and S. E. Kern: Detection of mitochondrial DNA mutations in pancreatic cancer offers a "mass"-ive advantage over detection of nuclear DNA mutations. *Cancer Res*, 61(4), 1299-304 (2001)
- T. Mizumachi, L. Muskhelishvili, A. Naito, J. Furusawa, C. Y. Fan, E. R. Siegel, F. F. Kadlubar, U. Kumar and M. Higuchi: Increased distributional variance of mitochondrial DNA content associated with prostate cancer cells as compared with normal prostate cells. *Prostate*, 68(4), 408-17 (2008) DOI: 10.1002/pros.20697
- H. Pelicano, R. H. Xu, M. Du, L. Feng, R. Sasaki, J. S. Carew, Y. Hu, L. Ramdas, L. Hu, M. J. Keating, W. Zhang, W. Plunkett and P. Huang: Mitochondrial respiration defects in cancer cells cause activation of Akt survival pathway through a redox-mediated mechanism. *J Cell Biol*, 175(6), 913-23 (2006) DOI: 10.1083/jcb.200512100
- 21. J. S. Carew and P. Huang: Mitochondrial defects in cancer. *Mol Cancer*, 1, 9 (2002) DOI: 10.1186/1476-4598-1-9
- G. Amuthan, G. Biswas, H. K. Ananadatheerthavarada, C. Vijayasarathy, H. M. Shephard and N. G. Avadhani: Mitochondrial stress-induced calcium signaling, phenotypic changes and invasive behavior in human lung carcinoma A549 cells. *Oncogene*, 21(51), 7839-49 (2002) DOI: 10.1038/sj.onc.1205983
- W. Qian, M. Nishikawa, A. M. Haque, M. Hirose, M. Mashimo, E. Sato and M. Inoue: Mitochondrial density determines the cellular sensitivity to cisplatin-induced cell death. *Am J Physiol Cell Physiol*, 289(6), C1466-75 (2005) DOI: 10.1152/ajpcell.00265.2005

- D. Hanahan and R. A. Weinberg: Hallmarks of cancer: the next generation. *Cell*, 144(5), 646-74 (2011) DOI: 10.1016/j.cell.2011.02.013
- 25. D. Chandra and K. K. Singh: Genetic insights into OXPHOS defect and its role in cancer. *Biochim Biophys Acta*, 1807(6), 620-5 (2011) DOI: 10.1016/j.bbabio.2010.10.023
- N. Yadav and D. Chandra: Mitochondrial DNA mutations and breast tumorigenesis. *Biochim Biophys Acta*, 1836(2), 336-44 (2013) DOI: 10.1016/j.bbcan.2013.10.002
- 27. A. Chatterjee, E. Mambo and D. Sidransky: Mitochondrial DNA mutations in human cancer. *Oncogene*, 25(34), 4663-74 (2006) DOI: 10.1038/sj.onc.1209604
- J.A. Petros, A. K. Baumann, E. Ruiz-Pesini, M. B. Amin, C. Q. Sun, J. Hall, S. Lim, M. M. Issa, W. D. Flanders, S. H. Hosseini, F. F. Marshall and D. C. Wallace: mtDNA mutations increase tumorigenicity in prostate cancer. *Proc Natl Acad Sci U S A*, 102(3), 719-24 (2005) DOI: 10.1073/pnas.0408894102
- K. Ishikawa, K. Takenaga, M. Akimoto, N. Koshikawa, A. Yamaguchi, H. Imanishi, K. Nakada, Y. Honma and J. Hayashi: ROS-generating mitochondrial DNA mutations can regulate tumor cell metastasis. *Science*, 320(5876), 661-4 (2008) DOI: 10.1126/science.1156906
- G. Amuthan, G. Biswas, S. Y. Zhang, A. Klein-Szanto, C. Vijayasarathy and N. G. Avadhani: Mitochondria-to-nucleus stress signaling induces phenotypic changes, tumor progression and cell invasion. *EMBO J*, 20(8), 1910-20 (2001) DOI: 10.1093/emboj/20.8.1910
- S. Anderson, A. T. Bankier, B. G. Barrell, M. H. de Bruijn, A. R. Coulson, J. Drouin, I. C. Eperon, D. P. Nierlich, B. A. Roe, F. Sanger, P. H. Schreier, A. J. Smith, R. Staden and I. G. Young: Sequence and organization of the human mitochondrial genome. *Nature*, 290(5806), 457-65 (1981) DOI: 10.1038/290457a0
- 32. C. C. Cook, A. Kim, S. Terao, A. Gotoh and M. Higuchi: Consumption of oxygen: a mitochondrial-generated progression signal of advanced cancer. *Cell Death Dis*, 3, e258 (2012)

DOI: 10.1038/cddis.2011.141

- H. Simonnet, N. Alazard, K. Pfeiffer, C. Gallou, C. Beroud, J. Demont, R. Bouvier, H. Schagger and C. Godinot: Low mitochondrial respiratory chain content correlates with tumor aggressiveness in renal cell carcinoma. *Carcinogenesis*, 23(5), 759-68 (2002) DOI: 10.1093/carcin/23.5.759
- 34. A. K. Chaudhary, T. A. Bhat, S. Kumar, A. Kumar, R. Kumar, W. Underwood, S. Koochekpour, M. Shourideh, N. Yadav, S. Dhar, D. Chandra. Mitochondrial dysfunctionmediated apoptosis resistance associates with defective heat shock protein response in African-American men with prostate cancer. *Br J Cancer*, 114(10, 1090-100 (2016) DOI: 10.1038/bjc.2016.88
- P. L. Chen, C. F. Chen, Y. Chen, X. E. Guo, C. K. Huang, J. Y. Shew, R. L. Reddick, D. C. Wallace and W. H. Lee: Mitochondrial genome instability resulting from SUV3 haploinsufficiency leads to tumorigenesis and shortened lifespan. *Oncogene*, 32(9), 1193-201 (2013) DOI: 10.1038/onc.2012.120
- 36. L. S. Kaguni: DNA polymerase gamma, the mitochondrial replicase. *Annu Rev Biochem*, 73, 293-320 (2004)
  DOI: 10.1146/annurev.biochem.72.121801.1 61455
- S. Rajakulendran, R. D. Pitceathly, J. W. Taanman, H. Costello, M. G. Sweeney, C. E. Woodward, Z. Jaunmuktane, J. L. Holton, T. S. Jacques, B. N. Harding, C. Fratter, M. G. Hanna and S. Rahman: A Clinical, Neuropathological and Genetic Study of Homozygous A467T POLG-Related Mitochondrial Disease. *PLoS One*, 11(1), e0145500 (2016) DOI: 10.1371/journal.pone.0145500
- S. Azrak, V. Ayyasamy, G. Zirpoli, C. Ambrosone, E. V. Bandera, D. H. Bovbjerg, L. Jandorf, G. Ciupak, W. Davis, K. S. Pawlish, P. Liang and K. Singh: CAG repeat variants in the POLG1 gene encoding mtDNA polymerase-gamma and risk of breast cancer in African-American women. *PLoS One*, 7(1), e29548 (2012) DOI: 10.1371/journal.pone.0029548
- K. K. Singh, V. Ayyasamy, K. M. Owens, M. S. Koul and M. Vujcic: Mutations in mitochondrial DNA polymerase-gamma promote breast

tumorigenesis. *J Hum Genet*, 54(9), 516-24 (2009) DOI: 10.1038/jhg.2009.71

- 40. W. T. Lee and J. St John: The control of mitochondrial DNA replication during development and tumorigenesis. *Ann N Y Acad Sci*, 1350, 95-106 (2015) DOI: 10.1111/nyas.12873
- Q. Dang, L. Li, H. Xie, D. He, J. Chen, W. Song, L. S. Chang, H. C. Chang, S. Yeh and C. Chang: Anti-androgen enzalutamide enhances prostate cancer neuroendocrine (NE) differentiation via altering the infiltrated mast cells --> androgen receptor (AR) --> miRNA32 signals. *Mol Oncol*, 9(7), 1241-51 (2015) DOI: 10.1016/j.molonc.2015.02.010
- 42. C. D. Hu, R. Choo and J. Huang: Neuroendocrine differentiation in prostate cancer: a mechanism of radioresistance and treatment failure. *Front Oncol*, 5, 90 (2015) DOI: 10.3389/fonc.2015.00090
- Z. Li, C. J. Chen, J. K. Wang, E. Hsia, W. Li, J. Squires, Y. Sun and J. Huang: Neuroendocrine differentiation of prostate cancer. *Asian J Androl*, 15(3), 328-32 (2013) DOI: 10.1038/aja.2013.7
- 44. M. Kulawiec, V. Ayyasamy and K. K. Singh: p53 regulates mtDNA copy number and mitocheckpoint pathway. *J Carcinog*, 8, 8 (2009) DOI: 10.4103/1477-3163.50893
- G. Achanta, R. Sasaki, L. Feng, J. S. Carew, W. Lu, H. Pelicano, M. J. Keating and P. Huang: Novel role of p53 in maintaining mitochondrial genetic stability through interaction with DNA Pol gamma. *EMBO J*, 24(19), 3482-92 (2005) DOI: 10.1038/sj.emboj.7600819
- 46. J. T. Dong: Prevalent mutations in prostate cancer. *J Cell Biochem*, 97(3), 433-47 (2006) DOI: 10.1002/jcb.20696
- D. J. Grignon, R. Caplan, F. H. Sarkar, C. A. Lawton, E. H. Hammond, M. V. Pilepich, J. D. Forman, J. Mesic, K. K. Fu, R. A. Abrams, T. F. Pajak, W. U. Shipley and J. D. Cox: p53 status and prognosis of locally advanced prostatic adenocarcinoma: a study based on RTOG 8610. *J Natl Cancer Inst*, 89(2), 158-65 (1997) DOI: 10.1093/jnci/89.2.158
- 48. F. J. Meyers, P. H. Gumerlock, S. G. Chi, H.

Borchers, A. D. Deitch and R. W. deVere White: Very frequent p53 mutations in metastatic prostate carcinoma and in matched primary tumors. *Cancer*, 83(12), 2534-9 (1998) DOI: 10.1002/(SICI)1097-0142(19981215)8 3:12<2534:AID-CNCR19>3.3.CO;2-M DOI: 10.1002/(SICI)1097-0142(19981215)83 :12<2534:AID-CNCR19>3.0.CO;2-V

- S. Compton, C. Kim, N. B. Griner, P. Potluri, I. E. Scheffler, S. Sen, D. J. Jerry, S. Schneider and N. Yadava: Mitochondrial dysfunction impairs tumor suppressor p53 expression/function. *J Biol Chem*, 286(23), 20297-312 (2011) DOI: 10.1074/jbc.M110.163063
- S. Matoba, J. G. Kang, W. D. Patino, A. Wragg, M. Boehm, O. Gavrilova, P. J. Hurley, F. Bunz and P. M. Hwang: p53 regulates mitochondrial respiration. *Science*, 312(5780), 1650-3 (2006)
   DOI: 10.1126/science.1126863
- 51. R. F. Johnson and N. D. Perkins: Nuclear factor-kappaB, p53, and mitochondria: regulation of cellular metabolism and the Warburg effect. *Trends Biochem Sci*, 37(8), 317-24 (2012) DOI: 10.1016/j.tibs.2012.04.002
- 52. C. Adrain, E. M. Creagh and S. J. Martin: Apoptosis-associated release of Smac/ DIABLO from mitochondria requires active caspases and is blocked by Bcl-2. *EMBO J*, 20(23), 6627-36 (2001) DOI: 10.1093/emboj/20.23.6627
- J. Cai, J. Yang and D. P. Jones: Mitochondrial control of apoptosis: the role of cytochrome c. *Biochim Biophys Acta*, 1366(1-2), 139-49 (1998) DOI: 10.1016/S0005-2728(98)00109-1
- 54. C. Cande, N. Vahsen, C. Garrido and G. Kroemer: Apoptosis-inducing factor (AIF): caspase-independent after all. *Cell Death Differ*, 11(6), 591-5 (2004) DOI: 10.1038/sj.cdd.4401400
- 55. S. Zinkel, A. Gross and E. Yang: BCL2 family in DNA damage and cell cycle control. *Cell Death Differ*, 13(8), 1351-9 (2006) DOI: 10.1038/sj.cdd.4401987
- 56. S. B. Bratton and G. S. Salvesen: Regulation of the Apaf-1-caspase-9 apoptosome. *J Cell Sci*, 123(Pt 19), 3209-14 (2010)

DOI: 10.1242/jcs.073643

- N. Yadav and D. Chandra: Mitochondrial and postmitochondrial survival signaling in cancer. *Mitochondrion*, 16, 18-25 (2014) DOI: 10.1016/j.mito.2013.11.005
- J. E. Chipuk and D. R. Green: How do BCL-2 proteins induce mitochondrial outer membrane permeabilization? *Trends Cell Biol*, 18(4), 157-64 (2008) DOI: 10.1016/j.tcb.2008.01.007
- M. D. Jacobson, J. F. Burne, M. P. King, T. Miyashita, J. C. Reed and M. C. Raff: Bcl-2 blocks apoptosis in cells lacking mitochondrial DNA. *Nature*, 361(6410), 365-9 (1993) DOI: 10.1038/361365a0
- N. Yadava, S. S. Schneider, D. J. Jerry and C. Kim: Impaired mitochondrial metabolism and mammary carcinogenesis. *J Mammary Gland Biol Neoplasia*, 18(1), 75-87 (2013) DOI: 10.1007/s10911-012-9271-3
- N.Yadav, S. Kumar, R. Kumar, P. Srivastava, L. Sun, P. Rapali, T. Marlowe, A. Schneider, J.R. Inigo, J. O'Malley, R. Londonkar, R. Gogada, A.K. Chaudhary, N. Yadava, Chandra D. Mechanism of neem limonoidsinduced cell death in cancer: Role of oxidative phosphorylation. *Free radical biology & medicine* 2016; 90:261-71. DOI: 10.1016/j.freeradbiomed.2015.11.028
- N. Yadav, S. Kumar, T. Marlowe, A. K. Chaudhary, R. Kumar, J. Wang, J. O'Malley, P. M. Boland, S. Jayanthi, T. K. Kumar, N. Yadava and D. Chandra: Oxidative phosphorylationdependent regulation of cancer cell apoptosis in response to anticancer agents. *Cell Death Dis*, 6, e1969 (2015)
- D. Chandra, J. W. Liu and D. G. Tang: Early mitochondrial activation and cytochrome c up-regulation during apoptosis. *J Biol Chem*, 277(52), 50842-54 (2002) DOI: 10.1074/jbc.M207622200
- M. Higuchi, B. B. Aggarwal and E. T. Yeh: Activation of CPP32-like protease in tumor necrosis factor-induced apoptosis is dependent on mitochondrial function. *J Clin Invest*, 99(7), 1751-8 (1997) DOI: 10.1172/JCI119339
- D. S. McClintock, M. T. Santore, V. Y. Lee, J. Brunelle, G. R. Budinger, W. X. Zong, C. B. Thompson, N. Hay and N. S. Chandel:

Bcl-2 family members and functional electron transport chain regulate oxygen deprivationinduced cell death. *Mol Cell Biol*, 22(1), 94-104 (2002)

DOI: 10.1128/MCB.22.1.94-104.2002

- C. Chauvin, F. De Oliveira, X. Ronot, M. Mousseau, X. Leverve and E. Fontaine: Rotenone inhibits the mitochondrial permeability transition-induced cell death in U937 and KB cells. *J Biol Chem*, 276(44), 41394-8 (2001) DOI: 10.1074/jbc.M106417200
- T. Mizumachi, S. Suzuki, A. Naito, J. Carcel-Trullols, T. T. Evans, P. M. Spring, N. Oridate, Y. Furuta, S. Fukuda and M. Higuchi: Increased mitochondrial DNA induces acquired docetaxel resistance in head and neck cancer cells. *Oncogene*, 27(6), 831-8 (2008) DOI: 10.1038/sj.onc.1210681
- M. L. Boland, A. H. Chourasia and K. F. Macleod: Mitochondrial dysfunction in cancer. *Front Oncol*, 3, 292 (2013) DOI: 10.3389/fonc.2013.00292
- S. W. Tait, G. Ichim and D. R. Green: Die another way--non-apoptotic mechanisms of cell death. *J Cell Sci*, 127(Pt 10), 2135-44 (2014) DOI: 10.1242/jcs.093575
- S. Ponnala, C. Chetty, K. K. Veeravalli, D. H. Dinh, J. D. Klopfenstein and J. S. Rao: Metabolic remodeling precedes mitochondrial outer membrane permeabilization in human glioma xenograft cells. *Int J Oncol*, 40(2), 509-18 (2012)
- I. J. Powell, M. Banerjee, M. Novallo, W. Sakr, D. Grignon, D. P. Wood and J. E. Pontes: Prostate cancer biochemical recurrence stage for stage is more frequent among African-American than white men with locally advanced but not organ-confined disease. Urology, 55(2), 246-51 (2000) DOI: 10.1016/S0090-4295(99)00436-7
- 72. I. J. Powell, L. K. Heilbrun, W. Sakr, D. Grignon, J. Montie, M. Novallo, D. Smith and J. E. Pontes: The predictive value of race as a clinical prognostic factor among patients with clinically localized prostate cancer: a multivariate analysis of positive surgical margins. *Urology*, 49(5), 726-31 (1997) DOI: 10.1016/S0090-4295(96)00618-8

- M. Yu, Y. Zhou, Y. Shi, L. Ning, Y. Yang, X. Wei, N. Zhang, X. Hao and R. Niu: Reduced mitochondrial DNA copy number is correlated with tumor progression and prognosis in Chinese breast cancer patients. *IUBMB Life*, 59(7), 450-7 (2007) DOI: 10.1080/15216540701509955
- 74. I. R. Indran, G. Tufo, S. Pervaiz and C. Brenner: Recent advances in apoptosis, mitochondria and drug resistance in cancer cells. *Biochim Biophys Acta*, 1807(6), 735-45 (2011) DOI: 10.1016/j.bbabio.2011.03.010
- N. Yadav, A. Pliss, A. Kuzmin, P. Rapali, L. Sun, P. Prasad and D. Chandra: Transformations of the macromolecular landscape at mitochondria during DNA-damage-induced apoptotic cell death. *Cell Death Dis*, 5, e1453 (2014) DOI: 10.1038/cddis.2014.405
- L. Moro, A. A. Arbini, E. Marra and M. Greco: Mitochondrial DNA depletion reduces PARP-1 levels and promotes progression of the neoplastic phenotype in prostate carcinoma. *Cell Oncol*, 30(4), 307-22 (2008)
- 77. L. Moro, A. A. Arbini, J. L. Yao, P. A. di Sant'Agnese, E. Marra and M. Greco: Mitochondrial DNA depletion in prostate epithelial cells promotes anoikis resistance and invasion through activation of PI3K/Akt2. *Cell Death Differ*, 16(4), 571-83 (2009) DOI: 10.1038/cdd.2008.178
- A. M. Ray, K. A. Zuhlke, A. M. Levin, J. A. Douglas, K. A. Cooney and J. A. Petros: Sequence variation in the mitochondrial gene cytochrome c oxidase subunit I and prostate cancer in African American men. *Prostate*, 69(9), 956-60 (2009) DOI: 10.1002/pros.20943
- L. Quintana-Murci, O. Semino, H. J. Bandelt, G. Passarino, K. McElreavey and A. S. Santachiara-Benerecetti: Genetic evidence of an early exit of Homo sapiens sapiens from Africa through eastern Africa. *Nat Genet*, 23(4), 437-41 (1999) DOI: 10.1038/70550
- R. S. Arnold, S. A. Fedewa, M. Goodman, A. O. Osunkoya, H. T. Kissick, C. Morrissey, L. D. True and J. A. Petros: Bone metastasis in prostate cancer: Recurring mitochondrial DNA mutation reveals selective pressure exerted by the bone microenvironment. *Bone*, 78, 81-6 (2015)

DOI: 10.1016/j.bone.2015.04.046

- A. T. Collins, P. A. Berry, C. Hyde, M. J. Stower and N. J. Maitland: Prospective identification of tumorigenic prostate cancer stem cells. *Cancer Res*, 65(23), 10946-51 (2005) DOI: 10.1158/0008-5472.CAN-05-2018
- Y. Liu, X. Wu, X. Li, G. Kvalheim, U. Axcrona, K. Axcrona and Z. Suo: Blocking mtDNA replication upregulates the expression of stemness-related genes in prostate cancer cell lines. *Ultrastruct Pathol*, 37(4), 258-66 (2013) DOI: 10.3109/01913123.2013.770112
- R. Huang, J. Wang, Y. Zhong, Y. Liu, T. Stokke, C. G. Trope, J. M. Nesland and Z. Suo: Mitochondrial DNA Deficiency in Ovarian Cancer Cells and Cancer Stem Cell-like Properties. *Anticancer Res*, 35(7), 3743-53 (2015)
- R. Lamb, H. Harrison, J. Hulit, D. L. Smith, M. P. Lisanti and F. Sotgia: Mitochondria as new therapeutic targets for eradicating cancer stem cells: Quantitative proteomics and functional validation via MCT1/2 inhibition. *Oncotarget*, 5(22), 11029-37 (2014) DOI: 10.18632/oncotarget.2789
- R. Lamb, B. Ozsvari, C. L. Lisanti, H. B. Tanowitz, A. Howell, U. E. Martinez-Outschoorn, F. Sotgia and M. P. Lisanti: Antibiotics that target mitochondria effectively eradicate cancer stem cells, across multiple tumor types: treating cancer like an infectious disease. *Oncotarget*, 6(7), 4569-84 (2015) DOI: 10.18632/oncotarget.3174
- M. Baumann, M. Krause and R. Hill: Exploring the role of cancer stem cells in radioresistance. *Nat Rev Cancer*, 8(7), 545-54 (2008) DOI: 10.1038/nrc2419

Key Words:ProstateCancerRacialDisparity,MitochondrialDysfunction,OxidativePhosphorylation,OXPHOS,Apoptosis,MitochondrialDna,mtDNA,Review

Send correspondence to: Dhyan Chandra, Department of Pharmacology and Therapeutics, Roswell Park Cancer Institute, Elm and Carlton Streets, Buffalo, NY 14263, Tel: 716-845-4882, Fax: 716-845-8857, E-mail: dhyan.chandra@ roswellpark.org