PAMP-DAMPS interactions mediates development and progression of multiple sclerosis

Norma Y. Hernandez-Pedro¹, Roxana Magana-Maldonado², Aleli Salazar Ramiro², Veronica Perez-De la Cruz³, Edgar Rangel-Lopez⁴, Julio Sotelo², Benjamin Pineda²

¹Experimental Oncology Laboratory, National Cancer Institute, Av. San Fernando 22, 14080 Mexico City, Mexico, ²Neuroimmunology and Neuro-Oncology Unit, National Neurology and Neurosurgery Institute (INNN), Insurgentes sur 3877, 14269, Mexico City, Mexico, ³Neurochemistry Unit, National Neurology and Neurosurgery Institute (INNN), Insurgentes sur 3877, 14269, Mexico City, Mexico, ⁴Excitatory Amino Acids Laboratory, National Neurology and Neurosurgery Institute (INNN), Insurgentes sur 3877, 14269, Mexico City, Mexico

TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. Immunomodulation of TLR by PAMP-DAMPs
- 4. Role of PAMP DAMPs in multiple sclerosis
- 5. Conclusions
- 6. Acknowledgments
- 7. References

1. ABSTRACT

Multiple sclerosis (MS) is a disease presumably associated with chronic immune stimulation promoted by either pathogens or autoimmune processes. It has been hypothesized that MS could be the result of previous viral infections rendering a permanent immune stimulation that could be triggered by molecular similarities, or by modulating the antigens expression of major histocompatibility complex (MHC) on target cells, which in turn act as super antigens. During immune stimulation occurs the recruitment of immunological cells, resulting in local tissue damage and leading to the release of damage- associated molecular patterns (DAMPs), which also act as inflammation inducers. Recently, it has been proposed that the association between pathogen-associated molecular patterns (PAMP's) with DAMPs constitutes an additional level of immune regulation. The properties of DAMPs to act as carriers of PAMPs and their role as enhancers or inhibitors of PAMPs could play a role during inflammatory responses triggered by infections. Here, we focused this review in outcomes which support the hypothesis that particular PAMP- DAMPs interactions could regulated the relapse and progressive disability observed in multiple sclerosis.

2. INTRODUCTION

Multiple sclerosis (MS) has been described as an inflammatory, demyelinating and neurodegenerative disease which cause damage in Central Nervous System (CNS). MS affects mainly young adults and it causes non-traumatic disability; therefore, it has a great socioeconomic impact in both family and patients. During the development of MS, several neurologic symptoms are originated and they are followed by neurologic disorders that remain latent in patients, increasing their physical disability and eliciting a complete socioeconomic dependence beyond 30 years, resulting in a plethora of neurological symptoms in the most productive stage of the patient's life (1). MS affects women two-fold more than men and has a peak onset between 20 and 40 years old; however, it also may be found in children and occasionally in individuals aged above 60 years old. (2).

During the natural course of the disease, more than 60% of patients develop the relapsing- remitting form of MS (known as RRMS). At this stage, some neurological symptoms may arise followed by a partial or complete recovery of the patient (3-5). However, a high percentage of patients affected by RRMS (up to 90% younger than 25 years old) develop secondary progressive multiple sclerosis (SPMS), which is characterized by neurological degeneration without any relapsing episodes (3-7). Approximately, 15% of MS patients are diagnosed with primary progressive MS (PPMS) which show neurological symptoms that advance progressively from the beginning of the disease (3-5,8).

Besides the complexity of the pathophysiology of MS disease, several factors have been postulated in its development, such as genetic susceptibility, environmental conditions, and an abnormal immunemediated response leading to autoimmunity which causes focal destruction of myelin, axonal loss and inflammatory cells infiltrate (9). MS is characterized by chronic inflammation of the CNS that led to auto-reactivity immune in cells that destroy many components of the CNS. Oligodendrocytes form glial scarce in the nervous tissue, and the myelin sheaths of axons are affected (5). Particularly, the demyelinated plaques observed in MS patients consist in a well-delimited area with scarce cells characterized by loss of myelin sheath in neurons with relative preservation of their axons and the formation of astrocytic scars (10).

The presence of active plaques is the result of brain blood barrier (BBB) leakage, glial scar formation and the presence of inflammatory infiltrates (10). These infiltrates consist of auto reactive lymphocyte T cells, macrophages, and mast cells, which could reach to the CNS inducing a pro-inflammatory response, followed by local tissue damage (11-13). The formation of active plaques initiates when mononuclear cell infiltrates, which are concentrated in perivascular spaces, induce the destruction of myelin sheath. These infiltrates are composed mainly by B and T cells, plasmatic cells, and macrophages (14). Poskanzer introduced the hypothesis that MS is a late consequence of an infectious disease which is frequently acquired in childhood (15). More than 30 viral antigens, including virus for rabies, herpes simplex virus, varicella zoster, measles, corona virus, canine distemper virus, HTLV-1, and Epstein-Barr virus (EBV) have been tested to demonstrate the presence of antibodies in serum from patients with MS (14,16-21).

Currently, the main candidate related with MS is Epstein-Barr virus (EBV). Although this has not been confirmed as specific trigger for this disease; several authors suggest that EBV is involved in the initial disease phase. (22,23). This virus might induce damage on myelin sheaths by autoimmune processes triggered by molecular mimicry, expression and overexpression of MHC antigens on target cells, and also, EBV might act as super antigens (24). During the beginning and the development of MS, the persistent cell infection by virus infiltrates the CNS and modulates the immune response, activating both T and B cells, similar process that occurs when a pathogenic infection is involved (25-27). Besides, there are other mechanisms of activation of TLR where pathogens-associated molecular patterns (PAMPs) are not involved, which consist in endogenous inflammatory mediators called damage-associated molecular patterns (DAMPs) that also regulate immune response. However, the role played by DAMPs in inflammation/immunity during virus infection has little been studied (28).

3. IMMUNOMODULATION OF TLR BY PAMP-DAMPS

The pattern recognition receptors family (PRRs) is known to be the first line of defense against foreign pathogens which detects distinct evolutionarily conserved structures. PRRs are comprised by TLRs, which are activated either by PAMPs, DAMPs or by the PAMP- DAMPs complex. This association is essential to

initiate the development of several autoimmune diseases such as experimental autoimmune encephalomyelitis (EAE) and MS (29-31). These receptors can be found in the outer of surface cells, into the membrane of endosomes of several cell types, or non-immune and immune cells, where TLRs mainly recognize macrophages, B cells (32) and other antigen presenting cells (APCs) such as dendritic cells (DCs) (32,33). Several TLRs including types 1, 2, 4, 6, and 10 are expressed in the outer membrane where they recognized and respond primarily to bacterial and viral antigens surface associated to PAMPs. Also, TLRs 3, 7, 8 and 9 are preferentially activated by viral antigens, being able to recognize specific nucleic acid (either DNA or RNA) based PAMPs of intracellular pathogens (34).

The interaction of TLRs with PAMPs and DAMPs causes the activation of genes encoding for proinflammatory cytokines, chemokines, and co-stimulatory molecules that consequently trigger innate immune responses and prime antigen-specific T cells (35). Also, PAMPs and DAMPs are able to activate NF $\kappa\beta$, which in turn may triggers PI3K/AKT and Ras/MAPK signaling that are involved in both cell survival and mitogenic process (36). Abnormal TLR stimulation contributes to the development of many inflammatory and autoimmune diseases (35,37).

The repertory of DAMPs is determined by some factors such as the type of cell death, cell- line as well as the damaged tissue (38). Cell death processes and biochemical pathways determine the kind of DAMPs exposed, released, and their function. The DAMPs that trigger the activation of inflammasome include the mammalian cytosolic double-stranded RNA, low intracellular K+ levels, some heat shock proteins (Hsp) and gp96 (39,40). Hsp90 and Hsp70 enhance immune responses through a chaperone activity: where the generation of Hsp- antigen complexes promotes the presentation of unlinked antigens, maturation of APC and stimulation of cytotoxic T lymphocytes through MHC class I (41). In the innate immune system, Hsp act as immune-stimulators, suggesting that those proteins can contributed to the development of the autoimmune response after cell damage (24,42) (Figure 1).

Sometimes, PAMPs and DAMPs bind to the same receptors modulating their activation through TLRs promoting the synthesis of cytokines (43,44). This PAMP–DAMPs association is currently proposed as a new modality of immune regulation which depends on the preexisting collection of PAMPs and DAMPs, the assembly of the PAMP–DAMPs complexes, and the repertoire of PAMPs and DAMPs. Finally, PAMPs can induce the release of DAMPs during an infectious process, eliciting the release of others DAMPs (45). Besides, it has been described that many DAMPs could act as adjuvants, for example, genomic double- stranded DNA enhanced

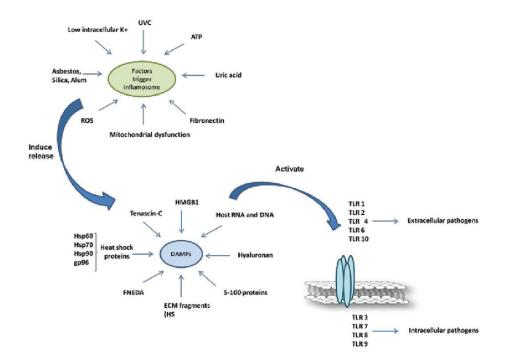


Figure 1. Proposed factors to induce the release of PAMPs/DAMPs, and their interaction with TLRs. Diverse exogenous and endogenous factors, such as asbestos, metabolic dysfunction, host RNA and DNA amongst others, promote the induction of DAMPs. These DAMPs/PAMPs could interact with different TLRs, which in turn, could recognize extracellular and intracellular pathogens.

both antibody and T cell responses in mice when they were administrated with these molecules (46). Likewise, lactoferrin, defensins, low molecular weight hyaluronic acid (HA), and high-mobility group 1 (HMGB1) showed adjuvant properties in animal models (47-49).

Diverse studies had described that PAMPs and/or DAMPs by themselves are able to modulate the course of inflammation and activate adaptive immunity response (50, 51). Piccini describes that lipopolysaccharide (LPS) from Gram-negative bacteria could act as a PAMP capable to bind to some DAMPs, such as HMGB1, Hsp70, Hsp90, surfactant protein A and α -defensins. (35). These complexes are recognized by TLR4 allowing the release of TNF- α and IL- 6 and thus perpetuating the damage to tissue due to inflammation (52).

4. ROLE OF PAMP DAMPS IN MULTIPLE SCLEROSIS

The family of herpes virus such as Herpes Simplex 1 and 6, Epstein-Bar, and Varicela Zoster has been suggested as the best candidates in the MS pathogenesis. Herpes viruses have the ability to cause lytic infection in permissive cells, and to remain latently in specific cell types, such as neurons, myeloid progenitor's cells and/or lymphocytes (53). In addition, they have the capacity to evade the innate immune system, given their slow replication cycle and their persistence of life-long latent infections, producing recurrent infections (53). Several reports suggest an association between infection and re-activation of Human Herpes Virus (HHV) during MS course. For example, the HHV type 1 has been isolated from the cerebrospinal samples of patients during the first manifestations of MS (54). Also, the reactivation of HHV-6 has been associated with MS exacerbation (55-60). On the other hand, EBV infection is frequently associated with autoimmune disorders, such as RA, MS, Sjogren's syndrome (SS), thyroiditis and hepatitis autoimmune (61-68). EBV is characterized for its ability to infect, and to persist in a silence mode inside B-lymphocytes for the lifetime in infected patients. It has been found the presence of EBV into infected cells in the vast majority of MS cases, although the association between EBV in MS remains in controversy.

During the infection with HHV, some conserved herpesvirus protein kinases (CHPKs) have been associated with Hsp 90, which participates in the viral replication and infection promoting the immune recognition of antigens in MS (69-71). Likewise, Hsp 90 regulates the immunomodulation of $\gamma\delta$ T cells in the acute phase of infection with EBV on B cell (72).

Also, it has been described that the number of circulating $\gamma\delta$ T cells are increased in patients with MS during the early onset of the disease; however, the depletion of $\gamma\delta$ T cells in the CNS before the onset of acute disease, or during the chronic stage, causes significant reduction in the severity of the clinical symptoms (73).

Particularly, it was demonstrated that the depletion of the $\gamma\delta$ T-cells during the acute stage of EAE in animals, resulted in a significant reduction of inflammatory and demyelination processes. According to this, it could be possible that during the early phase of HHV infection, Hsp 90 is able to interact with $\gamma\delta$ T cells eliciting the release of Th1-type cytokines that tend to produce the inflammatory responses to kill intracellular viruses. In spite that the numbers of $\gamma\delta$ T cells are decreased during chronic stages, and the production of Hsp90 is continuous, some pathways could be activated due their interactions with PPRs, thus perpetuating an immune response within active plaques in MS.

Previous studies have indicated that Hsp70 is a critical molecule in MS pathogenesis (74). In the EAE model induced by Myelin Oligodendrocyte Glycoprotein (MOG), indicated that Hsp70.1 could play a role in both the immune response and cytoprotection of CNS cells. In addition, extracellular Hsp70 can act as an adjuvant that promotes adaptive immune responses against specific antigens. Additionally, complexes of Hsp70 and MOG or PLP were present in the CNS of mice with EAE (75). In this context, complexes of Hsp70 and MOG or proteolipid protein (PLP) were found in MS lesions. Also, it has been shown an increase in the protein expression of inducible Hsp70 in T lymphocytes (CD4+ and CD8+) and monocytes from under basal conditions that may reflect the immunological activation (74). These data suggest that during the initial stages of MS/EAE the inflammatory response cts as a preconditioning stimulus to induce the expression of Hsp70 as well as the releasing f this protein from glial cells in order to avoid neuronal damage in the successive neurodegenerative stages (76).

In the Japanese encephalitis (JE) induced by HPV, Hsp70/90 produce a subsequent inflammatory reaction regulated by TLR4 during viral replication (77). These results indicate that deficient TLR4 (-/-) mice showed enhanced resistance to JE, however some data revealed that TLR4 reduction also provided potent type I IFN innate responses through enhanced induction of antiviral ISG genes by alternative activation of IRF-3 and NF-KB in myeloid-derived DCs and macrophages. Also, TLR4-/- mice showed an alteration of plasmacytoid DCs subpopulation and CD4+Foxp3+ regulatory T cells, which were closely associated with enhanced type I IFN innate immune and JEV-specific CD4+ and CD8+ T cell responses (77). These Hsp proteins act as "protectors" to eliminate viruses during early stage of the infection; however, if this damage remains constant, the releasing of Hsp promotes a persistent inflammation, thus contributing to the MS development.

Nowadays, it is unclear the role-played by HMGB1 during the establishment of MS, or its contribution in boost progressive autoimmune disease. HMGB1 is a ubiquitous nuclear protein which is continuously released from necrotic cells or it is unrestrictedly secreted by monocytes, macrophages and DC's (78,79). HMGB1 contributes to the nuclear homeostasis by acting as an extracellular alert signal when tissues are injured; therefore HMGB1 could act as a typical DAMP molecule (79,80). Even when some authors suggest that HMGB1 participates during non-pathogenic inflammatory processes, but it is required to initiates sterile inflammation following tissues injury (81), there is enough evidence that implicates to this nuclear protein during infections. It has been showed that the releasing of HMGB1 ispromoted by pro inflammatory cytokines generated during HIV-1 infection, and also it is able to activate the replication in latent HIV-1 (82). Besides, during early response of infectious or some damage process, extracellular HMGB1 is able to trigger inflammation, (83). HMGB1 has been detected in the nuclei of nervous cells (84), and there are significant extracellular levels of HMGB1 as well as the receptors for advanced glycation end products (RAGE), TLR2, and TLR4 in cerebrospinal samples of MS patients (85). Furthermore, microglia and macrophages express cytosolic HMGB1 and this expression is increased in active plagues of MS patients and in the EAE model (85). HMGB1-mediated TLR2, TLR4, TLR9 and RAGE signaling pathways are involved in the NF-KB modulation, thus participating in the releasing of pro-inflammatory molecules from macrophages and promoting the recruitment of inflammatory mediators through endothelial barriers of damaged tissue such as it has been observed in EAE mice model (86).

Neutralization of HMGB1 ameliorates experimental autoimmune encephalomyelitis, and this amelioration was associated with defective systemic T cell activation and decreased T cell recruitment into the CNS (87). Also, in patients with amyotrophic lateral sclerosis (ALS) the levels of HMGB1 are increased in spinal cord tissues when they were compared with Alzheimer's disease, Parkinson's disease, and healthy control subjects. Also the amount of auto-antibodies against HMGB1 correlated with the severity of the disease (88). On the other hand, it has been observed that high levels of HMGB1 and tenascin C are present in the serum of septic patients (78,89), while in systemic lupus erythematosus (SLE) high levels of DNA have been associated with immune complexes in serum, including the nucleosome-HMGB1 complex (90-92). In several cases, increased levels of endogenous TLR are considered as markers of the progress disease; also, high levels of extracellular HMGB1 recognized active lesions in MS patients correlating with a dynamic inflammatory process (85).

Other DAMPs associated with MS are beta amyloid (β -AP) and S100 proteins which have been involved in tissue homeostasis and regeneration/ repairing processes above of all acute injury. These molecules are proposed as important neural damage

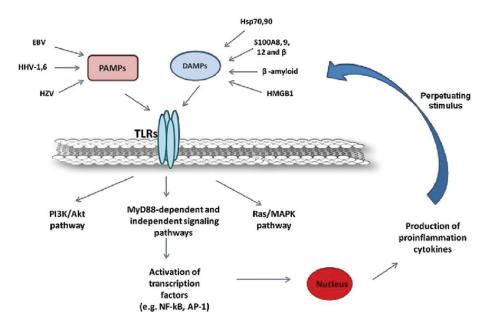


Figure 2. PAMPs/DAMPs complex promotes inflammation on multiple sclerosis. The family of herpes virus such as Herpes Simplex 1 and 6, Epstein-Bar, and Varicela Zoster are considered the best candidates in the MS pathogenesis. These pathogens could represent a source of PAMPs mediating the progression of the disease. The presence of diverse PAMPs or DAMPs during the progression of multiple sclerosis could lead to a continuous inflammation, through their recognition by TLRs. In MS have been described some DAMPs such as HMGB1; S100A8, 9, 12 and β ; HSP70, HSP90 and β -amyloid.

markers during diverse courses of MS however their relationship with virus is still unknown (93-95). The β -AP is generated by proteolytic cleavage of its precursor beta amyloid precursor protein (β -APP), and it is frequently found in brain active plaques. β-APP is a multifunctional protein which expression is induced in several nervous cells in response the acute injury (96,97). The overexpression of β -APP is promoted by S100B proteins which are members of calgranulins proteins. These S100 proteins are prominent molecular mediators in several diseases, including microbial infections, degenerative and autoimmune disease and cancer. (98). S-100 β protein is highly expressed during brain injury, ischemia, neurodegenerative, inflammatory, and psychiatric disorders and it has been implicated in driving the progression of MS (99). In cerebrospinal fluid of patient with MS during relapse disease high levels of S-100B have been found (93). Furthermore, S100 proteins inhibit kinases activity, regulate the enzymes associated with energetic metabolism, stimulate Ca2+ release induced by calcium in sarcoplasmic reticulum membrane, and contribute to the stability of cytoskeleton constituents (100).

S100A9 is another member of S100 proteins and it is implicated in leukocyte migration and chemotaxis, leukocyte activation, and it has potent anti-oxidant activity (101,102). S100A9 is a consistent marker of inflammation and a pro-inflammatory molecule during the primary immune response. Increased plasma levels of S100A9 are related with diseases that involve inflammatory processes such as cystic fibrosis, chronic bronchitis, and RA (103,104). Recent reports have suggested that S100A9 acts as an extra antimicrobial peptide during the primary immune response, by limiting microbial propagation and by avoiding tissue inflammation (105,106). In addition, S100A9 acts as a critical host-derived molecular pattern to regulate inflammatory response outcome and disease during infection by exacerbating the pro- inflammatory cell-death. and viral pathogenesis. response. Extracellular S100A9 regulates two key mechanisms that contribute to inflammation during Influenza A Virus (IAV) infection. These processes are the proinflammatory cytokine releasing during early infection, and the other mechanism is the induction of apoptosis, being an independent T response of virus replication. In addition, S100A9 alone can directly activate TLR4/ MyD88 pathway (in the absence of LPS) and it contributes to the regulation of inflammatory process during IAV infection (28). The S100A9 gene is activated by the DDX21-TRIF, which regulates inflammation trough DDX21-TRIF-S100A9-TLR4-MyD88 (28).

Both S100A9 and S100A8 have been implicated in activation of TLR4 pathway during LPS stimulation (107,108). These myeloid-related proteins are constitutively expressed in neutrophils (109), and they are released at the sites of injury. By binding to RAGE and TLR4, these receptors trigger a proinflammatory response (110). This effect has been documented during the alveolar inflammatory response after endotoxemia and could be implicated in MS (111) (Figure 2).

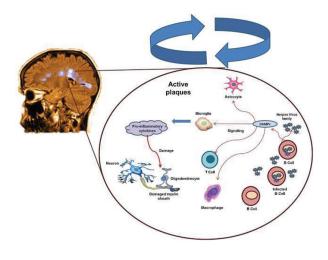


Figure 3. Perpetual generation of active plaques by PAMP-DAMP's in MS. Once the Herpes virus family reaches the CNS tissue, B cells internalize them and the viral replicative cycle begins. Infected cells release new viral particles (PAMP's) and DAMP's which in turn signal through microglia, macrophages, astrocytes, and T-cells. These cells generate pro-inflammatory cytokines which eventually can disturb the protective function of oligodendrocytes leading to the damage of myelin sheets on neurons, named active plaques in MS.

Finally, the detection of oligoclonal bands in the cerebrospinal fluid (CSF) of MS patients, the presence of B cells and plasma cells in MS plaques, and the occurrence in MS lesions of ubstantial depositions of antibodies and complement (112,113) suggest that B cells and antibodies are crucial in the progression and pathogenesis of MS.

B lymphocyte activating factor of the tumor necrosis factor superfamily (BAFF) is a fundamental cytokine for B cell homeostasis that can play a dual role in immunity, by regulating both innate and adaptive immune responses and by sustaining autoimmunity. BAFF is highly expressed in spleen and lymph nodes and it is mainly produced by macrophages, monocytes and dendritic cells. Also, it is produced by astrocytes in CNS (114). In several autoimmune diseases, including MS, high levels of BAFF have been detected in serum (115-117). Overexpression of BAFF is associated with the pathogenesis of MS when EBV infection is involved. It has been shown that BAFF overexpression leads to an expansion of the B-cell compartment and autoimmunity in mice (118) Also, high amounts of BAFF remain elevated in mice that developed relapsingremitting and chronic- relapsing EAE (119).

In brain samples of MS harboring large EBV deposits revealed that most of the B cells in white matter lesions, meninges, and ectopic B-cell follicles are CD27+ and co-express latent membrane protein 1, latent membrane protein 2A and 2 EBV-encoded proteins that provide survival and maturation signals to B cells (120). In MS plaques, BAFF expression has

been found that it is up-regulated at levels comparable to those detected in lymphatic tissues (121). Similarly, mRNA levels of BAFF were increased in monocytes, and in B and T cells of MS patients (117). However, MS patients and headache controls had the same levels of BAFF protein in CSF and plasma (117,122). These findings support the idea that the up-regulated BAFF expression in autoimmune diseases, such as EAE and MS is able to contributing to CNS tissue damage, due to its ability of sustain the survival of auto reactive B cells (119).

During a viral infection several intracellular functions are affected. This damage could be related with the viral prevalence in patients and the development of brain damage. For example, when HIV-1 infects cells, Tat protein induce an increase in the 1-amyloid generation, promotes its accumulation in endolysosomes, enhances the enlargement of these organelles, elevates their internal pH, and increases the activity of the enzyme BACE-1, which converts the precursor of β -amyloid protein to its active form (123). It is known that neurons are cells which possess elaborated endolysosomes and by disturbing the generation of β - amyloid and its precursor, it is possible to alter the endocytic pathway followed for the intracellular traffic of this protein with its consequent accumulation. This process has been related whit cognitive disorders seen in Alzheimer's disease, associated with patients infected with HIV and also it could be present during the generation of active plaques in MS (93,123,124). Particularly, when EBV particles are internalized by B-cells, the activity of diverse proteins such as EBV-determined nuclear antigen (EBNA2), EBER, BZLF-1 and latent membrane- proteins (LMP) play a key role not only during viral replication, but also in the progression of the disease (125). After infection, EBNA2 is released and it is followed by others EBNAs. LMP and EBERs. The expression of EBER is related with the viral life cycle and it plays an important role in antiviral innate immunity. EBER constitutively activates RIG1, leading to the activation of downstream molecules such as NF_K beta and IRF-3, which induce type-1 interferon and IL-10. Furthermore, EBERs is released by EBV infected cells as complex with La, which promotes the release of type-1 IFN and inflammatory cytokines, thus leading to subsequent immune activation by TLR-3 (126-129). Additionally, it has been described that β -amyloid is also accumulated in the nuclei of human brain endothelial cells, which are the main components of the BBB. The breakage of this barrier has been involved in the infiltration of mononuclear cells to the CNS, which promotes inflammatory processes (9). LMP1 is localized in lipid micro-domains and designated lipid rafts. located on the plasma membrane. LMP1 has no intrinsic enzymatic activity but instead aggregates cellular proteins of the tumor necrosis factor receptor signaling pathway to activate transcription factor NF- kB. These proteins not only contributes to replication and immortalization of B-Cell, but also they are able to drive immune response through diverse pathways. In several pathologies associated with viruses, it has been described the role of viral proteins in the generation of DAMPs, being reported for HIV, EBV and HHV infections (69,71,123). In patients with VZV have been detected some proteins considered as markers for neuronal damage, such as S-100 β protein, neurofilament protein (NFL) and glial fibrillary acidic proteins (GFAp) in cerebrospinal fluid (130). Particularly, S100B has also been detected in MS patients (93). Therefore, it is possible that these proteins could be generated as the result of viral infections in CNS promoting a continuous inflammatory process and inducing a permanent release of several DAMP's such as β -amyloid and S-100B between others (Figure 3).

5. CONCLUSIONS

All mammalian cells are supplied with a remarkable numbers of strategies for protection from several cell intruders, which in turn possess a huge diversity of PAMPs for the proper establishment of the infection. Once these cellular alarms identify foreign antigens composed by PAMPs, several biological reactions, such as inflammation, are elicited in order to eradicate pathogens. The "danger theory" proposed by Matzinger (131) explains how the immune response could produce an inflammatory reaction thought DAMPs in response to exogenous and endogenous pathogens, especially following injury or cellular death (110,132). When a virus infection is initiated, the innate immunity is activated and an antiviral and inflammatory response occurs, triggering an incessant release of DAMP's as consequence of chronic infection or continuous injuries as it happens in MS. Unspecific identification of DAMPs by receptors involved in the recognizing of PAMPs, promotes the incoming and releasing of DAMPs in specific areas, which lead to an acute inflammation and continuous delivering of DAMPs; which in turn becomes a repetitive cycle that could elicit the progress of autoimmunity (133). We suggest that in MS, an initial stimulus produces scarce concentrations of DAMPs in tissue, which participate in an immune response to repair injured tissues. However, the relapsing stage of this disease is characterized by an extremely destructive tissue environment, where increased levels of DAMPs are produced during acute inflammation, leading to a chain reaction of damage. Increased rates of proinflammatory DAMPs could promote an extensive tissue injury, amplifying significantly the DAMPs levels in tissue in a local and/or systemical manner, which finally could create a perpetual state of tissue injury. Recently it has been proposed that suppressing DAMP's to avoid the activation of TLRs could be considered as a new potential target in the treatment of autoimmune diseases such as MS, which may offer feasible alternatives to improve current therapies. Recent studies suggest that the inhibition of these molecules might minimize the

symptoms related to the disease. Other strategies could be the blockage of DAMP's necessary for the activation of TLRs, in order to inhibit specific co-receptors or accessory molecules essentials for the activation of this pathway, however, more approaches are needed to give us insight into the MS development.

6. ACKNOWLEDGMENTS

This work was supported by the National Council of Science and Technology of Mexico (CONACyT CB-180851) from Doctor Benjamin Pineda and by CONACyT FOSSIS-182362 from Doctor Sergio Moreno. All authors contributed as the same manner in the design and discussion. All authors have read and approved the manuscript and concur with the submission.

7. REFERENCES

- O Stuve, SS Zamvil: Neurologic diseases. In: Medical Immunology. Parslow Stites D, Terr Almboden J. Eds: Lange Medical books McGraw-Hill Medical Publishing Division, New York, NY, USA, (2001)
- A Sadovnick, PA Baird: Sex ratio in offspring of patients with multiple sclerosis. N Engl J Med, 306, 1114-5 (1982)
- JH Noseworthy, C Lucchinetti, M Rodriguez, BG Weinshenker: Multiple sclerosis. N Engl J Med, 343 (13), 938-52 (2000) DOI: 10.1056/NEJM200009283431307 PMid:11006371
- R Dutta, BD Trapp: Pathogenesis of axonal and neuronal damage in multiple sclerosis. *Neurology*, 68, S22-31; discussion S43-54 (2007)
- H Lassmann, W Bruck, CF Lucchinetti: The immunopathology of multiple sclerosis: an overview. *Brain Pathol*, 17, 210-8 (2007) DOI: 10.1111/j.1750-3639.2007.00064.x PMid:17388952
- 6. J W Peterson, BD Trapp: Neuropathobiology of multiple sclerosis. *Neurol Clin*, 23, 107-29, vi-vii (2005)
- G C DeLuca, SV Ramagopalan, MZ Cader, DA Dyment, BM Herrera, S Orton, A Degenhardt, M Pugliatti, AD Sadovnick, S Sotgiu, GC Ebers: The role of hereditary spastic paraplegia related genes in multiple sclerosis. A study of disease susceptibility and clinical outcome. *J Neurol*, 254, 1221-6 (2007) DOI: 10.1007/s00415-006-0505-4 PMid:17420921

- MC Levin, S Lee, LA Gardner, Y Shin, JN Douglas *et al.*: Pathogenic mechanisms of neurodegeneration based on the phenotypic expression of progressive forms of immunemediated neurologic disease. *Degener Neurol Neuromuscul Dis* 2, 175-187 (2012) DOI: 10.2147/DNND.S38353
- 9. H Lassmann: Acute disseminated encephalomyelitis and multiple sclerosis. *Brain*, 133, 317-9 (2010).
- EM Frohman, MK Racke, CS Raine: Multiple sclerosis--the plaque and its pathogenesis. *N Engl J Med*, 354, 942-55 (2006) DOI: 10.1056/NEJMra052130 PMid:16510748
- A Bar-Or: Immunology of multiple sclerosis. *Neurol Clin*, 23, 149-75, vii (2005) DOI: 10.1016/j.ncl.2004.11.001 PMid:15661092
- YJ Morales, E Parisi, CF Lucchinetti: The pathology of multiple sclerosis: evidence for heterogeneity. *Adv Neurol*, 98, 27-45 (2006). PMid:16400825
- SJ Pittock, CF Lucchinetti: The pathology of MS: new insights and potential clinical applications. *Neurologist*, 13, 45-56 (2007) DOI: 10.1097/01.nrl.0000253065.31662.37 PMid:17351524
- GP Owens, D Gilden, MP Burgoon, X Yu, JL Bennett: Viruses and multiple sclerosis. *Neuroscientist*, 17, 659-76 (2011) DOI: 10.1177/1073858411386615 PMCid: PMC3293404
- DC Poskanzer, J Sever, L Sheridan, LB Prenney: Multiple sclerosis in the Orkney and Shetland Islands. IV: Viral antibody titres and viral infections. *J Epidemiol Community Health*, 34, 258-64 (1980) DOI: 10.1136/jech.34.4.258 DOI:10.1136/jech.34.4.265 PMid:7241024 PMCid: PMC1052088
- G Ordonez, B Pineda, R Garcia-Navarrete, J Sotelo: Brief presence of varicella-zoster vral DNA in mononuclear cells during relapses of multiple sclerosis. *Arch Neurol*, 61, 529-32 (2004) DOI: 10.1001/archneur.61.4.529 PMid:15096401
- 17. R Mancuso, S Delbue, E Borghi, E Pagani, MG Calvo, D Caputo, E Granieri, P Ferrante: Increased prevalence of varicella zoster virus

DNA in cerebrospinal fluid from patients with multiple sclerosis. *J Med Virol*, 79, 192-9 (2007) DOI: 10.1002/jmv.20777 PMid:17177306

- J Sotelo, G Ordonez, B Pineda: Varicellazoster virus at relapses of multiple sclerosis. *J Neurol*, 254, 493-500 (2007) DOI: 10.1007/s00415-006-0402-x PMid:17401519
- J Sotelo, A Martinez-Palomo, G Ordonez, B Pineda: Varicella-zoster virus in cerebrospinal fluid at relapses of multiple sclerosis. *Ann Neurol*, 63, 303-11(2008) DOI: 10.1002/ana.21316 PMid:18306233
- SV Ramagopalan, W Valdar, DA Dyment, GC DeLuca, IM Yee, G Giovannoni, GC Ebers, AD Sadovnick: Association of infectious mononucleosis with multiple sclerosis. Apopulation-based study. *Neuroepidemiology*, 32, 257-62 (2009) DOI: 10.1159/000201564 PMid:19209005
- C Ahlgren, K Toren, A Oden, O Andersen: A population-based case-control study on viral infections and vaccinations and subsequent multiple sclerosis risk. *Eur J Epidemiol*, 24, 541-52 (2009) DOI: 10.1007/s10654-009-9367-2 PMid:19633994
- 22. KL Munger, LI Levin, EJ O'Reilly, KI Falk, A Ascherio: Anti-Epstein-Barr virus antibodies as serological markers of multiple sclerosis: a prospective study among United States military personnel. *Mult Scler*, 17, 1185-93 (2011) DOI: 10.1177/1352458511408991 PMid:21685232 PMCid: PMC3179777
- J Pakpoor, G Disanto, JE Gerber, R Dobson, UC Meier, G Giovannoni,SV Ramagopalan: The risk of developing multiple sclerosis in individuals seronegative for Epstein-Barr virus: a meta-analysis. *Mult Scler*, 19, 162-6 (2013) DOI: 10.1177/1352458512449682 PMid:22740437
- NY Hernandez-Pedro, G Espinosa-Ramirez, VP de la Cruz, B Pineda, J Sotelo: Initial immunopathogenesis of multiple sclerosis: innate immune response. *Clin Dev Immunol*, 2013, 413465 (2013).
 PMid:24174969 PMCid: PMC3794540
- 25. M Sospedra, R Martin: Immunology of

multiple sclerosis. *Annu Rev Immunol*, 23, 683-747 (2005) DOI: 10.1146/annurev.immunol.23.021704. 115707 PMid:15771584

- HL Weiner: A shift from adaptive to innate immunity: a potential mechanism of disease progression in multiple sclerosis. *J Neurol*, 255 Suppl 1, 3-11 (2008) DOI: 10.1007/s00415-008-1002-8 PMid:18317671
- 27. R Gandhi, ALaroni, HL Weiner: Role of the innate immune system in the pathogenesis of multiple sclerosis. *J Neuroimmunol*, 221, 7-14 (2010) DOI: 10.1016/j.jneuroim.2009.10.015 PMid:19931190 PMCid: PMC285418928.
- SY Tsai, JA Segovia, TH Chang, IR Morris, MT Berton, PA Tessier, MR Tardif, A Cesaro,S Bose: DAMP molecule S100A9 acts as a molecular pattern to enhance inflammation during influenza A virus infection: role of DDX21-TRIF-TLR4-MyD88 pathway. *PLoS Pathog*, 10, e1003848 (2014) DOI: 10.1371/journal.ppat.1003848 PMid:24391503 PMCid: PMC3879357
- M Marta, A Andersson, M Isaksson, O Kampe, A Lobell: Unexpected regulatory roles of TLR4 and TLR9 in experimental autoimmune encephalomyelitis. *Eur J Immunol*, 38, 565-75 (2008) DOI: 10.1002/eji.200737187 PMid:18203139
- ZY Zhang, Z Zhang, HJ Schluesener: Toll-like receptor-2, CD14 and heat-shock protein 70 in inflammatory lesions of rat experimental autoimmune neuritis. *Neuroscience*, 159, 136-42 (2009) DOI: 10.1016/j.neuroscience.2008.12.034 PMid:19162137
- JM Reynolds, GJ Martinez, Y Chung, C Dong: Toll-like receptor 4 signaling in T cells promotes autoimmune inflammation. *Proc Natl Acad Sci U S A*, 109, 13064-9 (2012) DOI: 10.1073/pnas.1120585109 PMid:22826216 PMCid: PMC3420161
- M. Dorner, S Brandt, M Tinguely, F Zucol, JP Bourquin, L Zauner, C Berger, M Bernasconi, RF Speck, D Nadal: Plasma cell toll-like receptor (TLR) expression differs from that of B cells, and plasma cell TLR triggering enhances immunoglobulin production. *Immunology*, 128, 573-9 (2009)

DOI: 10.1111/j.1365-2567.2009.03143.x PMid:19950420 PMCid: PMC2792141

- D Jarrossay, G Napolitani, M Colonna, F Sallusto, A Lanzavecchia: Specialization and complementarity in microbial molecule recognition by human myeloid and plasmacytoid dendritic cells. *Eur J Immunol*, 31, 3388-93 (2001) DOI: 10.1002/1521-4141(200111)31:11 <3388:AID-IMMU3388>3.0.CO;2-Q
- 34. SB Rasmussen, LS Reinert, SR Paludan: Innate recognition of intracellular pathogens: detection and activation of the first line of defense. *APMIS*, 117, 323-37 (2009) DOI: 10.1111/j.1600-0463.2009.02456.x PMid:19400860
- 35. AM Piccinini, KS Midwood: DAMPening inflammation by modulating TLR signalling. *Mediators Inflamm*, 2010 (2010)
- 36. M Escamilla-Tilch, G Filio-Rodriguez, R Garcia-Rocha, I Mancilla-Herrera, NA Mitchison, JA Ruiz-Pacheco, FJ Sanchez-Garcia, D Sandoval-Borrego, EA Vazquez-Sanchez: The interplay between pathogenassociated and danger-associated molecular patterns: an inflammatory code in cancer? *Immunol Cell Biol*, 91, 601-10 (2013) DOI: 10.1038/icb.2013.58 PMid:24100386
- A. Marshak-Rothstein: Toll-like receptors in systemic autoimmune disease. Nat Rev Immunol, 6, 823-35 (2006) DOI: 10.1038/nri1957 PMid:17063184
- AD Garg, D Nowis, J Golab, P Vandenabeele, DV Krysko, P. Agostinis: Immunogenic cell death, DAMPs and anticancer therapeutics: an emerging amalgamation. *Biochim Biophys Acta*, 1805, 53-71(2010) DOI: 10.1016/j.bbcan.2009.08.003
- 39. V Petrilli, C Dostert, DA Muruve, J Tschopp: The inflammasome: a danger sensing complex triggering innate immunity. *Curr Opin Immunol*, 19, 615-22 (2007).
 DOI: 10.1016/j.coi.2007.09.002
 PMid:17977705
- S Chakraborty, DK Kaushik, M Gupta, A Basu: Inflammasome signaling at the heart of central nervous system pathology. *J Neurosci Res*, 88, 1615-31 (2010) DOI: 10.1002/jnr.22343

- DG Millar, KM Garza, B Odermatt, AR Elford, N Ono, Z Li, PS Ohashi: Hsp70 promotes antigen-presenting cell function and converts T-cell tolerance to autoimmunity *in vivo*. *Nat Med*, 9, 1469-76 (2003) DOI: 10.1038/nm962 PMid:14625545
- 42. S Yokota, S Minota, N Fujii: Anti-HSP autoantibodies enhance HSP-induced proinflammatory cytokine production in human monocytic cells via Toll-like receptors. *Int Immunol*, 18, 573-80 (2006) DOI: 10.1093/intimm/dxh399 PMid:16481340
- HS Hreggvidsdottir, T Ostberg, H Wahamaa, H Schierbeck, AC Aveberger, L Klevenvall, K Palmblad, L Ottosson, U Andersson, and HE Harris: The alarmin HMGB1 acts in synergy with endogenous and exogenous danger signals to promote inflammation. *J Leukoc Biol*, 86, 655-62 (2009) DOI: 10.1189/jlb.0908548 PMid:19564572
- 44. SK Drexler, BM Foxwell: The role of tolllike receptors in chronic inflammation. *Int J Biochem Cell Biol*, 42, 506-18 (2010) DOI: 10.1016/j.biocel.2009.10.009 PMid:19837184
- 45. DS Wheeler, MA Chase, AP Senft, SE Poynter, HR Wong, K Page: Extracellular Hsp72, an endogenous DAMP, is released by virally infected airway epithelial cells and activates neutrophils via Toll-like receptor (TLR)-4. *Respir Res*, 10, 31 (2009) DOI: 10.1186/1465-9921-10-31 PMid:19405961 PMCid: PMC2679007
- KJ Ishii, K Suzuki, C Coban, F Takeshita, Y Itoh, H Matoba, LD Kohn, DM Klinman: Genomic DNA released by dying cells induces the maturation of APCs. *J Immunol*, 167, 2602-7 (2001) DOI: 10.4049/jimmunol.167.5.2602 PMid:11509601
- 47. K Tani, WJ Murphy, O Chertov, R Salcedo, CY Koh, I Utsunomiya, S Funakoshi, O Asai, SH Herrmann, JM Wang, LW Kwak, JJ Oppenheim: Defensins act as potent adjuvants that promote cellular and humoral immune responses in mice to a lymphoma idiotype and carrier antigens. *Int Immunol*, 12, 691-700 (2000) DOI: 10.1093/intimm/12.5.691 PMid:10784615

- SA Hwang, ML Kruzel, JK Actor: Lactoferrin augments BCG vaccine efficacy to generate T helper response and subsequent protection against challenge with virulent Mycobacterium tuberculosis. *Int Immunopharmacol*, 5, 591-9 (2005)
- 49. KA Scheibner, MA Lutz, S Boodoo, MJ Fenton, JD Powell, MR Horton: Hyaluronan fragments act as an endogenous danger signal by engaging TLR2. *J Immunol*, 177, 1272-81 (2006) DOI: 10.4049/jimmunol.177.2.1272 PMid:16818787
- S Akira, S Sato: Toll-like receptors and their signaling mechanisms. *Scand J Infect Dis*, 35, 555-62 (2003)
 DOI: 10.1080/00365540310015683
 PMid:14620134
- 51. M Galdiero, E Finamore, F Rossano, M Gambuzza, MR Catania, G Teti, A Midiri, G Mancuso: Haemophilus influenzae porin induces Toll-like receptor 2-mediated cytokine production in human monocytes and mouse macrophages. *Infect Immun*, 72, 1204-9 (2004) DOI: 10.1128/IAI.72.2.1204-1209.2004 PMid:14742577 PMCid: PMC321594
- JH Youn, YJ Oh, ES Kim, JE Choi, JS Shin: High mobility group box 1 protein binding to lipopolysaccharide facilitates transfer of lipopolysaccharide to CD14 and enhances lipopolysaccharide-mediated TNF-alpha production in human monocytes. *J Immunol*, 180, 5067-74 (2008) DOI: 10.4049/jimmunol.180.7.5067 PMid:18354232
- 53. SR Saludan, AG Bowie, KA Horan, KA Fitzgerald: Recognition of herpesviruses by the innate immune system. *Nat Rev Immunol*, 11, 143-54 (2011) DOI: 10.1038/nri2937 PMid:21267015 PMCid: PMC3686362 54
- T Bergstrom, O Andersen, A Vahlne: Isolation of herpes simplex virus type 1 during first attack of multiple sclerosis. *Ann Neurol*, 26, 283-5 (1989) DOI: 10.1002/ana.410260218 PMid:2549851
- 55. R Berti, SS Soldan, N Akhyani, HF McFarland, S Jacobson: Extended observations on the association of HHV-6 and multiple sclerosis. *J Neurovirol*, 6 Suppl 2, S85-7 (2000)
- 56. R Berti, MB Brennan, SS Soldan, JM Ohayon,

L Casareto, HF McFarland, S Jacobson: Increased detection of serum HHV-6 DNA sequences during multiple sclerosis (MS) exacerbations and correlation with parameters of MS disease progression. *J Neurovirol*, 8, 250-6 (2002) DOI: 10.1080/13550280290049615-1 PMid:12053279

- 57. M Cirone, L Cuomo, C Zompetta, S Ruggieri, L Frati, A Faggioni, G Ragona: Human herpesvirus 6 and multiple sclerosis: a study of T cell cross-reactivity to viral and myelin basic protein antigens. *J Med Virol*, 68, 268-72 (2002) DOI: 10.1002/jmv.10190 PMid:12210418
- 58. R Alvarez-Lafuente, V De las Heras, M Bartolome, JJ Picazo, R. Arroyo: Relapsingremitting multiple sclerosis and human herpesvirus 6 active infection. *Arch Neurol*, 61, 1523-7(2004) DOI: 10.1001/archneur.61.10.1523 PMid:15477505
- 59. K Voumvourakis, KI, DK Kitsos, S Tsiodras, G Petrikkos, E Stamboulis: Human herpesvirus 6 infection as a trigger of multiple sclerosis. *Mayo Clin Proc*, 85, 1023-30 (2010) DOI: 10.4065/mcp.2010.0350 PMid:20926836 PMCid: PMC2966366
- 60. ABehzad-Behbahani, MH Mikaeili, M Entezam, A Mojiri, GY Pour, MM Arasteh, M Rahsaz, M Banihashemi, B Khadang, A Moaddeb, Z Nematollahi, N Azarpira: Human herpesvirus-6 viral load and antibody titer in serum samples of patients with multiple sclerosis. *J Microbiol Immunol Infect*, 44, 247-51 (2011 DOI: 10.1016/j.jmii.2010.08.002 PMid:21524958
- S Vento, L Guella, F Mirandola, F Cainelli, G Di Perri, M Solbiati, T Ferraro, E Concia: Epstein-Barr virus as a trigger for autoimmune hepatitis in susceptible individuals. *Lancet*, 346, 608-9 (1995) DOI: 10.1016/S0140-6736(95)91438-2
- 62. J Vrbikova, I Janatkova, V Zamrazil, F Tomiska, T Fucikova: Epstein-Barr virus serology in patients with autoimmune thyroiditis. *Exp Clin Endocrinol Diabetes*, 104, 89-92 (1996) DOI: 10.1055/s-0029-1211428 PMid:8750577
- 63. EY Padalko, X Bossuyt: Anti-dsDNA antibodies associated with acute EBV

infection in Sjogren's syndrome. *Ann Rheum Dis*, 60, 992 (2001) DOI: 10.1136/ard.60.10.992 PMid:11589186 PMCid: PMC1753389

- 64. MP Pender: Infection of autoreactive B lymphocytes with EBV, causing chronic autoimmune diseases. *Trends Immunol*, 24, 584-8 (2003) DOI: 10.1016/j.it.2003.09.005 PMid:14596882
- 65. A Ascherio, M Rubertone, D Spiegelman, L Levin, K Munger, C Peck, E Lennette: Notice of retraction: "Multiple sclerosis and Epstein-Barr virus". (JAMA. 2003;289:1533-1536). *JAMA*, 293, 2466 (2005)
- 66. KH Costenbader, EW Karlson: Epstein-Barr virus and rheumatoid arthritis: is there a link? *Arthritis Res Ther*, 8, 204 (2006) DOI: 10.1186/ar1893 PMid:16542469 PMCid: PMC1526553
- S Haahr, P Hollsberg: Multiple sclerosis is linked to Epstein-Barr virus infection. *Rev Med Virol*, 16, 297-310 (2006) DOI: 10.1002/rmv.503 PMid:16927411
- M Trimeche, S Ziadi, K Amara, M Khelifa, F Bahri, S Mestiri, H Braham, M Hachana, B Sriha, M Mokni, S Korbi: Prevalence of Epstein-Barr virus in Sjogren's syndrome in Tunisia. *Rev Med Interne*, 27, 519-23 (2006) DOI: 10.1016/j.revmed.2006.03.034 PMid:16806593
- 69. FC Purves, WO Ogle, B Roizman: Processing of the herpes simplex virus regulatory protein alpha 22 mediated by the UL13 protein kinase determines the accumulation of a subset of alpha and gamma mRNAs and proteins in infected cells. *Proc Natl Acad Sci U S A*, 90, 6701-5 (1993) DOI: 10.1073/pnas.90.14.6701 PMid:8393574 PMCid: PMC47000
- 70. E Gershburg, S Raffa, MR Torrisi, JS Pagano: Epstein-Barr virus-encoded protein kinase (BGLF4) is involved in production of infectious virus. *J Virol*, 81, 5407-12 (2007) DOI: 10.1128/JVI.02398-06 PMid:17360761 PMCid: PMC1900237
- 71. X Sun, JA Bristol, S Iwahori, SR Hagemeier, Q Meng, EA Barlow, JD Fingeroth, VL Tarakanova, RF Kalejta, SC Kenney: Hsp90 inhibitor 17-DMAG decreases expression of

conserved herpesvirus protein kinases and reduces virus production in Epstein-Barr virusinfected cells. *J Virol*, 87, 10126-38 (2013) DOI: 10.1128/JVI.01671-13 PMid:23843639 PMCid: PMC3754017

- 72. M Kotsiopriftis, JE Tanner, C Alfieri: Heat shock protein 90 expression in Epstein-Barr virusinfected B cells promotes gammadelta T-cell proliferation *in vitro. J Virol*, 79, 7255-61 (2005) DOI: 10.1128/JVI.79.11.7255-7261.2005 PMid:15890964 PMCid: PMC1112101
- 73. M Ejima, K Ota, H Tanaka, S Maruyama: Peripheral blood gamma delta T cells in multiple sclerosis. *Rinsho Shinkeigaku*, 33, 1131-4 (1993) PMid:8124869
- 74. MJ Mansilla, X Montalban, C Espejo: Heat shock protein 70: roles in multiple sclerosis. *Mol Med*, 18, 1018-28 (2012) DOI: 10.2119/molmed.2012.00119 PMid:22669475 PMCid: PMC3459486
- 75. H Cwiklinska, MP Mycko, O Luvsannorov, B Walkowiak, CF Brosnan, CS Raine, KW Selmaj: Heat shock protein 70 associations with myelin basic protein and proteolipid protein in multiple sclerosis brains. Int Immunol, 15, 241-9 (2003) DOI: 10.1093/intimm/dxg022 PMid:12578854
- SD D'Souza, JP Antel, MS Freedman: Cytokine induction of heat shock protein expression in human oligodendrocytes: an interleukin-1mediated mechanism. *J Neuroimmunol*, 50, 17-24 (1994) DOI: 10.1016/0165-5728(94)90210-0
- 77. YW Han, JY Choi, E Uyangaa, SB Kim, JH Kim, BS Kim, K Kim, SK Eo: Distinct dictation of Japanese encephalitis virus-induced neuroinflammation and lethality via triggering TLR3 and TLR4 signal pathways. *PLoS Pathog*, 10, e1004319 (2014) DOI: 10.1371/journal.ppat.1004319 PMid:25188232 PMCid: PMC4154777
- 78. H Wang, O Bloom, M Zhang, JM Vishnubhakat, M Ombrellino, J Che, A Frazier, H Yang, S Ivanova, L Borovikova, KR Manogue, E Faist, E Abraham, J Andersson, U Andersson, PE Molina, NN Abumrad, A Sama, KJ Tracey: HMG-1 as a late mediator of endotoxin lethality in mice. *Science*, 285, 248-51 (1999) DOI: 10.1126/science.285.5425.248 PMid:10398600

- 79. U Andersson, KJ Tracey: HMGB1 is a therapeutic target for sterile inflammation and infection. *Annu Rev Immunol*, 29, 139-62 (2011) DOI:10.1146annurev-immunol-030409-101323 PMid:21219181
- P Scaffidi, T Misteli, ME Bianchi: Release of chromatin protein HMGB1 by necrotic cells triggers inflammation. *Nature*, 418, 191-5 (2002) DOI: 10.1038/nature00858 PMid:12110890
- MJ Ray, GA Hawson: A comparison of two APTT reagents which use silica activators. *Clin Lab Haematol*, 11, 221-32 (1989) DOI: 10.1111/j.1365-2257.1989.tb00212.x PMid:2556232
- P Nowak, B Barqasho, CJ Treutiger, HE Harris, KJ Tracey, J Andersson, A Sonnerborg: HMGB1 activates replication of latent HIV-1 in a monocytic cell-line, but inhibits HIV-1 replication in primary macrophages. *Cytokine*, 34, 17-23 (2006) DOI: 10.1016/j.cyto.2006.03.010 PMid:16697213
- 83. IE Dumitriu, P Baruah, AA Manfredi, ME Bianchi, P Rovere-Querini: HMGB1: An immmune odyssey. *Discov Med*, 5, 388-92 (2005) PMid:20704878
- P Fang, M Schachner, YQ Shen: HMGB1 in development and diseases of the central nervous system. *Mol Neurobiol*, 45, 499-506 (2012) DOI: 10.1007/s12035-012-8264-y PMid:22580958
- 85. A Andersson, R Covacu, D Sunnemark, Al Danilov, ADal Bianco, M Khademi, E Wallstrom, A Lobell, L Brundin, H Lassmann, RA Harris: Pivotal advance: HMGB1 expression in active lesions of human and experimental multiple sclerosis. *J Leukoc Biol*, 84, 1248-55 (2008) DOI: 10.1189/jlb.1207844 PMid:18644848
- 86. O Hori, J Brett, T Slattery, R Cao, J Zhang, JX Chen, M Nagashima, ER Lundh, S Vijay, D Nitecki, *et al.*: The receptor for advanced glycation end products (RAGE) is a cellular binding site for amphoterin. Mediation of neurite outgrowth and co-expression of rage and amphoterin in the developing nervous system. *J Biol Chem*, 270, 25752-61 (1995) DOI: 10.1074/jbc.270.43.25752 PMid:7592757

- 87. AP Robinson, MW Caldis, CT Harp, GE Goings, SD Miller: High-mobility group box 1 protein (HMGB1) neutralization ameliorates experimental autoimmune encephalomyelitis. *J Autoimmun*, 43, 32-43 (2013) DOI: 10.1016/j.jaut.2013.02.005 PMid:23514872 PMCid: PMC3672339
- CS Hwang, GT Liu, MD Chang, IL Liao, HT Chang: Elevated serum autoantibody against high mobility group box 1 as a potent surrogate biomarker for amyotrophic lateral sclerosis. *Neurobiol Dis*, 58, 13-8 (2013) DOI: 10.1016/j.nbd.2013.04.013 PMid:23639787
- S Schenk, J Muser, G Vollmer, R Chiquet-Ehrismann: Tenascin-C in serum: a questionable tumor marker. *Int J Cancer*, 61, 443-9 (1995) DOI: 10.1002/ijc.2910610402 PMid:7538974
- 90. J Tian, AM Avalos, SY Mao, B Chen, K Senthil, H Wu, P Parroche, S Drabic, D Golenbock, C Sirois, J Hua, LL An, L Audoly, G La Rosa, A Bierhaus, P Naworth, A Marshak-Rothstein, MK Crow, KA Fitzgerald, E Latz, PA Kiener, AJ Coyle: Toll-like receptor 9-dependent activation by DNA-containing immune complexes is mediated by HMGB1 and RAGE. *Nat Immunol*, 8, 487-96 (2007) .DOI: 10.1038/ni1457 PMid:17417641
- 91. V Urbonaviciute, BG Furnrohr, S Meister, L Munoz, P Heyder, F De Marchis, ME Bianchi, C Kirschning, H Wagner, AA Manfredi, J. Kalden, G Schett, P Rovere-Querini, M Herrmann, RE Voll: Induction of inflammatory and immune responses by HMGB1-nucleosome complexes: implications for the pathogenesis of SLE. *J Exp Med*, 205, 3007-18 (2008) DOI: 10.1084/jem.20081165 PMid:19064698 PMCid: PMC2605236
- 92. V Urbonaviciute, BG Furnrohr, C Weber, M Haslbeck, S Wilhelm, M Herrmann, RE Voll: Factors masking HMGB1 in human serum and plasma. *J Leukoc Biol*, 81, 67-74 (2007) DOI: 10.1189/jlb.0306196 PMid:17060363
- 93. K Hein Nee Maier, A Kohler, R Diem, MB Sattler, I Demmer, P Lange, M Bahr, M Otto: Biological markers for axonal degeneration in CSF and blood of patients with the first event indicative for multiple sclerosis. *Neurosci Lett*,

436, 72-6 (2008) DOI: 10.1016/j.neulet.2008.02.064 PMid:18359164

- 94. H Tumani, C Teunissen, S Sussmuth, M Otto, AC Ludolph, J Brettschneider: Cerebrospinal fluid biomarkers of neurodegeneration in chronic neurological diseases. *Expert Rev Mol Diagn*, 8, 479-94 (2008) DOI: 10.1586/14737159.8.4.479 PMid:18598229
- 95. K Mitosek-Szewczyk, W Gordon-Krajcer, D Flis, Z Stelmasiak: Some markers of neuronal damage in cerebrospinal fluid of multiple sclerosis patients in relapse. *Folia Neuropathol*, 49, 191-6 (2011) PMid:22101952
- 96. RN Kalaria, SU Bhatti, EA Palatinsky, DH Pennington, ER Shelton, HW Chan, G Perry, WD Lust: Accumulation of the beta amyloid precursor protein at sites of ischemic injury in rat brain. *Neuroreport*, 4, 211-4(1993) DOI: 10.1097/00001756-199302000-00025 PMid:8453062
- J Gehrmann, RB Banati, ML Cuzner, GW Kreutzberg, J Newcombe: Amyloid precursor protein (APP) expression in multiple sclerosis lesions. *Glia*, 15, 141-51 (1995) DOI: 10.1002/glia.440150206 PMid:8567065
- 98. E Leclerc, G Fritz, SW Vetter, CW Heizmann: Binding of S100 proteins to RAGE: an update. *Biochim Biophys Acta*, 1793, 993-1007 (2009) DOI: 10.1016/j.bbamcr.2008.11.016 PMid:19121341
- 99. Rothermundt, M., M. Peters, J. H. Prehn & V. Arolt: S100B in brain damage and neurodegeneration. *Microsc Res Tech*, 60, 614-32 (2003) DOI: 10.1002/jemt.10303 PMid:12645009
- 100. R Donato: Intracellular and extracellular roles of S100 proteins. *Microsc Res Tech*, 60, 540-51 (2003)
 DOI: 10.1002/jemt.10296
 PMid:12645002
- 101. HY Sroussi, J Berline, JM Palefsky: Oxidation of methionine 63 and 83 regulates the effect of S100A9 on the migration of neutrophils *in vitro*. *J Leukoc Biol*, 81, 818-24 (2007) DOI: 10.1189/jlb.0706433 PMid:17138858

- 102. C Perera, HP McNeil, CL Geczy: S100 Calgranulins in inflammatory arthritis. *Immunol Cell Biol*, 88, 41-9 (2010) DOI: 10.1038/icb.2009.88 PMid:19935766
- 103. PL van Lent, L Grevers, AB Blom, A Sloetjes, JS Mort, T Vogl, W Nacken, WB van den Berg, J Roth: Myeloid-related proteins S100A8/ S100A9 regulate joint inflammation and cartilage destruction during antigen-induced arthritis. Ann Rheum Dis, 67, 1750-8 (2008) DOI: 10.1136/ard.2007.077800 PMid:18055478
- 104. EO Lee, JH Yang, KAChang, YH Suh, YH Chong: Amyloid-beta peptide-induced extracellular S100A9 depletion is associated with decrease of antimicrobial peptide activity in human THP-1 monocytes. *J Neuroinflammation*, 10, 68 (2013) DOI: 10.1186/1742-2094-10-68 PMid:23721320 PMCid: PMC3693929
- 105. O Erez, R Romero, AL Tarca, T Chaiworapongsa, YM Kim, NG Than, EVaisbuch, S Draghici, G Tromp: Differential expression pattern of genes encoding for antimicrobial peptides in the fetal membranes of patients with spontaneous preterm labor and intact membranes and those with preterm prelabor rupture of the membranes. *J Matern Fetal Neonatal Med*, 22, 1103-15 (2009) DOI: 10.3109/14767050902994796 PMid:19916708 PMCid: PMC3560925
- 106. Hiroshima, Y., M. Bando, M. Kataoka, Y. Inagaki, M. C. Herzberg, K. F. Ross, K. Hosoi, T. Nagata & J. Kido: Regulation of antimicrobial peptide expression in human gingival keratinocytes by interleukin-1alpha. *Arch Oral Biol*, 56, 761-7 (2011) DOI: 10.1016/j.archoralbio.2011.01.004 PMid:21316034 PMCid: PMC3412402
- 107. T Vogl, K Tenbrock, S Ludwig, N Leukert, C Ehrhardt, MA. van Zoelen, W Nacken, D Foell, T van der Poll, C Sorg, J Roth: Mrp8 and Mrp14 are endogenous activators of Tolllike receptor 4, promoting lethal, endotoxininduced shock. *Nat Med*, 13, 1042-9 (2007) DOI: 10.1038/nm1638 PMid:17767165
- 108. P Sun, Q Li, Q Zhang, L Xu, JY Han: Upregulated expression of S100A8 in mice brain after focal cerebral ischemia reperfusion. *World J Emerg Med*, 4, 210-4 (2013) DOI: 10.5847/wjem.j.issn.1920-8642.2013.03.010 PMid:25215121 PMCid: PMC4129849

- 109. PA Hessian, J Edgeworth, N Hogg: MRP-8 and MRP-14, two abundant Ca(2+)-binding proteins of neutrophils and monocytes. *J Leukoc Biol*, 53, 197-204 (1993) PMid:8445331
- 110. ME Bianchi: DAMPs, PAMPs and alarmins: all we need to know about danger. *J Leukoc Biol*, 81, 1-5 (2007) DOI: 10.1189/jlb.0306164 PMid:17032697
- 111. A Gonzalez-Lopez, A Aguirre, I Lopez-Alonso, L Amado, A Astudillo, MS Fernandez-Garcia, MF Suarez, E Batalla-Solis, E Colado, GM Albaiceta: MMP-8 deficiency increases TLR/RAGE ligands S100A8 and S100A9 and exacerbates lung inflammation during endotoxemia. *PLoS One*, 7, e39940 (2012) DOI: 10.1371/journal.pone.0039940 PMid:22768176 PMCid: PMC3386945
- 112. JJ Archelos, MK Storch, HP Hartung: The role of B cells and autoantibodies in multiple sclerosis. *Ann Neurol*, 47, 694-706 (2000) DOI: 10.1002/1531-8249(200006)47:6<694: AID-ANA2>3.3.CO;2-N DOI: 10.1002/1531-8249(200006)47:6<694: AID-ANA2>3.0.CO;2-W
- 113. C Lucchinetti, W Bruck, J Parisi, B Scheithauer, M Rodriguez, H Lassmann: Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. *Ann Neurol*, 47, 707-17 (2000)
 DOI: 10.1002/1531-8249(200006)47:6<707: AID-ANA3>3.0.CO;2-Q
- 114. F Mackay, P Schneider: Cracking the BAFF code. *Nat Rev Immunol*, 9, 491-502 (2009)
 DOI: 10.1038/nri2572
 PMid:19521398
- 115. GS Cheema, V Roschke, DM Hilbert, W Stohl: Elevated serum B lymphocyte stimulator levels in patients with systemic immune-based rheumatic diseases. *Arthritis Rheum*, 44, 1313-9 (2001) DOI: 10.1002/1529-0131(200106)44:6<1313: AID-ART223>3.0.CO;2-S
- 116. M Thangarajh, A Gomes, T Masterman, J Hillert, P Hjelmstrom: Expression of B-cellactivating factor of the TNF family (BAFF) and its receptors in multiple sclerosis. *J Neuroimmunol*, 152, 183-90 (2004) DOI: 10.1016/j.jneuroim.2004.03.017 PMid:15223251
- 117. A Vaknin-Dembinsky, L Brill, N Orpaz, O

Abramsky, D Karussis: Preferential increase of B-cell activating factor in the cerebrospinal fluid of neuromyelitis optica in a white population. *Mult Scler*, 16, 1453-7 (2010) DOI: 10.1177/1352458510380416 PMid:20935029

- 118. F Mackay, SA Woodcock, P Lawton, C Ambrose, M Baetscher, P Schneider, J Tschopp, JL Browning: Mice transgenic for BAFF develop lymphocytic disorders along with autoimmune manifestations. *J Exp Med*, 190, 1697-710 (1999) DOI: 10.1084/jem.190.11.1697 PMid:10587360 PMCid: PMC2195729
- 119. R Magliozzi, S Columba-Cabezas, B Serafini, F Aloisi: Intracerebral expression of CXCL13 and BAFF is accompanied by formation of lymphoid follicle-like structures in the meninges of mice with relapsing experimental autoimmune encephalomyelitis. *J Neuroimmunol*, 148, 11-23 (2004) DOI: 10.1016/j.jneuroim.2003.10.056 PMid:14975582
- 120. B Serafini, M Severa, S Columba-Cabezas, B Rosicarelli, C Veroni, G Chiappetta, R Magliozzi, R Reynolds, EM Coccia, F Aloisi: Epstein-Barr virus latent infection and BAFF expression in B cells in the multiple sclerosis brain: implications for viral persistence and intrathecal B-cell activation. *J Neuropathol Exp Neurol*, 69, 677-93 (2010) DOI: 10.1097/NEN.0b013e3181e332ec PMid:20535037
- 121. M Krumbholz, D Theil, S Cepok, B Hemmer, P Kivisakk, RM Ransohoff, M Hofbauer, C Farina, T Derfuss, C Hartle, J Newcombe, R Hohlfeld, E Meinl: Chemokines in multiple sclerosis: CXCL12 and CXCL13 up-regulation is differentially linked to CNS immune cell recruitment. *Brain*, 129, 200-11 (2006) PMid:16280350
- 122. F Piazza, JC DiFrancesco, ML Fusco, D Corti, L Pirovano, B Frigeni, L Mattavelli, S Andreoni, M Frigo, C Ferrarese, G Tredici, G Cavaletti: Cerebrospinal fluid levels of BAFF and APRIL in untreated multiple sclerosis. *J Neuroimmunol*, 220, 104-7 (2010) DOI: 10.1016/j.jneuroim.2010.01.011 PMid:20149932
- 123. X Chen, L Hui, NH Geiger, NJ Haughey, JD Geiger: Endolysosome involvement in HIV-1 transactivator protein-induced neuronal amyloid beta production. *Neurobiol Aging*, 34,

2370-8 (2013)

DOI: 10.1016/j.neurobiolaging.2013.04.015 PMid:23673310 PMCid: PMC3706576

- 124. SLDeshmane, RMukerjee, SFan, BE Sawaya: High-performance capillary electrophoresis for determining HIV-1 Tat protein in neurons. *PLoS One*, 6, e16148 (2011)
 DOI: 10.1371/journal.pone.0016148
 PMid:21249135 PMCid: PMC3017553
- 125. LS Young, JR Arrand, PG Murray: EBV gene expression and regulation. Cambridge (2007)
- 126. LS Young, AB Rickinson: Epstein-Barr virus: 40 years on. *Nat Rev Cancer*, 4, 757-68 (2004) DOI: 10.1038/nrc1452 PMid:15510157
- 127. E Freire, C Oddo, L Frappier, G de Prat-Gay: Kinetically driven refolding of the hyperstable EBNA1 origin DNA-binding dimeric betabarrel domain into amyloid-like spherical oligomers. *Proteins*, 70, 450-61 (2008) DOI: 10.1002/prot.21580 PMid:17680697
- 128. DF Angelini, B Serafini, E Piras, M Severa, EM Coccia, B Rosicarelli, S Ruggieri, C Gasperini, F Buttari, D Centonze, R Mechelli, M Salvetti, G Borsellino, F Aloisi, L Battistini: Increased CD8+ T cell response to Epstein-Barr virus lytic antigens in the active phase of multiple sclerosis. *PLoS Pathog*, 9, e1003220 (2013) DOI: 10.1371/journal.ppat.1003220 PMid:23592979 PMCid: PMC3623710
- 129. RM Leskowitz, XY Zhou, F Villinger, MH Fogg, A Kaur, PM Lieberman, F Wang, HC Ertl: CD4+ and CD8+ T-cell responses to latent antigen EBNA-1 and lytic antigen BZLF-1 during persistent lymphocryptovirus infection of rhesus macaques. J Virol, 87, 8351-62 (2013) DOI: 10.1128/JVI.00852-13 PMid:23698300 PMCid: PMC3719796
- 130. A Grahn, L Hagberg, S Nilsson, K Blennow, H Zetterberg, M Studahl: Cerebrospinal fluid biomarkers in patients with varicellazoster virus CNS infections. *J Neurol*, 260, 1813-21 (2013) DOI: 10.1007/s00415-013-6883-5 PMid:23471614
- 131. P Matzinger: Tolerance, danger, and the extended family. *Annu Rev Immunol*, 12,

991-1045 (1994) DOI: 10.1146/annurev.immunol.12.1.991 DOI: 10.1146/annurev.iy.12.040194.005015 PMid:8011301

- 132. Z Zhang, C Wang, L Nie, G Soon: Assessing the heterogeneity of treatment effects via potential outcomes of individual patients. *J R Stat Soc Ser C Appl Stat*, 62, 687-704 (2013). DOI: 10.1111/rssc.12012
- 133. N Jounai, K Kobiyama, F Takeshita, KJ Ishii: Recognition of damage-associated molecular patterns related to nucleic acids during inflammation and vaccination. *Front Cell Infect Microbiol*, 2, 168 (2012) PMid:23316484 PMCid: PMC3539075

Key Words: Pathogen-Associated Molecular Patterns, Damage-Associated Molecular Patterns, Multiple Sclerosis, Immune Response, Inflammation, Review

Send correspondence to: Benjamin Pineda, Neuroimmunology and Neuro-Oncology Unit, Instituto Nacional de Neurologia y Neurocirugia (INNN), Insurgentes sur 3877, 14269, Mexico City, Mexico, Tel: 52-55-56064040, Fax: 52-55-56064040, E-mail: benpio76@hotmail.com