Discovery of new anticancer agents from higher plants

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1. ABSTRACT

Small organic molecules derived from higher plants have been one of the mainstays of cancer chemotherapy for approximately the past half a century. In the present review, selected single chemical entity natural products of plant origin and their semi-synthetic derivatives currently in clinical trials are featured as examples of new cancer chemotherapeutic drug candidates. Several more recently isolated compounds obtained from plants showing promising in vivo biological activity are also discussed in terms of their potential as anticancer agents, with many of these obtained from species that grow in tropical regions. Since extracts of only a relatively small proportion of the ca. 300,000 higher plants on earth have been screened biologically to date, bioactive compounds from plants should play an important role in future anticancer drug discovery efforts.

2. INTRODUCTION

Natural products are generally defined as small-molecule secondary metabolites that originate from marine organisms, microorganisms, and plants. They occur as distinctive chemical types in taxonomically different organisms, for which they serve multiple biological functions related to organism survival. These naturally derived substances exhibit considerable structural diversity. and tend to adopt the preferred conformation and necessary steric complexity to exert varied activities in biological test When compared with synthetic organic systems. compounds as a whole, natural product molecules typically have more chiral centers, less hetero atoms, less heavy atoms, and more varied ring systems. Overall, natural products are generally regarded as possessing "drug-like" pharmacological qualities and "biologically friendly" molecular properties (1-3). These attributes make natural

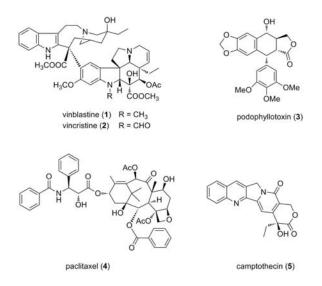


Figure 1. Structures of plant-derived compounds used as cancer chemotherapeutic agents.

products an invaluable resource of chemical diversity and hence they have acted as excellent lead compounds for optimization by synthetic organic chemistry methods in anticancer agent discovery.

Natural products have proved to be very useful in anticancer chemotherapy drug development over last five decades, particularly those derived from terrestrial microbes and higher plants. An important review of the anticancer drugs introduced to the market in North America, western Europe, and Japan since the 1940s has indicated that some 47% of a total of the 155 anticancer drugs approved up to 2006 were either natural products or directly derived from natural product lead compounds by semi-synthesis (4). However, a reduced emphasis on natural products in terms of new drug development has become evident by large pharmaceutical companies during approximately the last 20 years (5-7). One of the major reasons underlying this decline is that the pharmaceutical industry has largely changed its strategy of drug discovery to the rapid high-throughput screening of molecular target-based pure compound chemical libraries, which have been generated particularly using combinatorial chemistry. Also, it is considered too time-consuming and labor-intensive to perform bioassay-guided isolation of natural products from crude extracts. Moreover, there is a tendency when screening natural product samples to rediscover bioactive compounds of already known structure. However, in recent vears, new technologies have been developed to enhance natural product drug discovery in an industrial setting, including streamlined screening procedures and enhanced organism sourcing mechanisms (5-7). These changes actually bode well for the additional inclusion of natural product samples in anticancer drug discovery in the future. Indeed, several new anticancer agents of natural origin have been introduced to the market recently (8), and there is a promising pipeline of natural products in oncology-related clinical trials from various types of organisms (9-11).

The approved plant-derived oncology cancer chemotherapeutic agents used clinically in the United States

may be structurally classified into four major groups: the vinca alkaloids, the epipodophyllotoxin lignans, the taxane diterpenoids, and the camptothecin quinoline alkaloid derivatives (Figure 1). The discoveries of the five antineoplastic lead compounds: vinblastine (vincaleukoblastine, 1) (12, 13), vincristine (leurocristine, 2) (13, 14), podophyllotoxin (3) (15) paclitaxel (taxol, 4) (16), and camptothecin (5) (17), represent significant contributions to natural products isolation accomplished by pioneering natural product chemists, and these have all led subsequently to considerable advances in cancer chemotherapy. These compounds have been found to act on important biochemical targets, tubulin two and topoisomerase. According to our present knowledge, vinca alkaloids and taxanes both target cellular microtubulin, but they lead to cancer cell death through different specific mechanisms. Thus, vinca alkaloids such as vinblastine and vincristine bind to the microtubulin "vinca domain" site in the β -subunit, and disrupt the assembly of microtubules in mitosis (18), while paclitaxel acts as a microtubule stabilizer, by binding to the taxane site, and, as a result, interfering with the normal breakdown of microtubules during cell division (18, 19). Camptothecin and its clinically used analogues, irinotecan and topotecan, arrest the cell cycle at the S-phase by inhibiting the activity of topoisomerase I, leading to the inhibition of DNA replication and transcription (20, 21). Podophyllotoxin binds to tubulin and interferes with the formation of spindles in mitosis (22). In contrast, the two clinically used derivatives of podophyllotoxin. etoposide, and teniposide are topoisomerase II inhibitors, which arrest the cell cycle in the metaphase by stabilizing the covalent DNA-enzyme cleavable complex, and inducing topoisomerase II-mediated DNA breakage (22). Since there is already much published information on the above-mentioned four classes of plant-derived anticancer compounds (e.g., 23-25), these agents will not be further discussed in the present review.

Much of the progress in discovering and developing natural product drugs for the oncology market has been made by pharmaceutical and biotechnology companies. There has also been a long-standing commitment to this same end by the U.S. National Cancer Institute (NCI). Through the use of extramural and intramural funding mechanisms, the NCI has focused on the extensive collection and biological screening of extracts from plants and other organisms for their potential anticancer activity. The NCI has also played an important role in terms of antineoplastic drug development (26-29). During the period 1960 to 2004, some 174,000 plant samples were collected, including many from tropical regions, and these were screened through a systematic combination of in vitro and in vivo screens (25-27). In this process, extracts of small-scale plant samples were subjected to a standard solvent extraction protocol. Initially, the extracts were evaluated against a three cell-line panel, inclusive of MCF-7 breast, NCI-H460 lung, and SF-268 CNS cancer cells, with active samples then tested against a 60-cell line panel derived from leukemia and eight solid tumors including breast cancer, CNS tumor, colon cancer, non-small cell lung cancer, postage cancer, renal cell cancer and ovarian cancer (25-27). Compounds of interest are then

evaluated in the *in vivo* hollow fiber assay, a rapid and economic murine model developed at NCI in order to prioritize candidate compounds for possible further *in vivo* testing. In this assay, human cancer cells are grown within fibers that are implanted at subcutaneous or intraperitoneal sites in immunodeficient mice (30, 31). Further *in vivo* evaluation in murine xenograft systems is conducted for compounds deemed active in the *in vivo* hollow fiber assay (25-27).

In the following paragraphs of this review, a number of selected plant-derived antineoplastic single chemical entities currently under clinical trials as oncology drug candidates, and several promising lead compounds purified from plants, will be discussed in term of their plant origin, modes of action, as well as their potential use as anticancer agents.

3. SELECTED ANTINEOPLASTIC PLANT-DERIVED SINGLE CHEMICAL ENTITIES IN CLINICAL TRIALS

A relatively large number of chemical entities of plant origin are currently in clinical trials (9-11). However, a considerable percentage of the plant-derived oncology drug candidates in clinical trials are based on paclitaxel [ABI-007, DHA-paclitaxel, paclitaxel poliglumex. RPR-116278A, XRP9881 (RPR109881A)], camptothecin [9-aminocamptothecin, exatecan mesylate, oral topotecan (hycamptin). rubitecan (9-nitrocamptothecin, OrathecinTM)], vinblastine and vincristine (vinflunine ditartrate, vinorelbine, anhydrovinblastine, vincristine sulfate TCS), and epipodophyllotoxin (NK-611 and tafluposide 105) (9-11). Members of these already established classes of candidate drugs will not be discussed in this section, with compounds of other structural types referred to instead. In this manner, it is hoped to better emphasize the overall structural diversity of promising chemical entities from plants as potential cancer chemotherapeutic agents (Figure 2).

Betulinic acid (6), a pentacyclic triterpenoid with a lupane skeleton, is widespread in the plant kingdom. This compound was purified as the active principle from Ziziphus mauritiana Lam. (Rhamnaceae), collected in Zimbabwe, using the bioassay-guided fractionation in earlier work by our group when at the University of Illinois at Chicago (32). In this investigation, betulinic acid was found to show in vivo selective growth inhibitory activity in athymic mice bearing human melanoma xenografts (32). This triterpenoid has been shown also to exhibit cytotoxicity against neuroectodermal and brain tumor cells (33). It induces apoptosis through regulation of the intrinsic pathway by changing mitochondrial membrane potential and activation of p38 MAPK and SAP/JNK by initiating ROS (reactive oxygen species) generation (34). This compound can be semi-synthesized by oxidation of betulin (7), a more abundant naturally occurring analogue (35). A betulinic acid-containing ointment is undergoing Phase I/II clinical evaluation for the treatment of dysplastic nevi with moderate to severe dysplasia (36).

Combretastatin A1 (8) and combretastatin A4 (9) are representative of several *cis*-stilbenes from *Combretum*

caffrum Kuntze (Combretaceae), a shrub from South Africa, that were isolated in the 1980s by Pettit and colleagues at Arizona State University (37, 38). Substances in the combretastatin class are antineoplastic agents that also act as tubulin-binding and vascular disrupting agents (VDAs), and cause morphological changes within endothelial cells, which can lead to rapid and selective vascular collapse in solid tumors and result in the shutdown of the nutrient supply for malignant cells (39, 40). Combretastatin A4 phosphate (10, CA4P) is a phosphate prodrug of combretastatin A4 (9), which inhibits the polymerization of tubulin by binding on tubulin at the cochicine-site with highly affinity, and leads to the failure of microtubule formation (41). CA4P (10), both alone, and in combination with conventional oncological agents, is currently in several Phase I/II clinical trials for the treatment of anaplastic thyroid carcinoma and other advanced solid tumors in the United States (42, 43). AVE8062 (AC7700, 11), a propanamide derivative of compound 9, exhibits even an more potent antitumor effect than CA4P by inducing an irreversible blockage of tumor blood flow, and is now in Phase I clinical studies in Europe and the United States (44, 45). CA1P (Oxi4503, 12), a bisphosphate prodrug of combretastatin A1 (12) also reported to be more potent than CA4P (10), is undergoing Phase I anticancer clinical trials in the U.K. (45).

Curcumin (13) is a phenolic diarylheptanoid isolated from turmeric, the roots of Curcuma longa L. (Zingiberaceae). Turmeric is a common spice widely consumed in many Asian countries, and has a long history of use in the traditional medicinal systems of India and China as a remedy for the treatment of ailments such as arthritis, digestive disorders, gallbladder and liver problems, and eye and skin infections (46, 47). The types of biological activities attributed to curcumin cover a wide range, and include antioxidant, anti-inflammatory, antimicrobial, immunomodulatory, and potential cancer chemopreventive effects (46, 47). Curcumin can induce apoptosis and inhibits the proliferation of a wide variety of malignant cells. The mechanisms underlying the activities exhibited by curcumin are complex, and involve the regulation of combined signaling pathways at multiple levels by acting on various targets. These include the modulation of gene transcription factors (NFkB, p53, AP-1), growth factors and their receptors (PDGF, EGF, VEGF), cell surface adhesion molecules (E-cadherin, \beta-cadenin), and protein kinases (CDKs, EGFR, PKC, p38 MAPK) (46, 47). Clinical studies of curcumin alone or in combination with other chemotherapeutic agents have been carried out in the United States and Israel for patients with colorectal and pancreatic cancers (48).

Flavopiridol (alvocidib, 14) is a semisynthetic substance derived from rohitukine (15), an *N*-methylpiperidine alkaloid isolated initially from *Amoora rohituka* (Roxb.) Wight & Arn. (Meliaceae), and then identified as an anti-inflammatory and immunomodulatory agent from the stem bark of a native plant from India, *Dysoxylum binectariferum* Hiern (Meliaceae) (49, 50). Flavopiridol was found to exhibit cytotoxity for a wide range of cancer cell lines and has demonstrated *in vivo*

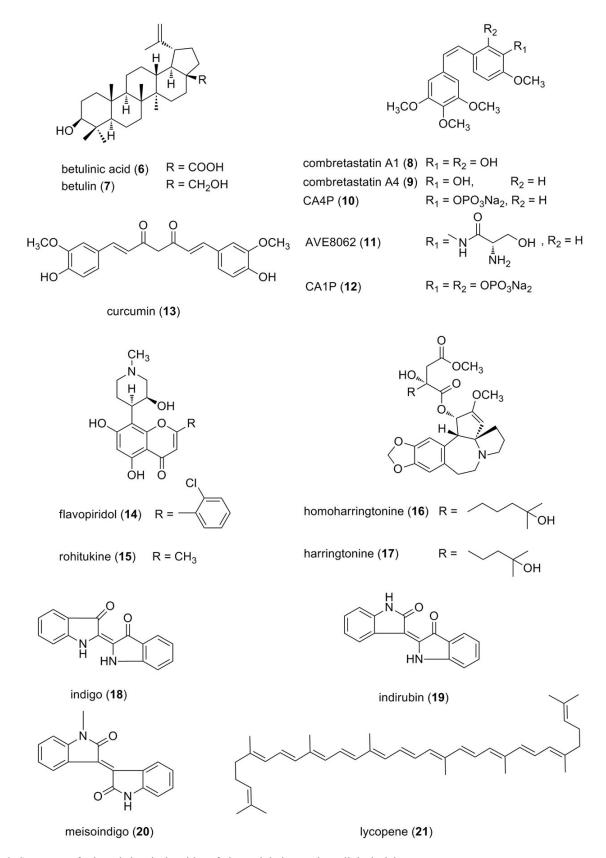


Figure 2. Structures of selected chemical entities of plant origin in oncologyclinical trials. activity against prostate cancer, head and neck cancer, hematopoietic neoplasia, leukemia, and lymphoma

xenograft murine models (51, 52). Mechanistic studies revealed that flavopiridol inhibits the activity of cyclin-dependent kinases (CDKs) by competing with ATP at their nucleotide binding sites, and causes cell cycle arrest at either the G_1 or G_1/M phases. Further studies indicated that flavopiridol also exhibits apoptosis induction, and anti-angiogenic and antiproliferative effects, by interacting at multiple other targets besides CDK (53, 54). Flavopiridol is the first cyclin-dependent kinase inhibitor in clinical trials for the treatment of patients with non-Hodgkin's lymphoma, renal, prostate, colon and gastric cancers (53-56).

Homoharringtonine (16, HHT), a cephalotaxine alkaloid ester, along with its analogue harringtonine (17). was first isolated in the early 1970s from the bark of Cephalotaxus harringtonia (Knight ex J. Forbes) K. Koch (Cephalotaxaceae), an evergreen tree native to East Asia, by Powell, Weisleder, and Smith at the United States Department of Agriculture facility at Peoria, Illinois (57). The only structural difference between these two alkaloids is that homoharringtonine possesses an additional methylene group in the ester side-chain when compared to harringtonine. Homoharringtonine (16) may be obtained also from several other Cephalotaxus species. The mechanism of action of compounds 16 and 17 is unusual, and they promote apoptosis and inhibit protein synthesis at the ribosomal level (58, 59). Clinical studies of a mixture of homoharringtonine (16) and harringtonine (17) were initiated in 1970s by the Chinese Academy of Medical Sciences, for use in the treatment of acute myeloid leukemia (AML) and chronic myeloid leukemia (CML). Homoharringtonine (16) has entered various Phase II/III clinical trials for patients with CML in the United States and in Europe (58-60). A semi-synthetic version of homoharringtonine (sHHT), also known as omacetaxine mepesuccinate and prepared by esterification of the parent compound cephalotaxine obtained from the leaves of a Cephalotaxus species, is undergoing a clinical trial for the treatment of CML in patients with imatinib resistance and intolerance (61).

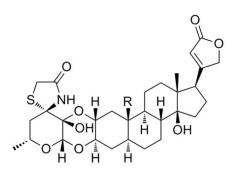
Indigo (18) and indirubin (19) are two bis-indole alkaloids obtained from the blue dye "Indigo Naturalis", which is also known as "Qing Dai" in the People's Republic of China. "Qing Dai" is a traditional Chinese medicinal herbal formula with antibacterial, anti-inflammatory, febrifuge, and hemostatic effects. It is composed of the dried residue derived from the leaves and/or stems of several plants that produce a dark blue dye, including Baphicacanthus cusia (Nees) Bremek. (Acanthaceae), Indigofera suffruticosa Mill. (Fabaceae), Indigofera tinctoria L. (Fabaceae), Isatis tinctoria L. (Brassicaceae), and Polygonum tinctorium Ait. (Polygonaceae) (62, 63). A mechanistic study has demonstrated that indirubin (19) exerts its antileukemic effect by competing with ATP for binding to the catalytic subunit of cyclin-dependent kinase (CDK), leading to the inhibition of this enzyme. The crystallized structure of a CDK2/indirubin analogue complex indicated an interaction with an ATP binding site via hydrogen bonds to key amino acids (64). Meisoindigo (1-methylisoindigo, 20) is a derivative developed to improve the solubility in water and other pharmaceutical

properties of indirubin. This compound showed significant activities against cancer cells through a multi-targeting profile including inhibition of DNA biosynthesis and assembly of microtubules, induction of cell differentiation, and down-regulation of c-myb gene expression (64, 65). Meisoindigo (**20**) has been subjected to clinical trial in the People's Republic of China for chronic myelogenous leukemia (CML) (66).

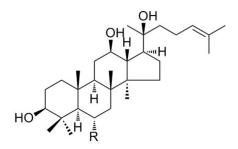
Lycopene (21), a well-known red pigment based on a β-carotene skeleton, is distributed widely in fruits and vegetables, and is especially abundant in tomatoes (Solanum lycopersicum L.; Solanaceae) and processed tomato products (67). The 40-carbon aliphatic chain of the molecule of lycopene contains thirteen *trans*- double bonds. with eleven of these being conjugated. Besides its antioxidant and anti-inflammatory activities, lycopene also exhibits anticarcinogenic peoperties in both in vitro and in vivo models (68, 69). Mechanism-of-action studies have indicated that lycopene exerts its anticancer and chemoprevention activities through the activation of the electrophile/antioxidant response element (EpRE/ARE) transcription system, inducing the expression of phase II detoxifying enzymes, and arresting the cell cycle at the G_0/G_1 phase by regulating cyclin D1 and the PI3K/Akt pathway (69). Lycopene has entered Phase II clinical trials in the United States for the prevention and treatment of prostate cancer (70).

2"-Oxovoruscharin (22), a cardenolide with a rare dihydrothiazole ring in its molecule, was isolated from a tropical evergreen shrub, Calotropis procera (Aiton) W.T. Aiton (Asclepiadaceae), and was demonstrated to have potent in vitro antitumor and Na⁺/K⁺-ATPase inhibitory UNBS1450 (23), derived activities (71). from 2"-oxovoruscharin by reducing the formyl group in the molecule to a hydroxymethyl group, exhibited an improved in vitro cytotoxicity profile activity when compared with the parent compound (72). A mechanistic investigation demonstrated that UNBS1450 induces the disruption of the actin cytoskeleton to affect multiple signaling pathways by binding to the sodium pump, and leads to non-apoptotic cell death (73). UNBS1450 has entered Phase I clinical studies in Europe for patients with solid tumors and lymphomas (74).

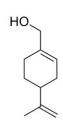
Perillyl alcohol (POH, **24**), is a monoterpenoid with a monocyclic carbon skeleton that is found in the essential oils in a number of plants such as lavender (*Lavendula x intermedia*; Lamiaceae) and cherries [*Prunus avium* (L.) L. (Rosaceae)] (75). Preclinical studies have indicated that perillyl alcohol exhibits cytotoxicity for cancer cell lines derived from lung cancer, pancreatic cancer, prostate cancer, breast cancer and leukemia, and also showed *in vivo* inhibitory effects against UVB-induced skin carcinogenesis and DMBA-induced murine melanoma models (76, 77). Mechanistically, perillyl alcohol induces cell cycle arrest at the G_0/G_1 phase, by modulating the protein levels of cyclin-dependent kinases and



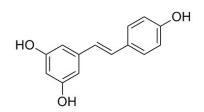
2"-oxovoruscharin (22) R = CHOUNBS1450 (23) $R = CH_2OH$



protopanaxadiol (25) R = H protopanaxatriol (26) R = OH



perillyl alcohol (24)



resveratrol (27)

Figure 3. Structures of selected compounds derived from plants with potential anticancer activity.

cyclin-dependent kinase inhibitors (78). Perillyl alcohol is undergoing Phase I/II clinical trials in patients with breast cancer, ovarian cancer and glioblastoma multiform (79).

Protopanaxadiol [25; 20S-protopanaxadiol (PPD)] and protopanaxatriol [26; 20S-protopanaxatriol (PPT)] are two dammarane-type triterpenoids prepared by the hydrolysis of certain saponins obtained from Asian ginseng (Panax ginseng C.A. Mey.; Araliaceae) and related species (80). Protopanaxadiol induces cancer cell apoptosis and inhibits proliferation through targeting the Wnt/ β-catenin signaling pathway, down-regulating AKt activity, and inhibiting the effects of P-glycoprotein (P-gp) (81-83). Protopanaxadiol and protopanaxatriol are reported to have immunomodulating activity (84). A mixture of protopanaxadiol (25) and protopanaxatriol (26)(PandimexTM) has been approved conditionally in mainland China for the treatment of advanced cancers of the breast, colon-rectum, lung, and pancreas, and is ongoing a Phase I clinical trial in the United States for advanced lung, gastric, breast, and pancreatic cancers in combination with paclitaxel or alone (85, 86).

Resveratrol (27, 3,5,4'-trihydroxy-*trans*-stilbene) is a phenolic compound found in several dietary items, such

as grapes (Vitis vinifera L.; Vitaceae), white mulberries (Morus alba L.; Moraceae), and peanuts (Arachis hypogaea L.; Fabaceae). This compound is also considered as a major constituent responsible for the cardioprotective activity of red wine (87, 88). Intensive studies have revealed that resveratrol possesses antioxidant, anti-inflammatory, and anticarcinogenetic activities, and can prevent and slow a wide range of illnesses, including age-related diseases, cancer, cardiovascular problems, diabetes, and ischemic injuries (87-89). Resveratrol inhibits the growth of cancer cells and induces apoptosis by acting at multiple cellular targets, including activation of p53, inhibiting cycloxygenase and cytochrome P450 enzymes, and activating AMP-activated kinase (AMPK) (87-89). Resveratrol has been also reported to show sensitization effects on drug-resistant tumor cells and to result in a synergistic cytotoxicity when combined with established anticancer therapies (90). This compound is now undergoing Phase I/II clinical trials for the prevention and treatment of colon cancer in the United States (91).

4. SOME PLANT-DERIVED COMPOUNDS WITH POTENTIAL ANTICANCER ACTIVITY (Figure 3)

A series of anthracenone C-glycosides,

alvaradoins E-N, was isolated from the leaves of Alvaradoa haitiensis Urb. (Picramniaceae), by Mansukh Wani and colleagues at Research Triangle Institute in North Carolina (92). Among these compounds, alvaradoin E (28), and its 10(R) isomer, alvaradoin F (29), were found to be the most potent cytotoxic agents against the KB cell line, exhibiting IC_{50} values of 0.050 and 0.065 μ M, respectively (92). Alvaradoins E and F were also evaluated in *in vivo* hollow fiber assay using KB, LNCaP, and Col2 cells, and both demonstrated significant inhibitory activity at the intraperitoneal site, while being less active when administered subcutaneously (93). In an in vivo P388 murine lymphocytic leukemia model study, alvaradoin E exhibited discernible activity (92). This compound was studied further for its mechanism of action. After treatment of LNCaP human prostate cancer cells with alvaradoin E. early signs of apoptosis including chromatin condensation and dose-dependent membrane depolarization were noticed. A significant decrease in cell viability accompanied by an increase in DNA breakage was observed using HL-60 leukemia cells (93).

Quassinoids are highly oxygenated degraded triterpenoid derivatives typically with a bitter taste, which are found in many species in the family Simaroubaceae. As the major active principles of these plants, quassinoids have been documented with a wide spectrum of biological activities, including anti-HIV, antimalarial, antiparasitic, antitumor, and insecticidal effects (94). Bruceantin (30), a quassinoid with potent cytotoxicity, was brought to clinical trials as an anticancer candidate by the U.S. NCI in the 1980s, but was eventually dropped because of a lack of demonstrated efficacy (95). Very recently, a new antitumor quassinoid. 2'-(R)-O-acetylglaucarubinone (31) was isolated by Usami et al. from the bark of Odvendyea gabonensis (Pierre) Engler (96). Compound 31 exhibited potent cytotoxicity against a small panel of human cancer cell lines, including prostate (DU145), lung (A549), and oral epidermoid carcinoma (KB) cells, with ED₅₀ values around 0.05 µg/mL in each case. This compound was further evaluated using multiple breast cancer and ovarian cancer cell lines. Compound 31 demonstrated strong inhibitory effects against several ER- or/and PR-negative cell lines, suggesting that neither the estrogen receptor (ER) nor the progesterone receptor (PR) is the major target of this compound. Moreover, the HER2-overexpressing ER/PR-negative SKBR3 cell line was hypersensitive to this quassinoid, which implied that the HER2 signaling pathway might be involved in mediating its cytotoxicity (96). The in vivo mammary epithelial proliferation inhibitory activity of compound 31 was evaluated using a Brca1/p53-deficient mouse model. The mammary duct branching points were reduced to 32% after a daily intraperitoneal injection of 0.1/mg of compound 31 for one week, and the reduction effect on such branching was even more significant than observes for the control compounds, bruceantin (30) and paclitaxel (96). Further mechanism studies of action on this promising antineoplastic agent are being carried out.

More than 30 species in the family Amaryllidaceae have been documented as having a history of use as folk medicines for the treatment of tumors. Two

isocarbostyril alkaloids, narciclasine (32) and pancratistatin (33), first isolated from bulbs of a *Narcissus* species (97) and from Hymenocallis littoralis Salisb. (formerly known as Pancratium littorale Jacq.) (98), respectively, are two of the most promising antineoplastic compounds from this plant family, and are effective and selective anticancer agents with the potential for clinical development. These two compounds exhibited in vivo growth inhibitory activity, in turn, in M5076 sarcoma (99) and P388 lymphocytic leukemia models (98). A study has indicated that normal human fibroblasts were nearly 250-fold less sensitive to narciclasine than MCF-7 cells and PC-3 cells (100). Pancratistatin selectively induced apoptosis of leukemia cells from patients with minimal affect on normal peripheral blood mononuclear cells (101), and induced apoptosis specifically in breast cancer cells in comparison to non-cancerous cells (102). Pyridinum narciclasine (34) and sodium pancratistatin 3,4-O-cyclic phosphate (35) are prodrugs of narciclasine and pancratistatin, respectively, which have been produced to improve the water solubility of these two compounds (99, 103). Compound 35 was shown to retain the potency of cancer cell line inhibition when compared to the parent compound (33), and has been selected for preclinical development (103).

Tanshinone I (36), tanshinone IIA (37), and cryptotanshinone (38), are three major diterpene quinone derivatives isolated from the rhizomes of Salvia miltiorrhiza Bunge (Lamiaceae), which is known as "Tanshen" in traditional Chinese medicine and used as a herbal remedy with multiple therapeutic effects (104). These three tanshinone derivatives exhibited significant in vitro cytotoxicity against several human carcinoma cell lines (104). Tanshinone I (36) was found to inhibit the growth and invasion of breast cancer cells both in vitro and in vivo through regulation of adhesion molecules including ICAM-1 and VCAM-1 (105), and induce apoptosis of leukemia cells by interfering with the mitochondrial transmembrane potential ($\Delta \Psi$ m), increasing the expression of Bax, as well as activating caspase-3 (106). Tanshinone IIA (37) has been reported to inhibit the growth of cervical cancer cells through disrupting the assembly of microtubules, and induces G₂/M phase arrest and apoptosis (107). This compound (37) can also inhibit invasion and metastasis of hepatocellular carcinoma (HCC) cells both in vitro and in vivo, by suppressing the expression of the metalloproteinases, MMP2 and MMP9 and interfering with the NF κ B signaling pathway (108). Cryptotanshione (38) was reported to induce cell-cycle arrest at the G₁-G₀ phase, which was accompanied by the inhibition of cyclin D1 expression, retinoblastoma (Rb) protein phosphorylation, and of the rapamycin (mTOR) signaling pathway (109). Compound 38 can sensitize DU-345 breast cancer cells by suppressing the expression of the apoptosis inhibitory protein, Bcl-2 (110). Neo-tanshinlactone (39) is a new lead antineoplastic compound isolated from S. miltiorrhiza by Dr. Kuo-Hsiung Lee's research group at the University of North Carolina (111). Unlike the tanshinones, neo-tanshinlactone (39) possesses a lactone feature rather than an ortho-quinone moiety in ring C. This compound exhibited selectivity against two estrogen receptor-positive (ER⁺) breast cancer cell lines, MCF-7 and ZR-75-1, with ED₅₀

values, in turn, of 0.6 and 0.3 μ g/mL (111). Compound **40**, a synthetic analogue of neo-tanshinlactone (**39**), was found to inhibit the growth of the estrogen-dependent breast cancer cells, ZR-75-1, in an *in vivo* xenograft model (112). These investigations suggest that neo-tanshinlactone (**39**) and its derivatives might be promising candidates for the treatment of hormone-dependent breast cancers.

Silvestrol (41), and its 5"S epimer, episilvestrol (42), were isolated by Kinghorn et al. from the tropical plant Aglaia foveolata Pannell (Meliaceae), with the structure and absolute configuration elucidated using single-crystal X-ray crystallography analysis (113). Silvestrol has also been documented as an antineoplastic constituent of Aglaia leptantha, collected in Sarawak, Malaysia, but with the stereochemistry of the 1.4-dioxanyloxy ring undetermined (114). This "flavagline" derivative was found to possess potent cytotoxicity for a small panel of human cancer cell lines and exhibited activity in both in vivo hollow fiber and P-388 lymphocytic leukemia assays (113). The total synthesis of silvestrol (41) has been accomplished by two independent groups (115-117). Preliminary mechanistic studies conducted by Dr. Steven Swanson and co-workers at the University of Illinois at Chicago showed that in LNCaP human prostate cancer cells, silvestrol produces a p53-independent cell-cycle blockage at the G2/M check-point, and induces apoptosis through the regulation of caspases-2, -9 and -10 but not caspases-3 and -7 (118, 119). Additional investigation at The Ohio State University Medical Center, in the laboratory of Dr. Michael Grever, demonstrated that silvestrol exhibited B-cell selective activity and showed efficacy in both chronic lymphocytic leukemia and acute lymphocytic leukemia models (120). Silvestrol can lead an early reduction in Mcl-1 expression in chronic lymphocytic leukemia cells (120), and inhibits the translation of malignancy-related mRNA by regulating initiation factor elF4A (121). The compound demonstrated synergy with other agents using acute mylegenous leukemia cells (122) and modulated sensitivity to doxorubicin in a pre-clinical mouse lymphoma model (123). Further preclinical evaluation of silvestrol (41) as a potential antileukemic agent is ongoing.

5. CONCLUSION

Although cancer is one of the most pressing heath concerns worldwide, the success rate in oncology new drug development is more than three times less than that for cardiovascular diseases (124). Cancer drug discovery is difficult because of many inherent obstacles such as tumor heterogeneity (124). Chemotherapy is an important option in modern cancer treatment, and plant-derived chemotherapeutic agents have contributed greatly to the progress of oncology chemotherapy development and to clinical practice. From the examples of antineoplastic candidates described in this review, it is evident that small molecules of plant origin continue to be valuable as sources of potential lead compounds in anticancer drug discovery. According one recent estimate, more than 85% of higher plants have not been evaluated systematically for the presence of bioactive principles (125). There are more than 60 compounds from plant sources in the pipeline as

potential anticancer agents (9-11). Innovations in the multidisciplinary investigative methods offer great promise for plant-derived drug discovery and development. These include new techniques related to compound isolation and structure elucidation, enhanced high-throughput biological screening procedures using novel biological targets, and continuing improvements in synthetic chemistry to make the optimization of the lead compounds more efficient. Therefore, there remains great potential to explore the plant kingdom for new antineoplastic lead compound by using novel targets and newly developed technologies, so a promising future for anticancer drug development based on these agents can be predicted.

6. ACKNOWLEGMENT

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7. REFERENCES

1. Kai U. Bindseil, Jasmin Jakupovica, Dietmar Wolf, Jacques Lavayre, Jean Leboul and Didier van der Pyl: Pure compound libraries: a new perspective for natural product based drug discovery. *Drug. Discov. Today* 6, 840-847 (2001)

2. Richard D. Firn and Clive G. Jones: Natural products - a simple model to explain chemical diversity. *Nat. Prod. Rep.* 20, 382-391 (2003)

3. P. Vuorela, M. Leinonen, P. Saikku, P. Tammela, J.-P. Rauha, T. Wennberg and H. Vuorela: Natural products in the process of finding new drug candidates. *Curr. Med. Chem.* 11, 1375-1389 (2004)

4. David J. Newman and Gordon M. Cragg: Natural products as sources of new drugs over the last 25 years. *J. Nat. Prod.* 70, 461-477 (2007)

5. Frank E. Koehn, Guy T. Carter: The evolving role of natural products in drug discovery. *Nature Rev. Drug Disc.* 4, 206-220 (2005)

6. Kin S. Lam: New aspects of natural products in drug discovery. *Trends Microbiol.* 15, 279-289 (2007)

7. Jesse W.-H. Li, John C. Vederas: Drug discovery and natural products: end of an era or an endless frontier? *Science* 325, 161-165 (2009)

8. Christian Bailly: Ready for a comeback of natural products in oncology. *Biochem. Pharmacol.* 77, 1447-1457 (2009)

9. Arvind Saklani, Samuel K. Kutty: Plant-derived compounds in clinical trials. *Drug Disc. Today* 13, 161-171 (2008)

10. Alan L. Harvey: Natural products in drug discovery.

Drug Disc. Today 13, 894-901 (2008).

11. Mark S. Butler: Natural products to drugs: natural product-derived compounds in clinical trials. Nat. Prod. Rep. 25, 475-516 (2008)

12. R. H. Noble, J. H. Cutts, C. H. Beer: Further biological activities of vincaleukoblastine – an alkaloid isolated from *Vinca rosea* (L.). Biochem. Pharmacol. 1347-1348 (1958)

13. Norbert Neuss, Marvin Gorman, Harold E. Boaz, Nancy J. Cone: Vinca alkaloids. XI. Structures of leurocristine and vincaleukoblastine. J. Am. Chem. Soc. 84, 1509-1510 (1962)

14. Gordon H. Svoboda: Alkaloids of *Vinca rosea* (*Catharanthus roseus*). IX. Extraction and characterization of leurosidine and leurocristine. Lloydia 24, 173-178 (1961)

15. Jonathan L. Hartwell, Anthony W. Schrecker: Components of podophyllin. V. The constitution of podophyllotoxin. J. Am. Chem. Soc. 73, 2909-2916 (1951)

16. Mansukh C. Wani, Harold Lawrence Taylor, Monroe E. Wall, Philip Coggon, Andrew T. McPhail: Plant antitumor agents. VI. Isolation and structure of taxol, a novel antileukemic and antitumor agent from *Taxus brevifolia*. J. Am. Chem. Soc. 93, 2325-2327 (1971)

17. Monroe E. Wall, M. C. Wani, C. E. Cook, Keith H. Palmer, A. T. McPhail, G. A. Sim: Plant antitumor agents. I. The isolation and structure of camptothecin, a novel alkaloidal leukemia and tumor inhibitor from *Camptotheca acuminata*. J. Am. Chem. Soc. 88, 3888-3890 (1966)

18. Mary Ann Jordan, Leslie Wilson: Microtubules as a target for anticancer drugs. *Nat. Rev.* 4, 253-265 (2004)

19. David G. I. Kingston: Tubulin-interactive natural products as anticancer agents. *J. Nat. Prod.* 72, 507-515 (2009).

20. Gordon M. Cragg, David J. Newman: A tale of two tumor targets: topoisomerase I and tubulin. the Wall and Wani contribution to cancer chemotherapy. *J. Nat. Prod.* 67, 232-244 (2004)

21. Yaw-Huei Hsiang, Michelle G Lihou, Leroy F. Liu: Arrest of replication forks by drug-stabilized topoisomerase I-DNA cleavable complex as a mechanism of cell killing by camptochecin. *Cancer. Res.* 49, 5077-5082 (1989)

22. K. R. Hande: Etoposide: four decades of development of a topoisomerase II inhibitor. *Eur. J. Cancer* 34, 1514-1521 (1998)

23. Gordon M. Cragg, David G. I. Kingston, David J. Newman, Eds.: Anticancer Agents from Natural Products, CRC/Taylor & Francis, Boca Raton, FL (2005)

24. M. Gordaliza: Natural products as leads to anticancer

drugs. Clin. Transl. Oncol. 9, 767-776 (2007).

25. Gordon M. Cragg, Paul G. Grothaus, David J. Newman: Impact on natural products on developing new anticancer agents. *Chem. Rev.* 109, 3012-3043 (2009).

26. Matthew Suffness, John Douros: Current status of the NCI plant and animal program. *J. Nat. Prod.* 45, 1-14 (1982)

27. Gordon M. Cragg, Michael R. Boyd, Rita Khanna, Robert Kneller, Thomas D. Mays, Kate D. Mazan, David J. Newman, Edward A. Sausville: International collaboration in drug discovery and development: the NCI experience. *Pure Appl. Chem.* 71, 1619-1633 (1999)

28. Gordon M. Cragg, David J. Newman, Stringner S. Yang: Natural product extracts of plant and marine origin having antileukemia potential. The NCI experience. *J. Nat. Prod.* 69, 488-498 (2006)

29. Yali F. Hallock, Gordon M. Cragg: National Cooperative Drug Discovery Groups (NCDDGs): a successful model for public private partnerships in cancer drug discovery. *Pharm. Biol.* 41 (Suppl.), 78-91 (2003)

30. Melinda G. Hollingshead, Michael C. Alley, Richard F. Camalier, Betty J. Abbott, Joseph G. Mayo, Louis Malspeis, Michael R. Grever: *In vivo* cultivation of tumor cells in hollow fibers. *Life Sci.* 57, 131-141 (1995).

31. Qiuwen Mi, John M. Pezzuto, Norman R. Farnsworth, Mansukh C. Wani, A. Douglas Kinghorn, Steven M. Swanson: Use of the *in vivo* hollow fiber assay in natural products drug discovery. *J. Nat. Prod.* 72, 573-580 (2009)

32. Emily Pisha, Heebyung Chai, Ik-Soo Lee, Tangai E. Chagwedera, Norman R. Farnsworth, Geoffrey A. Cordell, Christopher W.W. Beecher, Harry H. S. Fong, A. Douglas Kinghorn, Daniel M. Brown, Mansukh C. Wani, Monroe E. Wall, Tina E. Heiken, Tapas K. Das Gupta, John M. Pezzuto: Discovery of betulinic acid as a selective inhibitor of human melanoma that functions by induction of apoptosis. *Nat. Med.* 1, 1046-1051 (1995)

33. Valentina Zuco, Rosanna Supino, Sabina C. Righetti, Loredana Cleris, Edoardo Marchesi, Carlo Gambacorti-Passerini, Franca Formelli: Selective cytotoxicity of betulinic acid on tumor cell lines, but not on normal cells. *Cancer Lett.* 175: 17-25 (2002)

34. Melanie N. Laszczyk: Pentacyclic triterpenes of the lupane, oleanane and ursane group as tools in cancer therapy. *Planta Med.* 75, 1549-1560 (2009)

35. Alakurtti Sami, Ma kela Tarua, Koskimies Salmea, Yli-Kauhaluoma Jari: Pharmacological properties of the ubiquitous natural product betulin. *Eur. J. Pharm. Sci.* 29, 1-13 (2006).

36. U.S. National Institutes of Health: Evaluation of 20% betulinic acid ointment for treatment of dysplastic nevi

(moderate to severe dysplasia), 2010. Further information available at:

http://www.cancer.gov/search/ViewClinicalTrials.aspx?cdri d=494341&version=HealthProfessional&protocolsearchid= 3664931

37. George R. Pettit, Gordon M. Cragg, Delbert L. Herald, Jean M. Schmidt, Prasert Lohavanijaya: Isolation and structure of combretastatin. *Can. J. Chem.* 60, 1374-1376 (1982)

38. George R. Pettit, Sheo Bux Singh, Margaret L. Niven, Ernest Hamel, Jean M. Schmidt: Isolation, structure, and synthesis of combretastatins A-1 and B-1, potent new inhibitors of tubulin assembly, derived from *Combretum caffrum. J. Nat. Prod.* 50, 119-131 (1987)

39. Scott L. Young, David J. Chaplin: Combretastatin A4 phosphate: background and current clinical status. *Expert. Opin. Invest. Drugs* 13, 171-1182 (2004)

40. Dietmar W. Siemann, David J. Chaplin, Patricia A Walicke: A review and update of the current status of the vasculature-disabling agent combretastatin-A4 phosphate (CA4P). *Expert. Opin. Investig. Drugs* 18, 189-197 (2009)

41. Kevin G. Pinney, Christopher Jelinek, Klaus Edvardsen, David J. Chaplin, George R. Pettit: The discovery and development of the combretastatins. In Anticancer Agents from Natural Products, Gordon M. Cragg, David G. I. Kingston and David J. Newman, Eds., CRC Taylor & Francis, Boca Raton, FL, pp. 23-46 (2005)

42. Catherine M. L. West, Pat Price: Combretastatin A4 phosphate. *Anti-Cancer Drugs* 5, 179-187 (2004)

43. ClinicalTrials.gov.: Study of Combretastatin and Paclitaxel/Carboplatin in the Treatment of Anaplastic Thyroid Cancer. Further information available at: http://clinicaltrials.gov/ct2/show/NCT00507429 (accessed October 2009).

44. Angelo Delmonte, Cristiana Sessa: AVE8062: a new combretastatin derivative vascular disrupting agent. *Expert. Opin. Invest. Drugs* 18, 1541-1548 (2009)

45. John W. Lippert III: Vascular disrupting agents. *Bioorg. Med. Chem.* 5, 605-615 (2007)

46. Gaurisankar Sa, Tanya Das, Shuvomoy Banerjee, Juni Chakraborty: Curcumin: from exotic spice to modern anticancer drug. *Al Ameen J. Med. Sci.* 3, 21-37 (2010)

47. Bharat B. Aggarwal, Chitra Sundaram, Nikita Malani, Haruyo Ichikawa: Curcumin: the Indian solid gold. *Adv. Exp. Med. Biol.* 595, 1-75 (2007)

48. Navneet Dhillon, Bharat B. Aggarwal, Robert A. Newman, Robert A.Wolff, Ajaikumar B. Kunnumakkara, James L. Abbruzzese, Chaan S. Ng, Vladimir Badmaev, Razelle Kurzrock: Phase II trial of curcumin in patients with advanced pancreatic cancer. *Clin. Cancer Res.* 14,

4491-4499 (2008)

49. Alan D. Harmon, Ulrich Weiss, J. V. Silverton: The structure of rohitukine, the main alkaloid of *Amoora rohituka* (syn. *Aphanamixis polystachya*) (Meliaceae). *Tetrahedron Lett.* 20, 721-724 (1979)

50. A. D. Lakdawala, M. V. Shirole, S. S. Mandrekar, A. N. Dohadwalla: Immunopharmacological potential of rohitukine: a novel compound isolated from the plant *Dysoxylum binectariferum. Asia Pac. J. Pharmcol.* 3, 91-98 (1988)

51. Keith C. Bible, Scott H. Kaufmann: Flavopiridol: a cytotoxic flavone that induces cell death in noncycling A549 human lung carcinoma cells. *Cancer Res.* 56, 4856-4861 (1996)

52. Francisco Arguello, Mark Alexander, Judith A. Sterry, Gabriela Tudor, Erik M. Smith, Naina T. Kalavar, John F. Greene Jr., William Koss, C. David Morgan, Sherman F. Stinson, Timothy J. Siford, W. Gregory Alvord, Richard L. Klabansky, Edward A. Sausville: Flavopiridol induces apoptosis of normal lymphoid cells, causes immunosuppression, and has potent antitumor activity *in vivo* against human leukemia and lymphoma xenografts. *Blood* 91, 2482-2490 (1998)

53. Adrian M. Senderowicz: Flavopiridol: the first cyclin-dependent kinase inhibitor in human clinical trials. *Invest. New Drugs* 17, 313-320 (1999)

54. Vladimir Krystof, Stjepan Uldrijan: Cyclin-dependent kinase inhibitors as anticancer drugs. *Curr. Drug Targets* 11, 291-302 (2010)

55. Gary K. Schwartz, David Ilson, Leonard Saltz, Eileen O'Reilly, William Tong, Peter Maslak, Jeanine Werner, Pam Perkins, Maxine Stoltz, David Kelsen: Phase II study of the cyclin-dependent kinase inhibitor flavopiridol administered to patients with advanced gastric carcinoma. *J. Clin. Oncol.* 19, 1985-1992 (2001)

56. Judith E. Karp, Amanda Blackford, B. Douglas Smith, Katrina Alino, Amy Hatfield Seung, Javier Bolanos-Meade, Jacqueline M. Greer, Hetty E. Carraway, Steven D. Gore, Richard J. Jones, Mark J. Levis, Michael A. McDevitt, L. Austin Doyle, John J. Wright: Clinical activity of sequential flavopiridol, cytosine arabinoside, and mitoxantrone for adults with newly diagnosed, poor-risk acute myelogenous leukemia. *Leukemia Res.* 34, 877-882 (2010)

57. R. G. Powell, D. Weisleder, C. R. Smith Jr: Antitumor alkaloids from *Cephalotaxus harringtonia*: structure and activity. *J. Pharm. Sci.* 61, 1227-1230 (1972)

58. Hideji Itokawa, Xihong Wang, Kuo-Hsiung Lee: Homoharringtonine and related compounds. In Anticancer Agents from Natural Products, Gordon M. Cragg, David G. I. Kingston and David J. Newman, Eds., CRC/Taylor & Francis, Boca Raton, FL, pp. 47-70 (2005) 59. Alfonso Quintas-Cardama, Hagop Kantarjian, Jorge Cortes: Homoharringtonine, omacetaxine mepesuccinate, and chronic myeloid leukemia circa 2009. Cancer 115, 5382-5393 (2009)

60. The Rocky Mountain Blood and Marrow Transplant Program (RMBMTP), 2009: Initial patients treated in multinational Phase 2/3 study of ceflatonin 09-20-2006. Further information available at: http://www.rockymountainbmt.com/news/Initial-Patients-T reated-in-Multinational-Phase-23-Study-of-Ceflatonin-468 2.html (accessed October 2009)

61. Alfonso Quintas-Cardama, Jorge Cortes: Omacetaxine mepesuccinate - a semisynthetic formulation of the natural antitumoral alkaloid homoharringtonine, for chronic myelocytic leukemia and other myeloid malignancies. *IDrugs* 11, 356-372 (2008)

62. Bai-lin Deng: Direct colorimetric method for determination of indigo and indirubin in Qingdai. Zhong Cao Yao 17, 163-164 (1986)

63. Ralph Hoessel, Sophie Leclerc, Jane A. Endicott, Martin E. M. Nobel, Alison Lawrie, Paul Tunnah, Maryse Leost, Eve Damiens, Dominique Marie, Doris Marko, Ellen Niederberger, Weici Tang, Gerhard Eisenbrand, Laurent Meijer: Indirubin, the active constituent of a Chinese antileukaemia medicine, inhibits cyclin-dependent kinases. *Nat. Cell Biol.* 1, 60-67 (1999)

64. Zhijian Xiao, Yushu Hao, Bingcheng Liu, Lingsheng Qian: Indirubin and meisoindigo in the treatment of chronic myelogenous leukemia in China. *Leuk. Lymph.* 43, 1763-1768 (2002)

65. Mingxin. Zuo, Yan Li, Hongbo Wang, Jianhua Zhou, Hongyan Li, He Liu, Hongqi Xin, Sen Zhang, Xiaoguo Chen. The antitumor activity of meisoindigo against human colorectal cancer HT-29 cells *in vitro* and *in vivo*. J. Chemother. 20, 728-733 (2008)

66. Cooperative Study Group of Phase III Clinical Trial on Meisoindigo. Phase II clinical trial on meisoindigo in the treatment of chronic myelogenous leukemia. *Zhonghua Xueyexue Zazhi* 18, 69-72 (1997).

67. Kin-Weng Kong, Hock-Eng Khoo, K. Nagendra Prasad, Amin Ismail, Chin-Ping Tan, Nor Fadilah Rajab: Revealing the power of the natural red pigment lycopene. *Molecules* 15, 959-987 (2010)

68. Yoav Sharoni, Michael Danilenko, Joseph Levy: Molecular mechanisms for the anticancer activity of the carotenoid lycopene. *Drug Dev. Res.* 50, 448-456 (2000)

69. V. Bhuvaneswari, S. Nagini: Lycopene: A review of its potential as an anticancer agent. *Curr. Med. Chem.*-*Anti-Cancer Agents* 5, 627-635 (2005)

70. U.S. National Institutes of Health: Lycopene in treating patients with metastatic prostate cancer. (2009)

http://clinicaltrials.gov/ct2/show/NCT00068731

71. Eric Van Quaquebeke, Gentiane Simon, Aurelie Andre, Janique Dewelle, Mohamed El Yazidi, Frederic Bruyneel, Jerome Tuti, Odile Nacoulma, Pierre Guissou, Christine Decaestecker, Jean-Claude Braekman, Robert Kiss, Francis Darro: Identification of a novel cardenolide (2"-oxovoruscharin) from *Calotropis procera* and the hemisynthesis of novel derivatives displaying potent *in vitro* antitumor activities and high *in vivo* tolerance: structure-activity relationship analyses. *J. Med. Chem.* 48, 849-856 (2005)

72. Tatjana Mijatovic, Florence Lefranc, Eric Van Quaquebeke, Frank Van Vynckt, Francis Darro, Robert Kiss: UNBS1450: a new hemi-synthetic cardenolide with promising anti-cancer activity. *Drug Dev. Res.* 68, 164-173 (2007)

73. Tom Juncker, Marc Schumacher, Mario Dicato, Marc Diederich: UNBS1450 from *Calotropis procera* as a regulator of signaling pathways involved in proliferation and cell death. *Biochem. Pharmacol.* 78, 1-10 (2009)

74. Unibioscreen: The piperline – UNBS 1450, 2009. Further information available at http://www.unibioscreen.com/randd/pipeline_unbs1450.html (accessed August 2010)

75. James T. Belanger: Perillyl alcohol: applications in oncology. *Altern. Med. Rev.* 3, 448-457 (1998)

76. Margaret Barthelman, Weixing Chen, Helen L. Gensler, Chuanshu Huang, Zigang Dong, G. Tim Bowden: Inhibitory effects of perillyl alcohol on UVB - induced murine skin cancer and AP-1 transactivation. *Cancer Res.* 58, 11-716 (1998)

77. Maria Lluria-Prevatt, Jeanne Morreale, Jacie Gregus, David S. Alberts, Fiona Kaper, Amato Giaccia, Marianne Broome Powell: Effects of perillyl alcohol on melanoma in the TPras mouse model. *Cancer Epidemiol. Biomark. Prev.* 11, 573-579 (2002)

78. Dean A. Wiseman, Sean R. Werner, Pamela L. Crowell: Cell cycle arrest by the isoprenoids perillyl alcohol, geraniol, and farnesol is mediated by p21Cip1 and p27Kip1 in human pancreatic adenocarcinoma cells. *J. Pharmacol. Exp. Ther.* 320, 1163-1170 (2007)

79. Juliana de Saldanha da Gama Fischer, Paulo Costa Carvalho, Ana Gisele da Costa Neves-Ferreira, Clovts Orlando da Fonseca, Jonas Perales, Maria da Gloria da Costa Carvalho, Gilberto Barbosa Domont: Anti-thrombin as a prognostic biomarker candidate for patients with recurrent glioblastoma multiform under treatment with perillyl alcohol. *J. Exp. Ther. Oncol*. 7, 285-290 (2008)

80. Shi-Fen Chu, Jun-Tian Zhang: New achievements in ginseng research and its future prospects. *Chin. J. Integr. Med.* 15, 403-408 (2009)

81. David G. Popovich, David D. Kitts: Structure-function relationship exists for ginsenosides in reducing cell proliferation and inducing apoptosis in the human leukemia

(THP-1) cell line. Arch. Biochem. Biophys. 406, 1-8 (2002)

82. Gang Li, Zhenhua Wang, Yaxuan Sun, Ke Liu, Ziren Wang: Ginsenoside 20(*S*)-protopanaxadiol inhibits the proliferation and invasion of human fibrosarcoma HT1080 cells. *Basic Clin. Pharmacol. Toxicol.* 98, 588-592 (2006)

83. Guo-Yu Liu, Xuexian Bu, Hang Yan, William W.-G. Jia: 20S-Protopanaxadiol-induced programmed cell death in glioma cells through caspase-dependent and -independent pathways. J. Nat. Prod. 70, 259-265 (2007)

84. Jianhua Sun, Songhua Hu, Xiaoming Song: Adjuvant effects of protopanaxadiol and protopanaxatriol saponins from ginseng roots on the immune responses to ovalbumin in mice. *Vaccine* 25, 1114-1120 (2007)

85. X. Ouyang, Z. Yu, Z. Chen, F. Xie, W. Fang, Y. Peng, X. Chen, W. Chen, W. Wang, P. Qi, W. Jia: A pilot study of safety and efficacy of Pandimex with or without paclitaxel in the treatment of advanced solid tumors. *J. Clin. Oncol.* 23, 3188 (2005)

86. PanaGin Pharmaceuticals, Inc.: Clinical trials sponsored by pegasus pharmaceuticals group Inc., 2009. Further information available at http://www.panagin.com/clinical.html (accessed Agust 2010)

87. Bharat B. Aggarwal, Anjana Bhardwaj, Rishi S. Aggarwal, Navindra P. Seeram, Shishir Shishodia, Yasunari Takada: Role of resveratrol in prevention and therapy of cancer: preclinical and clinical studies. *Anticancer Res.* 24, 2783-2840 (2004)

88. Anupam Bishayee, Themos Politis, Altaf S. Darvesh: Resveratrol in the chemoprevention and treatment of hepatocellular carcinoma. *Cancer Treat. Rev.* 36, 43-53 (2010)

89. Mohammad Athar, Jung Ho Back, Xiuwei Tang, Kwang Ho Kim, Levy Kopelovich, David R. Bickers, Arianna L. Kim: Resveratrol: A review of preclinical studies for human cancer prevention. *Toxicol. Appl. Pharmacol.* 224, 274-283 (2007)

90. Simone Fulda, Klaus-Michael Debatin: Sensitization for anticancer drug-induced apoptosis by the chemopreventive agent resveratrol. *Oncogene* 23, 6702–6711 (2004)

91. ClinicalTrials.gov: Resveratrol for patients with colon cancer, 2009. Further information available at: http://clinicaltrials.gov/ct2/show/NCT00256334 (accessed October 2010).

92. Sharnelle S. Phifer, Dongho Lee, Eun-Kyoung Seo, Nam-Cheol Kim, Tyler N. Graf, David J. Kroll, Hernan A. Navarro, Robert A. Izydore, Francisco Jiménez, Ricardo Garcia, William C. Rose, Craig R. Fairchild, Robert Wild, Djaja D. Soejarto, Norman R. Farnsworth, A. Douglas Kinghorn, Nicholas H. Oberlies, Monroe E. Wall, Mansukh C. Wani: Alvaradoins E-N, antitumor and cytotoxic anthracenone *C*-glycosides from the leaves of *Alvaradoa* haitiensis. J. Nat. Prod. 70, 954-961 (2007)

93. Qiuwen Mi, Daniel Lantvit, Eulenia Reyes-Lim, Heebyung Chai,Sharnelle S. Phifer, Mansukh C. Wani, Monroe E. Wall, Ghee T. Tan, Geoffrey A. Cordell, Norman R. Farnsworth, A. Douglas Kinghorn, John M. Pezzuto: Apoptotic anticancer effect of alvaradoin E isolated from *Alvaradoa haitiensis*. *Anticancer Res.* 779-787 (2005)

94. Ivo J. Curcino Vieira, Raimundo Braz-Felho: Quassinoids. Structural diversity, biological activity and synthetic studies. *Stud. Nat. Prod. Chem.* 33, Part M, 433-492 (2006)

95. Gordon M. Gragg, David J. Newman. Plants as a source of anticancer agents. J. Ethnopharmacol. 100, 72-79 (2005)

96. Yoshihide Usami, Kyoko Nakagawa-Goto, Jing-Yu Lang, Yoon Kim, Chin-Yu Lai, Masuo Goto, Nobuko Sakurai, Masahiko Taniguchi, Toshiyuki Akiyama, Susan L. Morris-Natschke, Kenneth F. Bastow, Gordon Cragg, David J. Newman, Mihoyo Fujitakeo, Koichi Takeya, Mien-Chie Hung, Eva Y.-H. P. Lee, and Kuo-Hsiung Lee: Antitumor Agents. 282. 2'-(*R*)-*O*-acetylglaucarubinone, a quassinoid from *Odyendyea gabonensis* as a potential anti-breast and anti-ovarian cancer agent. *J. Nat. Prod.* 73, 1553-1558 (2010)

97. Giovanni Ceriotti, Luigi Spandrio, Annibale Gazzaniga: Demonstration, isolation and physical and chemical characteristics of narciclasine, a new antimitotic agent of plant origin. *Tumori*, 53, 359-371 (1967)

98. George R. Pettit, Venkatswamy Gaddamidi, Gordon M. Cragg, Delbert L. Herald and Yoneo Sagawa: Isolation and structure of pancratistatin. *J. Chem. Soc., Chem. Commun.* 1693-1694 (1984)

99. George R. Pettit, Noeleen Melody, Michael Simpson, Michael Thompson, Delbert L. Herald, and John C. Knight: Antineoplastic agents 500. Narcistatin. *J. Nat. Prod.* 66, 92-96 (2003)

100. Patrick Dumont, Laurent Ingrassia, Sebastien Rouzeau, Fabrice Ribaucour, Stephanie Thomas, Isabelle Roland, Francis Darro, Florence Lefrancy, Robert Kiss: The Amaryllidaceae isocarbostyril narciclasine induces apoptosis by activation of the death receptor and/or mitochondrial pathways in cancer cells but not in normal fibroblasts. *Neoplasia* 9, 766-776 (2007)

101. Carly Griffin, Caroline Hamm, James McNulty, Siyaram Pandey: Pancratistatin induces apoptosis in clinical leukemia samples with minimal effect on non-cancerous peripheral blood mononuclear cells. *Cancer Cell Intern.* 10:6 (2010) doi:10.1186/1475-2867-10-6.

102. Peter Siedlakowski, Amanda McLachlan-Burgess, Carly Griffin, Sridhar S. Tirumalai, James McNulty, Siyaram Pandey: Synergy of pancratistatin and tamoxifen on breast cancer cells in inducing apoptosis by targeting mitochondria. *Cancer Biol. Ther.* 7, 376-384 (2008)

103. George R. Pettit, Noeleen Melody, Delbert L. Herald: Antineoplastic agents. 511. Direct phosphorylation of phenpanstatin and pancratistatin. *J. Nat. Prod.* 67, 322-327 (2004)

105. Xihong Wang, Susan L. Morris-Natschke, Kuo-Hsiung Lee: New developments in the chemistry and biology of the bioactive constituents of Tanshen. *Med. Res. Rev.* 27, 133-148 (2007)

105. Irina Tsoy Nizamutdinova, Gyeong Won Lee, Jong Sil Lee, Min Kyung Cho, Kun Ho Son, Su Jin Jeon, Sam Sik Kang, Yeong Shik Kim, Jae Heun Lee, Han Geuk Seo, Ki Churl Chang, Hye Jung Kim: Tanshinone I suppresses growth and invasion of human breast cancer cells, MDA-MB-231, through regulation of adhesion molecules. *Carcinogenesis* 29, 1885-1892 (2008)

106. Jia-Jun Liu, Wen-Da Liu, Hong-Zhi Yang, Yong Zhang, Zhi-Gang Fang, Pei-Qing Liu, Dong-Jun Lin, Ruo-Zhi Xiao, Yuan Hu, Chun-Zhi Wang, Xu-Dong Li, Yi He, Ren-Wei Huang: Inactivation of PI3k/Akt signaling pathway and activation of caspase-3 are involved in tanshinone I-induced apoptosis in myeloid leukemia cells *in vitro. Ann. Hematol.* 89, 1089-1097 (2010)

107. Tai-Long Pan, Yu-Chiang Hung, Pei-Wen Wang, Shui-Ten Chen, Teng-Kuei Hsu, Nardnisa Sintupisut, Chao-Sheng Cheng, Ping-Chiang Lyu: Functional proteomic and structural insights into molecular targets related to the growth inhibitory effect of tanshinone IIA on HeLa cells. *Proteomics* 10, 914-929 (2010).

108. Xu Yuxian, Tian Feng, Li Ren, Liu Zhengcai: Tanshinone II-A inhibits invasion and metastasis of human hepatocellular carcinoma cells *in vitro* and *in vivo*. *Tumori* 95, 789-795 (2009)

109. Wenxing Chen, Yan Luo, Lei Liu, Hongyu Zhou, Baoshan Xu, Xiuzhen Han, Tao Shen, Zhijun Liu, Yin Lu, Shile Huang: Cryptotanshinone inhibits cancer cell proliferation by suppressing mammalian target of rapamycin-mediated cyclin D1 expression and Rb phosphorylation. *Cancer Prev. Res.* 3, 1015-1025 (2010)

110. In-Ja Park, Min-Jung Kim, Ock Jin Park, Myoung Gyu Park, Wonchae Choe, Insug Kang, Sung-Soo Kim, Joohun Ha: Cryptotanshinone sensitizes DU145 prostate cancer cells to Fas(APO1/CD95)-mediated apoptosis through Bcl-2 and MAPK regulation. *Cancer Lett.* 298, 88-98 (2010)

111. Xihong Wang, Kenneth F. Bastow, Chang-Ming Sun, Yun-Lian Lin, Shi-Jung Yu, Ming-Jaw Don, Tian-Shung Wu, Seikou Nakamura, Kuo-Hsiung Lee: Antitumor agents. 239. Isolation, structure elucidation, total synthesis, and anti-breast cancer activity of neo-tanshinlactone from *Salvia miltiorrhiza*. *J. Med. Chem.* 47, 5816-5819 (2004)

112. Yizhou Dong, Qian Shi, Huei-Chen Pai, Chieh-Yu Peng, Shiow-Lin Pan, Che-Ming Teng, Kyoko Nakagawa-Goto, Donglei Yu, Yi-Nan Liu, Pei-Chi Wu, Kenneth F. Bastow, Susan L. Morris-Natschke, Arnold Brossi, Jing-Yu Lang, Jennifer L. Hsu, Mien-Chie Hung, Eva Y.-H. P. Lee, Kuo-Hsiung Lee: Antitumor agents. 272. Structure-activity relationships and *in vivo* selective anti-breast cancer activity of novel neo-tanshinlactone analogues. *J. Med. Chem.* 53, 2299-2308 (2010)

113. Bang Yeon Hwang, Bao-Ning Su, Heebyung Chai, Qiuwen Mi, Leonardus B. S. Kardono, Johar J. Afriastini, Soedarsono Riswan, Bernard D. Santarsiero, Andrew D. Mesecar, Robert Wild, Craig R. Fairchild, Gregory D. Vite, William C. Rose, Norman R. Farnsworth, Geoffrey A. Cordell, John M. Pezzuto, Steven M. Swanson, A. Douglas Kinghorn: Silvestrol and episilvestrol, potential anticancer rocaglate derivatives from *Aglaia silvestris. J. Org. Chem.* 69, 3350-3358, *ibid.* 6156 (2004)

114. Barbara Martha Meurer-Grimes, Jin Yu, Gino Luigi Vairo: Therapeutic compounds and methods. U.S patent 6710075 B2 (2004)

115. Baudouin Gerard, Regina Cencic, Jerry Pelletier, John A. Porco: Enantioselective synthesis of the complex rocaglate (-)- silvestrol. *Angew. Chem. Int. Ed.* 46, 7831-7834 (2007)

116. Mariana El Sous, Mui Ling Khoo, Georgina Holloway, David Owen, Peter J. Scammells, Mark A. Rizzacasa: Total synthesis of (-)-episilvestrol and (-)-silvestrol. *Angew. Chem. Int. Ed.* 46, 7835-7838 (2007)

117. Tim E. Adams, Mariana El Sous, Bill C. Hawkins, Sebastian Hirner, Georgina Holloway, Mui Ling Khoo, David J. Owen, G. Paul Savage, Peter J. Scammells, Mark A. Rizzacasa: Total synthesis of the potent anticancer aglaia metabolites (-)-silvestrol and (-)-episilvestrol and the active analogue (-)-4'-desmethoxyepisilvestrol. J. Am. Chem. Soc. 131, 1607-1616 (2009)

118. Qiuwen Mi, Soyoung Kim, Bang Yeon Hwang, Bao-Ning Su, Heebyung Chai, Zarema H. Arbieva, A. Douglas Kinghorn, Steven M. Swanson: Silvestrol regulates G2/M checkpoint genes independent of p53 activity. *Anticancer Res.* 26, 3349-3356 (2006)

119. Soyoung Kim, Bang Yeon Hwang, Bao-Ning Su, Heebyung Chai, Qiuwen Mi, A. Douglas Kinghorn, Robert Wild, Steven M. Swanson: Silvestrol, a potential anticancer rocaglate derivative from *Aglaia foveolata*, induces apoptosis in LNCaP cells through the mitochondrial/apoptosome pathway without activation of executioner caspase-3 or -7. *Anticancer Res.* 27, 2175-2183 (2007).

120. David M. Lucas, Ryan B. Edwards, Gerard Lozanski, Derek A. West, Jungook D. Shin, Melissa A. Vargo, Melanie E. Davis, Darlene M. Rozewski, Amy J. Johnson, Bao-Ning Su, Virginia M. Goettl, Nyla A. Heerema, Thomas S. Lin, Amy Lehman, Xiaoli Zhang, David Jarjoura, David J. Newman, John C. Byrd, A. Douglas Kinghorn, Michael R. Grever: The novel plant-derived agent silvestrol has B-cell selective activity in chronic lymphocytic leukemia and acute lymphoblastic leukemia *in vitro* and *in vivo*. *Blood* 113, 4656-4666 (2009)

121. Regina Cencic, Marilyn Carrier, Gabriela Galicia-Vázquez, Marie-Eve Bordeleau, Rami Sukarieh, Annie Bourdeau, Brigitte Brem, Jose G. Teodoro, Harald Greger, Michel L. Tremblay, John A. Porco Jr., Jerry Pelletier: Antitumor activity and mechanism of action of the cyclopenta[b]benzofuran, silvestrol. *PLoS ONE 4*, e5223 (2009)

122. Regina Cencica, Marilyn Carriera, Amanda Trnkusa, John A. Porco Jr.b, Mark Mindenc, Jerry Pelletier. Synergistic effect of inhibiting translation initiation in combination with cytotoxic agents in acute myelogenous leukemia cells. *Leukemia Res.* 34, 535-541 (2010).

123. Marie-Eve Bordeleau, Francis Robert, Baudouin Gerard, Lisa Lindqvist, Samuel M. H. Chen, Hans-Guido Wende, Brigitte Brem, Harald Greger, Scott W. Lowe, John A. Porco, Jr., Jerry Pelletier: Therapeutic suppression of translation initiation modulates chemosensitivity in a mouse lymphoma model. *J. Clin. Invest.* 118, 1-11(2008)

124. Alexander Kamb, Susan Wee, Christoph Lengauer: Why is cancer drug discovery so difficult? *Nat. Rev. Drug Disc.* 6, 115-120 (2007)

125. Luc Pieters, Arnold J. Vlietinck: Bioguided isolation of pharmacologically active plant components, still a valuable strategy for the finding of new lead compounds? *J. Ethnopharmacol.* 100, 57-60 (2005)

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