Persistent low-grade inflammation and regular exercise

Maj-Briit Astrom, Michael Feigh, Bente Klarlund Pedersen

The Centre of Inflammation and Metabolism, Department of Infectious Diseases and CMRC Copenhagen University Hospital, Rigshospitalet, University of Copenhagen, Faculty of Health Sciences, Denmark

TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. Exercise and chronic diseases
- 4. The effects of exercise on systemic and local inflammation
- 5. Exercise and abdominal adiposity
- 6. Acute exercise and anti-inflammation
- 7. The link between inflammation, insulin resistance and atherosclerosis
- 8. Conclusion
- 9. Acknowledgement
- 10. References

1. ABSTRACT

Persistent low-grade systemic inflammation is a feature of chronic diseases such as cardiovascular disease (CVD), type 2 diabetes and dementia and evidence exists that inflammation is a causal factor in the development of insulin resistance and atherosclerosis. Regular exercise offers protection against all of these diseases and recent evidence suggests that the protective effect of exercise may to some extent be ascribed to an anti-inflammatory effect of regular exercise. Visceral adiposity contributes to systemic inflammation and is independently associated with the occurrence of CVD, type 2 diabetes and dementia. We suggest that the anti-inflammatory effects of exercise may be mediated via a long-term effect of exercise leading to a reduction in visceral fat mass and/or by induction of anti-inflammatory cytokines with each bout of exercise.

2. INTRODUCTION

Over the past decade, there has been an increasing focus on the role of persistent low-grade inflammation in the pathogenesis of atherosclerosis (1-3). Further, inflammation has been suggested to be a key factor in insulin resistance (4, 5) and neurodegeneration (6, 7).

Persistent low-grade systemic inflammation is reflected by increased systemic levels of some cytokines (8), as well as C-reactive protein (CRP). Numerous reports investigating various markers of inflammation have confirmed an association between low-grade systemic inflammation on one hand and several chronic diseases on the other (9, 10).

Infiltration of immune cells into white adipose tissue, and subsequently local inflammation in fat tissue, is

correlated with the development of insulin resistance and type 2 diabetes (11). Similarly, activated immune cells and inflammation have an important role in cardiovascular diseases (12, 13) and local inflammation in the brain is a feature of Alzheimer's disease. In addition, it appears that systemic inflammation may further stimulate the progression of these neurological disorders (14).

3. EXERCISE AND CHRONIC DISEASES

Over the past several decades, numerous large cohort studies have attempted to quantify the protective effect of physical activity on cardiovascular and all-cause mortality. A recent meta-analysis included a total of 33 studies with 883.372 participants with a follow-up time of up to more than 20 years. Concerning cardiovascular mortality, physical activity was associated with a risk reduction of 35%, whereas all-cause mortality was reduced by 33% (15). The risk reduction is, at least in part, attributed to the favourable effect of physical activity on the cardiovascular risk factors. Increased physical activity lowers blood pressure in hypertensive individuals, increases high-density lipoprotein cholesterol in a doseresponse fashion, and reduces the incidence of diabetes (16). Taken together, there is no doubt that physical activity is independently associated with a marked decrease in risk of cardiovascular disease (CVD) as well as CVD mortality in both men and women.

Randomised controlled trials including people with impaired glucose tolerance have found that lifestyle modification (diet and moderate physical activity) protects against the development of type 2 diabetes. A Finnish trial randomised 522 overweight middle-aged people with impaired glucose tolerance to physical training combined with diet or to control and followed them for 3.2 years (17). The risk of type 2 diabetes was reduced by 58% in the intervention group. The effect was greatest in the patients who made the greatest lifestyle modification. An American trial randomised 3,234 people with impaired glucose tolerance to either treatment with metformin, lifestyle modification entailing dietary change and at least 150 minutes of physical exercise weekly, or placebo, and followed them for 2.8 years (18). The lifestyle modification reduced the risk of type 2 diabetes by 58%. The reduction was thus the same as in the Finnish trial (17), whereas treatment with metformin only reduced the risk of diabetes by 31%. After a median of 4 years of active intervention period, participants in the Finish study who were still free of diabetes were further followed up for a median of 3 years, with median total follow-up of 7 years. During the total follow-up, the incidence of type 2 diabetes was 4.3 and 7.4 per 100 person-years in the intervention and control group, respectively, indicating 43% reduction in relative risk. The risk reduction was related to the success in achieving the intervention goals of weight loss, reduced intake of total and saturated fat and increased intake of dietary fibre, and increased physical activity (19).

Also, the beneficial effect of training in patients, who have been diagnosed with type 2 diabetes, is well documented (20). Post-intervention HbA1c was lower in the exercise groups than in the control groups (7.65% versus 8.31%). In comparison, intensive glycaemic control with metformin reduced HbA1c by 0.6% (21). A metaanalysis encompassing 95,783 non-diabetic individuals showed that cardiovascular morbidity is strongly correlated to fasting blood glucose (22). The effect of physical training on HbA1c is thus highly clinically relevant.

In humans, type 2 diabetes is associated with impaired cognitive function, including learning, memory, and processing speed (23). Large longitudinal populationbased studies show that the rate of cognitive decline is accelerated in elderly people with type 2 diabetes (24). A recent review (25) showed that the incidence of 'any dementia' was higher in individuals with type 2 diabetes than in those without. This high risk included both Alzheimer's disease and vascular dementia.

Interestingly, a couple of studies suggest that regular exercise also protects against dementia (26-28).

Chronic inflammation accompanies CVD, type 2 diabetes and dementia, potentially explaining the clustering of these conditions in epidemiological studies. The fact that regular exercise offers protection against these diseases further suggest that the beneficial effects of exercise may at least in part be mediated by anti-inflammatory mechanisms.

4. THE EFFECTS OF EXERCISE ON SYSTEMIC AND LOCAL INFLAMMATION

Regular exercise appears to induce antiinflammatory effects. An association between physical inactivity and low-grade systemic inflammation has been demonstrated in cross-sectional studies, including healthy younger individuals (29-34), elderly people (35), as well as in patients with intermittent claudication (36). However, also longitudinal studies show that regular training induces a reduction in CRP level (37, 38), suggesting that physical activity *per se* may suppress systemic low-grade inflammation. Several studies have shown that markers of inflammation are reduced following longer-term behavioural changes involving both reduced energy intake and increased physical activity (39-46).

In the Finish diabetes prevention study, lifestyle interventions reduced circulating levels of CRP and IL-6. Increases in fibre intake and moderate to vigorous leisure time physical activity but not total leisure time physical activity, predicted decreases in CRP and/or IL-6 and remained associated even after adjustment for baseline BMI or changes in BMI during the first year of the study. Changes in carbohydrate or fat intake were either weakly or not linked to reductions in CRP and IL-6 (47).

It appears that exercise may have antiinflammatory effects, which are independent of weightloss. However, the mediators of this effect are unclear. A number of mechanisms have been identified. Exercise increases the release of epinephrine, cortisol, growth hormone, prolactin and other factors that have immunomodulatory effects (48). Furthermore, exercise results in decreased expression of Toll-like receptor on monocytes suggested being involved in mediating whole body inflammation (49).

A recent study demonstrates that exercise training decreases expression of inflammation-related adipokines through reduction of oxidative stress in rat white adipose tissue (50).

In the latter study, it was shown that the levels of inflammation-related adipokines, such as tumor necrosis factor-alpha and monocytes chemo attractant protein-1, in white adipose tissue of trained rats were lower than those in sedentary rats. Interestingly, the effects of exercise training were more remarkable in visceral fat than in subcutaneous fat. Another experimental study showed that training increased the IL-10/TNF-alpha ratio in rat adipose tissue (51). They furthermore showed that the mesenteric depot was more responsive to moderate intensity exercise training than the retroperitoneal pad, and that such heterogeneity of response was present also in healthy animals.

Furthermore, it was demonstrated that the expression of TNF-, MCP-1, PAI-1 and IKK β was increased in adipose tissue from mice on high fat/high sucrose diet compared with chow mice, whereas exercise training reversed the increased expression of these inflammatory cytokines (52).

A study evaluated the effect of 12 wk of exercise (aerobic and resistance) or 12 wk of weight loss (7% reduction) in obese individuals. They found that exercise resulted in a 37% decrease in TLR-4 mRNA while weight loss had no significant effect. Additionally, exercise led to a 50% decrease in IL-6 and TNF- mRNA in muscle, while weight loss had no effect (53). The latter study confirmed that exercise but not weight loss had a beneficial effect on markers of muscle inflammation in frail obese elderly individuals.

5. EXERCISE AND ABDOMINAL ADIPOSITY

Abdominal adiposity is associated with cardiovascular disease (54), type 2 diabetes (55), and dementia (56). Moreover, abdominal adiposity is directly associated with all-cause and cardiovascular disease mortality independently of body mass index, even in people with a normal body weight (57). Abdominal adiposity reflects most often accumulation of visceral fat, which is more inflamed than subcutaneous fat tissue (58, 59). Moreover, the inflammation of the visceral fat is thought to be a major cause of the systemic low-grade inflammation (60). It is therefore obvious that regular exercise may mediate anti-inflammatory effects simply by reducing visceral adipose tissue mass.

A number of studies suggest that physical activity reduces abdominal and particularly visceral fat in healthy overweight and obese men and women, independently of changes in dietary energy intake (61). In accordance, a recent longitudinal study showed that when women go through the menopausal transition, they have deleterious changes in inflammatory markers and adipokines that correlate with increased visceral adiposity (62).

Importantly, however, another study concluded that in early postmenopausal women, the level of physical activity accounts for the variability in abdominal fat distribution observed, while menopausal status and age do not play a significant role (63).

Our group recently conducted a longitudinal study, in which a group of young healthy men decreased their daily stepping for 2 weeks to 1500 steps from the range recommended for US adults of around 10 000 steps. In this time, they experienced a 7% increase in intraabdominal fat mass without a change in total fat mass and while total fat-free mass decreased. Moreover, they developed metabolic changes suggestive of decreased insulin sensitivity and attenuation of postprandial lipid metabolism. Accordingly, the anti-inflammatory effects of regular exercise may be mediated, at least in part, by an effect of exercise on visceral fat.

6. ACUTE EXERCISE AND ANTI-INFLAMMATION

To study whether acute exercise induces a true anti-inflammatory response, a model of "low grade inflammation" was established in which a low dose of E. Coli endotoxin to healthy volunteers, who had been randomised to either rest or exercise prior to endotoxin administration. In resting subjects, endotoxin induced a 2 to 3 fold increase in circulating levels of TNF- α . In contrast, when the subjects performed 3 h of ergo meter cycling and received the endotoxin bolus at 2.5 h, the TNF- α response was totally blunted (64). This study provides some evidence that acute exercise may inhibit TNF production.

The cytokine response to exercise differs from that elicited by severe infections (65, 66). The fact that the classical pro-inflammatory cytokines, TNF- α and IL-1 β , in general do not increase with exercise indicates that the cytokine cascade induced by exercise markedly differs from the cytokine cascade induced by infections. Typically, IL-6 is the first cytokine released into the circulation during exercise. The level of circulating IL-6 increases in an exponential fashion (up to 100 fold) in response to exercise, and declines in the post-exercise period (67, 68).

The circulating levels of well-known antiinflammatory cytokines such as IL-1ra and IL-10 also increase after exercise. Taken together, exercise provokes an increase primarily in IL-6, followed by an increase in IL-1ra and IL-10. The appearance of IL-6 in the circulation is by far the most marked and its appearance precedes that of the other cytokines and a number of studies have demonstrated that contracting skeletal muscle fibers per se produce and release IL-6. Moreover, it appears that musclederived IL-6 can account for most of the systemic IL-6 response to exercise (67-69). Recent work has shown that both up-stream and down-stream signalling pathways for IL-6 differ markedly between myocytes and macrophages. It appears that unlike IL-6 signalling in macrophages, which are dependent upon activation of the NF κ B signalling pathway, intramuscular IL-6 expression is regulated by a network of signalling cascades that among other pathways are likely to involve cross talk between the Ca2+/NFAT and glycogen/p38 MAPK pathways (70).

The possibility exists that with regular exercise, the anti-inflammatory effects of an acute bout of exercise will protect against chronic systemic low-grade inflammation, but such a direct link between the acute effects of exercise and the long-term benefits has not yet been proven.

7. THE LINK BETWEEN INFLAMMATION, INSULIN RESISTANCE AND ATHEROSCLEROSIS

Mounting evidence suggests that TNF- α plays a direct role in the metabolic syndrome. Patients with diabetes demonstrate high protein expression of TNF- α in skeletal muscle (71) and increased TNF- α levels in plasma (72-74), and it is likely that adipose tissue, which produces TNF- α , is the main source of the circulating TNF- α (75, 76). Mounting evidence points to an effect of TNF- α on insulin signaling. TNF- α impairs insulin-stimulated rates of glucose storage in cultured human muscle cells (77) and impairs insulin mediated glucose uptake in rats (78). Obese mice with a gene knockout of TNF- α are protected from insulin resistance (79) and inhibition of TNF- α with an anti-TNF- α antibody treatment improves the insulin sensitivity in the insulin resistance rat model (80). In vitro studies demonstrate that TNF- α has direct inhibitory effects on insulin signalling (81-83). Recently, it was demonstrated that TNF- α infusion in healthy humans induces insulin resistance in skeletal muscle, without an effect on endogenous glucose production. TNF- α directly impaired glucose uptake and metabolism by altering insulin signal transduction. These data provide a *direct* molecular link between low-grade systemic inflammation and insulin resistance (84).

It has also been proposed that TNF- α indirectly causes insulin resistance *in vivo* by increasing the release of free fatty acids (FFA) from adipose tissue (85-90). TNF- α increases lipolysis in human (90, 91), rat (87, 92) and 3T3-L1 adipocytes (93-95). Recently, it was found that TNF- α had no effect on muscle fatty acid oxidation, but increased fatty acid incorporation into diacylglycerol, which may be involved in the development of TNF- α -induced insulin resistance in skeletal muscle (96).

Recent evidence suggests that TNF- α plays a key role in linking insulin resistance to vascular disease (97, 98). Several downstream mediators and signalling pathways seem to provide the crosstalk between inflammatory and metabolic signalling. These include the discovery of c-Jun N-terminal kinase (JNK) and I kappa beta kinase (IkK) as critical regulators of insulin action activated by TNF- α (99). In human TNF- α infusion studies, TNF- α increases phosphorylation of p70 S6 kinase, extra cellular signal-regulated kinase-1/2, and c-Jun NH (2)-terminal kinase, concomitant with increased serine and reduced tyrosine phosphorylation of insulin receptor substrate-1. These signalling effects are associated with impaired phosphorylation of Akt substrate 160, the most proximal step identified in the insulin signalling cascade regulating GLUT4 translocation and glucose uptake (84).

With regard to IL-6, its role in insulin resistance is highly controversial. In humans, circulating IL-6 levels may (100) or may not (101, 102) be associated with insulin resistance. Infusion of rhIL-6 into resting healthy humans does not impair whole body, lower limb or subcutaneous adipose tissue glucose uptake or endogenous glucose production (EGP) (103), although IL-6 contributes to the contraction-induced increase in endogenous glucose production (104, 105). When diabetes patients were given recombinant human (rh) IL6-infusion, plasma concentrations of insulin declined to levels comparable with that in age and BMI-matched healthy controls, indicating that the IL-6 enhanced insulin sensitivity (106). In vitro studies demonstrate that IL-6 can induce insulin resistance in isolated 3T3-L1 adipocytes (107) and in mice (108). Interestingly, IL-6 knockout mice develop impaired glucose tolerance, which is reverted by IL-6 (109). Thus, accumulating data suggest that IL-6 enhances glucose uptake in myocytes.

AMP-activated protein kinase (AMPK) activity stimulates a variety of processes that increases ATP generation including fatty acid oxidation and glucose transport in skeletal muscle (110). Incubation with IL-6 increases the phosphorylation of AMPK (an indicator of its activation) and that of its target molecule, acetyl CoA carboxylase (ACC) in skeletal muscles. In addition, AMPK activity and ACC levels are very low in IL-6 knockout mice, suggesting a role of IL-6 in the regulation of AMPK activity. These data suggest that IL-6 activation of AMPK is dependent on the presence of IL-6 (111).

A number of studies indicate that IL-6 enhances lipolysis (112-115), as well as fat oxidation (115). Consistent with this idea, Wallenius *et al* (109) demonstrated that IL-6 deficient mice developed matureonset obesity and insulin resistance. In addition, when the mice were treated with IL-6, there was a significant decrease in body fat mass in the IL-6 knockout, but not in the wild-type mice. To determine whether physiological concentrations of IL-6 affected lipid metabolism, our group administered physiological concentrations of rhIL-6 to healthy young and elderly humans as well as patients with type 2 diabetes (103, 106). The latter studies identified IL-6 as a potent modulator of fat metabolism in humans, increasing lipolysis as well as fat oxidation without causing hypertriacylglycerolemia.

Of note, whereas it is known that both TNF- α and IL-6 induce lipolysis, only IL-6 appears to induce fat oxidation. High levels of IL-6 and TNF- α in patients with the metabolic syndrome is associated with truncal fat mass (116) and both TNF- α and IL-6 are produced in adipose tissue (117-120). Given the different biological profiles of TNF- α and IL-6 and given that TNF- α can trigger IL-6

release, one theory holds that it is TNF- α derived from adipose tissue that actually is the "driver" behind insulin resistance and cardiovascular diseases and that locally produced TNF- α cause's increased systemic levels of IL-6.

8. CONCLUSION

Several chronic diseases are associated with chronic low-grade inflammation and evidence exists that inflammation is a causal factor in the development of insulin resistance and atherosclerosis.

Visceral fat is inflamed and appears to contribute to systemic inflammation. On the other hand, evidence suggests that regular exercise protects against systemic inflammation and induces anti-inflammatory effects. We suggest that anti-inflammatory effects of exercise may be mediated via a long-term effect on abdominal adiposity and/or by elevation of anti-inflammatory cytokines following acute bouts of exercise.

9. ACKNOWLEDGEMENTS

The Centre of Inflammation and Metabolism is supported by a grant from the Danish National Research Foundation (# 02-512-55). The study was further supported by the Danish Medical Research Council (# 22-01-009) and from the Commission of the European Communities (Contract No LSHM-CT-2004-005272 EXGENESIS). The Copenhagen Muscle Research Centre is supported by grants from The Copenhagen Hospital Corporation, The University of Copenhagen, The Faculties of Science and of Health Sciences at this University.

10. REFERENCES

1. Libby P.: Inflammation in atherosclerosis. *Nature* 420, 868-74 (2002)

2. Libby P., P.M. Ridker & A. Maseri: Inflammation and atherosclerosis. *Circulation* 105, 1135-43 (2002)

3. Shishehbor M.H. & D.L. Bhatt: Inflammation and atherosclerosis. *Curr Atheroscler Rep* 6, 131-9 (2004)

4. Dandona P., A. Aljada & A. Bandyopadhyay: Inflammation: the link between insulin resistance, obesity and diabetes. *Trends Immunol* 25, 4-7 (2004)

5. Sjoholm A., & T. Nystrom: Inflammation and the aetiology of type 2 diabetes. *Diabetes Metab Res Rev* 22, 4-10 (2006)

6. Zipp F., & O. Aktas: The brain as a target of inflammation: common pathways link inflammatory and neurodegenerative diseases. *Trends in Neurosciences* 29, 518-27 (2006)

7. Cotman C.W., N.C. Berchtold & L.A. Christie: Exercise builds brain health: key roles of growth factor cascades and inflammation. *Trends in Neurosciences* 30, 464-72 (2007)

8. Ross R.: Atherosclerosis - an inflammatory disease. *New Engl J med* 340, 115-26 (1999)

9. Festa A., R. D'Agostino, R.P. Jr. Tracy & S.M. Haffner: Elevated levels of acute-phase proteins and plasminogen activator inhibitor-1 predict the development of type 2 diabetes: the insulin resistance atherosclerosis study. *Diabetes* 51, 1131-7, (2002)

10. Handschin C., & B.M. Spiegelman: The role of exercise and PGC1 (alpha) in inflammation and chronic disease. *Nature* 454, 463-9 (2008)

11. Hotamisligil G.S.: Inflammation and metabolic disorders. *Nature* 444, 860-7 (2006)

12. Haffner S.M.: The Metabolic Syndrome: Inflammation, Diabetes Mellitus, and Cardiovascular Disease. *The American Journal of Cardiology* 97, 3-11 (2006)

13. Matter C.M., & C. Handschin: RANTES (Regulated on Activation, Normal T Cell Expressed and Secreted), Inflammation, Obesity, and the Metabolic Syndrome. *Circulation* 115, 946-8 (2007)

14. Perry V.H., C. Cunningham & C. Holmes: Systemic infections and inflammation affect chronic neurodegeneration. *Nat Rev Immunol* 7, 161-7 (2007)

15. Nocon M., T. Hiemann, F. Muller-Riemenschneider, F. Thalau, S. Roll & S.N. Willich: Association of physical activity with all-cause and cardiovascular mortality: a systematic review and meta-analysis. *Eur J Cardiovasc Prev Rehabil* 15, 239-46 (2008)

16. Kokkinos P.: Physical Activity and Cardiovascular Disease Prevention: Current Recommendations. *Angiology* 59, 26S-29S (2008)

17. Tuomilehto J., J. Lindstrom, J.G. Eriksson, T.T. Valle, H. Hamalainen, P. Ilanne-Parikka, S. Keinänen-Kiukaanniemi, M. Laakso, A. Louheranta, M. Rastas, V. Salminen & M. Uusitupa: Prevention of type 2 diabetes mellitus by changes in lifestyle among subjects with impaired glucose tolerance. *N Engl J Med* 344, 1343-50 (2001)

18. Knowler W.C., E. Barrett-Connor, S.E. Fowler, R.F. Hamman, J.M. Lachin, E.A. Walker & D.M. Nathan: Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. *N Engl J Med* 346, 393-403 (2002)

19. Lindstrom J., P. Ilanne-Parikka, M. Peltonen, S. Aunola, J.G. Eriksson, K. Hemio, H. Hämäläinen, P. Härkönen, S. Keinänen-Kiukaanniemi, M. Laakso, A. Louheranta, M. Mannelin, M. Paturi, J. Sundvall, T.T. Valle, M. Uusitupa & J. Tuomilehto: Sustained reduction in the incidence of type 2 diabetes by lifestyle intervention: follow-up of the Finnish Diabetes Prevention Study. *Lancet* 368, 1673-9 (2006)

20. Boule N.G., E. Haddad, G.P. Kenny, G.A. Wells & R.J. Sigal: Effects of exercise on glycaemic control and body mass in type 2 diabetes mellitus: a meta-analysis of controlled clinical trials. *JAMA* 286, 1218-27 (2001)

21. UK Prospective Diabetes Study (UKPDS) Group. Effect of intensive blood-glucose control with metformin

on complications in overweight patients with type 2 diabetes (UKPDS 34). *Lancet* 352, 854-65 (1998)

22. Coutinho M., H.C. Gerstein, Y. Wang & S. Yusuf: The relationship between glucose and incident cardiovascular events. A metaregression analysis of published data from 20 studies of 95,783 individuals followed for 12.4 years. *Diabetes Care* 22, 233-40 (1999)

23. Bruunsgaard H., M. Pedersen & B.K. Pedersen: Aging and proinflammatory cytokines. *Curr Opin Hematol* 8, 131-6 (2001)

24. Allen K.V., B.M. Frier, & M.W. Strachan: The relationship between type 2 diabetes and cognitive dysfunction: longitudinal studies and their methodological limitations. *Eur J Pharmacol* 490, 169-75 (2004)

25. Biessels G.J., S. Staekenborg, E. Brunner, C. Brayne & P. Scheltens: Risk of dementia in diabetes mellitus: a systematic review. *Lancet Neurol* 5, 64-74 (2006)

26. Rovio S., I. Kareholt, E.L. Helkala, M. Viitanen, B. Winblad, J. Tuomilehto, H. Soininen, A. Nissinen & M. Kivipelto: Leisure-time physical activity at midlife and the risk of dementia and Alzheimer's disease. *Lancet Neurol* 4, 705-11 (2005)

27. Andel R., M. Crowe, N.L. Pedersen, L. Fratiglioni, B. Johansson & M. Gatz: Physical Exercise at Midlife and Risk of Dementia Three Decades Later: a population-based study of Swedish twins. *J Gerontol A Biol Sci Med Sci* 63, 62-6 (2008)

28. Lautenschlager N.T., K.L. Cox, L. Flicker, J.K. Foster, F.M. van Bockxmeer, J. Xiao, K.R. Greenop & O.P. Almeida: Effect of physical activity on cognitive function in older adults at risk for Alzheimer disease: a randomized trial. *JAMA* 300, 1027-37 (2008)

29. Abramson J.L. & V. Vaccarino: Relationship between physical activity and inflammation among apparently healthy middle-aged and older US adults. *Arch Intern Med* 162, 1286-92 (2002)

30. Geffken D.F., M. Cushman, G.L. Burke, J.F. Polak, P.A. Sakkinen & R.P. Tracy: Association between physical activity and markers of inflammation in a healthy elderly population. *Am J Epidemiol* 153, 242-50 (2001)

31. King D.E., P. Carek, A.G. Mainous III & W.S. Pearson: Inflammatory markers and exercise: differences related to exercise type. *Med Sci Sports Exerc* 35, 575-81 (2003)

32. Smith J.K., R. Dykes, J.E. Douglas, G. Krishnaswamy & S. Berk: Long-term exercise and atherogenic activity of blood mononuclear cells in persons at risk of developing ischemic heart disease. *JAMA* 281, 1722-7 (1999)

33. Taaffe D.R., T.B. Harris, L. Ferrucci, J. Rowe & T.E. Seeman: Cross-sectional and prospective relationships of interleukin-6 and C-reactive protein with physical

performance in elderly persons: MacArthur studies of successful aging. *J Gerontol A Biol Sci Med Sci* 55, M709-M715 (2000)

34. Wannamethee S.G., G.D. Lowe, P.H. Whincup, A. Rumley, M. Walker & L. Lennon: Physical activity and haemostatic and inflammatory variables in elderly men. *Circulation* 105, 1785-90 (2002)

35. Bruunsgaard H., S. Ladelund, A.N. Pedersen, M. Schroll, T. Jorgensen & B.K. Pedersen: Predicting death from tumour necrosis factor-alpha and interleukin-6 in 80-year-old people. Clin Exp Immunol 132, 24-31 (2003)

36. Tisi P.V., M. Hulse, A. Chulakadabba, P. Gosling & C.P. Shearman: Exercise training for intermittent claudication: does it adversely affect biochemical markers of the exercise-induced inflammatory response? Eur J Vasc Endovasc Surg 14, 344-50 (1997)

37. Fallon K.E., S.K. Fallon & T. Boston: The acute phase response and exercise: court and field sports. Br J Sports Med 35, 170-3 (2001)

38. Mattusch F., B. Dufaux, O. Heine, I. Mertens & R. Rost: Reduction of the plasma concentration of C-reactive protein following nine months of endurance training. Int J Sports Med 21, 21-4 (2000)

39. Tchernof A., A. Nolan, C.K. Sites, P.A. Ades & E.T. Poehlman: Weight Loss Reduces C-reactive protein Levels in Obese Postmenopausal Women. Circulation 105, 564-9 (2002)

40. Ziccardi P., F. Nappo, G. Giugliano, K. Esposito, R. Marfella, M. Cioffi, F. D'Andrea, A.M. Molinari & D. Giugliano: Reduction of Inflammatory Cytokine Concentrations and Improvement of Endothelial Functions in Obese Women After Weight Loss Over One Year. Circulation 105, 804-9 (2002)

41. Roytblat L., M. Rachinsky, A. Fisher, L. Greemberg, Y. Shapira, A. Douvdevani & S. Gelman: Raised Interleukin-6 Levels in Obese Patients. Obesity 8, 673-5 (2000)

42. Marfella R., K. Esposito, M. Siniscalchi, F. Cacciapuoti, F. Giugliano, D. Labriola, M. Ciotola, C. Di Palo, L. Misso & D. Giugliano: Effect of Weight Loss on Cardiac Synchronization and Proinflammatory Cytokines in Premenopausal Obese Women. Diabetes Care 27, 47-52 (2004)

43. Dandona P., R. Weinstock, K. Thusu, E. bdel-Rahman, A. Aljada & T. Wadden: Tumor Necrosis Factor-alpha in Sera of Obese Patients: Fall with Weight Loss. Journal of Clinical Endocrinology Metabolism 83, 2907-10 (1998)

44. Esposito K., A. Pontillo, M. Ciotola, C. Di Palo, E. Grella, G. Nicoletti & D. Giugliano: Weight loss reduces interleukin-

18 levels in obese women. *J Clin Endocrinol Metab* 87, 3864-6 (2002)

45. Monzillo L.U., O. Hamdy, E.S. Horton, S. Ledbury, C. Mullooly, C. Jarema, S. Porter, K. Ovalle, A. Moussa & C.S. Mantzoros: Effect of Lifestyle Modification on Adipokine Levels in Obese Subjects with Insulin Resistance. *Obesity* 11, 1048-54 (2003)

46. Seshadri P., N. Iqbal, L. Stern, M. Williams, K.L. Chicano, D.A. Daily, J. McGrory, E.J. Gracely, D.J. Rader & F.F. Samaha: A randomized study comparing the effects of a low-carbohydrate diet and a conventional diet on lipoprotein subfractions and C-reactive protein levels in patients with severe obesity. *The American Journal of Medicine* 117, 398-405 (2004)

47. Herder C., M. Peltonen, W. Koenig, K. Sutfels, J. Lindstrom, S. Martin, P. Ilanne-Parikka, J.G. Eriksson, S. Aunola, S. Keinänen-Kiukaanniemi, T.T. Valle, M. Uusitupa, H. Kolb & J. Tuomilehto: Finnish Diabetes Prevention Study Group: Anti-inflammatory effect of lifestyle changes in the Finnish Diabetes Prevention Study. *Diabetologia* 52, 433-42 (2009)

48. Nieman D.C.: Current perspective on exercise immunology. *Curr Sports Med Rep* 2, 239-42 (2003)

49. Gleeson M.: Mucosal immune responses and risk of respiratory illness in elite athletes. *Exerc Immunol Rev* 6, 5-42 (2000)

50. Sakurai T, T. Izawa, T. Kizaki, J.E. Ogasawara, K. Shirato, K. Imaizumi, K. Takahashi, H. Ishida & H. Ohno : Exercise training decreases expression of inflammation-related adipokines through reduction of oxidative stress in rat white adipose tissue. *Biochemical and Biophysical Research Communications* 379, 605-9 (2009)

51. Lira F.S., J.C. Rosa, A.S. Yamashita, C.H. Koyama, J. Batista & M. Seelaender: Endurance training induces depot-specific changes in IL-10/TNF- (alpha) ratio in rat adipose tissue. *Cytokine* 45, 80-5 (2009)

52. Bradley R.L., J.Y. Jeon, F.F. Liu & E. Maratos-Flier: Voluntary exercise improves insulin sensitivity and adipose tissue inflammation in diet-induced obese mice. *Am J Physiol Endocrinol Metab* 295, E586-E594 (2008)

53. Lambert C.P., N.R. Wright, B.N. Finck & D.T. Villareal: Exercise but not diet-induced weight loss decreases skeletal muscle inflammatory gene expression in frail obese elderly persons. *Journal of Applied Physiology* 105, 473-8 (2008)

54. Zhang C., K.M. Rexrode, R.M. van Dam, T.Y. Li & F.B. Hu: Abdominal Obesity and the Risk of All-Cause, Cardiovascular, and Cancer Mortality: Sixteen Years of Follow-Up in US Women. *Circulation* 117, 1658-67 (2008)

55. Castell L.M., J.R. Poortmans, R. Leclercq, M. Brasseur, J. Duchateau & E.A. Newsholme: Some aspects of the

acute phase response after a marathon race, and the effects of glutamine supplementation. *Eur J Appl Physiol Occup Physiol* 75, 47-53 (1997)

56. Beydoun M.A., H.A. Beydoun & Y. Wang: Obesity and central obesity as risk factors for incident dementia and its subtypes: a systematic review and meta-analysis. *Obes Rev* 9, 204-18 (2008)

57. Pischon T., H. Boeing, K. Hoffmann, M. Bergmann, M.B. Schulze, K. Overvad, Y.T. van der Schouw, E. Spencer, K.G. Moons, A. Tjønneland, J. Halkjaer, M.K. Jensen, J. Stegger, F. Clavel-Chapelon, M.C. Boutron-Ruault, V. Chajes, J. Linseisen, R. Kaaks, A. Trichopoulou, D. Trichopoulos, C. Bamia, S. Sieri, D. Palli, R. Tumino, P. Vineis, S. Panico, P.H. Peeters, A.M. May, H.B. Bueno-de-Mesquita, F.J. van Duijnhoven, G. Hallmans, L. Weinehall, J. Manjer, B. Hedblad, E. Lund, A. Agudo, L. Arriola, A. Barricarte, C. Navarro, C. Martinez, J.R. Quirós, T. Key, S. Bingham, K.T. Khaw, P. Boffetta, M. Jenab, P. Ferrari & E. Riboli: General and Abdominal Adiposity and Risk of Death in Europe. *The New England Journal of Medicine* 359, 2105-20 (2008)

58. You T. & B.J. Nicklas: Chronic inflammation: role of adipose tissue and modulation by weight loss. *Curr Diabetes Rev* 2, 29-37 (2006)

59. Yudkin J.S.: Inflammation, obesity, and the metabolic syndrome. *Horm Metab Res* 39, 707-9 (2007)

60. Mohamed-Ali V., S. Goodrick, A. Rawesh, D.R. Katz, J.M. Miles & J.S. Yudkin, S. Klein & S.W. Coppack: Subcutaneous adipose tissue releases interleukin-6, but not tumor necrosis factor-alpha, in vivo. *J Clin Endocrinol Metab* 82, 4196-200 (1997)

61. Kay S.J., M.A. Fiatarone Singh: The influence of physical activity on abdominal fat: a systematic review of the literature. *Obes Rev* 7, 183-200 (2006)

62. Lee C.G., M.C. Carr, S.J. Murdoch, E. Mitchell, N.F. Woods, M.H. Wener, W.L. Chandler, E.J. Boyko & J.D. Brunzell: Adipokines, Inflammation, and Visceral Adiposity Across The Menopausal Transition: A Prospective Study. *Journal of Clinical Endocrinology Metabolism* Jan 6 (2009) Epub ahead.

63. Kanaley J.A., C. Sames, L. Swisher, A.G. Swick, L.L. Ploutz-Snyder, C.M. Steppan, K.S. Sagendorf, D. Feiglin, E.B. Jaynes, R.A. Meyer & R.S. Weinstock: Abdominal fat distribution in pre- and postmenopausal women: The impact of physical activity, age, and menopausal status. *Metabolism* 50, 976-82 (2001)

64. Starkie R., S.R. Ostrowski, S. Jauffred, M. Febbraio, B.K. Pedersen: Exercise and IL-6 infusion inhibit endotoxininduced TNF-alpha production in humans. *FASEB J* 17, 884-6 (2003)

65. Pedersen B.K. & L. Hoffman-Goetz: Exercise and the immune system: regulation, integration and adaptation. *Physiol Rev* 80, 1055-81 (2000)

66. Suzuki K., S. Nakaji, M. Yamada, M. Totsuka, K. Sato & K. Sugawara: Systemic inflammatory response to exhaustive exercise. Cytokine kinetics. *Exerc Immunol Rev* 8, 6-48 (2002)

67. Febbraio M.A. & B.K. Pedersen: Muscle-derived interleukin-6: mechanisms for activation and possible biological roles. *FASEB J* 16, 1335-47 (2002)

68. Pedersen B.K., A. Steensberg & P. Schjerling: Musclederived interleukin-6: possible biological effects. *J Physiol* 536, 329-37 (2001)

69. Pedersen B.K. & A.D. Toft: Effects of exercise on lymphocytes and cytokines. Br J Sports Med 34, 246-51 (2000)

70. Pedersen B.K. & M.A. Febbraio: Muscle as an endocrine organ - focus on muscle-derived IL-6. *Physiol Rev* 88, 1379-406 (2008)

71. Saghizadeh M., J.M. Ong, W.T. Garvey, R.R. Henry, & P.A. Kern: The expression of TNF alpha by human muscle. Relationship to insulin resistance. *J Clin Invest* 97, 1111-6 (1996)

72. Feingold K.R. & C. Grunfeld: Role of cytokines in inducing hyperlipidemia. *Diabetes* 41, 97-101 (1992)

73. Mishima Y., A. Kuyama, A. Tada, K. Takahashi, T. Ishioka & M. Kibata: Relationship between serum tumor necrosis factor-alpha and insulin resistance in obese men with Type 2 diabetes mellitus. *Diabetes Res Clin Pract* 52, 119-23 (2001)

74. Winkler G., F. Salamon, G. Harmos, D. Salamon, G. Speer, O. Szekeres, P. Hajós, M. Kovács, K. Simon & K. Cseh: Elevated serum tumor necrosis factor-alpha concentrations and bioactivity in Type 2 diabetics and patients with android type obesity. *Diabetes Res Clin Pract* 42, 169-74 (1998)

75. Coppack S.W.: Pro-inflammatory cytokines and adipose tissue. *Proc Nutr Soc* 60, 349-56 (2001)

76. Hotamisligil G.S., N.S. Shargill & B.M. Spiegelman: Adipose expression of tumor necrosis factor-alpha: direct role in obesity-linked insulin resistance. *Science* 259, 87-91 (1993)

77. Halse R., S.L. Pearson, J.G. McCormack, S.J. Yeaman & R. Taylor: Effects of tumor necrosis factor-alpha on insulin action in cultured human muscle cells. *Diabetes* 50, 1102-9 (2001)

78. Youd J.M., S. Rattigan & M.G. Clark: Acute impairment of insulin-mediated capillary recruitment and glucose uptake in rat skeletal muscle *in vivo* by TNF-alpha. *Diabetes* 49, 1904-9 (2000)

79. Staels B., W. Koenig, A. Habib, R. Merval, M. Lebret, I.P. Torra, P. Delerive, A. Fadel, G. Chinetti, J.C. Fruchart,

J. Najib, J. Maclouf & A. Tedgui: Activation of human aortic smooth-muscle cells is inhibited by PPARalpha but not by PPARgamma activators. *Nature* 393, 790-3 (1998)

80. Borst S.E. & G.J. Bagby: Neutralization of tumor necrosis factor reverses age-induced impairment of insulin responsiveness in skeletal muscle of Sprague-Dawley rats. *Metabolism* 51, 1061-4 (2002)

81. Hotamisligil G.S.: Mechanisms of TNF-alpha-induced insulin resistance. *Exp Clin Endocrinol Diabetes* 107, 119-25 (1999)

82. Hotamisligil G.S., P. Peraldi, A. Budavari, R. Ellis, M.F. White & B.M. Spiegelman: IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF-alpha- and obesity-induced insulin resistance. *Science* 271, 665-8 (1996)

83. Peraldi P., G.S. Hotamisligil, W.A. Buurman, M.F. White & B.M. Spiegelman: Tumor necrosis factor (TNF)-alpha inhibits insulin signalling through stimulation of the p55 TNF receptor and activation of sphingomyelinase. *J Biol Chem* 271, 13018-22 (1996)

84. Plomgaard P., K. Bouzakri, R. Krogh-Madsen, J.R. Zierath & B.K. Pedersen: TNF-alpha induces skeletal muscle insulin resistance in healthy human subjects via inhibition of AS160 phosphorylation. *Diabetes* 54, 2939-45 (2005)

85. Botion L.M., A.R. Brasier, B. Tian, V. Udupi & A. Green: Inhibition of proteasome activity blocks the ability of TNF alpha to down-regulate G (i) proteins and stimulate lipolysis. *Endocrinology* 142, 5069-75 (2001)

86. Gasic S., B. Tian & A. Green: Tumor necrosis factor alpha stimulates lipolysis in adipocytes by decreasing Gi protein concentrations. *J Biol Chem* 274, 6770-5 (1999)

87. Green A., S.B. Dobias, D.J. Walters & A.R. Brasier: Tumor necrosis factor increases the rate of lipolysis in primary cultures of adipocytes without altering levels of hormonesensitive lipase. *Endocrinology* 134, 2581-8 (1994)

88. Souza S.C., M.T. Yamamoto, M.D. Franciosa, P. Lien & A.S. Greenberg: BRL 49653 blocks the lipolytic actions of tumor necrosis factor-alpha: a potential new insulin-sensitizing mechanism for thiazolidinediones. *Diabetes* 47, 691-5 (1998)

89. Souza S.C., L.M. de Vargas, M.T. Yamamoto, P. Lien, M.D. Franciosa, L.G. Moss, A.S. Greenberg: Over expression of perilipin A and B blocks the ability of tumor necrosis factor alpha to increase lipolysis in 3T3-L1 adipocytes. *J Biol Chem* 273, 24665-9 (1998)

90. Ryden M., A. Dicker, V. van Harmelen, H. Hauner, M. Brunnberg, L. Perbeck, F. Lonnqvist & P. Arner: Mapping of early signalling events in tumor necrosis factor-alpha - mediated lipolysis in human fat cells. *J Biol Chem* 277, 1085-91 (2002)

91. Zhang H.H., M. Halbleib, F. Ahmad, V.C. Manganiello & A.S. Greenberg: Tumor necrosis factor-alpha stimulates

lipolysis in differentiated human adipocytes through activation of extracellular signal-related kinase and elevation of intracellular cAMP. *Diabetes* 51, 2929-35 (2002)

92. Botion L.M., A.R. Brasier, B. Tian, V. Udupi & A. Green: Inhibition of proteasome activity blocks the ability of TNF alpha to down-regulate G (i) proteins and stimulate lipolysis. *Endocrinology* 142, 5069-75 (2001)

93. Souza S.C., L.M. de Vargas, M.T. Yamamoto, P. Lien, M.D. Franciosa, L.G. Moss, A.S. Greenberg: Over expression of perilipin A and B blocks the ability of tumor necrosis factor alpha to increase lipolysis in 3T3-L1 adipocytes. *J Biol Chem* 273, 24665-9 (1998)

94. Ogawa H., S. Nielsen & M. Kawakami: Cachectin/tumor necrosis factor and interleukin-1 show different modes of combined effect on lipoprotein lipase activity and intracellular lipolysis in 3T3-L1 cells. *Biochim Biophys Acta* 1003, 131-5 (1989)

95. Rahn-Landstrom T., J. Mei, M. Karlsson, V. Manganiello & E. Degerman: Down-regulation of cyclicnucleotide phosphodiesterase 3B in 3T3-L1 adipocytes induced by tumour necrosis factor alpha and cAMP. *Biochem J* 346, 337-43 (2000)

96. Bruce C.R. & D.J. Dyck: Cytokine regulation of skeletal muscle fatty acid metabolism: effect of interleukin-6 and tumor necrosis factor-alpha. *Am J Physiol Endocrinol Metab* 287, E616-E621 (2004)

97. Yudkin J.S., E. Eringa & C.D. Stehouwer: "Vasocrine" signalling from perivascular fat: a mechanism linking insulin resistance to vascular disease. *Lancet* 365, 1817-20 (2005)

98. Plomgaard P., P. Keller, C. Keller & B.K. Pedersen: TNF-alpha, but not IL-6, stimulates plasminogen activator inhibitor-1 expression in human subcutaneous adipose tissue. *J Appl Physiol* 98, 2019-23 (2005)

99. Hotamisligil G.S.: Inflammatory pathways and insulin action. *Int J Obes Relat Metab Disord* 27, S53-S55 (2003)

100. Pickup J.C., M.B. Mattock, G.D. Chusney & D. Burt: NIDDM as a disease of the innate immune system: association of acute- phase reactants and interleukin-6 with metabolic syndrome X. *Diabetologia* 40, 1286-92 (1997)

101. Petersen K.F., S. Dufour, D. Befroy, R. Garcia & G.I. Shulman: Impaired mitochondrial activity in the insulinresistant offspring of patients with type 2 diabetes. *N Engl J Med* 350, 664-71 (2004)

102. Carey A.L., C.R. Bruce, M. Sacchetti, M.J. Anderson, D. Olson, B. Saltin, J.A. Hawley & M.A. Febbraio: Interleukin-6 and tumor necrosis factor-alpha are not increased in patients with type 2 diabetes: evidence that plasma IL-6 is related to fat mass and not insulin responsiveness. *Diabetologia* 47, 1029-37 (2004)

103. Steensberg A., C.P. Fischer, M. Sacchetti, C. Keller, T. Osada, P. Schjerling, G. van Hall, M.A. Febbraio & B.K. Pedersen: Acute interleukin-6 administration does not impair muscle glucose uptake or whole body glucose disposal in healthy humans. *J Physiol* 548, 631-8 (2003)

104. Lyngso D., L. Simonsen & J. Bulow: Interleukin-6 production in human subcutaneous abdominal adipose tissue: the effect of exercise. *J Physiol* 543, 373-8 (2002)

105. Febbraio M.A., N. Hiscock, M. Sacchetti, C.P. Fischer & B.K. Pedersen: Interleukin-6 is a novel factor mediating glucose homeostasis during skeletal muscle contraction. *Diabetes* 53, 1643-8 (2004)

106. Petersen E.W., A.L. Carey, M. Sacchetti, G.R. Steinberg, S.L. Macaulay, M.A. Febbraio, B.K. Pedersen: Acute IL-6 treatment increases fatty acid turnover in elderly humans *in vivo* and in tissue culture *in vitro*. *Am J Physiol* 288, E155-E162 (2005)

107. Rotter V., I. Nagaev & U. Smith: Interleukin-6 (IL-6) induces insulin resistance in 3T3-L1 adipocytes and is, like IL-8 and tumor necrosis factor-alpha, overexpressed in human fat cells from insulin-resistant subjects. *J Biol Chem* 278, 45777-84 (2003)

108. Kim H.J., T. Higashimori, S.Y. Park, H. Choi, J. Dong, Y.J. Kim, H.L. Noh, Y.R. Cho, G. Cline, Y.B. Kim & J.K. Kim: Differential effects of interleukin-6 and -10 on skeletal muscle and liver insulin action *in vivo*. *Diabetes* 53, 1060-7 (2004)

109. Wallenius V., K. Wallenius, B. Ahren, M. Rudling, H. Carlsten, S.L. Dickson, C. Ohlsson, J.O. Jansson: Interleukin-6-deficient mice develop mature-onset obesity. *Nat Med* 8, 75-9 (2002)

110. Carling D.: The AMP-activated protein kinase cascade--a unifying system for energy control. *Trends Biochem Sci* 29, 18-24 (2004)

111. Kelly M., C. Keller, P.R. Avilucea, P. Keller, Z. Luo, X. Xiang, M. Giralt, J. Hidalgo, A.K. Saha, B.K. Pedersen & N.B. Ruderman: AMPK activity is diminished in tissues of the IL-6 knockout mice: the effect of exercise. *Biochem Biophys Res Commun* 320, 449-54 (2004)

112. Stouthard J.M., J.A. Romijn, T. Van der Poll, E. Endert, S. Klein, P.J. Bakker, C.H. Veenhof & H.P. Sauerwein: Endocrinologic and metabolic effects of interleukin-6 in humans. *Am J Physiol* 268, E813-E819 (1995)

113. Path G., S.R. Bornstein, M. Gurniak, G.P. Chrousos, W.A. Scherbaum & H. Hauner: Human breast adipocytes express interleukin-6 (IL-6) and its receptor system: increased IL-6 production by beta-adrenergic activation and effects of IL-6 on adipocyte function. *J Clin Endocrinol Metab* 86, 2281-8 (2001)

114. Nonogaki K., G.M. Fuller, N.L. Fuentes, A.H. Moser, I. Staprans, C. Grunfeld & K.R. Feingold: Interleukin-6 stimulates hepatic triglyceride secretion in rats. *Endocrinology* 136, 2143-9 (1995)

115. van Hall G., A. Steensberg, M. Sacchetti, C. Fischer, C. Keller, P. Schjerling, N. Hiscock, K. Møller, B. Saltin B, M.A. Febbraio & B.K. Pedersen: Interleukin-6 stimulates lipolysis and fat oxidation in humans. *J Clin Endocrinol Metab* 88, 3005-10 (2003)

116. Pedersen M., H. Bruunsgaard, N. Weis, H.W. Hendel, B.U. Andreassen, E. Eldrup, F. Dela & B.K. Pedersen: Circulating levels of TNF-alpha and IL-6-relation to truncal fat mass and muscle mass in healthy elderly individuals and in patients with type-2 diabetes. *Mech Ageing Dev* 124, 495-502 (2003)

117. Mohamed-Ali V., S. Goodrick, K. Bulmer, J.M. Holly, J.S. Yudkin & S.W. Coppack: Production of soluble tumor necrosis factor receptors by human subcutaneous adipose tissue *in vivo. Am J Physiol* 277, E971-E975 (1999)

118. Fried S.K., D.A. Bunkin & A.S. Greenberg: Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. *J Clin Endocrinol Metab* 83, 847-50 (1998)

119. Coppack S.W.: Pro-inflammatory cytokines and adipose tissue. *Proc Nutr Soc* 60, 349-56 (2001)

120. Tsigos C., I. Kyrou, E. Chala, P. Tsapogas, J.C. Stavridis & S.A. Raptis: Circulating tumour necrosis factor alpha concentrations are higher in abdominal versus peripheral obesity. *Metabolism* 48, 1332-5 (1999)

Key Words: Anti-inflammation, Cytokines, Exercise, Atherosclerosis, Review

Send correspondence to: Bente Klarlund Pedersen, Centre of Inflammation and Metabolism – 7641, Rigshospitalet, Blegdamsvej 9, DK-2100 Copenhagen, Denmark, Tel: 4535457797, Fax: 4535457644, E-mail: bkp@rh.dk

http://www.bioscience.org/current/vol2S.htm