TUMOR-SUPPRESSIVE AND PROMOTING FUNCTION OF TRANSFORMING GROWTH FACTOR BETA

LuZhe Sun

Department of Cellular & Structural Biology, MC 7762, University of Texas Health Science Center, 7703 Floyd Curl Drive, San Antonio, TX 78229-3900

TABLE OF CONTENTS

1. Abstract

2. Introduction

*3. Tumor suppressive activity of TGF***b**

3.1. Disruption of TGFb signaling pathway promotes early stage tumor progression

3.2. The tumor suppressive activity of TGFb is mediated by its effect on cell cycle progression and apoptosis

4. Switch from tumor suppression to tumor promotion

4.1. Tumor cells often evade the growth inhibition by TGFb

4.2. TGF**b** signal promotes late stage tumor progression

5. Inhibition of tumor progression with TGFb antagonists

6. Perspectives

7. Acknowledgements

8. References

1. ABSTRACT

Transforming growth factor beta (TGF β) is a multifunctional polypeptide. Its role in carcinogenesis can be either suppressive or promoting depending on tumor developmental stages and cellular context. During the early phase of epithelial tumorigenesis, TGF^β inhibits primary tumor development and growth by inducing cell cycle arrest and possibly apoptosis. However, in late stages of progression, as tumor cells evade the growth inhibition by TGFB due to inactivation of its signaling pathway or aberrant regulation of cell cycle machinery, the role of TGFβ signaling is often switched from tumor suppression to promotion. TGF β can apparently act in tumor stroma as well as tumor cells to inhibit host immune surveillance and stimulate invasion, angiogenesis, and metastasis. Studies have shown that antagonizing TGFB activity can inhibit tumor progression, especially metastasis, in certain tumor models. However, the molecular markers that can indicate the feasibility of the use of TGF β antagonists as cancer therapeutics remain to be determined.

2. INTRODUCTION

Transforming growth factor beta (TGF β) is a homodimeric polypeptide of 25 kDa. Three TGF β isoforms termed TGF β_1 , β_2 , and β_3 have been identified in mammals. The mature TGF β s are derived from the Cterminal 112 amino acids of their respective precursors by proteolytic cleavage. All three TGF β isoforms are secreted in a latent form in that the mature TGF β s are noncovalently associated with the N-terminal remainder of their processed precursors. This latent complex cannot interact with the cell surface TGF β receptors and thus requires activation to release the mature, active TGF β through its interaction with various extracellular proteins (1). The active TGF β binds to three different cell surface receptors called type I (RI), type II (RII) and type III (RIII) receptors. RIII (also called betaglycan) has two TGF β binding sites in its extracellular domain and can sequester and present TGF β s to RII to augment their activity when it is membrane-bound (2-6). RI and RII are serine/threonine kinase receptors. Binding of TGF β to RII recruits and transphosphorylates RI (7). The activated RI phosphorylates intracellular Smad2 and Smad3, which then interact with Smad4 protein to form an oligomeric complex (8). Once transported into nuclei, Smad2/Smad4 and Smad3/Smad4 complexes bind to specific DNA sequences and can act as transcription activator or repressor depending on DNA sequence and cellular context (1, 9).

TGFBs are multifunctional polypeptides involved in the regulation of cell proliferation, differentiation, extracellular matrix formation, and immune response (8, 10, 11). TGFB was initially identified as a **transforming** growth factor in that it stimulated anchorage-independent growth of the NRK fibroblasts in the presence of epidermal growth factor (12). Later, it was found to be a potent growth inhibitor in various types of cells including epithelial cells (13). More recently, many studies have shown that TGF β and its signaling components can act to inhibit or promote tumor progression depending on stages of carcinogenesis or model systems used. This minireview is intended to furnish the readers current information on the research findings of the role of TGF β signaling in epithelial tumor progression and the utilization of TGF β antagonists in suppressing TGFB-induced tumor progression.

3. TUMOR SUPPRESSIVE ACTIVITY OF TGFb

3.1. Disruption of TGF**b** signaling pathway promotes early stage tumor progression

When TGF β was shown to act as an autocrine negative growth regulator in various epithelial cell lines (14-16), it was quickly hypothesized that TGF β isoforms may act as inhibitors of tumor progression. Indeed, antisense inhibition of TGFB expression was shown to stimulate tumor formation and growth of two welldifferentiated human colon adenocarcinoma xenograft models in mice (17, 18). Cloning of TGFB receptors and identification of Smad proteins as intracellular $TGF\beta$ signaling mediators have dramatically expanded our understanding of the tumor suppressive role of TGF β signaling pathway. TGFBR2, the gene that encodes RII, was found to be mutated in colon carcinoma cells from hereditary non-polyposis colorectal cancer patients (19). These patients are predisposed to frequent insertion or deletion of repeated mono- or di-nucleotide sequences due to germline mutations of their DNA mismatch repair enzymes, a phenomenon called microsatellite instability. The mutation of TGFBR2 occurs in its coding sequence containing a repeated segment of adenines and is also observed in gastric cancer cells with microsatellite instability (20). In other types of carcinoma with microsatellite instability such as breast, lung, pancreatic, and endometrial carcinoma, RII is not frequently mutated (20-22). However, its expression can be down-regulated during carcinogenesis in some cases (23-25). On the other hand, re-expression of RII in human carcinoma cells with loss of or reduced RII expression can inhibit their malignancy (26-28). Mutation of RI gene has been reported in ovarian cancer with a high frequency (29, 30), whereas its expression is transcriptionally repressed by DNA methylation in gastric cancer (31). Like RII, ectopic expression of RI in human carcinoma cells with a low level of cell surface RI also inhibited tumor progression (32).

Deletions and mutations of the genes of the Smad proteins that mediate TGF β signal transduction have also been observed in human carcinomas. Most notable is the mutational inactivation of *SMAD4*, also known as *DPC4* (deleted in pancreatic carcinomas), in human pancreatic cancer (33, 34). Mutations of *SMAD4* have also been described in human colorectal cancer, especially in late stage, metastatic colon cancer patients (35-37). In contrast, mutational inactivation of *SMAD2* have only been reported, whereas mutations of *SMAD2* have only been observed in a limited number of human colon and lung carcinomas (38, 39) and are apparently uncommon. These observations suggest a pivotal role of Smad4 in TGF β -induced tumor suppression.

The tumor suppressive activity of TGFB signaling pathway during the early stage of tumorigenesis has also been extensively demonstrated in experimental mice. Transgenic expression of active $TGF\beta_1$ in the mammary gland was shown to inhibit TGF α - or chemical carcinogen-induced tumor formation (40). Converselv, TGF β_1 heterozygous null mice expressed 10-30% of the wild-type $TGF\beta_1$ level and were more susceptible to chemical carcinogen-induced tumorigenesis (41). Similarly, disruption of TGF β signaling by the expression of a dominant negative RII in the mammary gland or the skin also enhanced chemical carcinogen-induced tumor formation and progression (42-44). Smad4 heterozygous null mice was shown to develop gastric and duodenal polyps, which can progress into invasive tumors with the loss of the other copy of SMAD4 (45, 46). Compound

Smad4 and APC (adenomatous polyposis coli) heterozygous null mice was shown to develop more malignant intestinal tumors than APC heterozygous null mice and the remaining copy of both *SMAD4* and *APC* was found to be lost in the malignant tumors (47). Thus, TGF β signaling components are necessary to protect epithelial tissues from tumorigenesis.

3.2. The tumor suppressive activity of TGF**b** is mediated by its effect on cell cycle progression and apoptosis

Although TGF β is a multifunctional cytokine involved in the regulation of many genes and cellular phenotypes, its tumor suppressive activity is widely attributed to its ability to regulate the expression of a number of key proteins in the control of cell cycle progression from G1 to S phase. One of them is the protooncogene, *c-myc*, which is known to promote cell cycle entry into S phase by regulating the transcription of various cell cycle related genes (48). TGF β treatment can rapidly inhibit the transcription of *c-myc* in epithelial cells (49-51). The inhibition is apparently accomplished by TGFBstimulated nuclear translocation of a transcription repression complex containing Smad3, E2F4/5 and the corepressor p107, and subsequent docking of this complex in association with Smad4 on a Smad/E2F binding site of cmyc promoter (52). On the other hand, TGF β can also stimulate the transcription of the cyclin-dependent kinase (CDK) inhibitors, $p15^{ink4b}$ and $p21^{cip1}$ (53, 54). The former interacts with CDK4 and CDK6 to inhibit their kinase activity and association with cyclin D, whereas the latter mainly inhibits the activities of cyclin A-CDK2 and cyclin E-CDK2. Both cyclin D-CDK4/6 and cycline E-CDK2 can phosphorylate retinoblastoma gene product pRB to inactivate its ability to block G1 to S phase transition. The transcriptional activation of $p15^{ink4b}$ and $p21^{cip1}$ by TGF β is through a synergistic interaction between Smad proteins and the Sp1 transcription factor at the promoter region of $p15^{ink4b}$ or $p21^{cip1}$ genes (54-57). It should also be noted that the binding of $p15^{ink4b}$ can also displace $p21^{cip1}$ and another CDK inhibitor p27kip1 associated with cyclin D-CDK4/6 resulting in more p21^{cip1} and p27^{kip1} to inhibit cyclin A/E-CDK2. Thus, the stimulation of p15^{ink4b} by TGFβ can lead to the inhibition of both cyclin D-CDK4/6 and cyclin A/E-CDK2 (58, 59).

Another mechanism that is believed to mediate the tumor suppressive activity of TGFB is its stimulation of programmed cell death called apoptosis. Treatment with exogenous TGF β has been shown to stimulate apoptosis in various cell types including epithelial cells (60, 61). However, how TGF β regulate apoptosis at the molecular level is not well defined although several apoptosis-related proteins including Bcl-xL, caspases, Smad7 and p38 MAP kinase have been implicated (60, 62). The extent of apoptosis induced by TGF β and the effectors involved appears cell context-dependent. Of note, significantly higher concentrations of TGF β are usually required to induce apoptosis than to inhibit cell proliferation. Furthermore, few reports have shown a regulatory role of autocrine TGF β signaling in controlling apoptosis. Thus, the role of TGF β -mediated apoptosis in tumor suppression

remains elusive although it is known to play an important role in tissue development and remodeling (63).

While the inhibition of cell cycle progression is apparently the major mechanism of tumor suppression by TGFB, recent studies suggest that the inhibition of telomerase activity by TGFB may also contribute to its tumor suppressive function. TGF β was shown to inhibit telomerase activity by suppressing the transcription of human telomerase reverse transcriptase (64, 65). The inhibition can lead to attrition of telomere and cell senescence as observed in human lung adenocarcinoma cells (66). These activities of TGF β should place its signaling pathway as a "gate keeper" in preventing tumorigenesis and inhibiting the growth of primary tumors. However, mutational inactivation of TGFB signaling components appears to occur at late stages of carcinogenesis. For example, loss of DPC4 expression occurs during the transformation of human pancreatic neoplasm from non-invasive to invasive stage (67). Similarly, mutation of TGFBR2 is associated with progression of human colon adenomas to malignant carcinomas (68). Thus, TGF β signaling appears necessary for the suppression of tumor invasion and metastasis. Indeed, targeted expression of a dominant negative RII in mouse prostate was shown to promote metastasis of SV40 large T antigen-induced prostate tumors with little effect on the sizes of the neoplastic prostates (69). Further studies are needed to determine whether loss of control of cell proliferation, apoptosis and senescence by TGFB is sufficient to promote tumor invasion and metastasis or whether TGF β signaling specifically regulate a set of metastasis-related genes.

4. SWITCH FROM TUMOR SUPPRESSION TO TUMOR PROMOTION

4.1. Tumor cells often evade the growth inhibition by TGF**b**

Because of its potent growth inhibitory activity in normal epithelial cell, TGF β signaling pathway is often disrupted or modulated in tumor cells such that they are resistant to its growth inhibition. In some cases, the loss of TGF β sensitivity is categorical due to the loss of TGF β signaling receptors as mentioned above. Loss of Smad proteins, especially Smad4, can also abrogate a majority of TGF^β signaling activities although TGF^β has been shown to inhibit cell proliferation or stimulate apoptosis independent of Smad proteins (70, 71). However, complete inactivation of TGF β signaling through mutations of RII or Smad proteins is restricted to certain types of cancer and is rare in other types of cancers with or without microsatellite instability. For example, breast, endometrial, pancreatic and lung carcinomas with microsatellite instability showed few or no RII mutation (20-22) and mutation of Smad4 is uncommon in breast and ovarian carcinomas (72). These observations suggest that autocrine TGF β signaling may be needed for the progression of certain types of cancer.

As mentioned earlier, TGF β inhibits cell cycle progression mainly by inhibiting the expression of c-Myc,

inducing the expression of p15^{ink4b} and p21^{cip1}, and consequently causing hyperphosphorylation of pRB. Since tumor progression is often driven by inactivation of growth inhibitory genes such as *pRB* and overexpression of growth promoting genes such as *c-myc*, carcinoma cells are often resistant to TGF β 's growth inhibitory activity while retaining a functional TGFB signaling pathway. For example, the human carcinoma DU145 cells are resistant to TGFβ's growth inhibition due to mutational inactivation of *pRB*, but are sensitive to its regulation of gene expression (73, 74). Similarly, failure to inhibit c-Myc expression by TGF β was shown to cause the resistance to its growth inhibition in the human breast carcinoma MDA-MB-231 cells, which are responsive to $TGF\beta$ with respect to the regulation of the expression of several other genes (75). Thus, carcinoma cells with an operational TGF β signaling pathway are invariably less sensitive to the growth inhibition by TGF β than their normal counterparts.

4.2. TGF**b** signal promotes late stage tumor progression

The loss of sensitivity to TGFB's growth inhibition gives tumor cells a selective advantage over normal cells to proliferate. This is further exacerbated by increased TGF β expression and/or activation that are generally associated with tumorigenesis (76). For example, TGF β isoforms have been shown to be upregulated during neoplastic development and progression in breast (77-81), colon (82, 83), prostate (84, 85), and bladder cancers (86). Furthermore, a number of studies reported that increased expression of TGF β could actually promote tumor progression in carcinoma cells. Overexpression of $TGF\beta_1$ in human breast cancer MCF-7 cells led to increased, estrogen-independent tumor formation in athymic nude mice (87). In certain carcinoma cells, overexpression of TGF β_1 can enhance its growth inhibitory activity *in vitro*, yet stimulate tumor growth and progression when they are inoculated in vivo (88-90). In animal models, transgenic expression of $TGF\beta_1$ in the skin was shown to inhibit carcinogen-induced tumor incidence, however it promoted malignant progression to invasive carcinomas (91).

The mechanisms of tumor promotion by $TGF\beta$ were initially attributed to its action in tumor stroma (76). Indeed, TGF β is a potent immune suppressor in that they can inhibit proliferation, activation and differentiation of various types of lymphocytes (92). Overexpression of $TGF\beta_1$ in highly immunogenic murine tumor cells was shown to stimulate tumor growth by escaping immune surveillance (93). TGF β has also been shown to be angiogenic *in vivo* (94-96). Overexpression of $TGFB_1$ in Chinese hamster ovary cells and human prostate cancer cells was shown to significantly stimulate tumor growth and angiogenesis when they are inoculated in mice (97, 98) and the effects could be attenuated by peritumoral injection of a TGF β_1 neutralizing antibody (98). Thus, tumor stromal cells appear to be major targets of the tumor promoting activity of TGF β . However, treatment with exogenous TGFβ *in vitro* was shown to stimulate metastatic potential of a mammary adenocarcinoma cell line in vivo suggesting that TGF β can also directly act on tumor cells to enhance their malignancy (99). More recently, several studies have

shown that blockade of TGF β signaling in late-stage tumor cells can suppress their malignancy. For example, abrogation of TGF β signaling in mammary and colon cancer cells by the expression of a dominant negative RII was shown to inhibit their in vivo growth and metastasis (100). Dominant negative RII expression in human breast carcinoma MDA-MB-231 cell was also shown to inhibit its bone metastatic potential by blocking TGFB-induced tumor production of parathyroid hormone-related protein, which stimulates osteolytic activity (101). In genetically related progression models of a human mammary epithelial cell line, dominant negative blockade of TGFB signaling by RII or Smad was shown to promote tumorigenicity of a lowgrade pre-malignant cell, but inhibited metastasis of a highgrade tumorigenic cell (102, 103). Interestingly, the blockade of TGF β signaling did not affect primary tumorigenesis of the high-grade tumorigenic cell (102). These observations demonstrate that TGFB signaling in late stage, malignant cancer cells is necessary for their metastatic behavior.

The switch of TGFB signaling pathway from a tumor suppressor to a tumor promoter is likely accomplished by an alteration of expression and function of multiple gene products. Phenotypically, the attenuation of sensitivity to the growth inhibition by TGFB as observed in most malignant tumor cells may be a prerequisite for the switch. The ability of tumor cells to undergo epithelial to mesenchymal transdifferentiation (EMT) in the presence of TGF β signal activation may also be necessary for the switch since the process of EMT contributes to tumor cell migration and invasion, and TGFB has been shown to promote EMT in various transformed epithelial cells (104). The aberrant regulation of extracellular proteolytic activity in tumor cells by TGFB (105) is another important mechanism that can mediate TGFB-induced invasion and metastasis. Finally, both exogenous and autocrine $TGF\beta$ have been shown to generate a cell survival signal in various epithelial cells (106, 107), which is also likely to contribute to the tumor-promoting activity of TGFB.

5. INHIBITION OF TUMOR PROGRESSION WITH TGF**b** ANTAGONISTS

The observations that TGF β can promote malignant progression at late stages of carcinogenesis have stimulated investigations to target TGFB as a novel therapeutic strategy to suppress tumor progression. Various approaches have been utilized to antagonize the tumorpromoting activity of TGFB. For example, intraperitoneal injection of an anti-TGFB antibody that neutralized all three TGFB isoforms inhibited the tumorigenicity of the human breast carcinoma MDA-MB-231 cell (108). Antisense inhibition of the expression of $TGF\beta_1$ in a murine mammary tumor cell line was shown to abrogate the suppression of cytotoxic T cell by of TGF β_1 secreted from the tumor cells and resulted in the inhibition of tumor growth in syngeneic mice (109). Administration of TGFB neutralizing antibodies or $TGF\beta_2$ antisense oligonucleotides stimulated the activity of natural killer cell and restored the inhibition of in vivo growth of human

breast cancer cells by tamoxifen in mice with proficient natural killer cells (110). Several reports have shown that ectopic expression of TGFB binding proteins including decorin and extracellular domains of TGFB RII and RIII can inhibit tumorigenicity, tumor growth, and/or metastasis of xenograft models of glioma, hepatoma, and carcinomas of breast, colon and pancreas (111-116). Furthermore, transgenic expression of a soluble TGFB RII:Fc fusion protein was shown to reduce metastatic incidence in various organs when the mice were inoculated intravenously with an isogenic melanoma cell line (117). When the RII:Fc transgenic mouse was crossed with the MMTV-neu transgenic mouse that develops metastatic breast cancer, RII:Fc expression also inhibited metastasis from endogenous mammary tumors, but did not enhance primary tumor incidence (117). These observations were consistent with a separate study demonstrating that systemic administration of a recombinant RII:Fc fusion protein inhibited lung metastasis produced bv orthotopically inoculated breast cancer cell lines or by mammary tumors in MMTV-Polyomavirus middle T antigen transgenic mice (118). Similarly, systemic administration of a recombinant soluble TGFB RIII was also shown to inhibit the growth, angiogenesis and lung metastasis of growing tumors formed orthotopically by human breast cancer cells in nude mice (119).

These findings point to potential utility of $TGF\beta$ antagonists as a novel class of therapeutic agents for cancer treatment. Indeed, antisense TGF β_2 is under clinical trial for the treatment of glioblastoma (120). A few small molecules that specifically block the kinase activity of TGFB RI have also been developed (120). It will be interesting to compare TGFB receptor blockers and TGFB binding proteins with respect to their efficacy in blocking malignant progression. While TGFβ binding proteins may neutralize excessive amount of TGFBs associated with tumorigenesis and metastasis, but spare TGFBs at physiological levels in normal tissues (117), one concern of the application of TGFB receptor blockers is that they may induce toxicity associated with the blockade of TGFB signaling in normal tissues. Furthermore, since TGFB signaling can be tumor-suppressive in low-grade, welldifferentiated adenocarcinoma cells, a blanket blockade of TGF β signaling with a TGF β receptor blocker in a patient with a heterogeneous population of tumor cells may induce the progression of low-grade tumor cells. In contrast, TGFB binding proteins may be administered at a certain dosage such that they only neutralize those $TGF\beta s$ acting in paracrine and endocrine fashion to foster a favorable microenvironment for tumor cell growth and metastasis. As such, TGF β sequesters may be more applicable as cancer therapeutics than TGF β receptor blockers.

6. PERSPECTIVES

The dogma that TGF β signaling can be either tumor suppressive or tumor promoting has been realized for sometime. However, the molecular markers that can be used to distinguish a tumor that is suppressed by TGF β from a tumor that is promoted by TGF β remains unclear. Published findings are mostly circumstantial and often contradictory. Clearly, the role of TGFB signaling in tumorigenesis is context and stage dependent. TGFB is generally believed to suppress the development and earlystage progression of epithelial tumors. However, why the expression or administration of the TGFB binding protein RII:Fc did not enhance primary mammary tumorigenesis in transgenic mice with endogenous tumors (117, 118) is intriguing. It will be interesting to determine whether the transgenic expression of neu or Polyomavirus middle T antigen in the two studies attenuated the tumor suppressive ability of TGF β in the mammary epithelial cells. TGF β signaling is also generally believed to promote metastasis during malignant transformation. However, TGFB signaling may also be necessary for inhibition of metastasis. For example, loss of TGFB signaling is associated with adenoma to carcinoma transition in colorectal cancer (68) and abrogation of TGFB signaling promoted prostate tumor metastasis (69). Future studies will need to elucidate molecular signature(s) in tumor cells that can reveal whether TGFB signaling has switched from tumor suppression to tumor promotion for successful therapeutic intervention with $TGF\beta$ antagonists.

Because TGF β inhibits host immune surveillance and stimulates tumor angiogenesis, its action in tumor stroma is generally believed to be tumor-promoting. As such, TGFB action would be solely tumor-promoting in carcinomas with mutational inactivation of TGF^β signaling receptors. Thus, loss of TGFB RI or RII expression in carcinoma cells should constitute a molecular signature for the use of TGFB antagonists to inhibit or prevent local or distance metastasis after resection of primary tumor. In fact, since the inhibition of cell cycle progression appears to be the major mechanism of tumor suppression by $TGF\beta$, loss of TGF β -induced regulation of gene products that control cell proliferation should also indicate a potential favorable outcome with the use of TGF β antagonist. For example, loss of *c-myc* repression resulted in a significant loss of cell cycle arrest by TGF β in the human breast carcinoma MDA-MB-231 cell (75). Neutralization of TGFB isoforms with administration of an antibody or a recombinant soluble RIII protein inhibited tumor growth and metastasis of this cell line in vivo (108, 119). Another potential utility of TGF β sequesters is to inhibit cancer bone metastasis. Active TGF β isoforms are produced not only by carcinoma cells metastasizing in the bone, but also from bone matrix during osteolysis as observed in breast cancer bone metastasis. These active TGFB isoforms can stimulate proliferation of osteoblasts as well as differentiation of osteoclasts through various signaling pathways (121). As such, TGFB is implicated in promoting both osteolytic and osteoblastic bone metastasis. Since active TGF β isoforms released from bone matrix should be readily accessible by its binding proteins, it appears highly feasible that TGFB sequesters such as its soluble receptors or neutralizing antibodies will inhibit cancer-induced bone lesions.

The observations that life-long exposure to the TGF β sequester RII:Fc protein showed little deleterious

effect on development and spontaneous tumorigenesis (117) and that systemic administration of a TGF β soluble RIII did not induce any noticeable side effect (119) point to the feasibility of *in vivo* application of TGF β sequesters. It is expected that future research will identify more TGF β binding molecules for experimental trials and specific carcinomas and/or processes of carcinogenesis that can be intervened by TGF β binding molecules.

7. ACKNOWLEDGEMENTS

Because of the enormous and ever-expanding body of literature in the field, the author apologizes for not referencing many relevant studies due to his oversight and space limitation. The related research work from the author's laboratory has been supported by NIH grants CA75253 and CA79683.

8. REFERENCES

1. Piek, E., C. H. Heldin and P. ten Dijke: Specificity, diversity, and regulation in TGF-beta superfamily signaling. *FASEB J* 13, 2105-2124 (1999)

2. Chen, C., X. F. Wang and L. Z. Sun: Expression of transforming growth factor beta type III receptor restores autocrine TGF beta1 activity in human breast cancer MCF-7 cells. *J Biol Chem* 272, 12862-12867 (1997)

3. Lopez-Casillas, F., J. L. Wrana and J. Massague: Betaglycan presents ligand to the TGF beta signaling receptor. *Cell* 73, 1435-1444 (1993)

4. Wang, X. F., H. Y. Lin, E. Ng-Eaton, J. Downward, H. F. Lodish and R. A. Weinberg: Expression cloning and characterization of the TGF-beta type III receptor. *Cell* 67, 797-805 (1991)

5. Fukushima, D., R. Butzow, A. Hildebrand and E. Ruoslahti: Localization of transforming growth factor beta binding site in betaglycan. Comparison with small extracellular matrix proteoglycans. *J Biol Chem* 268, 22710-22715 (1993)

6. Lopez-Casillas, F., H. M. Payne, J. L. Andres and J. Massague: Betaglycan can act as a dual modulator of TGFbeta access to signaling receptors: mapping of ligand binding and GAG attachment sites. *J Cell Biol* 124, 557-568 (1994)

7. Wrana, J. L., L. Attisano, R. Wieser, F. Ventura and J. Massague: Mechanism of activation of the TGF-beta receptor. *Nature* 370, 341-347 (1994)

8. Massagué, J: TGF-β signal transduction. *Annu Rev Biochem* 67, 753-791 (1998)

9. Massague, J. and Y. G. Chen: Controlling TGF-beta signaling. *Genes and Development* 14, 627-644 (2000)

10. Moses, H. L., E. Y. Yang and J. A. Pietenpol: TGF-beta stimulation and inhibition of cell proliferation: new

mechanistic insights. Cell 63, 245-247 (1990)

11. Roberts, A. B. and M. B. Sporn: The transforming growth factor-betas. In: Peptide growth factors and their receptors I. Eds: Sporn MB, Roberts AB, Springer-Verlag, Heidelberg 419-472 (1991)

12. Roberts, A. B., M. A. Anzano, L. C. Lamb, J. M. Smith and M. B. Sporn: New class of transforming growth factors potentiated by epidermal growth factor: isolation from nonneoplastic tissues. *Proc Natl Acad Sci USA* 78, 5339-5343 (1981)

13. Moses, H. L., R. F. Tucker, E. B. Leof, R.J. Coffey, Jr., J. Halper and G. D. Shipley: Type-beta transforming growth factor is a growth stimulator and a growth inhibitor. In: Cancer Cells. Eds: Feramisco J, Ozanne B, Stiles C, Cold Spring Harbor Laboratory Press, Cold Spring Harbor 65-71 (1985)

14. Arteaga, C. L., R. J. Coffey, Jr., T. C. Dugger, C. M. McCutchen, H. L. Moses and R. M. Lyons: Growth stimulation of human breast cancer cells with anti-transforming growth factor beta antibodies: evidence for negative autocrine regulation by transforming growth factor beta. *Cell Growth Differ* 1, 367-374 (1990)

15. Glick, A. B., K. C. Flanders, D. Danielpour, S. H. Yuspa and M. B. Sporn: Retinoic acid induces transforming growth factor-beta 2 in cultured keratinocytes and mouse epidermis. *Cell Regul* 1, 87-97 (1989)

16. Hafez, M. M., D. Infante, S. Winawer and E. Friedman: Transforming growth factor beta 1 acts as an autocrinenegative growth regulator in colon enterocytic differentiation but not in goblet cell maturation. *Cell Growth Differ* 1, 617-626 (1990)

17. Wu, S. P., D. Theodorescu, R. S. Kerbel, J. K. Willson, K. M. Mulder, L. E. Humphrey and M. G. Brattain: TGFbeta 1 is an autocrine-negative growth regulator of human colon carcinoma FET cells *in vivo* as revealed by transfection of an antisense expression vector. *J Cell Biol* 116, 187-196 (1992)

18. Wu, S. P., L. Z. Sun, J. K. Willson, L. Humphrey, R. Kerbel and M. G. Brattain: Repression of autocrine transforming growth factor beta 1 and beta 2 in quiescent CBS colon carcinoma cells leads to progression of tumorigenic properties. *Cell Growth Differ* 4, 115-123 (1993)

19. Markowitz, S., J. Wang, L. Myeroff, R. Parsons, L. Sun, J. Lutterbaugh, R. S. Fan, E. Zborowska, K. W. Kinzler, B. Vogelstein, M. G. Brattain and J. K. V. Willson: Inactivation of the type II TGF-beta receptor in colon cancer cells with microsatellite instability. *Science* 268, 1336-1338 (1995)

20. Myeroff, L. L., R. Parsons, S. J. Kim, L. Hedrick, K. R. Cho, K. Orth, M. Mathis, K. W. Kinzler, J. Lutterbaugh, K. Park, Y. J. Bang, H. Y. Lee, J. G. Park, H. T. Lynch, A. B.

Roberts, B. Vogelstein and S. D. Markowitz: A transforming growth factor beta receptor type II gene mutation common in colon and gastric but rare in endometrial cancers with microsatellite instability. *Cancer Res* 55, 5545-5547 (1995)

21. Abe, T., H. Ouyang, T. Migita, Y. Kato, M. Kimura, K. Shiiba, M. Sunamura, S. Matsuno and A. Horii: The somatic mutation frequency of the transforming growth factor beta receptor type II gene varies widely among different cancers with microsatellite instability. *Eur J Surg Oncol* 22, 474-477 (1996)

22. Tomita, S., S. Deguchi, T. Miyaguni, Y. Muto, T. Tamamoto and T. Toda: Analyses of microsatellite instability and the transforming growth factor-beta receptor type II gene mutation in sporadic breast cancer and their correlation with clinicopathological features. *Breast Cancer Res Treat* 53, 33-39 (1999)

23. Eisma, R. J., J. D. Spiro, S. E. von Biberstein, R. Lindquist and D. L. Kreutzer: Decreased expression of transforming growth factor beta receptors on head and neck squamous cell carcinoma tumor cells. *Am J Surg* 172, 641-645 (1996)

24. Franchi, A., O. Gallo, I. Sardi and M. Santucci: Downregulation of transforming growth factor beta type II receptor in laryngeal carcinogenesis. *J Clin Pathol* 54, 201-204 (2001)

25. Gobbi, H., W. D. Dupont, J. F. Simpson, W. D. Plummer, P. A. Schuyler, S. J. Olson, C. L. Arteaga and D. L. Page: Transforming growth factor-beta and breast cancer risk in women with mammary epithelial hyperplasia. *J Natl Cancer Inst* 91, 2096-2101 (1999)

26. Chang, J., K. Park, Y. J. Bang, W. S. Kim, D. Kim and S. J. Kim: Expression of transforming growth factor beta type II receptor reduces tumorigenicity in human gastric cancer cells. *Cancer Res* 57, 2856-2859 (1997)

27. Sun, L., G. Wu, J. K. Willson, E. Zborowska, J. Yang, I. Rajkarunanayake, J. Wang, L. E. Gentry, X. F. Wang and M. G. Brattain: Expression of transforming growth factor beta type II receptor leads to reduced malignancy in human breast cancer MCF-7 cells. *J Biol Chem* 269, 26449-26455 (1994)

28. Wang, J., L. Sun, L. Myeroff, X. Wang, L. E. Gentry, J. Yang, J. Liang, E. Zborowska, S. Markowitz, J. K. Willson and M. G. Brattain: Demonstration that mutation of the type II transforming growth factor beta receptor inactivates its tumor suppressor activity in replication error-positive colon carcinoma cells. *J Biol Chem* 270, 22044-22049 (1995)

29. Chen, T., J. Triplett, B. Dehner, B. Hurst, B. Colligan, J. Pemberton, J. R. Graff and J. H. Carter: Transforming growth factor-beta receptor type I gene is frequently mutated in ovarian carcinomas. *Cancer Res* 61, 4679-4682 (2001)

30. Wang, D., T. Kanuma, H. Mizunuma, F. Takama, Y.

Ibuki, N. Wake, A. Mogi, Y. Shitara and S. Takenoshita: Analysis of specific gene mutations in the transforming growth factor- beta signal transduction pathway in human ovarian cancer. *Cancer Res* 60, 4507-4512 (2000)

31. Kang, S. H., Y. J. Bang, Y. H. Im, H. K. Yang, D. A. Lee, H. Y. Lee, H. S. Lee, N. K. Kim and S. J. Kim: Transcriptional repression of the transforming growth factor-beta type I receptor gene by DNA methylation results in the development of TGF-beta resistance in human gastric cancer. *Oncogene* 18, 7280-7286 (1999)

32. Wang, J., W. Han, E. Zborowska, J. Liang, X. Wang, J. K. V. Willson, L. Sun and M. G. Brattain: Reduced expression of transforming growth factor beta type I receptor contributes to the malignancy of human colon carcinoma cells. *J Biol Chem* 271, 17366-17371 (1996)

33. Hahn, S. A., M. Schutte, A. T. Hoque, C. A. Moskaluk, L. T. da Costa, E. Rozenblum, C. L. Weinstein, A. Fischer, C. J. Yeo, R. H. Hruban and S. E. Kern: DPC4, a candidate tumor suppressor gene at human chromosome 18q21.1. *Science* 271, 350-353 (1996)

34. Hahn, S. A., A. T. Hoque, C. A. Moskaluk, L. T. da Costa, M. Schutte, E. Rozenblum, A. B. Seymour, C. L. Weinstein, C. J. Yeo, R. H. Hruban and S. E. Kern: Homozygous deletion map at 18q21.1 in pancreatic cancer. *Cancer Res* 56, 490-494 (1996)

35. Maitra, A., K. Molberg, J. Albores-Saavedra and G. Lindberg: Loss of Dpc4 expression in colonic adenocarcinomas correlates with the presence of metastatic disease. *Am J Pathol* 157, 1105-1111 (2000)

36. Miyaki, M., T. Iijima, M. Konishi, K. Sakai, A. Ishii, M. Yasuno, T. Hishima, M. Koike, N. Shitara, T. Iwama, J. Utsunomiya, T. Kuroki and T. Mori: Higher frequency of Smad4 gene mutation in human colorectal cancer with distant metastasis. *Oncogene* 18, 3098-3103 (1999)

37. Takagi, Y., H. Kohmura, M. Futamura, H. Kida, H. Tanemura, K. Shimokawa and S. Saji: Somatic alterations of the DPC4 gene in human colorectal cancers *in vivo*. *Gastroenterology* 111, 1369-1372 (1996)

38. Eppert, K., S. W. Scherer, H. Ozcelik, R. Pirone, P. Hoodless, H. Kim, L. C. Tsui, B. Bapat, S. Gallinger, I. L. Andrulis, G. H. Thomsen, J. L. Wrana and L. Attisano: MADR2 maps to 18q21 and encodes a TGFbeta-regulated MAD-related protein that is functionally mutated in colorectal carcinoma. *Cell* 86, 543-552 (1996)

39. Uchida, K., M. Nagatake, H. Osada, Y. Yatabe, M. Kondo, T. Mitsudomi, A. Masuda and T. Takahashi: Somatic *in vivo* alterations of the JV18-1 gene at 18q21 in human lung cancers. *Cancer Res* 56, 5583-5585 (1996)

40. Pierce, D. F., Jr., A. E. Gorska, A. Chytil, K. S. Meise, D. L. Page, R. J. Coffey, Jr. and H. L. Moses: Mammary tumor suppression by transforming growth factor beta 1

transgene expression. Proc Natl Acad Sci U S A 92, 4254-4258 (1995)

41. Tang, B., E. P. Bottinger, S. B. Jakowlew, K. M. Bagnall, J. Mariano, M. R. Anver, J. J. Letterio and L. M. Wakefield: Transforming growth factor-beta1 is a new form of tumor suppressor with true haploid insufficiency. *Nat Med* 4, 802-807 (1998)

42. Amendt, C., P. Schirmacher, H. Weber and M. Blessing: Expression of a dominant negative type II TGF- β receptor in mouse skin results in an increase in carcinoma incidence and an acceleration of carcinoma development. *Oncogene* 17, 25-34 (1998)

43. Bottinger, E. P., J. L. Jakubczak, I. S. Roberts, M. Mumy, P. Hemmati, K. Bagnall, G. Merlino and L. M. Wakefield: Expression of a dominant-negative mutant TGF-beta type II receptor in transgenic mice reveals essential roles for TGF-beta in regulation of growth and differentiation in the exocrine pancreas. *EMBO J* 16, 2621-2633 (1997)

44. Go, C., W. He, L. Zhong, P. Li, J. Huang, B. R. Brinkley and X. J. Wang: Aberrant cell cycle progression contributes to the early-stage accelerated carcinogenesis in transgenic epidermis expressing the dominant negative TGF beta RII. *Oncogene* 19, 3623-3631 (2000)

45. Takaku, K., H. Miyoshi, A. Matsunaga, M. Oshima, N. Sasaki and M. M. Taketo: Gastric and duodenal polyps in Smad4 (Dpc4) knockout mice. *Cancer Res* 59, 6113-6117 (1999)

46. Xu, X., S. G. Brodie, X. Yang, Y. H. Im, W. T. Parks, L. Chen, Y. X. Zhou, M. Weinstein, S. J. Kim and C. X. Deng: Haploid loss of the tumor suppressor Smad4/Dpc4 initiates gastric polyposis and cancer in mice. *Oncogene* 19, 1868-1874 (2000)

47. Takaku, K., M. Oshima, H. Miyoshi, M. Matsui, M. F. Seldin and M. M. Taketo: Intestinal tumorigenesis in compound mutant mice of both Dpc4 (Smad4) and Apc genes. *Cell* 92, 645-656 (1998)

48. Dang, C. V: c-myc target genes involved in cell growth, apoptosis, and metabolism. *Mol Cell Biol* 19, 1-11 (1999)

49. Coffey, R. J., Jr., C. C. Bascom, N. J. Sipes, R. Graves-Deal, B. E. Weissman and H. L. Moses: Selective inhibition of growth-related gene expression in murine keratinocytes by transforming growth factor beta. *Mol Cell Biol* 8, 3088-3093 (1988)

50. Mulder, K. M., A. E. Levine, X. Hernandez, M. K. McKnight, D. E. Brattain and M. G. Brattain: Modulation of c-myc by transforming growth factor-beta in human colon carcinoma cells. *Biochem Biophys Res Commun* 150, 711-716 (1988)

51. Pietenpol, J. A., R. W. Stein, E. Moran, P. Yaciuk, R. Schlegel, R. M. Lyons, M. R. Pittelkow, K. Munger, P. M.

Howley and H. L. Moses: TGF-beta 1 inhibition of c-myc transcription and growth in keratinocytes is abrogated by viral transforming proteins with pRB binding domains. *Cell* 61, 777-785 (1990)

52. Chen, C. R., Y. B. Kang, P. M. Siegel and J. Massague: E2F4/5 and p107 as Smad cofactors linking the TGF beta receptor to c-myc repression. *Cell* 110, 19-32 (2002)

53. Datto, M. B., Y. Li, J. F. Panus, D. J. Howe, Y. Xiong and X. F. Wang: Transforming growth factor beta induces the cyclin-dependent kinase inhibitor p21 through a p53-independent mechanism. *Proc Natl Acad Sci U S A* 92, 5545-5549 (1995)

54. Hannon, G. J. and D. Beach: p15INK4B is a potential effector of TGF-beta-induced cell cycle arrest. *Nature* 371, 257-261 (1994)

55. Datto, M. B., Y. Yu and X. F. Wang: Functional analysis of the transforming growth factor beta responsive elements in the WAF1/Cip1/p21 promoter. *J Biol Chem* 270, 28623-28628 (1995)

56. Feng, X. H., X. Lin and R. Derynck: Smad2, Smad3 and Smad4 cooperate with Sp1 to induce p15(Ink4B) transcription in response to TGF-beta. *EMBO J* 19, 5178-5193 (2000)

57. Pardali, K., A. Kurisaki, A. Moren, P. ten Dijke, D. Kardassis and A. Moustakas: Role of smad proteins and transcription factor Sp1 in p21(Wafl/Cip1) regulation by transforming growth factor-beta. *J Biol Chem* 275, 29244-29256 (2000)

58. Reynisdottir, I., K. Polyak, A. Iavarone and J. Massague: Kip/Cip and Ink4 Cdk inhibitors cooperate to induce cell cycle arrest in response to TGF-beta. *Genes Dev* 9, 1831-1845 (1995)

59. Sandhu, C., J. Garbe, N. Bhattacharya, J. Daksis, C. H. Pan, P. Yaswen, J. Koh, J. M. Slingerland and M. R. Stampfer: Transforming growth factor beta stabilizes p15INK4B protein, increases p15INK4B-cdk4 complexes, and inhibits cyclin D1-cdk4 association in human mammary epithelial cells. *Mol Cell Biol* 17, 2458-2467 (1997)

60. Chipuk, J. E., M. Bhat, A. Y. Hsing, J. J. Ma and D. Danielpour: Bcl-xL, blocks transforming growth factorbeta 1-induced apoptosis by inhibiting cytochrome c release and not by directly antagonizing Apaf-1-dependent caspase activation in prostate epithelial cells. *J Biol Chem* 276, 26614-26621 (2001)

61. Rotello, R. J., R. C. Lieberman, A. F. Purchio and L. E. Gerschenson: Coordinated regulation of apoptosis and cell proliferation by transforming growth factor beta 1 in cultured uterine epithelial cells. *Proc Natl Acad Sci U S A* 88, 3412-3415 (1991)

62. Edlund, S., S. H. Bu, N. Schuster, P. Aspenstrom, R.

Heuchel, N. E. Heldin, P. ten Dijke, C. H. Heldin and M. Landstrom: Transforming growth factor-beta 1 (TGF-beta)induced apoptosis of prostate cancer cells involves Smad7dependent activation of p38 by TGF-beta-activated kinase 1 and mitogen-activated protein kinase kinase 3. *Mol Biol Cell* 14, 529-544 (2003)

63. Schuster, N. and K. Krieglstein: Mechanisms of TGFbeta-mediated apoptosis. *Cell and Tissue Research* 307, 1-14 (2002)

64. Lin, S. Y. and S. J. Elledge: Multiple tumor suppressor pathways negatively regulate telomerase. *Cell* 113, 881-889 (2003)

65. Yang, H., S. Kyo, M. Takatura and L. Sun: Autocrine transforming growth factor beta suppresses telomerase activity and transcription of human telomerase reverse transcriptase in human cancer cells. *Cell Growth Differ* 12, 119-127 (2001)

66. Katakura, Y., E. Nakata, T. Miura and S. Shirahata: Transforming growth factor β triggers two independentsenescence programs in cancer cells. *Biochem Biophys Res Commun* 255, 110-115 (1999)

67. Wilentz, R. E., C. A. Iacobuzio-Donahue, P. Argani, D. M. McCarthy, J. L. Parsons, C. J. Yeo, S. E. Kern and R. H. Hruban: Loss of expression of Dpc4 in pancreatic intraepithelial neoplasia: evidence that DPC4 inactivation occurs late in neoplastic progression. *Cancer Res* 60, 2002-2006 (2000)

68. Grady, W. M., A. Rajput, L. Myeroff, D. F. Liu, K. H. Kwon, J. Willis and S. Markowitz: Mutation of the type II transforming growth factor- β receptor is coincident with the transformation of human colon adenomas to malignant carcinomas. *Cancer Res* 58, 3101-3104 (1998)

69. Tu, W. H., T. Z. Thomas, N. Masumori, N. A. Bhowmick, A. E. Gorska, Y. Shyr, S. Kasper, T. Case, R. L. Roberts, S. B. Shappell, H. L. Moses and R. J. Matusik: The loss of TGF-beta signaling promotes prostate cancer metastasis. *Neoplasia* 5, 267-277 (2003)

70. Fink, S. P., S. E. Swinler, J. D. Lutterbaugh, J. Massague, S. Thiagalingam, K. W. Kinzler, B. Vogelstein, J. K. V. Willson and S. Markowitz: Transforming growth factor-beta-induced growth inhibition in a smad4 mutant colon adenoma cell line. *Cancer Res* 61, 256-260 (2001)

71. Yu, L., M. C. Hebert and Y. E. Zhang: TGF-beta receptoractivated p38 MAP kinase mediates smad-independent TGFbeta responses. *EMBO J* 21, 3749-3759 (2002)

72. Schutte, M., R. H. Hruban, L. Hedrick, K. R. Cho, G. M. Nadasdy, C. L. Weinstein, G. S. Bova, W. B. Isaacs, P. Cairns, H. Nawroz, D. Sidransky, R. A. Casero, Jr., P. S. Meltzer, S. A. Hahn and S. E. Kern: DPC4 gene in various tumor types. *Cancer Res* 56, 2527-2530 (1996)

73. Murphy, C. S., J. A. Pietenpol, K. Munger, P. M.

Howley and H. L. Moses: c-myc and pRB: role in TGFbeta 1 inhibition of keratinocyte proliferation. *Cold Spring Harb Symp Quant Biol* 56, 129-135 (1991)

74. Zentella, A., F. M. Weis, D. A. Ralph, M. Laiho and J. Massague: Early gene responses to transforming growth factor-beta in cells lacking growth-suppressive RB function. *Mol Cell Biol* 11, 4952-4958 (1991)

75. Chen, C. R., Y. B. Kang and J. Massague: Defective repression of c-myc in breast cancer cells: A loss at the core of the transforming growth factor beta growth arrest program. *Proc Natl Acad Sci USA* 98, 992-999 (2001)

76. Reiss, M: TGF-beta and cancer. *Microbes and Infection* 1, 1327-1347 (1999)

77. Barrett-Lee, P., M. Travers, Y. Luqmani and R. C. Coombes: Transcripts for transforming growth factors in human breast cancer: clinical correlates. *Br J Cancer* 61, 612-617 (1990)

78. Coombes, R. C., P. Barrett-Lee and Y. Luqmani: Growth factor expression in breast tissue. *J Steroid Biochem Mol Biol* 37, 833-836 (1990)

79. Daly, R. J., R. J. King and P. D. Darbre: Interaction of growth factors during progression towards steroid independence in T-47-D human breast cancer cells. *J Cell Biochem* 43, 199-211 (1990)

80. Gorsch, S. M., V. A. Memoli, T. A. Stukel, L. I. Gold and B. A. Arrick: Immunohistochemical staining for transforming growth factor beta 1 associates with disease progression in human breast cancer. *Cancer Res* 52, 6949-6952 (1992)

81. Walker, R. A. and B. Gallacher: Determination of transforming growth factor beta 1 mRNA expression in breast carcinomas by *in situ* hybridization. *J Pathol* 177, 123-127 (1995)

82. Friedman, E., L. I. Gold, D. Klimstra, Z. S. Zeng, S. Winawer and A. Cohen: High levels of transforming growth factor beta 1 correlate with disease progression in human colon cancer. *Cancer Epidemiol Biomarkers Prev* 4, 549-554 (1995)

83. Tsushima, H., S. Kawata, S. Tamura, N. Ito, Y. Shirai, S. Kiso, Y. Imai, H. Shimomukai, Y. Nomura, Y. Matsuda and Y. Matsuzawa: High levels of transforming growth factor beta 1 in patients with colorectal cancer: association with disease progression. *Gastroenterology* 110, 375-382 (1996)

84. Steiner, M. S., Z. Z. Zhou, D. C. Tonb and E. R. Barrack: Expression of transforming growth factor-beta 1 in prostate cancer. *Endocrinology* 135, 2240-2247 (1994)

85. Wikström, P., P. Stattin, I. Franck-Lissbrant, J. E. Damber and A. Bergh: Transforming growth factor $\beta 1$ is associated with angiogenesis, metastasis, and poor clinical

outcome in prostate cancer. Prostate 37, 19-29 (1998)

86. Miyamoto, H., Y. Kubota, T. Shuin, S. Torigoe, Y. Dobashi and M. Hosaka: Expression of transforming growth factor-beta 1 in human bladder cancer. *Cancer* 75, 2565-2570 (1995)

87. Arteaga, C. L., T. Carty-Dugger, H. L. Moses, S. D. Hurd and J. A. Pietenpol: Transforming growth factor beta 1 can induce estrogen- independent tumorigenicity of human breast cancer cells in athymic mice. *Cell Growth Differ* 4, 193-201 (1993)

88. Steiner, M. S. and E. R. Barrack: Transforming growth factor-beta 1 overproduction in prostate cancer: effects on growth *in vivo* and *in vitro*. *Mol Endocrinol* 6, 15-25 (1992)

89. Ye, S. C., J. M. Foster, W. H. Li, J. R. Liang, E. Zborowska, S. Venkateswarlu, J. G. Gong, M. G. Brattain and J. V. Willson: Contextual effects of transforming growth factor beta on the tumorigenicity of human colon carcinoma cells. *Cancer Res* 59, 4725-4731 (1999)

90. Tobin, S. W., K. Douville, U. Benbow, C. E. Brinckerhoff, V. A. Memoli and B. A. Arrick: Consequences of altered TGF-beta expression and responsiveness in breast cancer: evidence for autocrine and paracrine effects. *Oncogene* 21, 108-118 (2002)

91. Cui, W., D. J. Fowlis, S. Bryson, E. Duffie, H. Ireland, A. Balmain and R. J. Akhurst: TGFbeta1 inhibits the formation of benign skin tumors, but enhances progression to invasive spindle carcinomas in transgenic mice. *Cell* 86, 531-542 (1996)

92. Sosroseno, W. and E. Herminajeng: The immunoregulatory roles of transforming growth factor beta. *Br J Biomed Sci* 52, 142-148 (1995)

93. Torre-Amione, G., R. D. Beauchamp, H. Koeppen, B. H. Park, H. Schreiber, H. L. Moses and D. A. Rowley: A highly immunogenic tumor transfected with a murine transforming growth factor type beta 1 cDNA escapes immune surveillance. *Proc Natl Acad Sci USA* 87, 1486-1490 (1990)

94. Fajardo, L. F., S. D. Prionas, H. H. Kwan, J. Kowalski and A. C. Allison: Transforming growth factor betal induces angiogenesis *in vivo* with a threshold pattern. *Lab Invest* 74, 600-608 (1996)

95. Yang, E. Y. and H. L. Moses: Transforming growth factor beta 1-induced changes in cell migration, proliferation, and angiogenesis in the chicken chorioallantoic membrane. *J Cell Biol* 111, 731-741 (1990)

96. Wang, X. J., K. M. Liefer, S. Tsai, B. W. O'Malley and D. R. Roop: Development of gene-switch transgenic mice that inducibly express transforming growth factor beta1 in the epidermis. *Proc Natl Acad Sci USA* 96, 8483-8488 (1999)

97. Stearns, M. E., F. U. Garcia, K. Fudge, J. Rhim and M.

Wang: Role of interleukin 10 and transforming growth factor beta1 in the angiogenesis and metastasis of human prostate primary tumor lines from orthotopic implants in severe combined immunodeficiency mice. *Clin Cancer Res* 5, 711-720 (1999)

98. Ueki, N., M. Nakazato, T. Ohkawa, T. Ikeda, Y. Amuro, T. Hada and K. Higashino: Excessive production of transforming growth-factor beta 1 can play an important role in the development of tumorigenesis by its action for angiogenesis: validity of neutralizing antibodies to block tumor growth. *Biochim Biophys Acta* 1137, 189-196 (1992)

99. Welch, D. R., A. Fabra and M. Nakajima: Transforming growth factor beta stimulates mammary adenocarcinoma cell invasion and metastatic potential. *Proc Natl Acad Sci U S A* 87, 7678-7682 (1990)

100. Oft, M., K. H. Heider and H. Beug: TGF β signaling is necessary for carcinoma cell invasiveness and metastasis. *Curr Biol* 8, 1243-1252 (1998)

101. Yin, J. J., K. Selander, J. M. Chirgwin, M. Dallas, B. G. Grubbs, R. Wieser, J. Massague, G. R. Mundy and T. A. Guise: TGF-beta signaling blockade inhibits PTHrP secretion by breast cancer cells and bone metastases development. *J Clin Invest* 103, 197-206 (1999)

102. Tang, B. W., M. Vu, T. Booker, S. J. Santner, F. R. Miller, M. R. Anver and L. M. Wakefield: TGF-beta switches from tumor suppressor to prometastatic factor in a model of breast cancer progression. *Journal of Clinical Investigation* 112, 1116-1124 (2003)

103. Tian, F., B. S. DaCosta, W. T. Parks, S. Yoo, A. Felici, B. Tang, E. Piek, L. M. Wakefield and A. B. Roberts: Reduction in smad2/3 signaling enhances tumorigenesis but suppresses metastasis of breast cancer cell lines. *Cancer Res* 63, 8284-8292 (2003)

104. Derynck, R., R. J. Akhurst and A. Balmain: TGF-beta signaling in tumor suppression and cancer progression. *Nature Genet* 29, 117-129 (2001)

105. Norgaard, P., S. Hougaard, H. S. Poulsen and M. Spang-Thomsen: Transforming growth factor beta and cancer. *Cancer Treat Rev* 21, 367-403 (1995)

106. Lei, X. F., A. Bandyopadhyay, T. Le and L. Z. Sun: Autocrine TGF beta supports growth and survival of human breast cancer MDA-MB-231 cells. *Oncogene* 21, 7514-7523 (2002)

107. Shin, I., A. V. Bakin, U. Rodeck, A. Brunet and C. L. Arteaga: Transforming growth factor beta enhances epithelial cell survival via Akt-dependent regulation of FKHRL1. *Mol Biol Cell* 12, 3328-3339 (2001)

108. Arteaga, C. L., S. D. Hurd, A. R. Winnier, M. D. Johnson, B. M. Fendly and J. T. Forbes: Anti-transforming growth factor (TGF)-beta antibodies inhibit breast cancer cell tumorigenicity and increase mouse spleen natural killer

cell activity. Implications for a possible role of tumor cell/host TGF-beta interactions in human breast cancer progression. *J Clin Invest* 92, 2569-2576 (1993)

109. Park, J. A., E. Wang, R. A. Kurt, S. F. Schluter, E. M. Hersh and E. T. Akporiaye: Expression of an antisense transforming growth factor-betal transgene reduces tumorigenicity of EMT6 mammary tumor cells. *Cancer Gene Ther* 4, 42-50 (1997)

110. Arteaga, C. L., K. M. Koli, T. C. Dugger and R. Clarke: Reversal of tamoxifen resistance of human breast carcinomas *in vivo* by neutralizing antibodies to transforming growth factor-beta. *J Natl Cancer Inst* 91, 46-53 (1999)

111. Bandyopadhyay, A., Y. Zhu, M. L. Cibull, L. W. Bao, C. G. Chen and L. Z. Sun: A soluble transforming growth factor beta type III receptor suppresses tumorigenicity and metastasis of human breast cancer MDA-MB-231 cells. *Cancer Res* 59, 5041-5046 (1999)

112. Bandyopadhyay, A., Y. Zhu, S. N. Malik, J. Kreisberg, M. G. Brattain, E. A. Sprague, J. Luo, F. Lopez-Casillas and L. Z. Sun: Extracellular domain of TGFbeta type III receptor inhibits angiogenesis and tumor growth in human cancer cells. *Oncogene* 21, 3541-3551 (2002)

113. Rowland-Goldsmith, M. A., H. Maruyama, T. Kusama, S. Ralli and M. Korc: Soluble type II transforming growth factor-beta (TGF-beta) receptor inhibits TGF-beta signaling in COLO-357 pancreatic cancer cells *in vitro* and attenuates tumor formation. *Clinical Cancer Research* 7, 2931-2940 (2001)

114. Rowland-Goldsmith, M. A., H. Maruyama, K. Matsuda, T. Idezawa, M. Ralli, S. Ralli and M. Korc: Soluble type II transforming growth factor-beta receptor attenuates expression of metastasis-associated genes and suppresses pancreatic cancer cell metastasis. *Molecular Cancer Therapeutics* 1, 161-167 (2002)

115. Stander, M., U. Naumann, L. Dumitrescu, M. Heneka, P. Loschmann, E. Gulbins, J. Dichgans and M. Weller: Decorin gene transfer-mediated suppression of TGF-beta synthesis abrogates experimental malignant glioma growth *in vivo. Gene Ther* 5, 1187-1194 (1998)

116. Zhao, W. L., M. Kobayashi, W. Ding, L. Yuan, P. Seth, S. Cornain, J. X. Wang, F. Okada and M. Hosokawa: Suppression of *in vivo* tumorigenicity of rat hepatoma cell line KDH-8 cells by soluble TGF-beta receptor type II. *Cancer Immunology Immunotherapy* 51, 381-388 (2002)

117. Yang, Y. A., O. Dukhanina, B. Tang, M. Mamura, J. J. Letterio, J. MacGregor, S. C. Patel, S. Khozin, Z. Y. Liu, J. Green, M. R. Anver, G. Merlino and L. M. Wakefield: Lifetime exposure to a soluble TGF-beta antagonist protects mice against metastasis without adverse side effects. *J Clin Invest* 109, 1607-1615 (2002)

118. Muraoka, R. S., N. Dumont, C. A. Ritter, T. C.

Dugger, D. M. Brantley, J. Chen, E. Easterly, L. R. Roebuck, S. Ryan, P. J. Gotwals, V. Koteliansky and C. L. Arteaga: Blockade of TGF-beta inhibits mammary tumor cell viability, migration, and metastases. *J Clin Invest* 109, 1551-1559 (2002)

119. Bandyopadhyay, A., F. Lopez-Casillas, S. N. Malik, J. L. Montiel, V. Mendoza, J. Yang and L. Z. Sun: Antitumor Activity of a Recombinant Soluble Betaglycan in Human Breast Cancer Xenograft. *Cancer Res* 62, 4690-4695 (2002)

120. Dumont, N. and C. L. Arteaga: Targeting the TGF beta signaling network in human neoplasia. *Cancer Cell* 3, 531-536 (2003)

121. Mundy, G. R: Metastasis to bone: Causes, consequences and therapeutic opportunities. *Nature Reviews Cancer* 2, 584-593 (2002)

Abbreviations: APC: adenomatous polyposis coli, CDK: cyclin-dependent kinase, DPC4: deleted in pancreatic carcinomas, EMT: epithelial to mesenchymal transdifferentiation, pRB: retinoblastoma gene product, TGF α : transforming growth factor alpha, TGF β : transforming growth factor beta, RI: the type I receptor of TGF β , RII: the type II receptor of TGF β , RIII: the type III receptor of TGF β

Key Words: TGF beta, Cancer, Tumor progression, Antagonists, Review

Send correspondence to: LuZhe Sun, Ph.D., Dept. of Cellular & Structural Biology, University of Texas Health Science Center, 7703 Floyd Curl Dr., MC 7762, San Antonio, TX 78229-3900, Tel: 210-567-5746, Fax: 210-567-3803, E-mail: sunl@uthscsa.edu