PROSPECTS FOR NEUROPROTECTION IN MULTIPLE SCLEROSIS

V. Wee Yong

Departments of Oncology & Clinical Neurosciences, The University of Calgary, 3330 Hospital Drive N.W., Calgary, Alberta T2N 4N1, Canada

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1. ABSTRACT

Axonal injury and neuronal loss are now recognised to be hallmarks of multiple sclerosis (MS) in addition to neuroinflammation and demyelination. This review discusses the factors that contribute to neural degeneration, and it emphasizes the need to confer neuroprotection in MS. The beneficial role of neuroinflammation is highlighted, and the possibility that glatiramer acetate enables neuroprotection in MS through beneficial inflammation is evaluated. Finally, the prospect of an experimental treatment, minocycline, in producing neuroprotection in MS is suggested.

2. INTRODUCTION

Just over ten years ago, there was no approved treatment for MS. Since then, with the introduction of the interferon- β 's (Betaseron®, Avonex® and Rebif®) and glatiramer acetate (Copaxone®), the treatment options for patients with relapsing-remitting MS have dramatically expanded. Other immunomodulators have also become available (e.g. cyclophosphamide and mitoxantrone) to add to the armamentarium and the notion of combining therapies is now realistic. The sudden wealth of effective agents in MS has engendered more forward-looking thoughts on further improving the prognosis of the disease, including the prospects of conferring neuroprotection and regeneration. It is the possibility of providing for neuroprotection in MS that will form the basis of this review. I will describe first the pathology of MS, focussing on the axonal degeneration and neuronal loss, and I will discuss the literature that detrimental aspects of neuroinflammation contribute to that pathology. I shall then review contrasting evidence that neuroinflammation has beneficial roles, including in MS, and I will further explore themes of neuroprotection in the disease. Finally,

the likelihood that currently used MS drugs impact upon neuroprotection will be discussed.

3. PATHOLOGY OF MS: PREVALENCE OF AXONAL AND NEURONAL INJURY AND LOSS

The classical view of MS is that it is a disease of oligodendrocyte and myelin loss with relative sparing of axons, even though axonal degeneration was recognised from the early descriptions of the disease. In the late 1990s, exquisite microscopy techniques enabled investigators to refocus attention to the fact that MS is accompanied by substantial axonal atrophy and loss (1,2). Indeed, the extent of axonal loss in the spinal cord of nonambulatory MS subjects can exceed 80% (3) and progression of disease may be best correlated with the continuing loss of axons (4). In addition, we now appreciate that neuronal loss in MS is also significant. This is illustrated by the report of a 30% reduction in volume of the sub-thalamic region, assessed using MRI, as well as by histological counts of autopsied specimens (5). Recently, a 19% decrease in MRI-defined bicaudate volume (6) as well as a 7% focal thinning of the cerebral cortex by MRI was also noted (7). Thus, it is clear that MS is not only an inflammatory demyelinating disease, but also a degenerative one.

Axonal injury occurs from early in the MS disease process. By N-acetyl-aspartate (NAA) spectroscopy in patients with definite MS who do not yet show clinically significant disability (EDSS <2), diffuse axonal injury in the normal appearing white matter is already evident (8). In clinically isolated syndrome (CIS), whole brain NAA is reduced compared to that in normal individuals (9).

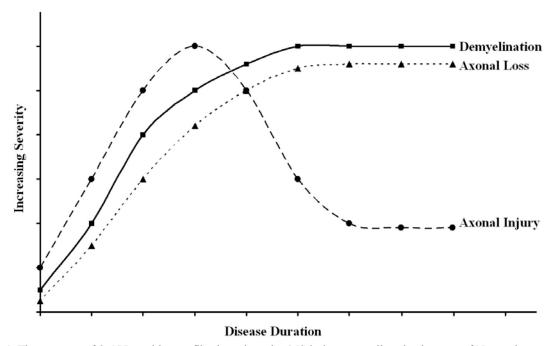


Figure 1. The presence of β -APP-positive profiles in early active MS lesions, as well as the decrease of N-acetylaspartate signal by spectroscopy in early MS, illustrate that axonal injury occurs from early on in the disease process. Some injured axons may recover but it is likely that most will go on to degenerate to result in progressive axonal loss. In this graph, demyelination is shown to precede axonal loss but it is possible that the reverse occurs in some lesions.

Histological investigations have also confirmed that axonal injury is seen in early evolving MS lesions. criterion of immunoreactivity for myelin oligodendrocyte glycoprotein (MOG) within macrophages in MS lesions (10) defines these as early active lesions, given that MOG is found on the outermost layer of myelin and would thus be one of the first myelin proteins to appear within phagocytic macrophages or microglia. In these early active lesions, high density of immunoreactive betaamyloid precursor protein (β-APP) in axons, a marker of axonal injury, can already be observed. B-APP is a normal constituent of neurons that is transported down axons by fast axoplasmic flow. When an axon is injured, it accumulates to levels that then become detectable by immunohistochemistry (11). Indeed, Kuhlmann et al. (12) have reported the surprising finding that β-APP immunoreactivity is most abundant in active lesions from patients with less than 1-year duration of disease, compared to those with longer disease duration. This finding indicates that either the immune cells migrating into the CNS is most rabid in the initial stages of MS, or that the reaction of the CNS to inflammation is exaggerated early in the disease course. Nonetheless, the results do indicate the need to confer neuroprotection in MS from the very early stages in the disease.

In summary, there is increasing appreciation that MS is not only a degenerative disease involving loss of axons and neurons in addition to myelin and oligodendrocytes, but that the axonal perturbations occur from early on in the disease process (Figure 1). Indeed, in some lesions, axonal loss may even precede the

demyelination (13). The long-term consequence of the early axonal injury is as yet unclear; some perturbed axons may have intrinsic repair capacity to recover fully but it is likely that most will go on to degenerate to account for the progressive axonal loss noted in MS.

4. FACTORS THAT CONTRIBUTE TO THE NEURODEGENERATIVE COMPONENTS OF MS

There are multiple factors that may result in the injury to axons and neurons in MS and almost all these are associated with neuroinflammation. The CNS is normally a region where immune reactivity is kept at a low basal surveillance level, so that a substantial elevation of various inflammatory components as occurs in MS then renders the tissue susceptible to injury. Several studies of autopsied specimens have indicated a good correlation between expression of β-APP or transected axons with the number of activated microglia/macrophages and T cells (1,2,10,14). The products of activated leukocytes include inflammatory cytokines such as interleukin-1 (IL-1) and tumor necrosis factor- α (TNF- α), which have been reported to be toxic to oligodendrocytes and neurons under some conditions (15-17). This is controversial, however, since protective effects of cytokines against neuronal injury have also been observed in certain situations (18,19). Other leukocyte products that may be damaging to neurons and axons radicals, nitric oxide. free metalloproteinases (MMPs) and cell surface associated molecules such as FasL (20-23). Complement activation and the deposition of membrane attack complex is also a potent means to induce axonal injury (24) and microglia

Table 1. A summary of mechanisms that may contribute to axonal and neuronal injury in MS

- Inflammatory cytokines
- Complement-mediated injury
- Direct injury by CD4+ and CD8+ T cells (may involve perforin, FasL, granzyme B)
- Free radicals
- Nitric oxide
- Matrix metalloproteinases and other proteases
- Persistent activation of microglia
- Physical factors including increased extracellular pressure caused by inflammatory edema

activation can result in neuronal death (25,26). A summary of mechanisms that can disrupt axonal and neuronal integrity is listed on Table 1.

Neurons in culture have been shown to be susceptible to toxicity by CD8-positive T cells in an MHC (major histocompatibility complex) Class I-restricted, antigen-specific manner (27,28). In some cases, this requires the forced induction of MHC Class 1 molecules on neurons, through electrical silencing with tetrodotoxin accompanied by interferon-gamma treatment (28,29). Whether this is a reasonable milieu $in\ vivo$ is unclear, as there is controversy on whether neurons $in\ situ$ can express MHC Class 1 molecules. Nonetheless, $\beta 2$ microglobulin, which is often associated with MHC Class 1, can be detected on neurons in brains from Rasmussen encephalomyelitis and these appear in close apposition to T cells, indicating that potential toxicity of T cells to neurons can occur in an MHC dependent manner $in\ vivo$ (30).

5. CAN T CELLS INJURE NEURONS THROUGH ANTIGEN NON-SPECIFIC MECHANISMS?

Although there is good evidence for the initiation of neuroinflammation through antigen-specific mechanisms in models of MS, including experimental autoimmune encephalomyelitis (EAE), many of the T cells that subsequently enter into the CNS may be antigen non-specific. In EAE where antigen-specific cells can be labelled and tracked, Cross *et al.* (31,32) had indicated that less than 2% of perivascular inflammatory cells were antigen-specific and these remained in the perivascular space while the parenchyma was intensely infiltrated with leukocytes that were not labelled (i.e. presumed to be non antigen-specific). Similarly, most infiltrating T cells in MS are thought to be non antigen-specific (33).

More recently, it was demonstrated that the axonal injury in animal models need not depend upon an immune response directed against particular CNS antigens (e.g. an antigen located in axon membrane) (22). These authors first injected heat-killed BCG intracerebrally followed sometime later by a peripheral challenge with BCG. The resultant delayed type hypersensitivity reaction to BCG within the CNS led to axon injury (22).

We have further demonstrated the susceptibility of axons and neurons to antigen non-specific T cells. In

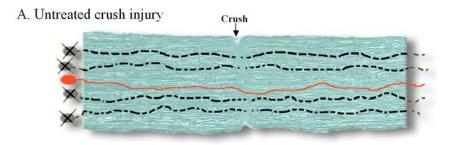
this regard, when T cells obtained from peripheral human blood are non-specifically activated (by anti-CD3 ligation). they acquire the capacity to kill human neurons in culture (34). Indeed, the toxicity of the activated T cells was remarkable, resulting in the loss of over 90% of neurons within 24 hours of co-culture. The mode of toxicity involves the alignment of T cells with axons, subsequently leading to the loss of the axon/neuron unit (34). The cell contact dependant mechanism for T cell killing of neurons was further supported by the observation that toxicity could be attenuated using function-blocking antibodies to FasL, LFA-1, and CD40. As there are no obvious antigens in culture, and given that function-blocking antibodies to MHC Class I or II molecules could not alleviate the toxicity, we would suggest that cell death occurs through antigen-non-specific mechanisms (34). These findings indicate that in some conditions, particularly when axons are exposed and when enough activated T cells accumulate in the CNS regardless of their specificity, axonal injury and subsequent neuronal death can occur.

6. BENEFICIAL ASPECTS OF NEUROINFLAMMATION

Despite the well-documented detrimental consequences of neuroinflammation, it has become evident in recent years that neuroinflammation has beneficial properties as well. In the situation of CNS trauma, such as in spinal cord injury models, the implantation of macrophages or microglia post-injury can alleviate the degenerative process to result in some regeneration of axons (35-37), although the converse has also been found (38,39). Similarly, an initial microglia activation in brain trauma could be a beneficial response, since microglial-derived IL-1β regulates the subsequent production of a growth factor, ciliary neurotrophic factor (CNTF) (40). Many cytokines are recognized to have direct neurotrophic properties, including the induction of neurite extension by neurons in culture (41).

Schwartz and colleagues have championed the concept that T cells can also have beneficial properties (42). The model that these investigators have used involves a crush injury to structures such as the optic nerve. In animals that are not treated following a crush injury to the optic nerve, there is the primary loss of cells in the immediate period at the epicentre of the injury. Over time, however, there is progressive loss of structures, including axons and projection neurons, presumably because these have not been able to recover from the original injury (Figure 2). This delayed death is referred to as secondary degeneration and its extent is substantial. Indeed, in the optic nerve, there is a 25% loss of projection retinal ganglion neurons at 1 week following the crush of the optic nerve, but this increases to 55% by the second week of injury (43). Because secondary degeneration occurs over a period of time, there is optimism that a time window of opportunity exists to alleviate secondary degeneration or to confer neuroprotection following an insult.

Schwartz et al. reported that following a crush injury to the optic nerve, mice injected with myelin-



B. Injected with myelin-reactive T cells after crush

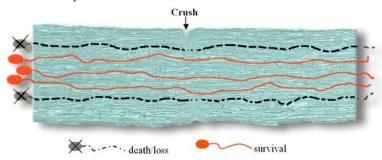


Figure 2. Secondary degeneration and the loss of axons and projection neurons is attenuated if a nerve or tissue (e.g. spinal cord) is treated with myelin-reactive T cells after a crush injury. This is the concept of protective autoimmunity which is also referred to as neuroprotective immunity.

reactive T cells had lower loss of retinal ganglion neurons resulting from the trauma compared to untreated animals (44) (Figure 2). Similar neuroprotection by myelin-reactive cells has also been observed after a spinal cord crush in rats (45). Presumably, these myelin reactive T cells undergo reactivation at the site of the injury by virtue of antigen (myelin components) presentation by microglia or macrophages that have engulfed degenerating myelin. The concept that some T cells can confer neuroprotection is referred to as protective autoimmunity (46) although some have preferred the term neuroprotective immunity (47). It highlights the concept that not all activated T cells are necessarily harmful, but that some of these are even neuroprotective in disease states. Indeed, the suggestion has been put forth that MS is contributed by the malfunctioning of a physiological autoimmune response whose purpose is protective (48). Other groups have since confirmed the beneficial properties of T cells in a variety of conditions, including in facial nerve resection (49), or in revascularization following an aseptic cerebral injury induced by a cold probe to the mouse cerebral cortex (50).

The mechanism of neuroprotective immunity is still unclear although a favored hypothesis is that these leukocytes bring with them neurotrophins and other growth factors into the CNS (Table 2). It is now clear that leukocytes that accumulate in human MS brain lesions (51) or in experimental models of injury (15) express brain derived neurotrophic factor (BDNF), neurotrophin-3 (NT-3) and nerve growth factor (NGF). These neurotrophins are survival factors that have the capacity to promote the well being of injured axons and neurons, and they may also facilitate the survival of oligodendrocytes and thereby

preserve myelin (47,52). Another advantage of neurotrophins is their ability to modulate immune reactivity (53). Some neurotrophins have been shown to decrease antigen presentation (54) and thereby potentially reduce detrimental inflammation in the CNS. NGF has been shown to reduce the transmigration of monocytes across a model of the blood-brain barrier (55) and also to promote a Th2 milieu within the CNS of marmosets (56).

The concept of neuroprotective immunity to treat diseases of the CNS is still in its infancy. It can be a dangerous manoeuvre, particularly since the arrival of many activated T cells can destroy neurons, as pointed out earlier. In the reports that the administration of autoreactive T cells conferred neuroprotection to some axonal tracts, a side effect was the development of EAE in animals (44,45). Others have also noted that the massive accumulation of MBP-specific T cells in experimental models of trauma is degenerative (57,58).

The postulate that neuroprotective immunity involves neurotrophic factors is by no means resolved. Also, whether particular T' cell subsets, including the T helper 1 or 2 effectors, are more neuroprotective than others, remain unclear. There have been other potential explanations for why leukocytes could be neuroprotective (Table 2), including that these cells deliver molecules such as anti-thrombin-3 into the CNS to detoxify thrombin with potential neurotoxic properties (59).

There has also been interest in other neurotrophic factors in the context of EAE and MS. In EAE, mice genetically deficient for CNTF (60) and leukemia

Table 2. Mechanisms of neuroprotective immunity

- Delivery of neurotrophins and other growth factors by leukocytes into the CNS
- Release of other beneficial molecules by leukocytes within the CNS, including anti-thrombin III
- The neurotrophins expressed by leukocytes have immunomodulatory properties
 - Decrease of antigen-presentation
 - Promotion of Th2 anti-inflammatory environment
 - Attenuation of monocyte migration

Please refer to the text for the relevant citations

inhibitory factory (LIFβ) (61) have a worse EAE disease course than wild type mice. Furthermore, many humans have a CNTF mutation that results in the production of biologically inactive truncated protein (62). Although this CNTF mutation does not predispose to MS, MS patients harbouring this mutation have an earlier onset of disease and a worse disease course with strong spinal cord involvement (62). These findings highlight the potential that growth factor therapy may have a role in MS, even though the mechanisms of neuroprotective immunity may not ultimately involve growth factors.

7. NEUROPROTECTIVE CAPACITY OF CURRENTLY USED MS THERAPEUTICS

It is pertinent to evaluate whether current MS treatments may confer neuroprotection in MS. Because glatiramer acetate treatment leads to the formation of glatiramer acetate-specific Th2 cells, which are then thought to migrate into the CNS to produce bystander suppression (63,64), there has been an interest in whether these cells might also be neuroprotective within the CNS As noted earlier, some T cells can confer neuroprotection although the myelin-reactive ones may also produce MS-like inflammatory consequences whilst conferring neuroprotection to some specific tracts. Glatiramer acetate-specific T cells, however, are not known to be pro-inflammatory in nature, and thereby might constitute cells that would be ideal to confer neuroprotective immunity. Indeed, this concept has been tested in experimental animals.

Kipnis *et al.* (65) reported that rats that were injected with glatiramer acetate-specific T cells had improved survival of retinal ganglion neurons after an optic nerve crush compared to untreated animals. Subsequently, it was shown that glatiramer acetate-specific T cells also conferred neuroprotection following facial nerve compression injury (66).

There is another relevant model of neurodegeneration in MS, and this is one that involves the injection of glutamate into the eye to result in death of retinal ganglion neurons. Although a neurotransmitter, glutamate excess and excitotoxicity is implicated in a variety of degenerative processes including in MS. Schori et al. (67) showed that the treatment of animals with glatiramer acetate or glatiramer acetate - reactive T cells reduced the death of retinal ganglion neurons in response to glutamate excitotoxicity. This was observed in conditions in which MOG or MBP-reactive T cells did not confer neuroprotection, indicating that not all T cells are necessarily endowed with neuroprotective functions. Further neuroprotective capacity of glatiramer acetate, at least in experimental animals, is demonstrated by the observation that the use of glatiramer acetate as a "vaccine" improves survival and motor function in a mouse model of amyotrophic lateral sclerosis (66). Finally, it was recently reported that axonal damage in EAE in mice was reduced following glatiramer acetate therapy (68).

Is there evidence that glatiramer acetate is neuroprotective in MS? Evidence for this may be emerging, as one published report notes that glatiramer acetate reduced the proportion of new T1 hypointense lesions that involved into "black holes", which are thought to be signs of irreversible injury (69). Human glatiramer acetate-specific cell lines are known to produce neurotrophins including BDNF (70).

Magnetic resonance spectroscopy studies of the ratio of N-acetylaspartate to creatine (NAA:Cr) offers another measure to address whether GA is neuroprotective in MS. The NAA:Cr ratio is a marker of axonal and neuronal metabolism and this declines in untreated MS individuals on a yearly basis. Using this measure, it was noted that treatment of MS patients with glatiramer acetate for 1 year increased the NAA:Cr ratio compared to pretreatment values (71). These are exciting data and they suggest that glatiramer acetate exerts neuroprotection when used to treat patients with MS.

Is there evidence for neuroprotection by interferon-β in MS? At first glance, it would be reasonable to suppose that by decreasing the infiltration of inflammatory cells into the CNS, a major mode of mechanism for this drug in MS (64), interferon-β would be expected to be neuroprotective, at least through indirect mechanisms (that is, keeping out the pathogenic intruders). In support, Narayanan et al. (72) reported that the mean NAA:Cr ratio by spectroscopy increased by 5.5% in 10 patients on interferon-\beta after 1 year of treatment compared to pre-treatment values. However, this suggestion of neuroprotection was not supported by a subsequent report by Parry et al. (73) in which the mean NAA:Cr ratio decreased by 6.2% after 1 year of treatment. Finally, with respect to the proportion of contrast enhancing lesions that involved into black holes, Bagnato et al. (74) found no evidence of a reduction in this parameter in 10 relapsingremitting MS patients given interferon-β.

On balance, it appears that glatiramer acetate may have the capacity to confer neuroprotection. The case for interferon- β is less strong, although it may be argued that decreasing pathogenic inflammation from entering the CNS (even at the cost of blocking the entry of some beneficial ones) would ultimately lead to resolution of damaging inflammation, and thereby neuroprotection. However, this indirect mode may not offer protection to neural elements already undergoing degeneration, as might be conferred by the physical presence of glatiramer acetate-reactive cells

Table 3. A summary of potential neuroprotection by established or experimental MS therapeutics and their

Mechanisms of protection	Glatiramer acetate	Interferon-β	Minocycline
Indirect Mechanisms (e.g.	Likely	Likely	Likely
decrease of CNS inflammation)			
Direct mechanisms within the	Likely	Not Likely	Likely
CNS (e.g. by production of		-	
growth factors within the CNS)			

that are directly within the CNS to elaborate their beneficial contents (Table 3).

8. OTHER POTENTIAL NEUROPROTECTIVE AGENTS FOR MS

As yet, there has been no evidence of neuroprotective roles for other immunomodulators that are employed in MS, including mitoxantrone. cyclophosphamide, IVIG and others. Experimental therapies offer a more optimistic future. Minocycline is a tetracycline antibiotic that is taken orally for the treatment of conditions such as acne. It is a small molecule that diffuses well into the CNS. It produces a low frequency of antibiotic resistance and therefore has been used in the long-term management of patients with acne. In the United Kingdom alone, over 6.5 million individuals have been treated for an average of 9 months with minocycline. It has many activities, including the potential to inhibit the activity of the matrix metalloproteinases (MMPs)

The MMPs are a family of proteolytic enzymes that are implicated in the physiological remodelling of the While having many extracellular matrix (ECM). physiological functions, the MMPs are also upregulated in a variety of CNS disorders including MS. In the context of MS, upregulated MMPs may be utilized by leukocytes to infiltrate into the CNS; MMPs may also produce encephalogens from myelin proteins or it may be directly or indirectly cytotoxic (76). We have used minocycline, originally because of its capacity to inhibit MMP activity as described by others. In this regard, we have shown that minocycline decreases the activity and level of MMPs produced by T cells, and that it attenuates T cell transmigration across a model of the blood brain barrier in vitro. In EAE, minocycline administration reduces the clinical severity, neuroinflammation and demyelination (77). Similar efficacy of minocycline in EAE was also reported by Popovic et al. (78).

It turns out that minocycline has a myriad of activities, including the inhibition of microglial activation, the inhibition of activity of caspases-1 and -3, which are involved in the regulation of apoptosis (79-81). Minocycline has also been reported to inhibit glutamate excitotoxicity (82). Because of these multiple activities, we have asked whether minocycline might be able to confer neuroprotection. Because EAE is a variable disease in which the inflammation across a section of the spinal cord is not consistent across animals, it is difficult to address possible neuroprotection of minocycline in EAE. For this reason, we have used minocycline in a

compression injury to the spinal cord of mice. We have determined that the administration of minocycline from 1 hour after the injury resulted in the attenuation of the lesion size in the spinal cord, axonal sparing, and also improves the motor behaviour of animals (83). These results suggest the prospects of minocycline not only as an immunomodulator in MS, but also as a potential neuroprotective agent in the disease (Table 3).

A Phase I trial of Minocycline in MS has been concluded and the results have been encouraging (84; Metz et al., manuscript in preparation). Future studies will clarify whether this potentially useful agent will have utility in MS, either when used in isolation, or when combined with existing MS therapeutics.

Finally, the sodium channel blocker, phenytoin, has been found to be neuroprotective in EAE (85); whether this will have utility in MS deserves attention. It is likely that the literature on neuroprotection in EAE, and hopefully in MS, will expand significantly in the near future.

9. PERSPECTIVE

As the pathology of MS becomes increasingly understood, it has become clear that this is not only an inflammatory demyelinating disease, but also a degenerative one. Thus, useful therapies must combine immunomodulatory actions with neuroprotective ones. Of the current available immunomodulators, there is stronger evidence for possible neuroprotective efficacy of glatiramer acetate compared to other drugs. It is also becoming clear that not all aspects of neuroinflammation are necessarily harmful and that the non-specific inhibition of inflammation may be detrimental. There are potential new therapies in MS and the example of minocycline has been noted. The prospect of conferring neuroprotection in MS is good, and this bodes well for the prognosis of the disease.

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11. REFERENCES

- 1. Ferguson B, MK Matyszak, MM Esiri & VH Perry: Axonal damage in acute multiple sclerosis lesions. *Brain* 120, 393-399 (1997)
- 2. Trapp BD, J Peterson, RM Ransohoff, R Rudick, S Mork & L Bo: Axonal transection in the lesions of multiple sclerosis. *N Engl J Med* 338, 278-285 (1998)
- 3. Bjartmar C, G Kidd & RM Ransohoff: A real-time insight into disease progression and the role of axonal injury in multiple sclerosis. *Arch Neurol* 58,37-39 (2001)
- 4. Trapp BD, RM Ransohoff, E Fisher & RA Rudick: Neurodegeneration in multiple sclerosis: relationship to neurological disability. *The Neuroscientist* 5, 48-57 (1999)
- 5. Cifelli A, M Arridge, P Jezzard, MM Esiri, J Palace & PM Matthews: Thalamic neurodegeneration in multiple sclerosis. *Ann Neurol* 52, 650-653 (2002)
- 6. Bermel RA, MD Innus, CW Tjoa & R Bakshi: Selective caudate atrophy in multiple sclerosis: a 3D MRI parcellation study. *Neuroreport* 14, 335-339 (2003)
- 7. Sailer M, B Fischl, D Salat, C Tempelmann, MA Schonfeld, E Busa, N Bodammer, HJ Heinze & A Dale: Focal thinning of the cerebral cortex in multiple sclerosis. *Brain* 126, 1734-1744 (2003)
- 8. De Stefano N, S Narayanan, SJ Francis, S Smith, M Mortilla, MC Tartaglia, ML Bartolozzi, L Guidi, A Federico & DL Arnold: Diffuse axonal and tissue injury in patients with multiple sclerosis with low cerebral lesion load and no disability. *Arch Neurol* 59, 1565-1571 (2002)
- 9. Filippi M, M Bozzali, M Rovaris, O Gonen, C Kesavadas, A Ghezzi, V Martinelli, RI Grossman, G Scotti, G Comi & A Falini: Evidence for widespread axonal damage at the earliest clinical stage of multiple sclerosis. *Brain* 126, 433-437 (2003)
- 10. Kornek B, MK Storch, R Weissert, E Wallstroem, A Stefferl, T Olsson, C Linington, M Schmidbauer & H Lassmann: Multiple sclerosis and chronic autoimmune encephalomyelitis: a comparative quantitative study of axonal injury in active, inactive, and remyelinated lesions. *Am J Pathol* 157, 267-276 (2000)
- 11. Gentleman SM, MJ Nash, CJ Sweeting, DI Graham & GGW Roberts: Beta-amyloid precursor protein as a marker of axonal injury after head injury. *Neurosci Lett* 160, 139-144 (1993)
- 12. Kuhlmann T, G Lingfeld, A Bitsch, J Schuchardt & W Bruck: Acute axonal damage in multiple sclerosis is most extensive in early disease stages and decreases over time. *Brain* 125, 2202-2212 (2002)
- 13. Bjartmar C, RP Kinkel, G Kidd, RA Rudick & BD Trapp: Axonal loss in normal-appearing white matter in a patient with acute MS. *Neurology* 57, 1248-1252 (2001)
- 14. Bitsch A, J Schuchardt, S Bunkowski, T Kuhlmann & W Bruck: Acute axonal injury in multiple sclerosis Correlation with demyelination and inflammation. *Brain* 123, 1174-1183 (2000)
- 15. Hammarberg H, O Lidman, C Lundberg, SY Eltayeb, AW Gielen, S Muhallab, A Svenningsson, H Linda, PH van Der Meide, S Cullheim, T Olsson, F Piehl: Neuroprotection by encephalomyelitis: rescue of mechanically injured neurons and neurotrophin production by CNS-infiltrating T and natural killer cells. *J Neurosci* 20, 5283-5291 (2000)

- 16. Allan SM & NJ Rothwell: Cytokines and acute neurodegeneration. *Nat Rev Neurosci* 2, 734-744 (2001)
- 17. Nadeau S & S Rivest: Glucocorticoids play a fundamental role in protecting the brain during innate immune response. *J Neurosci* 23, 5536 (2003)
- 18. Cheng B, S Christakos & MP Mattson: Tumor necrosis factors protect neurons against metabolic-excitotoxic insults and promote maintenance of calcium homeostasis. *Neuron* 12, 139-153 (1994)
- 19. Burger SW, D Horster, K Furukawa, Y Goodman, J Krieglstein & MP Mattson: Tumor necrosis factors alpha and beta protect neurons against amyloid beta-peptide toxicity: evidence for involvement of a kappa B-binding factor and attenuation of peroxide and Ca2+ accumulation. *PNAS* 92, 9328-9332 (1995)
- 20. Redford EJ, R Kapoor & KJ Smith: Nitric oxide donors reversibly block axonal conduction: demyelinated axons are especially susceptible. *Brain* 120, 2149-2157 (1997)
- 21. Kieseier BC, MK Storch, JJ Archelos, G Martino & HP Hartung: Effector pathways in immune mediated central nervous system demyelination. *Curr Opin Neurol* 12, 323-336 (1999)
- 22. Newman TA, ST Woolley, PM Hughes, NR Sibson, DC Anthony & VH Perry: T-cell- and macrophage-mediated axon damage in the absence of a CNS- specific immune response: involvement of metalloproteinases. *Brain* 124, 2203-2214 (2001)
- 23. Kapoor R, M Davies, PA Blaker, SM Hall & KJ Smith: Blockers of sodium and calcium entry potect axons from nitric oxide- mediated degeneration. *Ann Neurol* 53, 174-180 (2003)
- 24. Mead RJ, SK Singhrao, JW Neal, H Lassmann & BP Morgan: The membrane attack complex of complement causes severe demyelination associated with acute axonal injury. *J Immunol* 168, 458-465 (2002)
- 25. Chao CC, S Hu, TW Molitor, EG Saskan & PK Peterson: Activated microglia mediate neuronal cell injury via a nitric oxide mehanism. *J Immunol* 149, 2736-2741 (1992)
- 26. Choi SH, EH Joe, SU Kim & BK Jin: Thrombin-induced microglial activation produces degeneration of nigral dopaminergic neurons in vivo. *J Neurosci* 23, 5877-5886 (2003)
- 27. Johnson AJ, J Upshaw, KP Pavelko, M Rodriguez & LP Pease: Preservation of motor function by inhibition of CD8(+) virus peptide-specific T cells in Theiler's virus infection. *FASEB J* 15, 2760-2762 (2001)
- 28. Medana I, MA Martinic, H Wekerle & H Neumann: Transection of major histocompatibility complex class I-induced neurites by cytotoxic T lymphocytes. *Am J Pathol* 159, 809-815 (2001)
- 29. Neumann H, A Cavalie, DE Jenne & H Wekerle: Induction of MHC class I genes in neurons. *Science* 269, 549-552 (1995)
- 30. Bien CG, J Bauer, TL Deckwerth, H Wiendl, M Deckert, OD Wiestler, J Schramm, CE Elger & H Lassmann: Destruction of neurons by cytotoxic T cells: a new pathogenic mechanism in Rasmussen's encephalitis. *Ann Neurol* 51, 311-318 (2002)
- 31. Cross AH, B Cannella, CF Brosnan & CS Raine: Homing to central nervous system vasculature by antigen-

- specific lymphocytes I Localization of 14C-labeled cells during acute, chronic, and relapsing experimental allergic encephalomyelitis. *Lab Invest* 63, 162-170 (1990)
- 32. Cross AH, T O' Mara & CS Raine: Chronologic localization of myelin-reactive cells in the lesions of relapsing EAE. *Neurol* 43, 1028-1033 (1993)
- 33. Wucherpfennig, K W, J Newcombe, H Li, C Keddy, M L Cuzner & D A Hafler: T cell receptor V alpha-V beta repertoire and cytokine gene expression in active multiple sclerosis lesions. *J Exp Med* 175, 993 (1992)
- 34. Giuliani F, CG Goodyer & VW Yong: Selective and remarkable vulnerability of human neurons to T cell-mediated cytotoxicity. *J Immunol* 171, 368-379 (2003)
- 35. Lazaroc-Spiegler O, AS Solomon, AB Zeev-Brann, DL Hirschberg, V Lavie & M Schwartz: Transplantation of activated macrophages overcomes central nervous system regrowth failure. *FASEB J* 10, 1296-1302 (1996)
- 36. Prewitt CM, IR Niesman, CJ Kane & JD Houle: Activated macrophage/microglial cells can promote the regeneration of sensory axons into the injured spinal cord. *Exp Neurol* 148, 433-443 (1997)
- 37. Rabchevsky AG & WJ Streit: Grafting of cultured microglial cells into the lesioned spinal cord of adult rats enhances neurite outgrowth. *J Neurosci Res* 47, 34-48 (1997)
- 38. Popovich PG, Z Guan, P Wei, I Huitinga, N van Rooijen & BT Stokes: Depletion of hematogenous macrophages promotes partial hindlimb recovery and neuroanatomical repair after experimental spinal cord injury. *Exp Neurol* 158, 351-365 (1999)
- 39. Mabon PJ, LC Weaver & GA Dekaban: Inhibition of monocyte/macrophage migration to a spinal cord injury site by an antibody to the integrin alphaD: a potential new anti-inflammatory treatment. *Exp Neurol* 166, 52-64 (2000)
- 40. Herx LM, S Rivest & VW Yong: Central nervous system-initiated inflammation and neurotrophism in trauma: IL-1β is required for the production of ciliary neurotrophic factor. *J Immunol* 165, 2232-2239 (2000)
- 41. Yong VW: Cytokines, astrogliosis and neurotrophism following CNS trauma, in: Cytokines and the CNS: *Development, Defense and Disease*, Ed: E Benveniste, R Ransohoff, CRC Press, Boca Raton, Florida, pp 309-327 (1996)
- 42. Schwartz M & G Moalem: Beneficial immune activity after CNS injury: prospects for vaccination. *J Neuroimmunol* 113, 185-192 (2001)
- 43. Yoles E & M Schwartz: Degeneration of spared axons following partial white matter lesion: implications for optic nerve neuropathies. *Exp Neurol* 153, 1-7 (1998)
- 44. Moalem G, R Leibowitz-Amit, E Yoles, F Mor, IR Cohen & M Schwartz: Autoimmune T cells protect neurons from secondary degeneration after central nervous system axotomy. *Nat Med* 5, 49-55 (1999)
- 45. Hauben E, O Butovsky, U Nevo, E Yoles, G Moalem, E Agranov, F Mor, R Leibowitz-Amit, E Pevsner, S Akselrod, M Neeman, IR Cohen & M Schwartz: Passive or active immunization with myelin basic protein promotes recovery from spinal cord contusion. *J Neurosci* 20, 6421-6430 (2000)
- 46. Schwartz M & J Kipnis: Protective autoimmunity: regulation and prospects for vaccination after brain and spinal cord injuries. *Trends Mol Med* 7, 252-258 (2001)

- 47. Kerschensteiner M, C Stadelmann, G Dechant, H Wekerle & R Hohlfeld: Neurotrophic cross-talk between the nervous and immune systems: Implications for neurological diseases. *Ann Neurol* 53, 292-304 (2003)
- 48. Schwartz M & J Kipnis: Multiple sclerosis as a byproduct of the failure to sustain protective autoimmunity: a paradigm shift. *Neuroscientist* 8, 405-413 (2002)
- 49. Serpe CJ, AP Kohm, CB Huppenbauer, VM Sanders & KJ Jones: Exacerbation of facial motoneuron loss after facial nerve transection in severe combined immunodeficient (scid) mice. *J Neurosci* 19, RC7 (1999)
- 50. Hofstetter HH, DL Sewell, F Liu, M Sandor, T Forsthuber, PV Lehmann & Z Fabry: Autoreactive T cells promote post-traumatic healing in the central nervous system. *J Neuroimmunol* 2003, 134: 25-34
- 51. Kerschensteiner M, E Gallmeier, L Behrens, VV Leal, T Misgeld, WE Klinkert, R Kolbeck, E Hoppe, RL Oropeza-Wekerle, I Bartke, C Stadelmann, H Lassmann, H Wekerle & R Hohlfeld: Activated human T cells, B cells, and monocytes produce brain-derived neurotrophic factor in vitro and in inflammatory brain lesions: a neuroprotective role of inflammation? *J Exp Med* 189, 865-870 (1999)
- 52. McTigue DM, PJ Horner, BT Stokes, FHJ Gage: Neurotrophin-3 and brain-derived neurotrophic factor induce oligodendrocyte proliferation and myelination of regenerating axons in the contused adult rat spinal cord. *J Neurosci* 18, 5354-5365 (1998)
- 53. Hohlfeld R, M Kerschensteiner, C Stadelmann, H Lassmann, H Wekerle: The neuroprotective effect of inflammation: implications for the therapy of multiple sclerosis. *J Neuroimmunol* 107, 161-166 (2000)
- 54. Neumann H, T Misgeld, K Matsumuro & H Wekerle: Neurotrophins inhibit major histocompatibility class II inducibility of microglia: involvement of the p75 neurotrophin receptor. *Proc Natl Acad Sci* U S A 95, 5779-5784 (1998)
- 55. Flugel A, K Matsumuro, H Neumann, WE Klinkert, R Birnbacher, H Lassmann, U Otten & H Wekerle: Anti-inflammatory activity of nerve growth factor in experimental autoimmune encephalomyelitis: inhibition of monocyte transendothelial migration. *Eur J Immunol* 31, 11-22 (2001)
- 56. Villoslada P, SL Hauser, I Bartke, J Unger, N Heald, D Rosenberg, SW Cheung, WC Mobley, S Fisher & CP Genain: Human nerve growth factor protects common marmosets against autoimmune encephalomyelitis by switching the balance of T helper cell type 1 and 2 cytokines within the central nervous system. *J Exp Med* 191, 1799-1806 (2000)
- 57. Jones TB, DM Basso, A Sodhi, JZ Pan, RP Hart, RC MacCallum, S Lee, CC Whitacre & PG Popovich: Pathological CNS autoimmune disease triggered by traumatic spinal cord injury: implications for autoimmune vaccine therapy. *J Neurosci* 22, 2690-2700 (2002)
- 58. Fee D, A Crumbaugh, T Jacques, B Herdrich, D Sewell, D Auerbach, S Piaskowski, MN Hart, M Sandor & Z Fabry: Activated/effector CD4(+) T cells exacerbate acute damage in the central nervous system following traumatic injury. *J Neuroimmunol* 136, 54-66 (2003)
- 59. Friedmann I, E Hauben, E Yoles, L Kardash & T Schwartz: T cell-mediated neuroprotection involves antithrombin activity. *J Neuroimmunol* 121, 12-21 (2001)

- 60. Linker RA, M Maurer, S Gaupp, R Martini, B Holtmann, R Giess, P Rieckmann, H Lassmann, KV Toyka, M Sendtner & R Gold: CNTF is a major protective factor in demyelinating CNS disease: a neurotrophic cytokine as modulator in neuroinflammation. *Nat Med* 8, 620-624 (2002)
- 61. Butzkueven H, JG Zhang, M Soilu-Hanninen, H Hochrein, F Chionh, KA Shipham, B Emery, AM Turnley, S Petratos, M Ernst, PF Bartlett, TJ Kilpatrick: LIF receptor signaling limits immune-mediated demyelination by enhancing oligodendrocyte survival. *Nat Med* 8, 613-619 (2002)
- 62. Giess R, M Maurer, R Linker, R Gold, M Warmuth-Metz, KV Toyka, M Sendtner & P Rieckmann: Association of a null mutation in the CNTF gene with early onset of multiple sclerosis. *Arch Neurol* 59, 407-409 (2002)
- 63. Neuhaus O, C Farina, H Wekerle & R Hohlfeld: Mechanisms of action of glatiramer acetate in multiple sclerosis. *Neurology* 56, 702-708 (2001)
- 64. Yong VW: Differential mechanisms of action of interferon-β and glatiramer acetate in MS. *Neurology* 59, 802-808 (2002)
- 65. Kipnis J, E Yoles, Z Porat, A Cohen, F Mor, M Sela, IR Cohen & M Schwartz: T cell immunity to copolymer 1 confers neuroprotection on the damaged optic nerve: possible therapy for optic neuropathies. *Proc Natl Acad Sci U S A* 97, 7446-7451 (2000)
- 66. Angelov DN, S Waibel, O Guntinas-Lichius, M Lenzen, WF Neiss, TL Tomov, E Yoles, J Kipnis, H Schori, A Reuter, A Ludolph & M Schwartz: Therapeutic vaccine for acute and chronic motor neuron diseases: Implications for amyotrophic lateral sclerosis. *Proc Natl Acad Sci U S A* 100, 4790-4795 (2003)
- 67. Schori H, J Kipnis, E Yoles, E WoldeMussie, G Ruiz, LA Wheeler & M Schwartz: Vaccination for protection of retinal ganglion cells against death from glutamate cytotoxicity and ocular hypertension: Implications for glaucoma. *Proc Natl Acad Sci U S A* 98, 3398-3403 (2001)
- 68. Gilgun-Sherki Y, H Panet, V Holdengreber, R Mosberg-Galili & D Offen: Axonal damage is reduced following glatiramer acetate treatment in C57/b1 mice with chronig-induced experimental autoimmune encephalomyelitis. *Neurosci Res* 47, 201-207, 2003
- 69. Filippi M, M Rovaris, MA Rocca, MP Sormani, JS Wolinsky & G Comi: Glatiramer acetate reduces the proportion of new MS lesions evolving into "black holes". *Neurology* 57, 731-733 (2001)
- 70. Ziemssen T, T Kumpfel, WEF Klinkert, O Neuhaus & R Hohlfeld: Glatiramer acetate-specific T-helper 1- and 2-type cell lines produce BDNF: implications for multiple sclerosis therapy. *Brain* 125, 2381-2391 (2002)
- 71. Khan O, Y Shen, W Ching, C Caon, M Reznar, R Lisak, Z Latif, J Hu, V Sehgal: Combining immunomodulation and neuroprotection: Cerebral axonal recovery in relapsing-remitting multiple sclerosis patients treated with glatiramer acetate. *Multiple Sclerosis* 9 (Suppl 1), S63 (2003)
- 72. Narayanan S, N De Stefano, GS Francis, R Arnaoutelis, Z Caramanos & DL Collins: Axonal metabolic recovery in multiple sclerosis patients treated with interferon beta-1b. *J Neurol* 248, 979-986 (2001)
- 73. Parry A, R Corkill, AM Blamire, J Palace, S Narayanan, Arnold D, P Styles & PM Matthews: Beta-Interferon treatment

- does not always slow the progression of axonal injury in multiple sclerosis. *J Neurol* 250, 171-178 (2003)
- 74. Bagnato F, N Jeffries, N Richert, O Joan, S Roger, C Bash, HF McFarland & JA Frank: Interferon beta therapy does not affect duration in time of Gd-enhancing lesions and black holes in relapsing remitting Multiple Sclerosis. *Neurol* 60(Suppl 1), A300 (2003)
- 75. Paemen L, E Martens, K Norga, S Masure, E Roets, J Hoogmartens & G Opdenakker: The gelatinase inhibitory activity of tetracyclines and chemically modified tetracycline analogues as measured by a novel microtiter assay for inhibitors. *Biochem Pharmacol* 52, 105-111 (1996)
- 76. Yong VW, C Power, P Forsyth & DR Edwards: Metalloproteinases in biology and pathology of the nervous system. *Nat Rev Neurosci* 2, 502-511(2001)
- 77. Brundula V, NB Rewcastle, LM Metz, CC Bernard & VW Yong: Targeting leukocyte MMPs and transmigration: minocycline as a potential therapy for multiple sclerosis. *Brain* 125, 1297-1308 (2002)
- 78. Popovic N, A Schubart, BD Goetz, SC Zhang, C Linington & ID Duncan: Inhibition of autoimmune encephalomyelitis by a tetracycline. *Ann Neurol* 51, 215-223 (2002)
- 79. Yrjanheikki J, R Keinanen, M Pellikka, T Hokfelt & J Koistinaho: Tetracyclines inhibit microglial activation and are neuroprotective in global brain ischemia. *Proc Natl Acad Sci U S A* 95, 15769-15774 (1998)
- 80. Chen M, VO Ona, M Li, RJ Ferrante, KB Fink, S Zhu, J Bian, L Guo, LA Farrell, SM Hersch, W Hobbs, JP Vonsattel, JH Cha, RM Friedlander: Minocycline inhibits caspase-1 and caspase-3 expression and delays mortality in a transgenic mouse model of Huntington disease. *Nat Med* 6,797-801 (2000)
- 81. Zhu S, I Stavrovskaya, M Drozda, B Kim, V Ona, Li M, S Sarang, AS Liu, DM Hartley, C Wu du, S Gullans, RJ Ferrante, S Przedborski, BS Kristal & RM Friedlander: Minocycline inhibits cytochrome c release and delays progression of amyotrophic lateral sclerosis in mice. *Nature* 417, 74-78, 2002
- 82. Tikka T, BL Fiebich, G Goldsteins, R Keinanen & J Koistinaho: Minocycline, a tetracycline derivative, is neuroprotective against excitotoxicity by inhibiting activation and proliferation of microglia. *J Neurosci* 21, 2580-2588 (2001)
- 83. Wells J, J Hurlbert, M Fehlings & VW Yong: Neuroprotection by Minocycline Facilitates Significant Recovery from Spinal Cord Injury. *Brain* 126, 1628-1637 (2003)
- 84. Metz LM, Yong VW, Yeung M, Yan Y, Mitchell R, Openlabel trial of Minocycline in active relapsing-remitting multiple sclerosis. *Neurology* 2003;60(Suppl 1)A250(Abstract S31.001) 85. Lo AC, JA Black & SG Waxman: Neuroprotection of axons with phenytoin in experimental allergic encephalomyelitis. *Neuroreport* 13, 1909-1912 (2002)
- **Key Words:** Nervous system, Brain, Glatiramer Acetate, Interferon-β, Minocycline, Multiple sclerosis, Neuroprotection, Review
- **Send correspondence to:** Dr V. Wee Yong, University of Calgary, 3330 Hospital Drive N.W., Calgary, Alberta T2N 4N1, Canada, Tel:403-220-3544, Fax:403-283-8731, E-mail: vyong@ucalgary.ca