PATHOPHYSIOLOGIC ROLE OF SELECTINS AND THEIR LIGANDS IN ISCHEMIA REPERFUSION INJURY

Fady Chamoun, Melissa Burne, Michael O'Donnell and Hamid Rabb

Division of Nephrology, Hennepin County Medical Center, University of Minnesota Medical School, Minneapolis, MN 55415

TABLE OF CONTENTS

 Abstract.
Introduction
Pathophysiology of ischemia-reperfusion injury 3.1. Selectins 3.2. Selectin ligands
Clinical implications 4.1. Brain 4.2. Heart 4.3. Kidneys 4.4. Lungs 4.5. Liver
Novel perspectives
References

1. ABSTRACT

Research findings are unveiling the potential role of leukocytes and leukocyte adhesion molecules such as selectins in ischemia-reperfusion injury (IRI). "Antiadhesion" therapy using selectin blocking agents may represent a new approach to treatment of the many diverse clinical disorders in which ischemia-reperfusion occurs, including transplantation, reperfusion after thrombotic events and shock. In this paper we review the pathophysiology of IRI, the different types of selectins and selectin ligands, the clinical implications of selectin blockade in different organs with IRI, and new insights into mechanism of action.

2. INTRODUCTION

Recent developments in immunology and cell biology have demonstrated the importance of inflammation in the pathogenesis of post-ischemic organ dysfunction. Whereas prolonged ischemia causes anoxic cell death, recent evidence suggests that sublethal injury may be amplified by inflammatory and cytotoxic injury cascades activated during the reperfusion period.

Ischemia is a state of tissue oxygen deprivation accompanied by a reduced washout of the resulting metabolites (1). Reperfusion is the restoration of blood flow to the ischemic tissue. Despite the unequivocal benefit of reperfusion of blood to an ischemic tissue, reperfusion itself can elicit a cascade of adverse reactions that paradoxically injure tissue (2). Indeed, reperfusion injury has been well described in the literature to cause organ damage in the brain, heart, lungs, liver, kidneys and skeletal muscle. The susceptibility of tissue to ischemia reperfusion injury (IRI) is a major obstacle to both reperfusion after an infarct and successful organ transplantation.

The incidence and implications of ischemic injury are enormous: ischemia occurs in myocardial infarction, stroke, organ procurement injury, and many other situations. In 1995, ischemic cardiovascular events alone were the diagnosis in 5 million (16.2%) hospital patient discharge records and were the leading cause of death in the United States (38.7%) (3).

A growing body of evidence, primarily from animal models of IRI and preliminary human studies has revealed that inflammatory mechanisms play a major role in the pathogenesis of IRI. Interest in the inflammatory response to IRI has led to the identification of multiple inflammatory mediators, including leukocytes, leukocyte adhesion molecules and cytokines.

In the following review we will focus on the pathophysiology of ischemia-reperfusion, the role of leukocytes in the post-ischemic inflammatory cascade, and the crucial role of the selectin family and their ligands in mediating leukocyte induced post-ischemic injury.

3. PATHOPHYSIOLOGY OF ISCHEMIA-REPERFUSION INJURY

The pathophysiology of the IRI is complex. The inflammatory aspect of IRI includes both the cellular and humoral components (Figure 1). Moreover, mechanisms of IRI may be organ-dependent, with similar but distinct pathways involved in different organs. During the last decade there has been an explosion of research

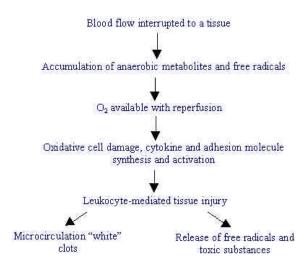


Figure 1. Proposed inflammatory cascade in ischemiareperfusion injury.

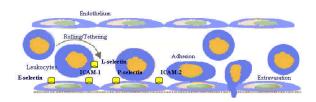


Figure 2. Current model of leukocyte migration to inflammed tissue.

documenting the role of leukocytes and leukocyte adhesion molecules in IRI (4). Multiple mechanisms have been postulated for the leukocyte-mediated tissue injury that occurs after ischemia-reperfusion. Microvascular occlusion (5), release of oxygen free radical (6), cytotoxic enzyme release (7), increased vascular permeability (8) and increased cytokine release (9) have all been demonstrated to contribute to leukocyte-induced tissue injury.

Leukocyte adhesion molecules (LAMs) are expressed on leukocytes and other cell types and regulate many leukocyte functions. LAMs function in various biological processes such as development, signaling, inflammation and apoptosis. There currently exists a multistep paradigm of leukocyte emigration to inflamed tissue that involves specific leukocyte adhesion molecules (Figure 2). To emigrate into tissue, leukocytes initially tether to and roll on the vascular endothelium. This relatively loose adhesion is mediated by the selectin family of adhesion molecules and their ligands (10-12). Subsequent activation of leukocytes and the endothelium leads to firmer adhesion mediated through integrin adhesion molecules (e.g., CD18) and their receptors (e.g., intercellular adhesion molecule-1, ICAM-1). Leukocyte transmigration is the final stage and occurs between endothelial cells. Leukocytes then travel through the extracellular matrix to the source of tissue injury, guided by a concentration gradient of cytokines and chemokines produced at the site of injury.

Because selectin engagement of leukocytes precedes CD11/CD18 binding to ICAM-1, targeting selectins is an attractive way to block inflammation at an even earlier step. In addition, selectin blockade might cause less susceptibility to bacterial infections than does CD11/CD18 blockade (13).

3.1. SELECTINS

The selectin family of leukocyte adhesion molecules consists of three known members: L-, P-, and Eselectin (table 1). These molecules are involved in the initial adhesion of leukocytes to activated endothelium at a site of tissue injury (14-19). L-selectin is a cell surface glycoprotein expressed constitutively on a wide variety of leukocytes (15). L-selectin plays a role in the emigration of lymphocytes into peripheral lymph nodes and sites of chronic inflammation and of neutrophils into acute inflammatory sites. P-selectin is a cell surface glycoprotein that also plays a critical role in the emigration of leukocytes into tissues (17). P-selectin is constitutively stored in the Weibel-Palade bodies of endothelial cells and in the alpha granules of platelets. It is expressed on the cell surface within minutes after exposure to stimuli such as thrombin. E-selectin expression is largely restricted to endothelial cells activated by different stimuli such as endotoxin and the pro-inflammatory cytokines interleukin-1 (IL-1) and tumor necrosis factor (TNF) (19). E-selectin expression peaks within 4-8 hours of tissue injury and returns to baseline by 24 hours (16). E-selectin expression, as with Pselectins, requires de novo messenger RNA and protein synthesis, and involves the translocation of the transcription factor NF-kappaB onto nuclear promoter sites (20).

In the first 10-20 minutes after tissue injury, leukocyte rolling on the vascular endothelium is mainly mediated by P-selectin, with minimal L-selectin contribution (14). This is consistent with the rapid mobilization of P-selectin from intracellular stores after tissue injury. After approximately 20 minutes, the role of Pselectin diminishes secondary to internal degradation, and L-selectin becomes the principal mediator of leukocyte rolling. There is little appreciable role for E-selectin in leukocyte rolling during the early response (<2 hrs) to tissue injury. While E-selectin expression on stimulated endothelial surfaces is detectable at 2 hrs after injury, the delay in peak surface expression precludes the contribution of E-selectin to the early rolling and leukocyte recruitment in acute inflammation. Rather, initial interactions between leukocytes and the vascular endothelium, mediated via Pand L-selectins, are followed by a stronger interactions, probably involving E-selectin and subsequently integrins, that lead eventually to extravasation through the blood vessel wall into lymphoid tissues and to sites of inflammation (16).

3.2. Selectin ligands

While initial studies suggested that all three selectins recognize carbohydratescontaining the sialyl Lewis x antigen (NeuAc $\alpha 2 \rightarrow 3$ Gal $\beta 1 \rightarrow 4$ [Fuc $\alpha 1 \rightarrow 3$] GlcNAc $\beta 1 \rightarrow R$) (sialy vl Le^x), however, recent studies have now shown that each selectin demonstrates higher affinity binding to specific macromolecular ligands expressing

Selectin	Expressing Cells	Ligands	Expression	Structure
L-selectin	Leukocytes	Sialylated	Constituent of cellular	NH2=L=E=CC==COOH
		Lewis X	surfaces	
P-selectin	Platelet &	PSGL-1, sialylated	Constituent of cellular	NH2=L=E=CC==COOH
	Endothelium	Lewis X	surface & induced	
E-selectin	Endothelium	ESGL-1, PSGL-1, sialylated	Induced	NH2=L=E=CC==COOH
		Lewis X		

Table 1. Structure and expression of selectins

PSGL-1 = platelet selectin glycoprotein ligand; ESGL-1 = endothelial selectin glycoprotein ligand; L= lectin; E= Epidermal growth factor-like molecule; C= Complement-binding protein-like.

sialylated and fucosylated glycans. To date the best characterized cell adhesion ligand for selectins is the P-selectin glycoprotein ligand-1 (PSGL-1). Although PSGL-1 is expressed on the membrane surfaces of all leukocytes, with respect to binding of P-selectin, it is only functional on granulocytes and subclasses of lymphocytes. Interestingly, PSGL-1 may also serve as a ligand for both E- and L-selectin (21).

Treatment of purified PSGL-1 with sialidase abolishes its binding to P-selectin, confirming cell studies that indicate a role for sialic acid in P-selectin recognition (22,23). Interestingly, treatment of neutrophil-derived PSGL-1 with peptide N-glycosidase F, which removes most of the Nglycans of the molecule, does not affect its recognition by Pselectin, suggesting that O-glycans, but not N-glycans, are important determinants of selectin binding (22). PSGL-1 may also be an important signaling molecule in neutrophils. Upon activation of polymorphonuclear leukocytes there is a redistribution of PSGL-1 resulting in a lowering of affinity of activated cells for P-selectin (23). Incubation of neutrophils with either P-selectin or the monoclonal antibody PL1 to PSGL-1 stimulates tyrosine phosphorylation of several proteins and production of IL-8 (24).

In vivo approaches to studying the importance of glycans in selectin ligand function have recently provided exciting new insights and have partly confirmed predictions about the importance of sialyl Le^x and core-2 O-glycans for PSGL-1 recognition by P-selectin. Neutrophils from null mice lacking the myeloid enzyme fucosyltransferase VII bind poorly to P-, E- or L-selectin, and neutrophil efflux in experimentally induced inflammation in such mice is dramatically reduced (25). Another approach to studying PSGL-1 function during in vivo inflammation has been to explore its role in ischemia/reperfusion injury models, in which blood flow is blocked and subsequently restored, thereby stimulating P-selectin expression by endothelial cells. In a rat model of hepatic ischemia/reperfusion injury (26), animals treated with 100 µg of recombinant PSGL-1 had significantly enhanced recovery of liver function and higher survival. PSGL-1 may also be important for lymphocyte recruitment to sites of inflammation in vivo, since intravenous administration of antibodies to the extreme N-terminus of mouse PSGL-1 blocks migration of Th1 T-lymphocytes into skin undergoing cutaneous delayed-type hypersensitivity reactions (27).

While all three selectins can bind to simple glycans containing the sialyl Le^x determinant, as demonstrated for P-selectin and PSGL-1, such binding is

relatively weak and macromolecular ligands bind with higher affinity. Although several glycoproteins are recognized by L- and E-selectin, whether these ligands serve physiologically to support selectin-mediated cell adhesion is still not clear. Interestingly, evidence is accumulating that indicates that PSGL-1 may be a physiological ligand for L-selectin and may participate in some E-selectin-dependent adhesion.

4. CLINICAL IMPLICATIONS

Adhesion molecules are vital for the physiological processes of leukocyte trafficking and are critically involved in the enhanced leukocyte emigration that is a key feature of all inflammatory and immune diseases. The studies on selectins and their ligands have yielded an insight to potential therapies for many diseases. Indeed, interference with selectin function is an attractive way to potentially block inflammation at a very early step. Although most intervention studies to date have been performed in animal models, human studies also have indicated a significant role for selectins and their ligands in the inflammatory response. In the following section we review select studies demonstrating a role for selectins in inflammatory injury in different organs.

4.1. Brain

Human studies have shown increased selectin levels in ventricular cerebrospinal fluid (CSF) from children with severe traumatic brain injury (Glasgow coma score < 8) (28) and in patients with relapsing-remitting multiple sclerosis (29). Middle cerebral artery occlusion in non-human primates is associated with upregulation of E-selectin (30). Selectins are also thought to contribute to tissue injury in stroke. In multiple murine models of stroke, the use of selectin ligands to block selectin function has reduced infarct size (31-33) (Table 2).

4.2. Heart

Leukocyte adhesion to damaged endothelium is enhanced in the presence of platelets by a mechanism involving platelet P-selectin. Thrombus formation may also be enhanced by this interaction. In human studies, P-selectin levels were shown to be significantly increased in plasma in patients with acute myocardial infarct (34). The use of a selectin blocker (CY-1503), an analogue of sialyl Lewis X selectin ligand, inhibited leukocyte and platelet interaction after arterial injury produced by angioplasty in pigs (35). Other studies have shown that the selectin blocker fucoidin provides cardioprotection in rats and dogs with coronary artery occlusion (36). These promising results may be the precursors of clinical trials of selectin blockers in humans (Table 3).

Specie	Selectin measured/targeted	Intervention	Injury/Model	Results	Ref
Human	P-selectin, E-selectin, L- selectin		Traumatic brain injury	P-selectin was increased	28
Human	L-selectin		Relapsing-remitting multiple sclerosis	Increase in soluble L- selectin	29
Non- human primate	E-selectin		Middle cerebral artery occlusion	E-selectin significantly upregulated	30
		sLex-glycosylated complement inhibitory protein	Stroke	Reduced cerebral infarct volumes.	31
Rats	E-selectin	Synthetic oligopeptide corresponding to lectin domain of selectin.	Transient cerebral ischemia	Decreases the size of ischemic injury.	32
Rats	P-selectin, E-selectin	tPA and anti- ICAM-1	Middle cerebral artery occlusion	Significant reduction in stroke volume	33

Table 2. Selectin studies in brain

4.3 Kidneys

Unlike in heart and muscle, L-selectin alone does not appear to mediate leukocyte recruitment to postischemic kidney (37). P-selectin, however, is upregulated in renal ischemia-reperfusion injury (38). Initial results with P-selectin antibody were protective in rats post renal ischemia (39). A soluble P-selectin glycoprotein ligand, which blocks P and Eselectins, has been shown to decrease renal injury secondary to ischemia-reperfusion in mice models (40). Blocking initial selectin-mediated events that accompany renal ischemiareperfusion has also been shown to reduce late renal dysfunction and tissue damage (41). Thus, selectin blockade might be a potential therapeutic intervention in the transplantation of kidneys from non-heart-beating donors and in kidneys subjected to prolonged ischemic times. Recently small molecule blockade of selectin ligands substantially reduced renal injury and improved mortality in rats. Interestingly, this occurred independent of neutrophils infiltration (42) (Table 4).

4.4. Lungs

In a sheep model of ischemia-reperfusion, antibodies directed against both L- and E-selectin significantly reduced pulmonary leakage and neutrophil accumulation (43). In a lung transplant model, anti-E- and anti-L-selectin antibodies improved post-transplant pulmonary function tests (44). Similarly, an analog of sialyl lewis X was found to reduce allograft rejection and reperfusion injury in a lung transplant model (45) (Table 5).

4.5. Liver

Studies of selectin involvement in liver ischemiareperfusion injury have focused primarily on P-selectin and its ligands. Rat studies have shown that P-selectin is the primary determinant of leukocyte adhesion to endothelial cells (46). Other studies have shown reduced liver injury and improved survival in rats treated with anti P-selectin antibodies during hepatic ischemia-reperfusion injury produced by uncontrolled hemorrhagic shock (47) (Table 6).

5. NOVEL PERSPECTIVES

There is now ample evidence that selectins are important mediators of ischemia-reperfusion injury in a number of different organs. Recent data also suggest that targeting selectin ligands may be a valuable therapeutic approach in the treatment of ischemia-reperfusion injury. The mechanisms underlying the protective effect of selectin antagonism in organ ischemia-reperfusion appear to be more complex than merely reducing neutrophil migration into reperfused tissue. Indeed, recent studies have suggested that blocking leukocyte adhesion molecules, including selectins, can protect against inflammatory injury independent of blocking tissue leukocyte infiltration. Interfering with the leukocyte adhesion molecule very late antigen-4 (VLA-4) on eosinophils and lymphocytes was found to attenuate late airway responses in an asthma model despite little effect on the migration of leukocytes into airway tissues (48).

It is possible that selectin antagonism may exert a protective effect during ischemia-reperfusion by interfering with cell signaling. Intriguingly, it has recently been reported that administration of glycyrrhizin, a licorice plant (*Glycyrrhiza radix*) derivative, decreased renal ischemia-reperfusion injury in a rabbit model (49). Moreover, the authors proposed that the protective effect of glycyrrhizin was due to selectin antagonism, which has been reported for glycyrrhizin (50). Glycyrrhizin inhibits 11 β -hydroxysteroid dehydrogenase, the enzyme that converts cortisol to cortisone. Thus, could the protective role of selectin blockade in organ ischemia-reperfusion injury be somehow related to modulation of glucocorticoid activity?

It is clear that further investigation is warranted to elucidate the mechanisms by selectins mediate ischemiareperfusion injury, beyond simply promoting leukocyte recruitment. In addition, given the ample evidence that now exists regarding efficacy of selectin antagonism in treating

Specie	Selectin measured/targeted	Intervention	Injury/Model	Results	Ref
Human	P-selectin		Coronary recanalization therapy after AMI.	P-selectin significantly higher in AMI than in stable angina pectoris.	34
Pigs	P-selectin	CY-1503	Arterial injury by angioplasty	CY-1503 reduced neutrophil adhesion but not platelets.	35
Rats		Fucoidin	No-flow ischemia to rats hearts.	Fucoidin significantly reduced leukocyte accumulation in capillaries and venules	36

Table 3. Selectin studies in Heart

Specie	Selectin measured/targeted	Intervention	Injury/Model	Results	Ref
Rats		P-selectin antibody	Renal artery cross-clamping for 60 min.	Role for P-selectin in renal ischemic injury.	39
Mice		P-selectin- deficient mice.	Renal artery cross-clamping.	Mice protected from I/R injury.	40
Rats		Soluble ligand for P- and E- selectin.	Clamping of renal pedicle 45 min.	Function and structure remained at relative baseline.	41

Specie	Selectin	Intervention	Injury/Model	Results	Ref
-	measured/targeted				
Sheep	L-selectin	Anti-L-selectin (EL-246)	Infrarenal aortic ischemia (3h) followed by reperfusion.	Significant reduction in pulmonary leakage by 59% and neutrophil accumulation by 84%	43
Sheep	L- and E-selectins	L- and E- selectins antibody	Left lung autotransplant	Improved pulmonary function tests	44
Rats	P-selectin	Analog of Sialyl-Lewis X (SLX).	Lung transplant.	Reduction in allograft rejection and reperfusion injury.	45
fable 6.	Selectin studies in Liver				
Speci e	Selectin measured/targeted	Intervention	Injury/Model	Results	Ref
Mice	P-selectin		Left hepatic lobe ischemia for 30 min	P-selectin is the primary determinant of leukocyte- endothelial cell adhesion	46
Rats		P-selectin antibody	Uncontrolled hemorrhagic shock	Decreased hepatocellular injury and increased survival	47
Rats		PSGL-1	Cold ischemia	Significant increase in liver isograft survival	26

organ ischemia-reperfusion injury, clinical trials are justified to translate these basic research findings to clinical utility. However, there are crucial questions to be answered before any human clinical trials, including: (1) What are the effects of selectin interventions on the immune system?, and (2) Is it possible to selectively block just the detrimental effects of leukocyte adhesion molecules without leaving an individual immunocompromised?

7. REFERENCES

1. Star RA.: Treatment of acute renal failure. *Kidney Int* 54, 1817-1831 (1998)

2. Bonventre JV: Mechanisms of ischemic acute renal failure. *Kidney Int* 43, 1160-1178 (1993)

3. Gillium BS, E. J. Graves & E. Wood: National Hospital Discharge Survey: Annual Summary. In: US Department of Health and Human Services, CDC, National Center for Health Statistics. MDHHS publication no. (PHS) 96-1232 (1996)

4. Albelda SM, C. W. Smith & P. A. Ward: Adhesion molecules and inflammatory injury. *FASEB J* 8, 504-512 (1994)

5. Del Zoppo GJ, G. W. Schmid-Schonbein, E. Mori, B. R. Copeland & C. M. Chang: Polymorphonuclear leukocytes occlude capillaries following middle cerebral artery

occlusion and reperfusion in baboons. *Stroke* 22, 1276-1283 (1991)

6. Murota S, H. Fujita, Y. Wakabayashi & I. Morita: Cell adhesion molecule mediates endothelial cell injury caused by activated neutrophils. *Keio Journal of Medicine* 45, 207-212 (1996)

7. Weiss SJ: Tissue destruction by neutrophils. *New Engl J Med* 320, 365-376 (1989)

8. Bjork J, P. Hedqvist & K. E. Arfors: Increase in vascular permeability induced by leukotriene B4 and the role of polymorphonuclear leukocytes. *Inflammation* 6, 189-200 (1982)

9. Abass AK, A. H. Lichtman & J. S. Pober : Cytokines. In: Cellular and molecular immunology. Eds: Abass AK, A. H.

Lichtman & J. S. Pober, WB Saunders, PA 249-277 (1997) 10. Springer TA: Adhesion receptors of the immune system. *Cell* 69, 11-25 (1992)

11. Lasky LA: Selectins: interpreters of cell-specific carbohydrate information during inflammation. *Science* 258, 964-969 (1992)

12. Ley K, D. C. Bullard, M. L. Arbones, R. Bosse, D. Vestweber, T. F. Tedder & A. L. Beaudet: Sequential contribution of L- and P-selectin to leukocyte rolling in vivo. *J Exp Med* 181, 669-675 (1995)

13. Sharar SR, S. S. Sasaki, L. C. Flaherty, J. C. Paulson, J. M. Harlan & R. K. Winn: P-selectin blockade does not impair leukocyte host defense against bacterial peritonitis and soft tissue infection in rabbits. *J Immunol* 151, 4982-4988 (1993)

14. Griffin JD, O. Spertini, T. J. Ernst, M. P. Belvin, H. B. Levine, Y. Kanakura & T. F. Tedder: Granulocytemacrophage colony-stimulation factor and other cytokines regulate surface expression of the leukocyte adhesion molecule-1 on human neutrophils, monocytes, and their precursors. *J Immunol* 145, 576-584 (1990)

15. Bevilacqua MP & R. M. Nelson: Selectins. J Clin Invest 91, 379-387 (1993)

16. Lasky LA: Selectin carbohydrate interactions and the initiation of the inflammatory response. *Ann Rev Biochem* 64, 113-139 (1995)

17. Tedder TF, D. A. Steeber, A. Chen & P. Engel: The selectins: vascular adhesion molecules. *FASEB J* 9, 866-873 (1995)

18. Kubes P, M. Jutila & D. Payne: Therapeutic potential of inhibiting leukocyte rolling in ishemia/reperfusion. *J Clin Invest* 95, 2510-2519 (1995)

19. Bevilacqua MP, S. Stengelin, M. A. Gimbrone Jr. & B. Seed: Endothelial leukocyte adhesion molecules: an inducible receptor for neutrophils related to complement regulatory proteins and lectins. *Science* 243, 1160-1165 (1989)

20. Anrather J, V. Csizmadia, C. Brostjan, M. P. Soares, F. H. Bach & H. Winkler: Inhibition of bovine endothelial cell activation in vitro by regulated expression of a transdominant inhibitor of NF-kappa B. *J Clin Invest* 99, 763-772 (1997)

21. Moore KL, N. L. Stults, S. Diaz, D. F. Smith, R. D. Cummings, A. Varki & R. P. McEver: Identification of a specific glycoprotein ligand for P-selectin (CD62) on myeloid cells. *J Cell Biol* 118, 445-456 (1992)

22. Norgard KE, K. L. Moore, S. Diaz, N. L. Stults, S. Ushiyama, R. P. McEver, R. D. Cummings & A. Varki:

Characterization of a specific ligand for P-selection on myeloid cells. A minor glycoprotein with sialylated O-linked oligosaccharides. *J Biol Chem* 268, 12764-12774 (1993)

23. Lorant DE, R. P. McEver, T. M. McIntyre, K. L. Moore, S. M. Prescott & G. A. Zimmerman: Activation of polymorphonuclear leukocytes reduces their adhesion to P-selectin and causes redistribution of ligands for P-selectin on their surfaces. *J Clin Invest* 96, 171-182 (1995)

24. Hidari KI, A. S. Weyrich, G. A. Zimmerman & R. P. McEver: Engagement of P-selectin glycoprotein ligand-1 enhances tyrosine phosphorylation and activates mitogenactivated protein kinases in human neutrophils. *J Biol Chem* 272, 28750-28756 (1997)

25. Maly P, A. Thall, B. Petryniak, C. E. Rogers, P. L. Smith, R. M. Marks, R. J. Kelly, K. M. Gersten, G. Cheng, T. L. Saunders, S. A. Camper, R. T. Camphausen, F. X. Sullivan, Y. Isogai, O. Hindsgaul, U. H. von Andrian & J. B. Lowe: The $\alpha(1,3)$ fucosyltransferase Fuc-TVII controls leukocyte trafficking through an essential role in L-, E-, and P-selectin ligand biosynthesis. *Cell* 86, 643-653 (1996) 26. Dulkanchainun TS, J. A. Goss, D. K. Imagawa, G. D. Shaw, D. M. Anselmo, F. Kaldas, T. Wang, D. Zhao, A. A. Busuttil, H. Kato, N. G. Murray, J. W. Kupiec-Weglinski & R. W. Busuttil: Reduction of hepatic ischemia/reperfusion injury by a soluble P-selectin glycoprotein ligand-1. *Ann Surg* 227, 832-840 (1998)

27. Borges E, W. Tietz, M. Steegmaier, T. Moll, R. Hallmann, A. Hamann & D. Vestweber: P-selectin glycoprotein ligand-1 (PSGL-1) on T helper 1 but not on T helper 2 cells binds to P-selectin and supports migration into inflamed skin. *J Exp Med* 185, 573-578 (1997)

28. Whalen MJ, T. M. Carlos, P. M. Kochanek, S. R. Wisniewski, M. J. Bell, J. A. Carcillo, R. S. Clark, S. T. Ke Kosky & P. D. Adelson: Soluble adhesion molecules in CSF are increased in children with severe head injury. *J Neurotrauma* 15, 777-787 (1998)

29. Duran I, E. M. Martinez-Caceres, J. Rio, N. Barbera, M. E. Marzo & X. Montalban: Immunological profile of patients with primary progressive multiple sclerosis. Expression of adhesion molecules. *Brain* 122, 2297-2307 (1999)

30. Haring EP, E. L. Berg, N. Tsurushita, M. Tagaya & G. J. Del Zoppo: E-selectin appears in nonischemic tissue during experimental focal cerebral ischemia. *Stroke* 27, 1386-1391 (1996)

31. Huang J, L. J. Kim, R. Mealey, H. C. Marsh Jr., Y. Zhang, A. J. Tenner, E. S. Connolly Jr. & D. J. Pinsky: Neuronal protection in stroke by an sLex-glycosylated complement inhibitory protein. *Science* 285, 595-599 (1999)

32. Morikawa E, S. M. Zhang, Y. Seko, T. Toyoda & T. Kirino: Treatment of focal cerebral ischemia with synthetic oligopeptide corresponding to lectin domain of selectin. *Stroke* 27, 951-955 (1996)

33. Zhang RL, Z. G. Zhang, M. Chopp & J. A. Zivin: Thrombolysis with tissue plasminogen activator alters adhesion molecule expression in the ischemic rat brain. *Stroke* 30, 624-629 (1999)

34. Tomoda H & N. Aoki: Plasma soluble P-selectin in acute myocardial infarction: effects of coronary recanalization therapy. *Angiology* 49, 807-813 (1998)

35. Merhi Y, P. Provost, P. Chauvet, J. F. Theoret, M. L. Phillips & J. G. Latour: Selectin blockade reduces neutrophil interaction with platelets at the site of deep arterial injury by angioplasty in pigs. *Arteriol Thromb Vasc Biol* 19, 372-373 (1999)

36. Ritter LS, J. G. Copeland & P. F. McDonagh: Fucoidin reduces coronary microvascular leukocyte accumulation early in reperfusion. *Ann Throc Surg* 66, 2063-2071 (1998) 37. Rabb H; Ramirez G; Saba SR; Reynolds D; Xu J;

Flavell R; Antonia S: Renal ischemic-reperfusion injury in L-selectin-deficient mice. *Am J Physiol* 271, 408-13 (1996)

38. Zizzi HC, G. B. Zibari, D. N. Granger, I. Singh, L. D. Cruz, F. Abreo, J. C. McDonald & M. F. Brown: Quantification of P-selectin expression after renal ischemia and reperfusion: *J Pediatr Surg* 32, 1010-1013 (1997)

39. Rabb H, C. Mendiola, S. R. Saba, J. Dietz, C. W. Smith, J. V. Bonventre & G. Ramirez: Antibodies to P-selectin and ICAM-1 protect kidneys from ischemic-reperfusion injury. *JASN* 5(3), 907 (1994)

40. Singbartl K; Green SA; Ley K: Blocking P-selectin protects from ischemia/reperfusion-induced acute renal

failure. FASEB J 14(1),48-54 (2000)

41. Takada M, K. C. Nadeau, G. D. Shaw & N. L. Tilney: Prevention of late renal changes after initial ischemia/reperfusion injury by blocking early selectin binding. *Transplantation* 64, 1520-1525 (1997)

42. Nemoto T, A. C. Issekutz, K. Berens, M. O'Donnell, B. Kasiske, W. Keane, H. Rabb: Small molecule inhibition of selectin ligands reduces mortality and improves renal recovery from ischemic reperfusion injury in rats. *JASN* 10, 637A (1999)

43. Seekamp A, G. Regel, K. Rother & M. Jutila: The effect of anti-L-selectin (EL-246) on remote lung injury after infrarenal ischemia/reperfusion. *Shock* 7, 447-454 (1997)

44. Brandt M, K. Boeke, M. L. Phillips, G. Steinhoff & A. Haverich: Effect of oligosaccharides on rejection and reperfusion injury after lung transplantation. *J Heart Lung Transplant* 16, 352-359 (1997)

45. Demertzis S, F. Langer, T. Graeter, A. Dwenger, T. Georg & H. J. Schafers: Amelioration of lung reperfusion injury by L- and E-selectin blockade. *Eur J Cardiothorac Surg* 16, 174-180 (1999)

46. Sawaya Jr D E, G. B. Zibari, A. Minardi, B. Bilton, D. Burney, D. N. Granger, J. C. McDonald & M. Brown: P-selectin contributes to the initial recruitment of rolling and adherent leukocytes in hepatic venules after ischemia/reperfusion. *Shock* 12, 227-232 (1999)

47. Rivera-Chavez F, L. H. Toledo-Pereyra, D. T. Nora, B. Bachulis, F. Ilgenfritz & R. E. Dean: P-selectin blockade is beneficial after uncontrolled hemorrhagic shock. *J Trauma* 45, 440-445 (1998)

48. Rabb HA; Olivenstein R; Issekutz TB; Renzi PM; Martin JG: The role of the leukocyte adhesion molecules VLA- 4, LFA-1, and Mac-1 in allergic airway responses in the rat. *Am J Respir Crit Care Med* 149(5), 1186-91 (1994) 49. Subramanian S; Bowyer MW; Egan JC; Knolmayer TJ: Attenuation of renal ischemia-reperfusion injury with selectin inhibition in a rabbit model. *Am J Surg* 178(6), 573-6 (1999)

50. Kim MK;. Brandley BK; Anderson MB; Bochner BS: Antagonism of selectin-dependent adhesion of human

eosinophils and neutrophils by glycomimetics and oligosaccharide compounds. *Am J Respir Cell Mol Biol.* 19(5), 836-41 (1998)

Key Words: Selectin, Selectin ligands, Leukocyte Adhesion molecules, Ischemia-reperfusion, Review

Send correspondence to: H. Rabb, M.D., Nephrology, Division, Hennepin County Medical Center, University of Minnesota, 701 Park Ave, Minneapolis, MN 55415, Tel:612-347-5871, Fax:612-347-2003, Email: rabbx003@tc.umn.edu

This manuscript is available on line at:

http://www.bioscience.org/2000/v5/e/chamoun/fulltext.htm