NATURALLY-OCCURRING ANTISPERM ANTIBODIES IN MEN: INTERFERENCE WITH FERTILITY AND IMPLICATIONS FOR TREATMENT

Felice Francavilla¹, Rossella Romano, Riccardo Santucci, Grazia La Verghetta, Piera D'Abrizio and Sandro Francavilla

Department of Internal Medicine, Andrologic Unit, University of L'Aquila, Via S.Sisto 22/E, 67100 L'Aquila, Italy

TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction: controversies on the significance of antisperm antibodies
 - 2.1. Antisperm antibody testing
 - 2.2. Prevalence of antisperm antibodies
 - 2.3. Prognostic studies
- 3. Mechanisms of fertility impairment
 - 3.1. Effect on semen quality
 - 3.2. Interference with cervical mucus penetration
 - 3.3. Complement-mediated cytotoxicity and opsonizing effect through the female genital tract
 - 3.4. Interference with the fertilization process
 - 3.4.1. In vitro fertilization (IVF) as model of study
 - 3.4.2. The hamster egg penetration (HEPT) as model of study
 - 3.4.3. Effects on capacitation and acrosome reaction
 - 3.4.4. Effects on the zona pellucida interaction
 - 3.5. Postfertilization effects
- 4. Implications for treatment
 - 4.1. Corticosteroid therapy
 - 4.2. Intrauterine insemination
 - 4.3. IVF-ET vs ICSI
- 5. Perspective
- 6. References

1. ABSTRACT

Naturally-occurring antisperm antibodies in men are a relative cause of infertility, being the fertility impairment related with the degree of autoimmunization. The impairment of sperm penetration through the cervical mucus represents the best established mechanism of the antibody interference with fertility. Another mechanism may involve complement-mediated sperm injury and opsonizing effect through the female genital tract. Finally, sperm-bound antibodies can interfere with sperm functions involved in the fertilization process, mainly in the sperm-zona pellucida interaction. While some mechanisms of the antibody-interference with fertility depend only on the degree of sperm autoimmunization (e.g., inhibition of cervical mucus penetration), other mechanisms (e.g., interference with gametes interaction) could or could not occur depending on the relevance in the fertilization process of the specific antigen(s) recognized by antisperm antibodies, which are policional in nature. Intrauterine insemination is an effective treatment when sperm autoimmunization is low or moderate, mainly if combined with corticosteroid treatment and superovulated cycles. On the contrary, its effectiveness in cases of high degree of sperm autoimmunization is controversial. The resort to "high tech" procedures is mandatory when other less invasive approaches have failed or they may also be chosen as a first-choice method in cases of high degree of sperm autoimmunization. Since in most reports the fertilization rate with in vitro fertilization and embryo transfer (IVF-ET) was significantly lower in the presence of sperm-bound antibodies than in the case of other indications, the likelihood of fertilization is higher with intracytoplasmatic sperm injection (ICSI), where the reported fertilization rates are similar to those in other indications, or even higher.

2. INTRODUCTION: CONTROVERSIES ON THE SIGNIFICANCE OF ANTISPERM ANTIBODIES

The role of naturally occurring antispermantibodies (ASA) as a cause of male infertility was recognized since Rumke (1) and Wilson (2) reported the presence of serum sperm-agglutinating antibodies in infertile men in 1954. Following those early observations, a large amount of reports has been directed to the investigation of the significance of ASA in male infertility including the aetiology of ASA formation, sites and mechanisms of antibody action, and possible treatments. However, since published studies used varying approaches for the recognition and treatment of male immunologic infertility, such as different antibodies assays, different drug regimes and lack of well-designed and controlled studies, it resulted in confusion of the prevalence as well as the actual role of ASA in male infertility. With the development and the encouraged use of accurate and easy assays for the screening of ejaculates for sperm-bound antibodies, the development of sperm function assays, and

Table 1. Diagnostic strategy for the detection of ASA in the men

the men			
Test*	Application		
IgG-MAR test ⁹ or	For routine screening of all		
SpermMAR ^{11,21}	semen analyses		
Direct immunobead	To be performed in case of		
binding test (IBT) ¹⁰	positive igg-MAR test, to		
	determine whether iga-ASA		
	are also bound to sperm		
	surface		
Sperm-agglutination test,	For the titration of ASA in		
expecially TAT ⁶	serum and seminal plasma		
	in patients with a positive		
	direct test, or as screening		
	test when it is impossible to		
	performe direct tests (e.g.,		
	Lack of motility)		
Indirect antiglobulin	As an alternative option to		
tests ^{10,23,27}	the sperm-agglutination		
	test, with the same purpose		

^{*:} reference

finally the progress in assisted reproductive medicine, the role of ASA in male infertility as well as the potential for their treatment is becoming better defined. In this article we will review current understanding of the interference of ASA with the male fertility and possible treatments, following a brief focusing on clinical aspects of the male immunologic infertility, including ASA assays, prevalence and prognostic evaluations.

2.1. Antisperm-antibody testing

Only ASA assays detecting antibodies directed towards surface antigens have a clinical significance, because subsuperficial antigens cannot be exposed to antibodies by living cells along the male genital tract (3). The first assays to be utilized were indirect tests detecting biological activities of circulating ASA, i. e., sperm agglutination techniques and complement dependent cytotoxicity techniques (see ref. 3 for review). Multicentric comparative studies (4,5) indicated that they determine largely the same antibody specificities but with different sensitivity which was higher for sperm agglutination techniques, especially for the Tray agglutination test-TAT (6).

However, circulating ASA may differ from sperm associated antibodies "in vivo" in their biological activity and affinity to sperm antigens, since locally produced secretory immunoglobulins occur in the genital tract in addition to serum-derived Ig (7,8). Therefore, during the last 2 decades widespread acceptance has been gained by direct tests developed for the detection of sperm-bound antibodies, including the mixed antiglobulin reaction (MAR) test (9), the immunobead test (IBT)(10) and the SpermMar test (11). Although MAR test detects only antibodies belonging to IgG class while IBT detects IgG,A,M antibodies, comparative studies have generally demonstrated that all these tests are suitable as effective screening direct tests (9, 11-18). In fact, sperm surface IgA are almost always found in association with IgG (12,15,19). Since MAR test and the commercially available SpermMAR are cheaper and quicker, they are generally considered as more suitable for routine screening of all semen analyses, with the IBT performed on samples with a positive former test (13,20), and indirect testing (especially TAT or indirect IBT) performed only in men with azoospermia or lack of sperm motility (table 1). Since IgA SperMar is now available and an its higher accuracy than IgA-IBT has been reported (21), this diagnostic strategy could be furter semplified; moreover, a rapid mixed immunobead screen has been recently proposed (22), but its accuracy must be confirmed.

Other antiglobulin-based tests, such as radiolabeled antiglobulin assay (23,24), enzyme-linked immunosorbent assay (ELISA) (25-28) immunofluorescent test on living sperm suspensions (29,19), flow cytometry assay (30-35) has been proposed both as indirect and direct tests. Radiolabeled antiglobulin assay, although highly sensitive and specific, does not detect the regional specificity of antibody link. ELISA suffers from the same limitation, and lacks specificity for surface antigens because internal antigens are exposed by fixation; to overcome this limitation, a new type of ELISA without fixation has been proposed (36). Using living sperm suspensions instead of fixed smears makes the immunofluorescent test highly specific for surface antigens-directed ASA; it is usually utilized in the direct form (19) in our lab for a better evaluation of the regional specificity of the antibody link in all samples positive at direct IgG-MAR test. Flow cytometry assay is very promising for its potential to quantify the antibody load on each sperm cell (31).

2.2. Prevalence of antisperm antibodies

The reported prevalence of ASA varied depending the modality of the immunological screening. Circulating ASA detected with indirect tests ranged from 8.1% to 30.3% in unselected men with infertile marriages (14,36-40). At low titres they were also reported in 2.4% to 10% of fertile men (36,40). Noteworthy, low titres of sperm-agglutinating activity can be due to nonimmunological factors (41), representing false positive results. When stricter criteria were used (i.e., the occurrence of sperm-immobilizing activity in addition to sperm-agglutinating activity (14,37) and/or occurrence of sperm-agglutinating activity in seminal plasma (14), the prevalence of ASA in men with infertile marriages was 4.7% to 7.5%. Immunological screening by means of direct tests gave positive results in 7.8% to 20.1% (14-16,42-47), with the occurrence of strong positive results in about 6-7% of patients (14,16).

Some clinical conditions associated with a high prevalence of ASA have been recognized as identifiable causes of their development, mainly acquired genital tract obstructions. Among them, vasectomy is the most common, with a prevalence of ASA of 34% to 74% (40,44,48-50), and their persistence in 38% to 60% following successful vasovasostomy (49,51,52). On the contrary, it is not yet well established the association of ASA with obstructive azoospermia due to congenital causes, since conflicting data have been reported (53-57). Antisperm antibodies have also been associated with acute and chronic genitourinary infections (58-62). Some studies

have focused on the relationship between ASA and asymptomatic Chlamydia trachomatis infections. Although a high prevalence of ASA was reported in the presence of Chlamydia in genital secretion as detected by means of culture (63) or the polymerase chain reaction (64), circulating chlamydial antibodies were not associated (65) or only weakly associated (66) with the presence of sperm bound antibodies, whereas both a strong correlation (66) as well as no association (67) were reported between the presence of seminal chlamydial antibodies and ASA. A high prevalence of ASA has been reported in men with testicular carcinoma (68-70), in adults with a previous cryptorchidism and a late ochidopexy (71), in homosexual men (72), and in human immunodeficiency virus (HIV) positive men (73,74). Finally, conflicting results have been reported on the association of ASA with varicocele (75-79) and spinal cord injury (80-84). In a recent multivariate analysis of men from infertile couples with and without ASA, only prior vas reversal and a history of genital tract infection were associated with the presence of sperm-bound antibodies (85).

2.3. Prognostic studies

Although the higher frequency of ASA in males with infertile marriages than in fertile controls could imply a negative effect of ASA on fertility, a cause-effect relationship needs to be validated by prospective studies of the relationship between the presence of ASA and subsequent fertility. Follow-up studies which compared negative and positive subjects for circulating ASA have produced conflicting results, since a significant association between antibody presence in the male and lower pregnancy rates was found in some studies (37,86,87) but not in others (39,88,89). A significant association was found when sperm bound antibodies were detected with direct tests (89,90). Furthermore, when the degree of the sperm autoimmunization was considered, a significant inverse correlation was found between eiher the titre of circulating ASA (37,91) or the percentage of sperm bound antibodies (92) and the incidence of pregnancies. A poor prognostic value of low to moderate levels of sperm-bound antibodies was also reported by Barrat et al. (93).

Altogether, the analysis of epidemiological and prognostic studies confirms the opinion of Bronson (2) that ASA are a *relative*, rather than absolute, cause of infertility. Fertility impairment is related with the degree of sperm autoimmunization, that is, the extent to which ASA are present in reproductive tract secretions and detected on sperm surface.

3. MECHANISMS OF FERTILITY IMPAIRMENT

Although the fertility impairment due to ASA is related with the degree of sperm autoimmunization, as suggested by epidemiological and prognostic studies, there are several evidences that also *qualitative* differences in the effects of ASA could have a role in the fertility impairment. In fact, firstly, different immunoglobulin isotypes of antibodies occurring on sperm surface can produce different biological effects (i.e., cytotoxic effects due to complement activation can be produced by IgG but not IgA

sperm bound antibodies). Secondly, due to their polyclonal nature, ASA are directed against more than one sperm antigen, which may differ among patients and may be more o less relevant to fertility.

3.1. Effect on semen quality

With some exceptions (39,45), most epidemiologic studies did not find any significant difference in the principal semen parameters (sperm count, motility and morphology) between infertile patients with and without ASA (14,16,94,95). In any case, there is little evidence that suggests a cause/effect relationship between ASA and abnormality of the principal semen parameters. An antibody effect on semen quality should involve a complement mediated sperm cytotoxicity occurring within the male genital tract. However, anticomplementary activity has been reported in human semen (96,97), and it was recovered in the low molecular weight of the seminal plasma (20-60.000 Daltons) using gel filtration chromatography (96). This fraction inhibited total complement activity as well as the activity of the early C components C1 and C3. Afterwards, a potent inhibitor of C5b-7 complexes was identified in human seminal plasma, where it was found in 5- to 10 -fold higher concentrations than in serum. A sulphated glycoprotein termed clusterin was also found on ram sperm (98), and purified human seminal clusterin was shown to inhibit C5b-6 mediated hemolysis (99). Finally, D'Cruz and Haas (100) demonstrated the lack of a detectable product of C activation (SC5b-9) in the seminal plasma of men with sperm bound antibodies (IgG were present in most cases). Taken together, these findings suggest that human seminal plasma contains inhibitors for both the initial and the terminal portions of the C cascade thereby protecting sperm from Cmediated injury in the male reproductive tract.

Since an increased sperm count in some oligozoospermic patients with ASA was reported in response to corticosteroid therapy, it was suggested that in those cases a cell-mediated immune reaction at the level of rete testis and/or epididymis responsive to the anti-inflammatory effect of corticosteroids might underlie the low sperm count (101). However, clinical and pathologic evidences of immune orchitis in men exhibiting natural autoimmunity to sperm has never been provided.

Sperm agglutination is the only semen alteration related to the presence of ASA. A significant increase in the proportion of motile sperm involved in agglutinations has been reported in the presence of ASA, whenever investigated (14,16,95,102). However, sperm agglutination, which is a time-dependent phenomenon, only rarely involves a large proportion of motile sperm soon after liquefaction, even when all ejaculated sperm are antibody-coated. Therefore, sperm agglutination, although extremely suggestive of sperm autoimmunization, does not represent an important mechanism of antibody-interference with fertility in most cases.

3.2 Interference with cervical mucus penetration

The impairment of sperm penetration through the cervical mucus represents the best known and established

mechanism of the antibody interference with fertility (2). Definitive clinical demonstrations of this impairment have been produced analysing the outcome of "in vivo" as well as "in vitro" tests of sperm cervical mucus interaction.

As far as the post coital testing is concerned, a significant association was reported between a poor sperm penetration into cervical mucus and sperm autoimmunization (103,104). Besides, the degree of impairment of sperm penetration into cervical mucus was found to correlate with the proportion of sperm exhibiting surface-bound antibodies (105), as well as with the titre of circulating sperm-immobilizing antibodies (104).

The outcome of the *in vitro* cervical mucus penetration test comparing men with and without ASA has largely confirmed this impairment (38,39,43). Finally, the demonstration of the actual responsibility of ASA in impairing cervical mucus penetration has been provided by matching donor sperm suspensions exposed to sera containing ASA against the same sperm suspensions exposed to control sera without ASA (106).

Although some reports suggested that IgA were more important than IgG in impairing sperm penetration of cervical mucus (107-109), other findings indicate that an abnormal interaction between the Fc portion of both IgA and IgG immunoglobulins bound to the sperm surface and constituents of the cervical mucus is responsible, almost in part, for the characteristic shaking phenomenon and the impairment of mucus penetration. Comparing the swimming ability of antibody-coated sperm within cervical mucus with that of sperm exposed to only the Fab fragments of the same antibodies, mucus penetration was abolished by complete antibody, whereas only reduced but not abolished by Fab (110). Bronson et al. (111) found an improvement of the ability of antibody-bound sperm to penetrate human cervical mucus in vitro, after exposure to a IgA₁ protease, which was expected to liberate Fc fragments of IgA₁ antibodies bound to the sperm surface. Moreover, this improvement varied inversely with the amount of remaining IgGs, not degraded by protease, indicating a role for both IgG and IgA sperm-bound antibodies in impairing cervical mucus penetration.

3.3 Complement-mediated cytotoxicity and opsonizing effect through the female genital tract

One mechanism of ASA-interference with fertility may involve sperm injury potentially mediated by complement and/or phagocytic cells in the female genital tract. While complement-mediated cytotoxic effect by complement-fixing ASA are prevented in semen, due to its anticomplementary activity (see 3.1), when antibody-bound sperm enter the female reproductive tract they might became liable to deleterious effects of complement activation, supposing that complement components are present in sufficient amount through the female genital tract. Full-complement component lytic activity has been documented in cervical mucus in amount enough to cause complement-dependent sperm immobilization (112). Also human follicular fluid exhibits complement activity, and IgG-antibodies bound to sperm were capable of activating

follicular fluid complement as detected by their ability to deposit terminal complement complexes (MC5b-9) on human sperm (113). In an elegant study, D'Cruz et al. (114) provided the direct evidence for the involvement of complement-fixing ASA and complement activation in exerting sperm injury. Using flow cytometry to evaluate simultaneously the binding of antibody and autologous complement to sperm cells, they demonstrated that incubation of donor sperm with sera containing IgG-ASA resulted in the activation of autologous complement in vitro as assessed by the deposition of the initial (C3d) and the terminal (C5b-9) complement complex on the sperm surface. Antisperm antibodies and complement deposition resulted in a dramatic loss of sperm motility, as well as in activation and aggregation (rosetting) polymorphonuclear leukocytes (PMN) to antibody- and complement-bound sperm. The inability of sera containing non-complement fixing IgG-ASA to promote sperm binding to PMN suggested that IgG alone is insufficient to initiate the interaction, that is, it would preclude a direct interaction between the Fc portion of sperm-bound Ig and the Fc-receptor on PMN.

An opsonizing effect exerted by IgG-ASA had been previously reported also by London *et al.*(115), who demonstrated that the incubation of donor sperm with sera containing IgG-ASA enhanced sperm phagocytosis and lysis by peritoneal macrophages. This effect had been hypothesised as mediated by Fc-receptor for IgG.

3.4. Interference with the fertilization process

A vast body of literature has focused on the possible interference of ASA with the fertilization process. Several experimental studies have demonstrated that antibodies raised against whole spermatozoa (116,117) or defined sperm antigens (118 for review) can interfere with sperm functions involved in the fertilization process, thereby blocking sperm-egg interactions. However, the actual role of naturally-occurring ASA in men in impairing sperm-egg interaction, as well as the level of this impairment, is not yet sufficiently known, because conflicting data have been produced. In this section we will attempt to analyse the reasons underlying these conflicting data, which could help the understanding of this debated matter.

Retrospective or prospective analyses of fertilization data from in vitro fertilization and embryo transfer (IVF-ET) programs provide a potential means of assessing possible effects of ASA on human gametes interaction. However, this model of study cannot give information about the level of this possible interference. Several studies in the last two decades have tried to determine the level as well as the actual occurrence of this interference. The hamster egg penetration test has mainly been employed to investigate the effect of ASA on sperm functions involved in the fusion with the oolemma, that is, the capability to complete capacitation and to exhibit the acrosome reaction as well as the fusogenic properties involved in the interaction with the oocyte membrane. Some studies have also been carried out to directly test the effect of ASA on each specific sperm function (i.e., capacitation or acrosome reaction). Other studies have

Table 2. Fertilization rate in IVF-ET programs in the presence of antisperm antibodies (ASA) in the male.

	ASA +		ASA -
Reference	Fertilized/total ova (Fertilization Rate)		Fertilized/total ova (Fertilization Rate)
Clarke <i>et al.</i> (1985) ¹²⁸	65/131 (50%)	IBT >20%	
	18/66 (27%)	IgA >80%	
	47/65 (72%)	IgA <80%	
Mandelbaum <i>et al.</i> (1987) ¹²⁵	8/14 (57%)	IBT >20%	118/180 (65%)
Matson et al.(1988) ¹¹⁹	37/70 (53%)	IBT >20%	156/201 (78%)
	15/39 (38%)	IgG+IgA	,
	22/31(71%)	IgG or IgA	
De Almeida <i>et al.</i> (1989) ¹³⁰	70/175 (40%)	IBT >10%	
	6/43 (14%)	>70%(IgG+IgA)	
	31/52 (60%)	<70%	
Palermo et al.(1989) ¹³²	132/273 (48%)	MAR test >10%	
,	33/80 (41%)	>90%	
	99/193 (51%)	<90%	
Chang et al.(1993) ¹²⁰	17/59 (30%)	IBT >10%	654/984 (66%)
Lahteenmaki (1993) ¹³¹	98/355 (28%)	MAR test >10%	,
, ,	28/170 (17%)	>90%	
	40/113 (35%)	>40% and <90%	
	30/72 (42%)	<40%	
Rajah <i>et al.</i> (1993) ¹²¹	53/105 (50%)	MAR test >20%	93/128 (73%)
Acosta <i>et al.</i> (1994) ¹²²	(42%)	MAR test >10%	(73%)
Sukcharoen & Keith (1995) ¹²⁶	124/165 (75%)	IBT >20%	978/1412 (69%)
Ford <i>et al.</i> (1996) ¹²³	209/544 (38%)	IBT >20%	380/558 (68%)
Vazquez-Levin et al. $(1997)^{124}$	46/104 (44%)	MAR test >20%	65/77 (84%)
Ombelet <i>et al.</i> $(1997)^{188}$	153/283 (54%)	MAR test >50%	` '
Culligan <i>et al.</i> (1998) ¹²⁷	(66%)	IBT >15%	(63%)
Total	1012/2278 (44%)		2444/3540 (69%)

focused on the interference of ASA with the sperm-zona pellucida (ZP) interaction.

Some considerations could be helpful in analyzing the results, often conflictual, which have been reported. The effects of natural ASA on fertilizing ability of human sperm have been studied either by matching donor sperm

suspensions exposed to sera from patients with circulating ASA against the same sperm suspensions exposed to control sera, or using spermatozoa coated "in vivo" with ASA. In both cases the results must be interpreted with caution. In fact, circulating ASA may differ from sperm associated antibodies "in vivo" in their biological activity and affinity to sperm antigens, since locally produced secretory immunoglobulins occur in the genital tract in addition to serum-derived Ig. On the other hand, using spermatozoa coated "in vivo" with ASA, the concomitant presence of non-immunological sperm abnormalities raises doubts about the responsibility of ASA in affecting sperm functions. Using antibodies eluted from autoimmune ejaculates instead of circulating ASA is another and potentially more demonstrative approach. Although used in some studies, its feasibility is hindered by the difficulty in eluting sufficient amounts of antibodies.

3.4.1. In vitro fertilization (IVF) as model of study

Table 2 shows the fertilization rates reported in series including couples with sperm autoimmunization in the male partner. In most reports the fertilization rate was

significantly lower in the presence of sperm-bound antibodies than in the case of other indications for IVF (119-124). However, in some other reports no significant difference was found (125-127). Although in a early report sperm head directed IgA- more than IgG-antibodies seemed to be associated with a reduced fertilization rate (128), in subsequent reports a significant reduction of the fertilization rate was found when sperm were covered both with IgG- and IgA-ASA (119,129, 130, and table 2). However, trying to analyse routine IFV results to determine the actual effect of sperm-bound antibodies on fertilization process is difficult because of some serious reasons. Firstly, nonimmunological sperm abnormalities may bias the results. Only in some reports the conventional semen parameters were taken into account in the comparison between patients with and without ASA (121,122,124,131). In these series, an independent impairment by sperm-bound antibodies was reported in the presence of normal semen parameters (121,124,131) as well as in the presence of asthenozoospermia (131) or teratozoospermia (122). Secondly, the criteria employed to define the occurrence of immunological infertility were different among series, and often inadequate. Also patients with low or moderate sperm autoimmunization were included in most series. The proportion of antibody-free sperm could be the determining factor of the fertilization outcome in those patients. Therefore, the inclusion of patients with low or moderate sperm autoimmunization could account for the normal overall fertilization rate found in some series, as well as for the variability of the fertilization outcome among

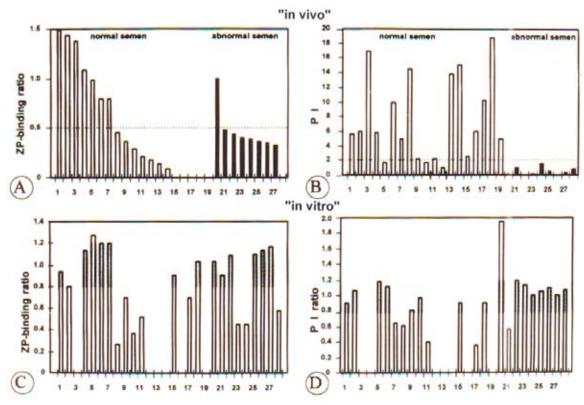


Figure 1. Results of the ZP-binding test (**A**) and the HEPT (**B**) exhibited by ejaculated sperm coated "in vivo" with antisperm antibodies from patients with normal and abnormal semen; PI = penetration Index (n° penetrations/oocyte); (----) indicates the cut off values for normal results; (**C**) and (**D**), effect of circulating antisperm antibodies from the same patients on the ZP-binding test (**C**) and on HEPT (**D**); the results are expressed as ZP-ratio and PI ratio, respectively, between antibody-exposed and non-exposed sperm suspensions from the same donor; *shaded area* represents the intra-assay variation. From Francavilla *et al.* (143) with permission.

individual cases of the same series. Indeed, when the degree of sperm autoimmunization was taken into account, it was always inversely correlated with the overall fertilization rate (130-132, and table 2). But, noteworthy, in some individual patients, a high fertilization rate was achieved even in the presence of a high degree of sperm autoimmunization (121,123,124,128).

In conclusion, the analysis of human IVF results seems to indicate that also on fertilization process sperm bound antibodies exert a relative impairment, which, to some extend, is related with the degree of sperm autoimmunization. However, the degree autoimmunization does not completely explain the variability of the antibody impairment. Seemingly, at the level of gamete interaction, more than at other levels (i.e., cervical mucus penetration) the interference of ASA exhibits qualitative apart from quantitative differences among patients, suggesting that this interference depends on the relevance of the specific antigens, targets of natural ASA, to the fertilization process.

3.4.2. The hamster egg penetration test (HEPT) as model of study

Conflicting results have been reported when the effect of circulating ASA on HEPT results was tested. In

fact either inhibition (133-135), or enhancement (136), both inhibition and enhancement (106,137), or no effect were found (138,139). Some discrepancies may be explained by procedural differences. Generally, when inhibitory effects were reported, donor sperm were exposed to ASA after capacitation or directly into the insemination medium: in any case, free antibodies were not removed before sperm/oocytes coincubation (106,134,135). Using this procedure, an interaction of ASA with internal sperm antigens, which can be revealed after the acrosomal loss, has been supposed to explain the inhibitory effect (140). However, although this possible interference may occur in the presence of ASA in the female, it cannot occur when ASA are detected in the male. When donor sperm were washed following exposure to ASA and then capacitated (procedure which is more suitable for studying the effects of naturally occurring ASA in the male), generally no effect (138,139) or even an enhancement of penetration was found (136).

Zouari and De Almeida (141) reported that antibodies eluted from eight autoimmune ejaculates and transferred onto donor sperm, reduced sperm penetration in three cases and in a case a modest increasing effect was exhibited. All samples with inhibitory effect contained both

IgG and IgA. The elimination of one of the two isotypes restored the ability of the sperm to penetrate the hamster oocytes.

Using spermatozoa coated in vivo with ASA Haas et al. (142) reported a variable degree of impairment in penetrating hamster oocytes, but the concomitant presence of astheno- and/or-teratozoospermia may bias the results. More recently, using the TEST-yolk buffer enhanced HEPT, our group (143) found that 12/28 patients with all ejaculated sperm coated with IgG- or IgG/IgA-antibodies exhibited a penetration index (PI = penetrations per oocyte) < 2 (the lowest value exhibited by fertile controls). But excluding from the analysis 9 patients exhibiting abnormal semen in terms of oligo/astheno-and/or-teratozoospermia. only in 16% of cases the PI was slightly less than 2 (figure 1B). Moreover, only 24% of the sera from the patients produced a reduction of the PI (figure1D), when absorbed on donor sperm suspensions, and in no case was the inhibitory effect great enough to cause a poor HEPT. It could be argued that after preincubation in TEST-yolk buffer antibody-coated sperm acquire the ability to fuse with oolemma by virtue of the spermatozoa being made highly fusogenic in spite of the presence of antibodies on their surface. This is not the authors opinion, who previously reported that ASA do not interfere also with the results of conventional HEPT (138).

3.4.3. Effects on capacitation and acrosome reaction

An interference of ASA with the acrosome reaction (AR) either as promoting or inhibitory effect was hypothesized since early experimental and clinical data suggested an antibody interference with gametes interaction. In some studies the effect of ASA on spontaneous acrosome reaction rate under capacitation conditions was evaluated. In most of them, no effect was found (137,138,144,145). In the most extensive of these studies (138), our group demonstrated that the exposure to ASA of all isotypes, even in association, from sera or seminal plasma samples, did not modify the spontaneous acrosome reaction rate of donor sperm used for HEPT, whose outcome similarly was not affected. On the contrary, testing ASA from female sera, an inhibitory effect was found in two reports (146,147). However, in the first report (146) the mean AR rate was 2.8% in antibody-exposed sperm and 5.8% in control sperm. The biological significance of this difference, although reported as significant, is doubtful. In the second report (147), the omission of vitality assessment, which permit to differentiate true from degenerative AR, may bias the results. Finally, in a recent report (148) in vivo antibodycoated sperm from infertile patients exhibited a massive acrosome loss not only in capacitating conditions but also in native preparations. The omission of vitality assessment and the inexplicable high rate of AR, which was also found in the control group, make these results hard to be accepted. However, high levels of spontaneous AR had also been reported by Lansfort et al.(149) in most patients with sperm coated in vivo with IgG plus IgA antibodies. However, neither in this report vitality assessment was carried out. Indeed, there is a general agreement that in human sperm, spontaneous AR represents a sporadic event (150) with

little biological efficacy, whereas, the sperm ability to undergo a complete AR in response to ionophore challenge has been reported as significantly related to the fertility status (151) and to the human IVF rates (152,153). Since the ionophore challenge bypasses the biological signalling that initiates the AR, and uncapacitated sperm respond poorly (154), this test may measure the capacitation status and the integrity of the chain of events between entry on calcium and exocytosis. In light of these considerations, in some studies the effect of ASA on AR induced by ionophore challenge was evaluated, but conflicting results were reported. Mahony et al. (144) reported an inhibitory effect exhibited by two sera with ASA also inhibiting ZPbinding, and Zouari et al. (145) reported an inhibitory effect exhibited by sperm-eluted antibodies. On the contrary, in a report of our group (139), circulating and seminal ASA caused a slight but constant and significant AR increase in response to ionophore challenge compared with that of the same donor sperm suspensions exposed to control sera. However the clinical relevance of this effect was considered as questionable, since it was not reflected, in most cases, in the results of HEPT also performed after ionophore challenge.

The effect of ASA on other aspects related to the capacitation process, such as hyperactivated motility (144,145) or calcium uptake after stimulation with follicular fluid (144) was also studied with inconclusive results. An interference with the capacitation-associated expression of sperm surface mannose receptors was reported by Benoff *et al.* (147) using ASA contained in female sera. This effect was related to an inhibition of the reduction of membrane cholesterol content associated with sperm capacitation, which prevented the membrane fluidity changes needed for mannose receptors expression.

3.4.4. Effects on the zona pellucida (ZP) interaction

There is general agreement that ASA can interfere with ZP-interaction. Circulating ASA have been shown to reduce sperm binding to (144,155-157) and penetration through (158) the ZP. Using spermatozoa coated in vivo with ASA, Liu et al.(159) found a reduced binding to saltstored human ZP as compared to donors' spermatozoa. However the low number of patients and the presence of concomitant non-immunologic semen abnormalities (even men with 5% normal sperms were included in the study) make difficult to infer the responsibility of ASA in this impairment. Zouari and De Almeida (141) reported that antibodies eluted from autoimmune ejaculates uniformly reduced the ZP-binding of spermatozoa from donors. Although this demonstration indicates the responsibility of sperm-coated antibodies in this impairment more appropriately, the very low number of samples (number=5) again does not permit the inference of information about the actual occurrence of this impairment. More recently our group (143) tested 22 patients with all ejaculated spermatozoa coated with antibodies against the sperm head, taking into account in the evaluation of the results the concomitant presence of non-immunological abnormalities in the conventional semen profile. An impairment of the ZP-binding was demonstrable in 50% of patients with normal semen profile (figure1A). Since a normal ZP-

binding was observed even when all ejaculated spermatozoa were coated with both IgG- and IgAantibodies, neither Ig class, even combined, appears to affect unavoidably this sperm function. Noteworthy, all normozoospermic patients with low ZP-binding showed circulating IgG-ASA with inhibitory effect, when transferred on donor sperm, while no patient with normal ZP-binding showed circulating ASA with inhibitory effect (figure1C). Therefore, IgG-ASA transuded from the blood into the genital tract can exert the inhibitory effect on ZPbinding in vivo. Zouari and De Almeide (141) reported that the removal of either IgG or IgA antibodies eluted from autoimmune ejaculates did not change the inhibitory effect on ZP-binding. Altogether, these observations suggest that both humoral and local sperm autoimmunizzation exhibit the same behaviour in impairing or not the ZP-binding. The fact that ZP-binding is not unavoidably inhibited by ASA, could explain the high fertilization rate in IVF programs, which was reported in some cases even in the presence of a high degree of sperm-autoimmunization (121,123,124,128). However, the ZP-binding is only the first step of the more complex interaction between spermatozoa and zona. Zona pellucida also triggers the AR of bound sperm, that is required by sperm for ZP penetration and fertilization. We recently demonstrated that ASA can interfere with sperm-ZP interaction not only by inhibiting ZP-binding but also by inhibiting the induction of AR by ZP (160). While the inhibition of ZP-binding is always associated with the inhibition of ZP-induced AR, this last interference can also occur in the absence of the inhibitory effect on ZP-binding. However, neither inhibitory effect may occur. The occurrence of different effects is not related with the ASAtitre, and it can be explained by the polyclonal nature of the naturally occurring ASA in men. Sperm-antibodies inhibiting ZP-binding could mask or prevent the expression of specific receptors on the plasma membrane over the sperm heads for the ZP3-O-linked oligosaccharides. Since ZP3 serves as both a ligand for sperm binding and a trigger for acrosome reaction (161), ASA, which inhibit ZP-binding, inhibit ZP-induced AR too. When an inhibition of ZP-induced AR occurs in the absence of interference on ZP-binding, ASA could interfere with cross-linking of several antigenic sites recognized by ZP3 resulting in a blockade of their aggregation which triggers the acrosome exocitosis (162). Another possibility is that ASA affect the fusogenic response to the biological signalling that triggers the AR. We are in favour of the first hypothesis since in our hands ASA did not affect the induction of AR by calcium-ionophore (139).

3.5. Postfertilization effects

Although a reduction in cleavage rate in the presence of ASA in the female was reported in one early IVF series (125), most clinical data indicate that spermbound antibodies do not interfere with postfertilization events. No reduction of cleavage and pregnancy rates have been generally reported in IVF programs where the presence of sperm-bound antibodies was associated to a reduced fertilization rate (120,121,131,132). However, in disagreement with previous data, a detrimental effect on early embryonic development was more recently claimed by Vazquez-Levin *et al.* (124), who reported a significant

reduction both in the cleavage rate and pregnancy rate in the presence of sperm-bound antibodies.

Data from intracytoplasmatic sperm injection (ICSI) available so far, confirm that ASA are not associated with a reduction of cleavage and pregnancy rate (163-165), even if a poorer embryo quality (163) and a higher rate of pregnancy loss (164) were reported. However, neither a poorer embryo quality nor a higher rate of pregnancy loss were born out by Clarke *et al.* (165).

Since in animals embryos share epitopes with sperm antigens (166), antibodies occurring in the female against sperm antigens could interact with embryonic antigens, providing an attractive reason to hypothesize that ASA could adversely affect embryonic development, when they occur in the female. Antibodies against the cleavage signal (CS-1), sperm derived protein which should function as an extranuclear cleavage signal for early division of fertilized zygotes, could represent an attractive explanation of a possible postfertilization effect of ASA when they occur also in the male (167). However the actual role and occurrence of these antibodies have to be determined.

4. IMPLICATIONS FOR TREATMENT

In the evaluation of possible therapeutic modalities for infertile patients with ASA, it is important to keep in mind two previous analysed considerations: 1) ASA are a relative cause of infertility, which is related to the degree of autoimmunization; 2) some mechanisms of the antibody-interference with fertility depend only on the degree of autoimmunization (e.g., sperm-agglutination, inhibition of cervical mucus penetration), while other mechanisms could or could not occur depending on the specific antigen(s) involved in the immune response (e.g., interference with gametes interaction); 3) It is not possible prevent or disrupt antigen/antibody complexes on sperm surface by means of "in vitro" sperm processing tecniques unless methods inconsistent with sperm vitality (heating at 56°C or lowering the pH to <3) are utilized (168.169)

4.1. Corticosteroid therapy

The rationale for this treatment is to reduce the production of ASA, thereby obtaining a proportion of antibody-free sperm sufficient for fertilization. Either longterm low dose treatment (e.g., prednisolone, 5 mg three times daily for at least 6 months)(170), even following soppression of spermatogenesis with testosterone (171), or intermittent high-doses of metilprednisolone (96 mg/day for 7 days) (172) have been widely used, with better results claimed with the latter approach (170). Unfortunately, most studies lack a placebo control (173 for review). In a doubleblind, placebo-controlled study, intermittent high-doses of metilprednisolone did not produce a favourable effect over placebo on the men's subsequent fertility (174). Because of the risk of serious adverse effect of high doses of corticosteroid treatment (175) an intermediate-dose cyclical regimen was evolved. In a double-blind crossover trial, prednisolone treatment, at 20 mg twice daily on days 1-10 of the female partner menstrual cycle, followed by 5 mg on days 11 and 12, was associated with a cumulative

pregnancy rate of 31% during 9 months, which was significantly higher than the rate of 9.5% for placebo (176). Unfortunately, with the same treatment and the same study design, no pregnancy was achieved during three months in a subsequent report (177). In both studies, the circulating ASA-titres were not significantly modified by steroid treatment, while a significant fall in antibody titres in seminal plasma was found in the former. With intermittent high-doses of metilprednisolone, a significant reduction of sperm-associated IgG had been reported radiolabeling antiglobulin assay, with no effect either on sperm-associated IgA or on circulating IgG-ASA levels (174). Finally, in a placebo-controlled flow cytometric study, the antibody levels measured before and after treatment with low-dose prednisolone (20 mg/day) or with placebo were not statistically different, but in 2/10 patients treated with prednisolone, a marked decrease in the proportion of spermatozoa positive both for IgG and IgA was observed (178).

In conclusion the efficacy of corticosteroid treatment has not been definitively proven.

4.2. Intrauterine insemination

Since the most established interference of ASA with fertility is represented by the impairment of cervical mucus penetration by antibody-coated sperm, intrauterine insemination (IUI) has been widely used for the treatment of the male immunological infertility. However, the usefulness of IUI for this indication remains controversial because conflicting results have been reported. In studies, which included patients with variable degree of spermautoimmunization (179-183), the reported pregnancy rate/couple ranged from 0%(181) to 37% (182). Indeed, there are two questions to be answered. The first one is: does overcoming the cervical mucus barrier remove the clinical interference of sperm-coated antibodies with fertility? In a study designed with this purpose(184), our group did not obtain any pregnancy with 110 IUIs in 19 couples, where all ejaculated spermatozoa were antibodycoated, while a pregnancy rate/couple of 25.6% (5.6% cycle fecundity) was obtained in the control group (n°=86) without ASA. The responsibility of ASA for the failure of IUI was inferred from the significant difference obtained by comparing the results in patients with and without ASA, homogeneous for both epidemiological data and seminal parameters, having excluded teratozoospermic patients (with and without ASA) from the analysis, since teratozoospermia had been proved as strongly impairing the outcome of IUI (185). The opsonizing effect through the female genital tract and/or the effects on fertilization process could account for this failure. In disagreement with this report, 6 out of 9 patients with ASA, whose wife conceived following IUI, exhibited a strong positive (>90%) IgG-MAR test in a crossover, randomized trial, where IUI (3 cycles/couple) was significantly more effective than cyclic, low dose prednisone treatment (1 pregnancy) in 40 couples (186).

The second question is: Can IUI improve the chance of achieving a conception when used as an adjuvant therapy to steroid treatment? In a randomized, cross-over

study comparing IUI in superovulated cycles with natural intercourse in men receiving cyclical intermediate dose steroid therapy for immunological subfertility (immunobead binding levels $\geq 50\%$ in either seminal plasma or serum), a cumulative pregnancy rate of 39.4% over four cycles of IUI was achieved, compared with only 4.8% over four cycles of timed intercourse with the same regimen of steroid therapy (187).

In conclusion the analysis of the reported data indicates that IUI is an effective treatment for low or moderate sperm autoimmunization, mainly if combined with corticosteroid treatment and superovulated cycles. On the contrary, its effectiveness in cases of high degree of sperm autoimmunization is controversial. In a recent pilot non-randomized study (188), where the effectiveness of IUI with ovarian stimulation was compared with that of IVF in 29 couples where the male partner had a positive (>50%) direct MAR test, 64.3% of patients conceived after a maximum of three IUI cycles, while 46.6% of patients conceived during the first IVF cycle. Cost benefit analysis favoured a course of four IUI cycles, indicating this treatment as a valuable first-choice method to use before starting more invasive and expensive techniques of assisted reproduction.

4.3. IVF-ET vs ICSI

The resort to IFV procedures (IVF-ET or ICSI) is mandatory when other less invasive approaches have failed or they may also be chosen as a first-choice method in cases of high degree of sperm autoimmunization. The question is: IVF-ET or ICSI? Since in most reports the fertilization rate was significantly lower in the presence of sperm-bound antibodies than in the case of other indications for IVF-ET (119-124; see also section 3.4.1.), the likelihood of fertilization is higher with ICSI, where the reported fertilization rates were similar to those in other indications (164,165), or even significantly higher (163). The inability of antibody-coated sperm to bind to the zona pellucida is apparently the main obstacle to fertilization (see section 3.4.), but it not unavoidably occurs (143). Therefore, the outcome of the ZP-binding test could identify those patients with immunological infertility who profit by conventional IVF and those who need micromanipulation techniques applied to IVF to achieve fertilization. However, the demonstration that ASA can inhibit the induction of AR by ZP in the absence of an inhibitory effect on ZP-binding (160) indicates that the ZPbinding test does not completely explore the interference of ASA with the sperm-ZP interaction. Therefore, testing ARinduction by ZP could be usefully included in the diagnostic screening before IVF. But simpler tests should be developed, and recombinant human ZP3 would be a convenient tool in such development.

5. PERSPECTIVE

Although many aspects of the infertility due to naturally-occurring ASA in men has been clarified, some controversies remain. Greater standardization of antibody testing, further development of sperm function assays, and more precise criteria to define the occurrence of

immunological infertility could help to throw light on the remaining debated aspects. Furthermore, the identification of the sperm antigens recognized by natural ASA, as well as the recognition of their relevance in the fertilization process, could shed new light upon this matter. Although a few definite sperm antigens relevant to fertilization have been characterized fairly well (189 for review), a role in human immunoinfertility has been suggested by some evidences only for two of them, fertilization antigen 1 (FA-1)(190-192) and cleavage signal (CS-1) protein,(167; see also section 3.5.).

The development of immunocontraceptive strategy has renewed the interest toward the study of clinical infertility mediated by ASA. Antisperm antibodies develop in post-vasectomized men or they spontaneously occur in infertile men without physiological complications, despite their persistence for years. Thus, ASA induced by immunization of men or women with antigens involved in natural immunoinfertility might similarly be without side effects. With this assumption, the study of clinical infertility due to ASA, regarded as "experiments of nature" in fertility reduction, has been approached to identify candidate sperm antigens for immunocontraceptive development (193,194). Numerous investigators have reported the identification of specific antigens using sera from WHO reference bank or from independent collections (106, 195-206). However, characterization of sperm antigens was limited to SDS-Page in most studies, and results varied widely due to the methods and reagents employed. With another approach, a two-dimensional protein database of human sperm proteins has been created as a means to identify sperm-surface proteins: vectorial labelling of the cell surface by biotinylation and iodination identified 98 dual-labelled sperm surface proteins (207). Only 3 of them was recognized by postvasectomy sera in a preliminary observation (208).

6. REFERENCES

- 1. P. Rumke: The presence of sperm antibodies in the serum of two patients with oligospermia. *Vox Sang* 4, 135-40 (1954)
- 2. L. Wilson: Sperm agglutinins in human semen and blood. *Proc Soc Exp Biol Med* 85, 652-55 (1954)
- 3 .R. Bronson, G. Cooper & D. Rosenfeld: Sperm antibodies: their role in infertility. *Fertil Steril* 42, 171-83 (1984)
- 4. WHO Reference Bank for Reproductive Immunology: Auto- and iso-antibodies to antigens of the human reproductive system. 1. Results of an international comparative study. *Clin Exp Immunol* 30, 173-80 (1977)
- 5. R. Bronson, G. Cooper, T. Hjort, R. Ing, W. R. Jones, S. X. Wang, S. Mathur, H. O. Williamson, P. F. Rust, H. H. Fudenberg, L. Mettler, A. B. Czuppon & N. Sudo: Antisperm antibodies, detected by agglutination, immobilization, microcytotoxicity and immunobead-binding assays. *J Reprod Immunol* 8, 279-99 (1985)
- 6. J. Friberg: A simple and sensitive micro-method for demonstration of sperm-agglutinating activity in serum

- from infertile men and women. Acta Obstet Gynecol Scand, suppl 36, 21-29 (1974)
- 7. D.T. Uehling: Secretory IgA in seminal fluid. Fertil Steril 22, 769-73 (1971)
- 8. Ph. Rumke: The origin of immunoglobulins in semen. *Clin Exp Immunol* 12, 287-97 (1974)
- 9. S. Jager, J. Kremer & T. van Slochteren-Draaisma: A simple method of screening for antisperm antibodies in the human male: detection of spermatozoan surface IgG with the direct mixed agglutination reaction carried out on untreated fresh human semen. *Int J Fertil* 23, 12-21 (1978)
- 10. R. Bronson, G. Cooper & D. Rosenfeld: Membrane-bound sperm-specific antibodies: their role in infertility. In "Bioregulators in Reproduction" Eds: Vogel H, Jagiello G. *Academic Press*, New York, p.521-27 (1981)
- 11. F. H. Comhaire, A. Hinting, L. Vermeulen, F. Schoonjans & I. Goethals: Evaluation of the direct and indirect mixed antiglobulin reaction with latex particles for the diagnosis of immunological infertility. *Int J Androl* 11, 37-44 (1987)
- 12. W. J. G. Hellstrom, J. W. Overstreet, S. J. Samuels & E. L. Lewis: The relationship of circulating antisperm antibodies to sperm surface antibodies in infertile men. J Urol 140,1039-44 (1988)
- 13. S. V. Rajah, J. M. Parslow, R. J. S. Howell & W. F. Hendry: Comparison of mixed antiglobulin reaction and direct immunobead test for detection of sperm-bound antibodies in subfertile males. *Fertil Steril* 57, 1300-303 (1992)
- 14. F. Francavilla, P. Catignani, R. Romano, R. Santucci, S. Francavilla, G. Poccia, V. Santiemma & A. Fabbrini: Immunological screening of a male population with infertile marriages. *Andrologia* 16, 578-86 (1984)
- 15. H. Meinertz & T. Hjort: Detection of autoimmunity to sperm: mixed antiglobulin reaction (MAR) test or sperm agglutination? A study on 537 men from infertile couples. *Fertil Steril* 46, 86-91 (1986)
- 16. G. N. Clarke, P. J. Elliott & C. Smaila: Detection of sperm antibodies in semen using the immunobead test: a survey of 813 consecutive patients. *Am J Reprod Immunol Microbiol* 7, 118-23 (1985)
- 17. S. Ackerman, G. McGuire, D. L. Fulgham & N. J. Alexander: An evaluation of a commercially available assay for the detection of antisperm antibodies. *Fertil Steril* 49, 732-34 (1988)
- 18. W. F. Hendry & J. Stedronska: Mixed erythrocyte-spermatozoa antiglobulin reaction (MAR test) for the detection of antibodies against spermatozoa in infertile males. *J Obstet Gynecol* 1, 59-62 (1980)
- 19. F. Francavilla, R. Santucci, R. Romano, S. Francavilla, L. Casasanta, & G. Properzi: A direct immunofluorescence test for the detection of sperm surface bound antibodies. Comparison with sperm agglutination test, indirect IF test and MAR test. *Andrologia* 20, 477-83 (1988)
- 20. D. A. Ohl & R. K. Naz: Infertility due to antisperm antibodies. *Urology* 46, 591-602 (1995)

- 21. E. Andreou, A. Mahmoud, L. Vermeulen, F. Schoonjans & F. Comhaire: Comparison of different methods for the investigation of antisperm antibodies on spermatozoa, in seminal plasma and in serum. *Hum Reprod* 10, 125-131 (1995)
- 22. E. Evans, P. J. Chan, W. C. Patton & A. King: A convenient mixed immunobeads screen for antisperm antibodies during routine semen analysis. Fertil Steril 70, 344-349 (1998)
- 23. G. G. Haas, D. B. Cines & A. D. Schreiber: Immunologic infertility: identification of patients with antisperm antibody. *N Engl J Med* 303, 722-27 (1980)
- 24. G.G. Haas, R. Weiss-Wik & D. P. Wolf: Identification of antisperm antibodies on sperm of infertile men. *Fertil Steril* 38, 54-61 (1982)
- 25. R. Zanchetta, F. Busolo & I. Mastrogiacomo: The enzyme-linked immunosorbent assay for detection of the antispermatozoal antibodies. *Fertil Steril* 38, 730-34 (1982)
- 26. S. Paul, V. Baukloh & L. Mettler: Enzyme-linked immunosorbent assays for sperm antibody detection and antigenic analysis. *J Immunol Methods* 56, 193-99 (1983)
- 27. R. Zanchetta & F. Busolo: A simplified method using enzyme-linked immunosorbent assay for titration of antisperm antibodies. *Am J Reprod Immunol* 5, 182-84 (1984)
- 28. D. M. Lynch & S. E. Howe: Comparison of a direct and indirect ELISA for quantitating antisperm antibody in semen. *J Androl* 8, 215-20 (1987)
- 29. F. Francavilla, R. Romano, R. Santucci, P. Catignani, A. Barone & A. Fabbrini: Detection of sperm surface related antibodies by direct immunofluorescence test on sperm suspensions. *Andrologia* 19, 69-75 (1987)
- 30. G. G. Haas & M. E. Cunningham: Identification of antibody-laden sperm by cytofluorometry. *Fertil Steril* 42, 606-13 (1984)
- 31. M. L. Rasanen, O. L. Hovatta, I. M. Penttila & Y. P. Agrawal: Detection and quantitation of sperm-bound antibodies by flow cytometry of human semen. *J Androl* 13, 55-64 (1992)
- 32. M. A. Nikolaeva, V. I. Kulakov, A. G. Ter, L. N. Terekhina, T. J. Pshenichnikova & G. T. Sukhikh: Detection of antisperm antibodies on the surface of living spermatozoa using flow cytometry: preliminary study. *Fertil Steril* 59, 639-44 (1993)
- 33. R. W. Ke, M. E. Dockter, G. Majumdar, J. E. Buster & S. A. Carson: Flow cytometry provides rapid and highly accurate detection of antisperm antibodies. *Fertil steril* 63, 902-6 (1995)
- 34. M. Rasanen, Y. P. Agrawal & S. Saarikoski: Seminal fluid antisperm antibodyes measured by direct flow cytometry do not correlate with those measured by indirect flow cytometry, the indirect immunobead test, and the indirect mixed antiglobulin reaction. *Fertil Steril* 65, 170-75 (1996)
- 35. S. C. Nicholson, J. N. Robinson, I. L. Sargent & D. H. Barlow: Detection of antisperm antibodies in seminal

- plasma by flow cytometry: comparison with the indirect immunobead binding test. *Fertil Steril* 68, 1114-19 (1997) 36. A. Heidenreich, R. Bonfig, D. M. Wilbert, W. L. Strohmaier & U. H. Engelmann: Risk factors for antisperm antibodies in infertile men. *Am J Reprod Immunol* 31, 69-76 (1994)
- 37. T. B. Hargreave, M. Haxton, J. Whitelaw, R. Elton & G. D. Chisholm: The significance of serum spermagglutinating antibodies in men with infertile marriages. *Brit J Urol* 52, 566-70 (1980)
- 38. A. C. Menge, N. E. Medley, C. M. Mangione & J. W. Dietrich: The incidence and influence of antisperm antibodies in infertile human couples on sperm-cervical mucus interactions and subsequent fertility. *Fertil Steril* 38, 439-46 (1982)
- 39. A. C. Menge & O. Beitner: Interrelationships among semen characteristics, antisperm antibodies, and cervical mucus penetration assays in infertile human couples. *Fertil Steril* 51, 486-92 (1989)
- 40. J. A. Collins, E. A. Burrows, J. Yeo & E. V. YoungLai: Frequency and predictive value of antisperm antibodies among infertile couples. *Hum Reprod* 8, 592-98 (1993)
- 41. H. J. Ingerslev: Characterization of sperm agglutinins in sera from infertile women. *Int J Fertil* 24, 1-12 (1979)
- 42. H. A. Pattinson & D. Mortimer: Prevalence of sperm surface antibodies in the male partners of infertile couples as determined by immunobead screening. *Fertil Steril* 48, 466-69 (1987)
- 43. M. Busacca, F. Fusi, C. Brigante, N. Doldi, M. Smid & P. Vigano: Evaluation of antisperm antibodies in infertile couples with immunobead test: prevalence and prognostic value. *Acta Eur Fertil* 20, 77-82 (1989)
- 44. J. P. Jarow & J. J. Sanzone: Risk factors for male partner antisperm antibodies. *J Urol* 148, 1805-7 (1992)
- 45. P. Devine, B. J. Sedensky, H. S. Jordan, A. J. Friedman & B. M. Berger. Detecting semen antisperm antibodies in the clinical laboratory. *Arch Pathol Lab Med* 117, 784-88 (1993)
- 46. A. A. Sinisi, B. Di Finizio, D. Pasquali, C. Scurini, A. D'Apuzzo & A. Bellastella: Prevalence of antisperm antibodies by SpermMAR test in subjects undergoing a routine sperm analysis for infertility. *Int J Androl* 16, 311-14 (1993)
- 47. L. Gandini, A. Lenzi, F. Culasso, F. Lombardo, D. Paoli & F. Dondero: Study of antisperm antibodies bound to the sperm cell surface and their relationship to circulating ASA. *Am J Reprod Immunol* 34, 375-80 (1995)
- 48. H. Fisch, E. Laor, N. BarChama, S. S. Witkin, B. M. Tolia & R. E. Reid: Detection of testicular endocrine abnormalities and their correlation with serum antisperm antibodies in men following vasectomy. *J Urol* 141, 1129-32 (1989)
- 49. G. A. Broderick, R. Tom & R. D. McClure: Immunological status of patients before and after vasovasostomy as determined by the immunobead antisperm antibody test. *J Urol* 142, 752-55 (1989)

- 50. J. P. Jarow, E. T. Goluboff, T. S. Chang & F. F. Marshall: Relationship between antisperm antibodies and testicular histologic changes in humans after vasectomy. *Urology* 43, 521-24 (1994)
- 51. R. A. Newton: IgG antisperm antibodies attached to sperm do not correlate with infertility following vasovasostomy. *Microsurgery* 9, 278-80 (1988)
- 52. P. L. Matson, S. M. Junk, J. R. Masters, J. P. Pryor & J. L. Yovich: The incidence and influence upon fertility of antisperm antibodies in seminal fluid following vasectomy reversal. *Int J Androl* 12, 98-103 (1989)
- 53. O. J. D'Cruz, G. G. Haas, R. De La Rocha & H. Lambert: Occurrence of serum antisperm antibodies in patients with cystic fibrosis. *Fertil Steril* 56, 519-27 (1991)
- 54. R. A. Bronson, W. J. O'Connor, T. A. Wilson, S. K. Bronson, F. I. Chasalow & K. Droesch: Correlation between puberty and the development of autoimmunity to spermatozoa in men with cystis fibrosis. *Fertil Steril* 58, 1199-204 (1992)
- 55. M. H.Vazques-Levin, G. S. Kupchik, Y. Torres, C. A. Chaparro, A. Shtainer, R. J. Bonforte & H. M. Nagler: Cystic fibrosis and congenital agenesis of the vas deferens, antisperm antibodies and CF-genotipe. *J Reprod Immunol* 27, 199-212 (1994)
- 56. P. Patrizio, I. Moretti-Rojas, J. Balmaceda, S. Silber & R. H. Asch: Low incidence of sperm antibodies in men with congenital absence of the vas deferens. *Fertil Steril* 52, 1018-21 (1989)
- 57. P. Patrizio, S. J. Silber, T. Ord, R. I. Moretti & R. H. Asch: Relationship of epididymal sperm antibodies to their in vitro fertilization capacity in men with congenital absence of the vas deferens. *Fertil Steril* 58, 1006-10 (1992)
- 58. G. N. Clarke: Sperm antibodies in normal men: association with a history of nongonococcal urethritis (NGU). *Am J Reprod Immunol Microbiol* 12, 31-32 (1986)
- 59. M. Shahmanesh, J. Stedronska & W. F. Hendry: Antispermatozoal antibodies in men with urethritis. *Fertil Steril* 46, 308-11 (1986)
- 60. H. J. Ingerslev, S. Walter, J. T. Andersen, P. Brandenhoff, J. Eldrup, J. P. Geerdsen, J. Scheibel, N. Tromholt, H. M. Jensen & T. Hjort: A prospective study of antisperm antibody development in acute epididymitis. *J Urol* 136, 162-64 (1986)
- 61. S. S. Witkin & G. Zelikovsky: Immunosuppression and sperm antibody formation in men with prostatitis. *J Clin Lab Immunol* 21, 7-10 (1986)
- 62. J. P. Jarrow, J. A. Jr Kirkland & D. G. Assimos: Association of antisperm antibodies with chronic nonbacterial prostatitis. *Urology* 36, 154-56 (1990)
- 63 Y. Soffer, R. Ron-El, A. Golan, A. Herman, E. Caspi & Z. Samra: Male genital mycoplasmas and Chlamydia trachomatis culture: its relationship with accessory gland function, sperm quality, and autoimmunity. *Fertil Steril* 53, 331-36 (1990)
- 64. S. S. Witkin, J. Jeremias, J. A. Grifo & W. J. Ledger: Detection of Chlamydia trachomatis in semen by the

- polymerase chain reaction in male members of infertile couples. *Am J Obstet Gynecol* 168, 1457-62 (1993)
- 65. W. Eggert-Kruse, G. Rohr, T. Demirakca, R. Rusu, H. Naher, D. Petzoldt & B. Runnebaum: Chamydial serology in 1303 asymptomatic subfertile couples. *Hum Reprod* 12, 1464-75 (1997)
- 66. S. S. Witkin, I. Kligman & A. M. Bongiovanni: Relationship between an asymtomatic male genital tract exposure to clamydia trachomatis and an autoimmune response to spermatozoa. *Hum Reprod* 10, 2952-55 (1995)
- 67. W. Eggert-Kruse, N. Buhlinger-Gopfarth, G. Rohr, S. Probst, J. Aufenanger, H. Naher & B. Runnebaum: Antibodies to clamydia trachomatis in semen and relationship with parameters of male infertility. *Hum Reprod* 11, 1408-17 (1996)
- 68. K. Hobarth, H. C. Klingler, U. Maier & H. Kollaritsch: Incidence of antisperm antibodies in patients with carcinoma of the testis and in subfertile men with normogonadotropic oligoasthenoteratozoospermia. *Urol Int* 52, 162-65 (1994)
- 69. R. S. Foster, L. R. Rubin, A. McNulty, R. Bihrle & J. P. Donohue: Detection of antisperm-antibodies in patients with primary testicular cancer. *Int J Androl* 14, 179-85 (1991)
- 70. S. Guazzieri, A. Lembo, G. Ferro, W. Artibani, F. Merlo, R. Zanchetta & F. Pagano: Sperm antibodies and infertility in patients with testicular cancer. *Urology* 26, 139-42 (1985)
- 71. R. L. Urry, D. T. Carrell, N. T. Starr, B. W. Snow & R. G. Middleton: The incidence of antisperm antibodies in infertility patients with a history of cryptorchidism. *J Urol* 151, 381-83 (1994)
- 72. H. Wolff & W. B. Schill: Antisperm antibodies in infertile and homosexual men: relationship to serologic and clinical findings. *Fertil Steril* 44, 673-77 (1985)
- 73. R. K. Naz, M. Ellaurie, T. M. Phillips & J. Hall: Antisperm antibodies in human immunodeficiency virus infection: effects on fertilization and embryonic development. *Biol Reprod* 42, 859-68 (1990)
- 74. L. E. Adams, R. Donovan-Brand, A. Friedman-Kien, el K. Ramahi & E. V. Hess: Sperm and seminal plasma antibodies in acquired immune deficiency (AIDS) and other associated syndromes. *Clin Immunol Immunopathol* 46, 442-49 (1988)
- 75. H. Ozen, G. Asar, S. Gungor & A. F. Peker: Varicocele and antisperm antibodies. *Int Urol Nephrol* 17, 97-101 (1985)
- 76. B. R. Gilbert, S. S. Witkin & M. Goldstein: Correlation of sperm-bound immunoglobulins with impaired semen analysis in infertile men with varicoceles. *Fertil Steril* 52, 469-73 (1989)
- 77. G. S. Oshinsky, M. V. Rodriguez & B. C. Mellinger: Varicocele-related infertility is not associated with increased sperm-bound antibody. *J Urol* 150, 871-73 (1993)
- 78. G. Knudson, L. Ross, D. Stuhldreher, D. Houlihan, E. Bruns & G. Prins: Prevalence of sperm bound antibodies in infertile men with varicocele: the effect of varicocele

- ligation on antibody levels and semen response. *J Urol* 151, 1260-62 (1994)
- 79. M. Cetinkaya, A. Memis, O. Adsan, S. Beyribey & B. Ozturk: Antispermatozoal antibody values after varicocelectomy. *Int Urol Nephrol* 26, 89-92 (1994)
- 80. G. Beretta, A. Zanollo, E. Chelo, C. Livi & G. Scarselli: Seminal parameters and auto-immunity in paraplegic/quadraplegic men. *Acta Eur Fertil* 18, 203-5 (1987)
- 81. A. Dahlberg & O. Hovatta: An ejaculation following spinal cord injury does not induce sperm-agglutinating antibodies. *Int J Androl* 12, 17-21 (1989)
- 82. A. C. Menge, D. A. Ohl, J. Denil, M. K. Korte, L. Keller & M. McCabe: Absence of antisperm antibodies in anejaculatory men. *J Androl* 11, 396-98 (1990)
- 83. I. H. Hirsch, J. Sedor, H. J. Callahan & W. E. Staas: Antisperm antibodies in seminal plasma of spinal cordinjured man. *Urology* 39, 243-47 (1992)
- 84. A. Siosteen, Y. Steen, L. Forssman & L. Sullivan: Auto-immunity to spermatozoa and quality of semen in men with spinal cord injury. *Int J Fertil* 38, 117-22 (1993)
- 85. D. A. Gubin, R. Dmochowski & W. H. Kutteh: Multivariant analysis of men from infertile couples with or without antisperm antibodies. *Am J Reprod Immunol* 39, 157-60 (1998)
- 86. R. Ansbacher, K. Keung-Yeung & S. J. Behrman: Clinical significant of sperm antibodies in infertile couples. *Fertil Steril* 24, 305-8 (1973)
- 87. H. W. Baker, G. N. Clarke, B. Hudson, J. C. McBain, M. P. McGowan & R. J. Pepperell: Treatment of sperm autoimmunity in men. *Clin Reprod Fertil* 2, 55-71 (1983)
- 88. W. Eggert-Kruse, M. Christmann, I. Gerhard, S. Pohl, K. Klinga & B. Runnebaum: Circulating antisperm antibodies and fertility prognosis: a prospective study. *Hum Reprod* 4, 513-20 (1989)
- 89. S. S. Witkin & S. S. David: Effect of sperm antibodies on pregnancy outcome in a subfertile population. *Am J Obstet Gynecol* 158, 59-62 (1988)
- 90. M. Busacca, F. Fusi, C. Brigante, N. Doldi, M. Smid & P. Vigano: Evaluation of antisperm antibodies in infertile couples with immunobead test: prevalence and prognostic value. *Acta Eur Fertil* 20, 77-82 (1989)
- 91. P. H. Rumke, Nanda Van Amstel, E. N. Messer & P. D. Bezemer: Prognosis of fertility of men with sperm agglutinins in the serum. *Fertil Steril* 25, 393-98 (1974)
- 92. B. Ayvaliotis, R. Bronson, D. Rosenfeld & G. Cooper: Conception rates in couples where autoimmunity to sperm is detected. *Fertil Steril* 43, 739-41 (1985)
- 93. C. L. Barratt, B. C. Dunphy, I. McLeod & I. D. Cooke: The poor prognostic value of low to moderate levels of sperm surface-bound antibodies. *Hum Reprod* 7, 95-98 (1992)
- 94. G. G. Jr Haas, A. D. Schreiber & L. Blasco: The incidence of sperm-associated immunoglobulins and C3, the third component of complement, in infertile men. *Fertil Steril* 39, 542-47 (1983)

- 95. M. Cerasaro, M. Valenti, A. Massacesi, A. Lenzi & F. Dondero: Correlation between the direct IgG MAR test (mixed antiglobulin reaction test) and seminal analysis in men from infertile couples. *Fertil Steril* 44, 390-95 (1985)
- 96. B. H. Petersen, C. J. Lammel, D. P. Stites & G. F. Brooks: Human seminal plasma inhibition of complement. *J Lab Clin Med* 96, 582-91 (1980)
- 97. R. J. Price, T. K. Robert, D. Green & B. Boettcher: Anticomplementary activity in human semen and its possible importance in reproduction. *Am J Reprod Immunol* 6, 92-98 (1984)
- 98. P. S. Tung & I. B. Fritz: Immuno localization of clusterin in the ram testis, rete testis and excurrent ducts. *Biol Reprod* 33, 177-86 (1985)
- 99. M. K. O' Bryan, H. W. G. Baker, J. R. Saunders et al: Human seminal clusterin (SP-40,40). Isolation and characterization. *J Clin Invest* 85, 1477-86 (1990).
- 100. O. J. D' Cruz & G. G. Haas Jr: Lack of complement activation in the seminal plasma of men with antisperm antibodies associated in vivo on their sperm. *Am J Reprod Immunol* 24, 51-57 (1990)
- 101. W. F. Hendry, J. Stedronska & L. Hughes: Steroid treatment of male subfertility caused by antisperm antibodies. *Lancet* 8, 498-500 (1979)
- 102. M. De Almeida, A. Soumah & P. Jouannet: Incidence of sperm-associated immunoglobulins in infertile men with suspected autoimmunity to sperm. *Int J Androl* 9, 321-30 (1986)
- 103. G. G. Haas: The inhibitory effect of sperm-associated immunoglobulins on cervical mucus penetration. *Fertil Steril* 46, 334-37 (1986)
- 104. S. Mathur, H. O. Williamson, M. E. Baker, P. F. Rust, G. L. Holtz & H. H. Fudenberg: Sperm motility on post coital testing correlates with male autoimmunity to sperm. *Fertil Steril* 41, 81-87 (1984)
- 105. R. A. Bronson, G. W. Cooper & D. L. Rosenfeld: Autoimmunity to spermatozoa: effect on sperm penetration of cervical mucus as reflected by post coital testing. *Fertil Steril* 41, 609-14 (1984)
- 106. R. J. Aitken, J. M. Parsow, T. B. Hargreave & W. F. Hendry: Influence of antisperm antibodies on human sperm function. *Br J Urol* 62, 367-73 (1988)
- 107. J. Kremer & S. Jeger: Characteristics of antispermatozoal antibodies responsible for the shaking phenomenon with special regard to immunoglobulin class and antigen-reactive sites. *Int J Androl* 3, 143 (1980)
- 108. C. Wang, H. W. G. Baker, M. G. Jennings, H. G. Burger & P. Lutjen: Interaction between human cervical mucus and sperm surface antibodies. *Fertil Steril* 44, 484-88 (1985)
- 109. G. N. Clarke: Immunoglobulin class and regional specificity of antispermatozoal autoantibodies blocking cervical mucus penetration by human spermatozoa. *Am J Reprod Immunol Microbiol* 16, 135-38 (1988)
- 110. S. Jager, J. Kramer, J. Kuiken & I. Mulder: The significance of the Fc part of antispermatozoal antibodies

- for the shaking phenomenon in the sperm-cervical mucus contact test. Fertil Steril 36, 792 (1981)
- 111. R. A. Bronson, G. W. Cooper, D. L. Rosenfeld, J. V. Gilbert & A. G. Plaut: The effect of an IgA₁ protease on immunoblobulins bound to the sperm surface and sperm cervical mucus penetrating ability. *Fertil Steril* 47, 985-91 (1987)
- 112. R. J. Price & B. Boettcher: The presence of complement in human cervical mucus and its possible relevance to infertility in women with complement-dependent sperm-immobilizing antibodies. *Fertil Steril* 32, 61-66 (1979)
- 113. O. J. D'Cruz, G. G. Haas Jr. & H. Lambert: Evaluation of antisperm complement-dependent immune mediators in human ovarian follicular fluid. *J Immunol* 144, 3841-48 (1990)
- 114. O. J. D'Cruz, G. G. Haas Jr., B. Wang & L. E. DeBault: Activation of human complement by IgG antisperm antibodies and the demonstration of C3 and C5b-9-mediated immune injury to human sperm. *J Immunol* 146, 611-20 (1991)
- 115. S. N. London, A. F. Haney & J. B. Weinberg: Macrophages and infertility: enhancement of human macrophage-mediated sperm killing by antisperm antibodies. *Fertil Steril* 43, 274-78 (1985)
- 116. K. S. K. Tung, A. Okada & R. Yanagimachi: Sperm autoantigens and fertilization. I. Effect of antisperm antibodies om rouleaux formation, viability, and acrosome reaction of guinea pig spermatozoa. *Biol Reprod* 23, 877-86 (1980)
- 117. R. Yanagimachi, A. Okada & K. S. K. Tung: Sperm autoantigens and fertilization. II. Effects of anti-guinea pig sperm autoantibodies on sperm ovum interactions. *Biol Reprod* 24, 512-18 (1981)
- 118. W. J. Snell & J. M. White: The molecules of mammalian fertilization. *Cell* 85, 629-37 (1996)
- 119. P. L. Matson, S. M. Junk, J. W. Spittle & J. L. Yovich: Effect of antispermatozoal antibodies in seminal plasma upon spermatozoal function. *Int J Androl* 11, 101-6 (1988)
- 120. T. H. Chang, M. H. Jih. & T. C. J. Wu: Relationship of sperm antibodies in women and men to human in vitro fertilization, cleavage, and pregnancy rate. *Am J Reprod Immunol* 30, 108-12 (1993)
- 121. S. V. Rajah, J. M. Parslow, R. J. Howell & W. F. Hendry: The effects on in-vitro fertilization of autoantibodies to spermatozoa in subfertile men. *Hum Reprod* 8, 1079-82 (1993)
- 122. A. A. Acosta, J. P. van der Merwe, G. Doncel, T. F. Kruger et al: Fertilization efficiency of morphologically abnormal spermatozoa in assisted reproduction is further impaired by antisperm antibodies on the male partner's sperm. *Fertil Steril* 62, 826-33 (1994)
- 123. W. C. L. Ford, K. M. Williams, E. A. McLaughlin, S. Harrison, B. Ray & M. G. R. Hull: The indirect immunobead test for seminal antisperm antibodies and fertilization rates at in-vitro fertilization. *Hum Reprod* 11, 1418-22 (1996)

- 124. M. H. Vazquez-Levin, J. A. Notrica & E. P. de Fried: Male immunologic infertility: sperm performance on in vitro fertilization. *Fertil Steril* 68, 675-81 (1997)
- 125. S. L. Mandelbaum, M. P. Diamond & A. H. DeCherney: Relationship of antisperm antibodies to oocyte fertilization in in vitro fertilization-embryo transfer. *Fertil Steril* 47, 644-51 (1987)
- 126. N. Sukcharoen & J. Keith: The effect of the antisperm auto-antibody-bound sperm on in vitro fertilization outcome. *Andrologia* 27, 281-89 (1995)
- 127. P. J. Culligan, M. M. Crane, W. R. Boone, T. C. Allen, T. M. Price & K. L. Blauer: Validity and cost-effectiveness of antisperm antibody testing before in vitro fertilization. *Fertil Steril* 69, 894-98 (1998)
- 128. G. N. Clarke, A. Lopata, J. C. McBain, H. W. Baker & W. I. Johnston: Effect of sperm antibodies in males on human in vitro fertilization (IVF). *Am J Reprod Immunol Microbiol* 8, 62-66 (1985)
- 129. S. M. Junk, P. L. Matson, J. M. Yovich, B. Bootsma & J. L. Yovich: The fertilization of human oocytes by spermatozoa from men with antispermatozoal antibodies in semen. *J In Vitro Fert Embryo Transf* 3, 350-52 (1986)
- 130. M. De Almeida, I. Gazagne, C. Jeulin, M. Herry, J. Belaisch-Allart, R. Frydman, P. Jouannet & J. Testart: Invirto processing of sperm with auto-antibodies and in-vitro fertilization results. *Hum Reprod* 4, 49-53 (1989)
- 131. A. Lähteenmäki: In-vitro fertilization in the presence of antisperm antibodies detected by the mixed antiglobulin reaction (MAR) and the tray agglutination test (TAT). *Hum Reprod* 8, 84-88 (1993)
- 132. G. Palermo, P. Devroey, M. Camus, I. Khan, A. Wisanto & A. C. Van Steirteghem: Assisted procreation in the presence of a positive direct mixed antiglobulin reaction test. *Fertil Steril* 52, 645-49 (1989)
- 133. G. G. Haas, J. E. Sokoloski & D. P. Wolf: The interfering effect of human IgG antisperm antibodies on human sperm penetration of zona-free hamster eggs. *Am J Reprod Immunol* 1, 40-43 (1980)
- 134. J. Dor, E. Rudak & R. J. Aitken: Antisperm antibodies: their effect on the process of fertilization studied in vitro. *Fertil Steril* 35, 535-41 (1981)
- 135. N. J. Alexander: Antibodies to human spermatozoa impede sperm penetration of cervical mucus or hamster eggs. *Fertil Steril* 41, 433-39 (1984)
- 136. R. Bronson, G. Cooper & D. Rosenfeld: Ability of antibody-bound human sperm to penetrate zona-free hamster ova in vitro. *Fertil Steril* 36, 778-83 (1981)
- 137. R. A. Bronson, G. W. Cooper & D. M. Phillips: Effect of anti-sperm antibodies on human sperm ultrastructure and function. *Hum Reprod* 4, 653-57 (1989)
- 138. F. Francavilla, R. Romano & R. Santucci: Effect of sperm-antibodies on acrosome reaction of human sperm used for the hamster egg penetration assay. *Am J Reprod Immunol* 25, 77-80 (1991)
- 139. R. Romano, R. Santucci, V. Marrone & F. Francavilla: Effect of ionophore challenge on hamster egg penetration

- and acrosome reaction of antibody-coated human sperm. Am J Reprod Immunol 29, 56-61 (1993)
- 140. G. N. Clarke: Sperm antibodies and human fertilization. *Am J Reprod Immunol Microbiol* 17, 65-70 (1988)
- 141. R. Zouari & M. De Almeida: Effect of spermassociated antibodies on human sperm ability to bind to zona pellucida and to penetrate zona-free hamster oocytes. *J Reprod Immunol* 24, 175-86 (1993)
- 142. G. G. Haas, M. Ausmanus, L. Culp, R. W. Tureck & L. Blasco: The effect of immunoglobulin occurring on human sperm in vivo on the human sperm/hamster ova penetration assay. *Am J Reprod Immunol Microbiol* 7, 109-12 (1985)
- 143. F. Francavilla, R. Romano, R. Santucci, V. Marrone, G. Properzi & G. Ruvolo: Occurrence of the interference of sperm-associated antibodies on sperm fertilizing ability as evaluated by the sperm-zona pellucida binding test and by the TEST-Yolk Buffer enhanced sperm penetration assay. *Am J Reprod Immunol* 37, 267-74 (1997)
- 144. M. C. Mahony, P. F. Blackmore, R. A. Bronson & N.J. Alexander: Inhibition of human sperm-zona pellucida tight binding in the presence of antisperm antibody positive polyclonal patient sera. *J Reprod Immunol* 19, 287-301 (1991)
- 145. R. Zouari, M. De Almeida & D. Feneux: Effect of sperm-associated antibodies on the dynamics of sperm movement and on the acrosome reaction of human spermatozoa. *J. Reprod Immunol* 22, 59-72 (1992)
- 146. R. Bandoh, S. Yamano, M. Kamada, T. Daitoh & T. Aono: Effect of sperm-immobilizing antibodies on the acrosome reaction of human spermatozoa. *Fertil Steril* 57, 387-92 (1992)
- 147. S. Benoff, G. W. Cooper, I. Hurley, F. S. Mandel & D. L. Rosenfeld: Antisperm antibody binding to human sperm inhibits capacitation induced changes in the levels of plasma membrane sterols. *Am J Reprod Immunol* 30, 113-30 (1993)
- 148. S. Harrison, G. Hull & S. Pillai: Sperm acrosome status and sperm antibodies in infertility. *J Urol* 159, 1554-58 (1998)
- 149. B. Lansford, G. G. Haas Jr., L. E. Debault & D.P. Wolf: Effect of sperm-associated antibodies on the acrosomal status of human sperm. *J Androl* 11, 532-38 (1990)
- 150. D. P. Wolf: Acrosomal status quantitation in human sperm. *Am J Reprod Immunol* 20, 106-13 (1989)
- 151. J. M. Cummins, S. M. Pember, A. M. Jequier, J. L. Yovich & P. E. Hartmann: A test of the human sperm acrosome reaction following ionophore challenge. Relationship to fertility and other seminal parameters. *J Androl* 12, 98-103 (1991)
- 152. P. Fenichel, M. Donzeau, D. Farahifar, B. Basteris, N. Ayraud & B. L. Hsi: Dynamics of human sperm acrosome reaction: relation with in vitro fertilization. *Fertil Steril* 55, 994-99 (1991)

- 153. J. S. Pampiglione, S. L. Tan & S. Campbell: The use of the stimulated acrosome reaction test as a test of fertilizing ability in human spermatozoa. *Fertil Steril* 59, 1280-84 (1993)
- 154. W. Byrd, J. Tsu & D. P. Wolf: Kinetics of spontaneous and induced acrosomal loss in human sperm incubated under capacitating and noncapacitating conditions. *Gam Res* 22, 109-22 (1989)
- 155. R. A. Bronson, G.W. Cooper & D. L. Rosenfeld: Sperm-specific isoantibodies and autoantibodies inhibit the binding of human sperm to the human zona pellucida. *Fertil Steril* 38, 724-29 (1982)
- 156. H. Shibahara, L. J. Burkman, S. Isojima & N. J. Alexander: Effects of sperm-immobilizing antibodies on sperm-zona pellucida tight binding. *Fertil Steril* 60, 533-39 (1993)
- 157. H. Shibahara, M. Shigeta, M. Inoue, A. Hasegawa, K. Koyama, N.J. Alexander & S. Isojima,: Diversity of the blocking effects of antisperm antibodies on fertilization in human and mouse. *Human Reprod* 11, 2595- 99 (1996)
- 158. S. Tsukui, Y. Noda, J. Yano, A. Fukuda & T. Mori: Inhibition of sperm penetration through human zona pellucida by antisperm antibodies. *Fertil Steril* 46, 92-96 (1986)
- 159. D.Y. Liu, G. Clarke & H. W. G. Baker: Inhibition of human sperm-zona pellucida and sperm-oolemma binding by antisperm antibodies. *Fertil Steril* 1991; 55:440-44
- 160. F. Francavilla, R. Romano, R. Santucci, V. Marrone, G. Properzi & G. Ruvolo: Interference of antisperm antibodies with the induction of the acrosome reaction by zona pellucida (ZP) and its relationship with the inhibition of ZP binding. *Fertil Steril* 67, 1128-33 (1997)
- 161. P. M. Wasserman. The biology and chemistry of fertilization. *Science* 235, 553-60 (1987)
- 162. L. Leyton & P. Saling: Evidence that aggregation of mouse sperm receptors by ZP3 triggers the acrosome reaction. *J Cell Biol* 108, 2163-68 (1989)
- 163. Z. P. Nagy, G. Verheyen, J. Liu, H. Joris, C. Janssenswillen, A. Wisanto, P. Devroey & A. C. Steirteghem: Results of 55 intracytoplasmic sperm injection cycle in the treatment of male-immunological infertility. *Human Reprod* 10, 1775-80 (1995)
- 164. A. Lähteenmäki, I. Reima & O. Hovatta: Treatment of severe male immunological infertility by intracytoplasmic sperm injection. *Hum Reprod* 10, 2824-28 (1995)
- 165. N. G. Clarke, H. Bourne & H. W. G. Baler: Intracytoplasmic sperm injection for treating infertility associated with sperm autoimmunity. *Fertil Steril* 68, 112-17 (1997)
- 166. M. G. O'Rand: The presence of sperm-specific surface isoantigens on the egg following fertilization. *J. Exp Zool* 202, 267-73 (1977)
- 167. R. K. Naz: Effects of antisperm antibodies on early cleavage of fertilized ova. *Biol Reprod* 46, 130-39 (1992)
- 168. A. Lenzi, L. Gandini, F. Lombardo, S. Morrone & F. Dondero: Immunological usefulness of semen manipulation for artificial insemination homologous (AIH) in subjects

- with antisperm antibodies bound to sperm surface. Andrologia 20, 314-321 (1988)
- 169. G. G. Haas, O. J. D'Cruz & B. M. Denum: Effect of repeated washing on sperm-bound immunoglobulin G. J Androl 9, 190-196 (1988)
- 170. W. F. Hendry, J. Stedronska & L. Hughes: Steroid treatment of male subfertility caused by antisperm antibodies. *Lancet* 8, 498-500 (1979)
- 171. F. Dondero, A. Isidori, A. Lenzi, M. Cerasaro, F. Mazzilli, P. Giovenco & C. Conti: Treatment and follow-up of patients with infertility due to spermagglutinins. *Fertil Steril* 31, 48-51 (1979)
- 172. S. Shulman, B. Harlin, P. Davis & J. V. Reyniak: Immune infertility and new approaches to treatment. *Fertil Steril* 29, 309-13 (1978)
- 173. P. J. Turek & L. I. Lipshultz: Immunologic infertility. *Urol Clin North Am* 21, 447-68 (1994)
- 174. G. G. Haas & P. Manganiello: A double-blind, placebo-controlled study of the use of methylprednisolone in infertile men with sperm-associated immunoglobulins. *Fertil Steril* 47, 295-301 (1987)
- 175. W. F. Hendry: Bilateral aseptic necrosis of femoral heads following intermittent high-dose steroid therapy. *Fertil Steril* 38: 120-22 (1982)
- 176. W.F. Hendry, L. Hughes, G. Scammell, J. P. Pryor & T. B. Hargreave: Comparison of prednisolone and placebo in subfertile men with antibodies to spermatozoa. *Lancet* 335, 85-88 (1990)
- 177. M. Bals-Pratsch, M. Doren, B. Karbowski, H. P. G. Schneider & E. Nieschlag: Cyclic corticosteroid immunosuppression is unsuccessful in the treatment of sperm antibody-related male infertily: a controlled study. *Human Reprod* 7, 99-104 (1992)
- 178. M. Rasanen, A. Lähteenmäki, Y. P. Agrawal, O. Hovatta: A placebo-controlled flow cytometric study of the effect of low-dose prednisolone treatment on sperm-bound antibody levels. *Int J Androl* 19, 150-56 (1996)
- 179. M. C. Wiltbank, T. S. Kosasa & B. J. Rogers: Treatment of infertile patients by intrauterine insemination of washed spermatozoa. *Andrologia* 17, 22-30 (1985)
- 180. E. Confino, J. Friberg, A. B. Dudkiewicz & N. Gleicher: Intrauterine inseminations with washed human spermatozoa. *Fertil Steril* 46, 55-60 (1986)
- 181. C. M. A. Glazener, C. Coulson, P. A. Lambert, E. M. Watt, R. A. Hinton & N. J. Kelly: The value of artificial insemination with husband's semen in infertility due to failure of postcoital sperm-mucus penetration- controlled trial of treatment. *Br J Obstet Gynaecol* 94, 774-78 (1987)
- 182. E. R. Velde, R. J. Kooj & J. J. H. Waterreus: Intrauterine insemination of washed human spermatozoa: a controlled study. *Fertil Steril* 51, 182-85 (1989)
- 183. S. L. Carson, F. R. Batzer, B. Gocial & G. Maislin: Intrauterine insemination and ovulation stimulation as treatment of infertility. *J Reprod Med* 34, 397-406 (1989)
- 184. F. Francavilla, R. Romano, R. Santucci, V. Marrone & G. Corrao: Failure of intrauterine insemination in male

- immunological infertility in cases in which all spermatozoa are antibody-coated. Fertil Steril 58, 587-91, 1992
- 185. F. Francavilla, R. Romano, R. Santucci & G. Poccia: Effect of sperm morphology and motile sperm count on outcome of intrauterine insemination in oligozoospermia and/or asthenozoospermia. *Fertil Steril* 53, 892-97 (1990)
- 186. A. Lahteenmaki, J. Veilati & O. Howaatta: Intrauterine insemination versus cyclic, low dose prednisolone in couples with male antisperm antibodies. *Hum Reprod* 10, 142-47 (1995)
- 187. J. N. Robinson, R. G. Forman, S. C. Nicholson, L. R. Maciocia & D. H. Barlow: A comparison of intrauterine insemination in superovulated cycles to intercourse in couples where the male is receiving steroids for the treatment of autoimmune infertility. *Fertil Steril* 63, 1260-66 (1995)
- 188. W. Ombelet, H. Vandeput, M. Janssen, A. Cox, C. Vossen, H. Pollet, O. Steeno & E. Bosmans: Treatment of male infertility due to sperm surface antibodies: IUI or IVF?. *Hum Reprod* 12, 1165-70 (1997)
- 189. W.J. Snell & J. M. White: The molecules of mammalian fertilization. *Cell* 85, 629-37 (1996)
- 190. R. K. Naz: Involvement of fertilization antigen (FA-1) in involuntary immunoinfertility in humans. *J Clin Invest* 80, 1375-83 (1987)
- 191. R. K. Naz, J. Deutsch, T. M. Phillips, A. C. Menge & Fisch H: Sperm antibodies in vasectomized men and their effects om fertilization. *Biol Reprod* 41, 163-73 (1989)
- 192. A. C. Menge & R. K. Naz: Immunoglobulin (Ig) G, IgA, and IgA subclass antibodies against fertilization antigen-1 in cervical secretions and sera of women of infertile couples. *Fertil Steril* 60, 658-63 (1993)
- 193. T. Hjort & P. D. Griffin: The identification of candidate antigens for the development of birth control vaccines. *J. Reprod Immunol* 8, 271-78 (1985)
- 194. A. B. Diekman & J. C. Herr: Sperm antigens and their use in the development of an immunocontraceptive. *Am J Reprod Immunol* 37, 111-17 (1997)
- 195. C. Y. G. Lee, V. Lum, E. Wong, A. Menge & Y. S. Huang: Identification of human sperm antigens to antisperm antibodies. *Am J Reprod Immunol* 3, 183-87 (1983)
- 196. D. Lehmann, B. Temminck, D. D. Rugna, B. Leibundgut & H. Muller: Blot-immunoblotting test for the detection of anti-sperm antibodies. *J Reprod Immunol* 8, 329-36 (1985)
- 197. S. Naaby-Hamsen & O. J. Bjerrum: Auto- and isoantigens of human spermatozoa detected by immunoblotting with human sera. *J Reprod Immunol* 7, 41-57 (1985)
- 198. F. Saji, K. Ohashi, T. Negoro & O. Tanizawa: Identification of human sperm antigen corrisponding to sperm-immobilizing antibodies. *Am J Reprod Immunol* 17, 128-33 (1988)
- 199. P. Primakoff, W. Lathrop & R. Bronson: Identification of human sperm surface glycoproteins recognized by

- autoantisera fro immune infertile men, women, and vasectomized men. *Biol Reprod* 42, 929-42 (1990)
- 200. S. Shai & Y. Noat: Identification of human sperm antigens reacting with antisperm antibodies from sera and genital tract secretions. *Fertil Steril* 58, 593-98 (1992)
- 201. K. Snow & G. D. Ball: Characterization of human sperm antigens and antisperm antibodies in infertile patients. *Fertil Steril* 58, 1011-19 (1992)
- 202. O. J. D'Cruz, G.G. Haas & H. Lambert: Heterogeneity of human sperm surface antigens identified by indirect imunoprecipitation of antisperm antibody bound to biotinylated sperm. *J. Immunol* 151, 1062-74 (1993)
- 203. E. W. Wingate, R. T. Patrick & S. Mathur: Antigens in capacitated spermatozoa eliciting autoimmune responses. *J. Urol* 149, 1331-37 (1993)
- 204. R. K. Naz, E. Gateva & C. Morte: Autoantigenicity of human sperm: molecular identities of sperm antigens recognized by sera of immunoinfertile men in the immunoprecipitation procedure. *Arch Androl* 35, 225-31 (1995)
- 205. J. Auer, I. Pignot-Paintrad & M. De Almeida: Identification of human sperm surface glycoproteins by sperm membrane-specific autoantibodies. *Hum Reprod* 10, 551-57 (1995)
- 206. R. Paradisi, E. Bellavia, A. Pession, S. Venturoli & C. Flamigni: Characterization of human sperm antigens reacting with sperm antibodies from autologous serum and seminal plasma in an infertile population. *Biol Reprod* 55, 54-61 (1996)
- 207. S. Naaby-Hansen, C. J. Flinckinger & J. C. Herr: Two-dimensional gel electrophoretic analysis of vectorially labeled surface proteins of human spermatozoa. *Biol Reprod* 56, 771-77 (1997)
- 208. J. C. Herr: Update on the center for recombinant gamete contraceptive vaccinogens. *Am J Reprod Immunol* 35, 184-89 (1996)
- **Key words:** Sperm Antibodies; Male Infertility; Fertilization; Spermatozoa Immunology; Semen Analysis

Send corespondence to: Professor Felice Francavilla, Dipartimento di Medicina Interna, Università de L'Aquila, Via S.Sisto 67100 L'Aquila, Italy, Tel: 0862.432854, Fax: 0862.432858, E-mail: francavillaf@yaxaq.cc.univaq.it

Received 9/15/98 Accepted 12/14/98