

Original Research

Increased Myeloperoxidase Concentrations in Saliva could Reflect Increased Body Mass and Oral Microinflammation

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Abstract

Background: Increased myeloperoxidase (MPO) levels in saliva are thought to reflect ongoing periodontal inflammation. Less clear is whether and to what extent salivary MPO is increased as a result of systemic inflammation. **Methods**: In the present study, we aimed to determine which demographic, anthropometric, biochemical, and dental parameters affect the level of MPO in whole mixed saliva in healthy adults with no apparent inflammatory lesions in the oral cavity. Thus, 113 individuals, aged 20–61 years (including 30.1% men and 23.9% smokers), were examined. **Results**: In the univariate analysis, higher levels of MPO in saliva were found to be associated with age, an increased body mass index (BMI), higher levels of cytokines tumour necrosis factor- α and interleukin-6, as well as poorer oral hygiene, gingival status, and lower saliva flow. Multivariate logistic regression analysis determined that the main predictors of MPO concentration in saliva were BMI and stimulated saliva flow rate. **Conclusions**: Overall, an increase in MPO in saliva could be related to an increase in BMI, possibly as a result of subclinical chronic microinflammation, which also involves the gingiva.

Keywords: myeloperoxidase; saliva; body mass index; inflammation; oral health; obesity

1. Introduction

Myeloperoxidase (MPO) is the most abundant proinflammatory enzyme stored in the azurophilic granules of neutrophils, accounting for 5% of their dry mass. It is also present to a lesser degree in the lysosomes of monocytes. MPO is a component of the intracellular microbicidal system of phagocytes and part of innate immunity [1,2]. MPO is a heme enzyme that is released during phagocytosis, which uses the superoxide (O_2^-) and hydrogen peroxide (H₂O₂) generated by the neutrophil oxidative burst (NADPH-oxidase) to produce highly reactive hypochlorous acid (HOCl) and other reactive oxidants (hypobromous, hypothiocyanous acids). Unlike other peroxidases, MPO has a chlorinating activity (hypochlorous acid production) in addition to its regular peroxidative activity, which means it can use hydrogen peroxide and superoxide to oxidize numerous biological compounds [3,4].

Saliva is a valuable yet complex diagnostic medium, as it is a mix of salivary gland secretions, serum transudate from gingival crevices, oral mucosa cells, and oral microorganisms [5]. Neutrophil-derived MPO constitutes a significant proportion of the total salivary peroxidase activity [6], while the number of neutrophils that migrate to the oral cavity determines the MPO activity in saliva. Due to its potentially defensive role, MPO is found to be increased in conditions associated with oxidative stress and inflammation [7,8]. Indeed, high salivary MPO levels have been reported in patients with oral inflammation [9]. However, much less

is known about the impact of systemic conditions and medication on salivary MPO. Some studies have reported that MPO could be a mediator of inflammatory processes, promoting tissue damage and contributing to the pathogenesis of diseases, such as cardiovascular disease, rheumatoid arthritis, and neurodegenerative disorders [10–12]. In this respect, Polizzi et al. [13] detected increased levels of MPO in saliva from patients with coronary artery disease. We have recently observed that, compared to healthy individuals, patients with ulcerative colitis that are not responding to combined immunosuppression treatment had significantly decreased salivary MPO levels [14], which subsequently increased when patients responded satisfactorily to biologic therapy [15]. However, changes in the salivary MPO levels from either oral or systemic factors have not currently been investigated in healthy subjects.

Thus, in the present study, we aimed to determine how oral health status and anthropometric factors affect salivary MPO levels in apparently healthy adults. Additionally, we aimed to assess whether biochemical alterations in saliva or demographic factors could be confounders in these relationships.

2. Materials and Methods

2.1 Study Group

The study group included 113 randomly selected adult patients (aged 20–61 years, median 32 years; 30.1% men) who presented for a routine dental examination in the De-



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Fig. 1. Comparison of MPO concentrations between (A) men (n = 34) and women (n = 79), between (B) smokers (n = 27) and non-smokers (n = 86), and between (C) underweight/normal weight (n = 50) and overweight/obese (n = 63) patients. The data were analysed using Welch's *t*-test (A,B) and the Mann–Whitney test (C) and presented as violin plots with medians and quartiles indicated. MPO, myeloperoxidase; BMI, body mass index.

partment of Conservative Dentistry and Endodontics at Poznan University of Medical Sciences. All patients provided informed consent to donate saliva for research purposes and were in generally good health, although a few patients had previously received therapy for hypertension (n = 11) and hypothyroidism (n = 7). The exclusion criteria included the presence of another systemic disease (e.g., diabetes mellitus), active oral diseases (e.g., periodontal disease), acute illnesses or infections, administering of drugs or treatment affecting salivary flow, and pregnancy. Smoking was declared by 27 patients.

2.2 Clinical Examination

The examinations of the patients were performed by routine methods and included the parameters of oral hygiene (approximal plaque index: API; plaque index: PII) and periodontal status (gingival index: GI; sulcus bleeding index: SBI; periodontal probing depth: PPD). The cleaning index was calculated as the product of self-reported brushing frequency and brushing time [16]. None of the included patients were found to have overt inflammatory lesions in the oral cavity.

All patients were measured and weighted to determine their body mass index (BMI).

2.3 Saliva Collection and Analysis

Unstimulated whole mixed saliva was collected over a period of 20 minutes and processed, as previously described in detail [17]. Salivary MPO concentrations were measured using the DuoSet Immunoassay (R&D Systems, Bio-Techne; Minneapolis, MN, USA). Moreover, two proinflammatory cytokines and two antioxidants were selected as the markers to reference any changes in body weight and oral microinflammation. The levels of interleukin-6 (IL-6) and tumour necrosis factor- α (TNF α) were measured using the Quantikine High Sensitivity kits (R&D Systems, Bio-Techne; Minneapolis, MN, USA). The uric acid concentration and superoxide dismutase activity were measured by assay kits from Cayman Chemical (Ann Arbor, MI, USA). All assays were performed as per the manufacturers' instructions.

2.4 Statistical Analysis

Statistical analysis was performed using Statistica 13.3 (StatSoft, Cracow, Poland) and GraphPad Prism 9.4.1 (GraphPad Software, San Diego, CA, USA). Continuous data were analysed for normality using the Shapiro–Wilk test. Data not normally distributed were analysed using non-parametric statistical tests, which are specified in each figure legend. To assess the relationships between MPO levels and other variables, Spearman's correlation coefficient analysis and logistic regression modelling, with V-fold cross-validation, were conducted. The significance level was set at $\alpha = 0.05$ for all analyses.

3. Results

The myeloperoxidase concentration in the saliva of the examined individuals ranged from 3 to 720 ng/mL (median of 297 ng/mL). No differences in salivary MPO concentrations were found between either men and women (Fig. 1A) or between smokers and non-smokers (Fig. 1B). Therefore, all patients were grouped, and all subsequent analyses were performed on all patients as a single group. In this group, the median BMI was 27.5 kg/m², while patients with a BMI above 25 kg/m² had significantly higher MPO levels

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Parameters	Median	01-03	MPO, ng/mL						
		Q1-Q3	R _s	<i>p</i> -value					
Oral health									
API, %	60.0	30.0-85.7	0.332	0.001*					
PlI	0.45	0.16-0.85	0.344	< 0.001*					
SBI, %	7.7	0.0-19.2	0.215	0.040*					
GI	0.23	0.00-0.75	0.272	0.009*					
PPD, mm	0.90	0.69-1.18	0.149	0.159					
Cleaning index	6	4-8	-0.163	0.134					
Salivary antioxidants									
SOD, U/mL	8.9	7.9–9.3	0.008	0.937					
Uric acid, µmol/L	182.1	120.7-246.9	-0.040	0.675					
Salivary pro-inflammatory cytokines									
IL-6, pg/mL	3.2	1.0-8.0	0.256	0.008*					
TNF, pg/mL	1.3	0.5–2.3	0.227	0.017*					
Salivation									
Saliva flow rate, mL/min	0.32	0.25 - 0.50	-0.272	0.005*					
Stimulated saliva flow rate, mL/min	1.25	0.75 - 1.75	-0.309	0.001*					
pH of unstimulated saliva	7.0	6.7–7.2	-0.175	0.078					
pH of stimulated saliva	7.4	7.0–7.7	0.006	0.950					

 Table 1. Clinical and laboratory parameters: descriptive statistics and Spearman's correlation coefficients (Rs) with salivary

 myeloperoxidase levels.

Q1–Q3, interquartile range; API, approximal plaque index; PII, plaque index; SBI, sulcus bleeding index; GI, gingival index; PPD, periodontal probing depth; SOD, superoxide dismutase; IL-6, interleukin 6; TNF, tumour necrosis factor; *, significant Spearman's correlation coefficient.

(Fig. 1C). Table 1 shows the recorded clinical and laboratory values. Then, these parameters were correlated with salivary MPO levels (Table 1).

3.1 Correlation Analysis

First, we observed that there was a moderate, positive correlation between salivary MPO levels and BMI ($R_{Spearman} = 0.397$, *p*-value < 0.001) and a weak correlation with age ($R_{Spearman} = 0.186$, *p*-value = 0.049). Moreover, predictably, there was a positive correlation between the patient's age and their BMI ($R_{Spearman} = 0.398$, *p*-value < 0.001).

As previously mentioned, clinical examinations revealed that the patients had generally good oral health, with a fairly good level of oral hygiene, and no evidence of periodontal inflammation. Nevertheless, there was a weak-tomoderate correlation between MPO levels and the parameters reflecting poorer oral hygiene (API and PII) and gingival status (GI and SBI). Interestingly, none of the other biochemical parameters correlated significantly with any indices of oral hygiene and gingival conditions, such as MPO in saliva (data not presented).

Moreover, MPO levels correlated positively with the levels of the two proinflammatory cytokines (IL-6 and $\text{TNF}\alpha$) in saliva. Of the salivary parameters of oxidative stress, MPO concentrations did not correlate with concentrations of superoxide dismutase, and uric acid.

Finally, the MPO levels showed an inverse correlation with both basal and stimulated salivation. Moreover, similar to the correlations with the dental indices, significant correlations were observed relating to saliva flow rates and MPO, although no correlations were noted for the other biochemical parameters (data not presented). Likewise, no correlation was found between MPO concentration and the pH of the saliva.

3.2 Logistic Regression Modelling

Since MPO levels significantly correlated with many potentially related parameters, logistic regression analysis was performed to clarify the nature of these relationships. Thus, the study group was divided into two subgroups according to the median MPO concentration (297 ng/mL), and all the variables were subjected to logistic regression analysis.

Table 2 shows the univariate analysis indicated that several of these variables were significant predictors of higher MPO, with BMI presenting the strongest correlation. Moreover, an increase in the levels of all predictors, except for stimulated salivation, was associated with higher salivary MPO concentrations. Furthermore, as shown in Fig. 2, many of the identified parameters showed levels of correlation, with oral hygiene and gingival indices presenting strong correlations, and moderately significant correlations between BMI and these indices. Again, the stimulated saliva flow rate presented inverse correlations.

Table 2. Results for the logistic regression modelling: univariate and multivariate models.

	0	0	0						
Parameter	β	Standard error	<i>p</i> -value	OR	-95% CI	+95% CI			
Univariate logistic regression models – statistically significant predictors									
BMI, kg/m ²	0.083	0.024	< 0.001	1.086	1.036	1.139			
API, %	0.014	0.007	0.039	1.015	1.001	1.029			
PlI	1.194	0.486	0.014	3.300	1.273	8.553			
SBI, %	0.025	0.011	0.025	1.025	1.003	1.048			
GI	1.250	0.547	0.022	3.492	1.196	10.197			
IL-6, pg/mL	0.058	0.025	0.018	1.060	1.010	1.112			
Stimulated SFR, mL/min	-0.667	0.284	0.019	0.513	0.294	0.895			
Multivariate logistic regression model – forward stepwise method									
Intercept	-1.682	0.887	0.058	0.186	0.033	1.057			
BMI, kg/m ²	0.085	0.026	0.001	1.089	1.035	1.146			
Stimulated SFR, mL/min	-0.555	0.298	0.062	0.574	0.320	1.029			

OR, odds ratio; CI, confidence interval; BMI, body mass index; API, approximal plaque index; PII,

plaque index; SBI, sulcus bleeding index; GI, gingival index; IL-6, interleukin 6; SFR, saliva flow rate.

Thus, next, a multivariate model of forward stepwise logistic regression was constructed. The calculated odds ratios indicated that an increased BMI and a lower stimulated saliva flow rate offered the best predictive marker values for high salivary MPO levels (Table 2).

The model was validated by the V-fold cross-validation method with the presented receiver operating characteristic (ROC) curves. The high area under the curve (AUC) values for the training and testing curves confirmed that the model provided good quality data (0.744 \pm 0.050 and 0.708 \pm 0.052, respectively).



Fig. 2. Correlation matrix between the shown parameters. The numbers represent Spearman's correlation coefficients (R). BMI, body mass index; API, approximal plaque index; PII, plaque index; SBI, sulcus bleeding index; GI, gingival index; SFR, saliva flow rate.

4. Discussion

The main observation in the present study was that the salivary MPO levels in apparently healthy adults predominantly correlate to their BMI. While this association does not imply causality, one may envisage a number of mechanisms through which an increased body mass can contribute toward an increase in saliva MPO levels.

Firstly, a higher BMI is associated with inferior gingival status, poorer oral hygiene, and increased levels of proinflammatory cytokines in the saliva. Although the impairment of oral health from the observed parameters appeared to be minimal in the population, the primary cause of these associations remains unknown; however, the resulting chronic microinflammation may lead to an increase in neutrophil migration to the oral cavity, their lysis in a hypotonic environment, and MPO release [18].

Secondly, the level of salivary MPO may be partly related to the level of systemic MPO, which has been reported to increase in accordance with obesity [19]. To the best of our knowledge, this represents the first study to clearly identify that the MPO levels found in saliva correlate to BMI in healthy subjects.

Previous studies on MPO and obesity only analysed the MPO levels found in the blood. For example, Qaddoumi *et al.* [20] found that plasma MPO concentrations were elevated in obese patients, both with and without diabetes, compared to non-obese subjects. These MPO levels correlated clearly with both BMI and inflammatory markers, such as CRP, TNF α , and IL-6. The authors speculated that MPO could regulate obesity independently of diabetes.

Moreover, Choromańska *et al.* [21] found that serum MPO concentrations were significantly elevated and could be used to differentiate obese patients with metabolic syndrome from those only suffering from obesity. These findings showed that MPO activity rose in parallel with the progression of metabolic disturbances in patients with obesity. Additionally, Sparks *et al.* [22] observed that a twelveweek aerobic exercise intervention promoted a reduction in BMI and a decrease in plasma MPO concentration.

Only a few previous studies have measured the level of MPO in saliva, and these have been performed primarily in the context of periodontium inflammation. In a threeweek experimental study by Nascimento *et al.* [23], the rapid development of gingival inflammation was associated with an increase in salivary MPO.

Klangprapan *et al.* [24] observed that compared to healthy controls, salivary MPO was significantly increased in patients with periodontitis, and to a lesser extent, in those with gingivitis. Nizam *et al.* [25] confirmed a close relationship between MPO levels in the saliva and periodontal indices of patients with generalised periodontitis.

Similarly, Lahdentausta *et al.* [26] showed that using salivary MPO concentrations could clearly distinguish patients with periodontitis from healthy subjects. Interestingly, Polizzi *et al.* [13] found that salivary MPO levels were higher in patients with periodontitis and coronary heart disease than in those solely with periodontitis. Based on the multivariate regression analysis, the authors concluded that plasma C-reactive protein and total cholesterol levels were the best predictors of salivary MPO levels in these patients.

Meschiari *et al.* [27] observed that salivary MPO activity was significantly higher in patients with periodontitis and decreased only marginally following three months of non-surgical therapy.

In contrast, Hernández *et al.* [28] found a significant decrease in MPO activity in the gingival crevicular fluid (GCF) after two months of treatment for chronic periodontitis. Over *et al.* [29] found that the highest MPO concentrations in patients with rapidly progressive periodontal disease were in the saliva and GCF. Moreover, Saloom *et al.* [30] observed an increase in GCF secretion during orthodontic treatment and a simultaneous increase in MPO levels, which were associated with a greater rate of tooth movements, especially in obese adolescents.

The strengths of the current study are that a crosssectional character was performed to investigate the potential relationships between local and systemic parameters and the antioxidant status in healthy adult subjects, while such data were determined in a non-invasive manner. Despite an absence in previously justifying the sample size used, this study determined the correlations and regressions with a high testing power. However, the limitations of the study include the relatively good periodontal condition of the included patients, who reported for regular routine oral examinations; however, this fact excluded the potential confounding of poor oral conditions on systemic dependencies. Thus, in order to investigate further relationships, such as considering body mass and age as concurrent confounders, the size of the subgroups should be increased to provide these analyses with higher statistical powers.

Moreover, additional studies should be performed to consider the correlations between salivary MPO levels and oral, demographic, and anthropometric factors and how these relate to potential alterations in serum MPO levels. Currently, salivary markers are a promising branch of laboratory diagnostics and offer the possibility of determining reference ranges in the future, since these have not yet been determined and large populations are required for testing.

5. Conclusions

Salivary levels of myeloperoxidase in healthy adults with no clinically detected signs of inflammation in the oral cavity correlated predominantly with the patient's body mass index. Additionally, a higher body mass index was also associated with reduced oral hygiene, lower gingival status, and less salivation. Therefore, these factors should be considered when interpreting increased salivary MPO concentrations.

Availability of Data and Materials

Data are available on request from the corresponding author.

Author Contributions

KN is responsible for conceptualisation, methodology, formal analysis, investigation and resources, writing the original draft, review and editing, visualisation. AL is responsible for investigation and resources. RR is responsible for investigation and resources. KK is responsible for conceptualisation and methodology. JW is responsible for conceptualisation, methodology, writing review and editing, visualisation and supervision. AS is responsible for conceptualisation, writing review and editing and supervision. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript.

Ethics Approval and Consent to Participate

The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Bioethics Committee of Poznan University of Medical Sciences (189/14). Informed consent was obtained from all subjects involved in the study.

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Conflict of Interest

Given the role as Guest Editor, Kacper Nijakowski had no involvement in the peer-review of this article and has no access to information regarding its peer-review. Full responsibility for the editorial process for this article was delegated to Pier Paolo Piccaluga.

References

- Arnhold J. The Dual Role of Myeloperoxidase in Immune Response. International Journal of Molecular Sciences. 2020; 21: 8057.
- [2] Davies MJ. Myeloperoxidase-derived oxidation: mechanisms of biological damage and its prevention. Journal of Clinical Biochemistry and Nutrition. 2011; 48: 8–19.
- [3] Aratani Y. Myeloperoxidase: its role for host defense, inflammation, and neutrophil function. Archives of Biochemistry and Biophysics. 2018; 640: 47–52.
- [4] Ashby MT. Inorganic Chemistry of Defensive Peroxidases in the Human Oral Cavity. Journal of Dental Research. 2008; 87: 900– 914.
- [5] Nijakowski K, Surdacka A. Salivary Biomarkers for Diagnosis of Inflammatory Bowel Diseases: A Systematic Review. International Journal of Molecular Sciences. 2020; 21: 7477.
- [6] Thomas EL, Jefferson MM, Joyner RE, Cook GS, King CC. Leukocyte Myeloperoxidase and Salivary Lactoperoxidase: Identification and Quantitation in Human Mixed Saliva. Journal of Dental Research. 1994; 73: 544–555.
- [7] van der Veen BS, de Winther MPJ, Heeringa P. Myeloperoxidase: molecular mechanisms of action and their relevance to human health and disease. Antioxidants & Redox Signaling. 2009; 11: 2899–2937.
- [8] Ndrepepa G. Myeloperoxidase a bridge linking inflammation and oxidative stress with cardiovascular disease. Clinica Chimica Acta. 2019; 493: 36–51.
- [9] Iannitti T, Rottigni V, Palmieri B. Role of free radicals and antioxidant defences in oral cavity-related pathologies. Journal of Oral Pathology & Medicine. 2012; 41: 649–661.
- [10] Stamp LK, Khalilova I, Tarr JM, Senthilmohan R, Turner R, Haigh RC, *et al.* Myeloperoxidase and oxidative stress in rheumatoid arthritis. Rheumatology. 2012; 51: 1796–1803.
- [11] Ramachandra CJA, Ja KPMM, Chua J, Cong S, Shim W, Hausenloy DJ. Myeloperoxidase as a Multifaceted Target for Cardiovascular Protection. Antioxidants and Redox Signaling. 2020; 32: 1135–1149.
- [12] Green PS, Mendez AJ, Jacob JS, Crowley JR, Growdon W, Hyman BT, *et al.* Neuronal expression of myeloperoxidase is increased in Alzheimer's disease. Journal of Neurochemistry. 2004; 90: 724–733.
- [13] Polizzi A, Torrisi S, Santonocito S, Stefano MD, Indelicato F, Giudice AL. Influence of myeloperoxidase levels on periodontal disease: An applied clinical study. Applied Sciences. 2020; 10: 1037.
- [14] Nijakowski K, Rutkowski R, Eder P, Simon M, Korybalska K, Witowski J, et al. Potential Salivary Markers for Differential Diagnosis of Crohn's Disease and Ulcerative Colitis. Life. 2021; 11: 943.
- [15] Nijakowski K, Rutkowski R, Eder P, Korybalska K, Witowski J, Surdacka A. Changes in Salivary Parameters of Oral Immunity after Biologic Therapy for Inflammatory Bowel Disease. Life. 2021; 11: 1409.
- [16] Nijakowski K, Lehmann A, Rutkowski R, Korybalska K, Witowski J, Surdacka A. Poor Oral Hygiene and High Levels of Inflammatory Cytokines in Saliva Predict the Risk of Overweight and Obesity. International Journal of Environmental Research and Public Health. 2020; 17: 6310.
- [17] Surdacka A, Ciężka E, Pioruńska-Stolzmann M, Wender-Ożegowska E, Korybalska K, Kawka E, et al. Relation of sali-

vary antioxidant status and cytokine levels to clinical parameters of oral health in pregnant women with diabetes. Archives of Oral Biology. 2011; 56: 428–436.

- [18] Rijkschroeff P, Loos BG, Nicu EA. Oral Polymorphonuclear Neutrophil Contributes to Oral Health. Current Oral Health Reports. 2018; 5: 211–220.
- [19] Nijhuis J, Rensen SS, Slaats Y, van Dielen FMH, Buurman WA, Greve JWM. Neutrophil Activation in Morbid Obesity, Chronic Activation of Acute Inflammation. Obesity. 2009; 17: 2014– 2018.
- [20] Qaddoumi MG, Alanbaei M, Hammad MM, Al Khairi I, Cherian P, Channanath A, *et al.* Investigating the Role of Myeloperoxidase and Angiopoietin-like Protein 6 in Obesity and Diabetes. Scientific Reports. 2020; 10: 6170.
- [21] Choromańska B, Myśliwiec P, Łuba M, Wojskowicz P, Myśliwiec H, Choromańska K, *et al.* The Impact of Hypertension and Metabolic Syndrome on Nitrosative Stress and Glutathione Metabolism in Patients with Morbid Obesity. Oxidative Medicine and Cellular Longevity. 2020; 2020: 1057570.
- [22] Sparks JR, Sarzynski MA, Davis JM, Grandjean PW, Wang X. Alterations in Glycemic Variability, Vascular Health, and Oxidative Stress following a 12-Week Aerobic Exercise Intervention-A Pilot Study. International Journal of Exercise Science. 2021; 14: 1334–1353.
- [23] Nascimento GG, Baelum V, Sorsa T, Tervahartiala T, Skottrup PD, López R. Salivary levels of MPO, MMP-8 and TIMP-1 are associated with gingival inflammation response patterns during experimental gingivitis. Cytokine. 2019; 115: 135–141.
- [24] Klangprapan S, Chaiyarit P, Hormdee D, Kampichai A, Khampitak T, Daduang J, *et al.* Salivary Myeloperoxidase, Assessed by 3,3'-Diaminobenzidine Colorimetry, can Differentiate Periodontal Patients from Nonperiodontal Subjects. Enzyme Research. 2016; 2016: 7517928.
- [25] Nizam N, Gümüş P, Pitkänen J, Tervahartiala T, Sorsa T, Buduneli N. Serum and Salivary Matrix Metalloproteinases, Neutrophil Elastase, Myeloperoxidase in Patients with Chronic or Aggressive Periodontitis. Inflammation. 2014; 37: 1771– 1778.
- [26] Lahdentausta L, Paju S, Mäntylä P, Buhlin K, Pietiäinen M, Tervahartiala T, *et al.* Smoking confounds the periodontal diagnostics using saliva biomarkers. Journal of Periodontology. 2019; 90: 475–483.
- [27] Meschiari CA, Marcaccini AM, Santos Moura BC, Zuardi LR, Tanus-Santos JE, Gerlach RF. Salivary MMPs, TIMPs, and MPO levels in periodontal disease patients and controls. Clinica Chimica Acta. 2013; 421: 140–146.
- [28] Hernández M, Gamonal J, Tervahartiala T, Mäntylä P, Rivera O, Dezerega A, et al. Associations between Matrix Metalloproteinase-8 and -14 and Myeloperoxidase in Gingival Crevicular Fluid from Subjects with Progressive Chronic Periodontitis: a Longitudinal Study. Journal of Periodontology. 2010; 81: 1644– 1652.
- [29] Over C, Yamalik N, Yavuzyilmaz E, Ersoy F, Eratalay K. Myeloperoxidase activity in peripheral blood, neutrophil crevicular fluid and whole saliva of patients with periodontal disease. The Journal of Nihon University School of Dentistry. 1993; 35: 235–240.
- [30] Saloom HF, Papageorgiou SN, Carpenter GH, Cobourne MT. Impact of Obesity on Orthodontic Tooth Movement in Adolescents: a Prospective Clinical Cohort Study. Journal of Dental Research. 2017; 96: 547–554.