Molecular insights into kinase mediated signaling pathways of chemokines and their cognate G protein coupled receptors

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1. ABSTRACT

Chemokines are small regulatory proteins that play a crucial role in the coordinated migration of cell populations to the site of infection/inflammation by binding to their cognate receptors. In principle, chemokine receptors, which are serpentine G protein-coupled receptors (GPCRs), mediate the series of downstream intracellular signaling events that occur inside the cells to resolve the pathogenicity. Intracellular signaling pathways regulated by the kinase protein sub-families are the center of attention for chemokine derived functional responses. Kinases potentially influence cell migration, cell growth, transcriptional activation, and other essential molecular events. The regulation and flow of the signals by the kinases are different for each physiological and pathological event and are tightly regulated by the nature and pairing of chemokine(s) with its receptor(s). For example, phosphoinositide 3-kinase (PI3K) is activated during the initial steps of the chemokine induced signaling cascade to regulate chemotaxis, transcription, and cell survival. G protein-coupled

receptor kinase (GRKs) plays a crucial role in the desensitization and internalization chemokine receptors. The regulation of chemokine receptor is also governed by kinases like protein kinase A (PKA), protein kinase C (PKC), mitogenactivated protein kinases / extracellular signalregulated kinases (MAPK/ERK), etc. It was also established that tyrosine-protein kinases (TECs) such as ITK and RLK play a significant role in chemokine signaling in T lymphocytes. On a similar note, many others like janus kinases (JAKs), Protein kinase B (PKB), PKC, etc. are also studied in chemokine mediated disease models. The present review elucidates the role of different kinases involved in the chemokine/chemokine receptor mediated signaling cascade during various pathophysiological processes.

2. INTRODUCTION

Chemokines are small (8-10 kDa) signaling proteins that regulate cell trafficking. They play many important roles in multiple physiopathological

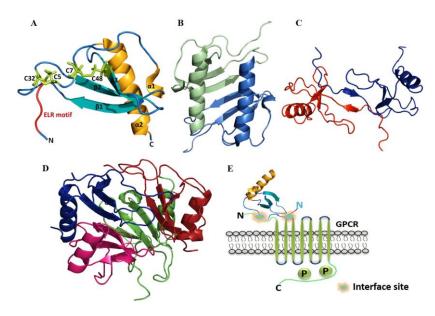


Figure 1. A) Structural components in the monomeric structure of ELR-CXC (IL8) chemokine (PDB ID: 5D14). B) Dimeric structure of Human CXCL8 (PDB ID: 1IL8). C) Dimeric structure of Human CCL2 (PDB ID: 1DOM). D) Tetrameric structure of CXCL7 (PDB ID: 1NAP). E) Schematic representation of the interaction between monomeric chemokine and GPCR.

activities like hematopoiesis, wound healing, organogenesis, inflammation, angiogenesis, tumorigenesis by regulated cell trafficking (1-5). Approximately 50 chemokines have been reported in humans to date. Depending upon the location of the cysteine residue at N-terminal, they are classified into two major (CXC and CC), and two minor (CX3C and C) families. Chemokines bind to cell surface glycosaminoglycans coupled and G-protein receptors (GPCRs) to perform their biological functions (5-8).

All the members of chemokine subgroups essentially form a similar monomeric structure. It comprises of a disordered N-terminal, followed by an extended N-loop, three-stranded βsheets forming the core of protein, and a helix at the C-terminal (Figure 1A) (9). However the oligomerization or dimerization of CXC and CC type chemokines vary significantly. In case of CXC chemokines such as CXCL8/IL8 (Interleukin-8), the intermolecular interactions between the Cterminal helix to the \beta-strand and the H-bonding between the β1-β1' sheets mediate dimerization (Figure 1B). Whereas, For CC chemokines such as CCL2, the extended N-loop forms the dimerization interface without involving

the C-terminal helix (Figure 1C). Indeed, chemokines such as CXCL7 forms higher order oligomers such as tetramers, where they use both the CC and CXC type dimer interfaces (Figure 1D).

G protein-coupled receptors (GPCRs) are the prevalent family of cell surface receptors which evolved to sense and communicate wide range of extracellular signals (10). GPCR based signaling is involved in various biochemical cascades such as immune, endocrine, cardiovascular, nervous, etc. Their existence on the plasma membrane makes GPCRs easily accessible to the drug molecules (agonist and antagonist) for the stimulation and blockade of the physiological processes. It is well manifested that defects in the receptor signaling triggered by genetic mutation or by overexpression of the receptors lead to various pathological conditions such as cancer, atherosclerosis, dwarfism, etc. (11, 12). The abundant presence of GPCRs on eukaryotic cells and its involvement in the human diseases accentuate the significance of studying the GPCRs. It is well elucidated that affinity of chemokines towards GPCR binding and activation significantly vary with their oligomerization (13). In general, the N-terminal domain of the chemokine binds to receptors at its N-terminal loop and first extracellular loop as shown in Figure 1E.

Binding of the chemokine to its cognate heptahelical GPCR induces a number of cellular responses like migration, adhesion, and different events of chemotaxis including changes in the shape of the cells, actin cytoskeleton reorganization, integrin expression, and activation, etc. (14, 15). Moreover, a particular chemokine can lead to a different response on the basis of the receptor to which it binds, and also the cells over which they are positioned (16). Chemokines and their GPCRs are promiscuous and can signal through multiple subtypes of G protein monomers, thus ensuring the regulated recruitment of the individual cells at a given period (7, 17). A wide range of proteins are involved in the synchronous administration of signaling pathways mediated by the chemokines and their receptors. Among them, chemokine activated intracellular "kinases" are the most crucial members (17, 18).

Kinases are the enzymes that catalyze the reaction in which the y-phosphate from adenosine triphosphate (ATP) is transferred to the substrate having a hydroxyl group. The catalytic fraction of the kinase is highly conserved and comprises of ATP binding sites. Kinases play a significant role in signal transductions, and regulate wide range of cellular like differentiation. transcriptional processes activation, apoptosis, metabolism, etc.(19). As protein phosphorylation plays a significant role in intracellular communication. abnormal phosphorylation leads to the generation of numerous diseases including neurodegenerative diseases, arthritis, rheumatoid immunodeficiency, cardiovascular diseases, endocrine disorders and cancer (20). The deregulated activity of kinase is an indicator of inflammation and cancer; thus targeting the activity of kinase enzyme has become a promising therapy for numerous diseases. The therapies comprise of kinase inhibitors that are being profiled in screening of kinome (21, 22).

Given the ubiquitous function of GPCRs, the activation, expression and administration of the GPCR based signaling are tightly regulated by the action of different kinases which include

phosphoinositide 3-kinase (PI3K), mitogen-activated kinases/extracellular signal-regulated protein kinases (MAPK/ERK), G protein-coupled receptor kinase (GRKs), janus kinases (JAKs) etc. The commencement of the chemokine mediated intracellular signal flow is fundamentally through the activation of inactive heterotrimeric G proteins (23). Once chemokine binds to the receptor, the stimulation facilitates the GDP/GTP exchange, which in turn activates many signaling molecules, secondary messengers, as well as phosphorylation reactions catalyzed by the intracellular kinases to regulate the flow of chemokine-mediated signal. Signaling cascade induced by chemoattractants and their cognate receptors comprises of multiple downstream pathways. Upon binding of chemokines to GPCRs, associated G proteins get activated, and are dissociated into the GTP-linked-Gα and Gβy subunits. Both these complexes are capable of activating different signaling cascades (Figure 2) (24). GTP-linked-Ga upon activation inhibits some isoforms of adenylyl cyclase, which in turn decrease the intracellular concentration of cAMP, and henceforth bring the reduction in the activity of cAMPdependent protein kinase. GBy subunit is significantly required for chemotaxis and plays crucial role in the activation of different signaling effectors such as phosphatidylinositol specific phospholipase (PLCβ2 and β3), and phosphatidylinositol-3-OH kinase (PI3Ky). PI3Ky catalyzes the rapid generation of phosphatidylinositol (3, 4, 5)-trisphosphate and also activates other kinases like protein kinase B (PKB/Akt) and MAPK (25, 26).

The regulated flow of kinase mediated multiple signaling pathways are essential for the recruitment of immune cells to generate immune responses. However, indecorous signaling pathways are implicated in the development and progression of a broad range of pathophysiological conditions. There are several diseases that are incidental with the inadequate activation of the chemokine mediated signaling cascade; especially those characterized by an excessive cellular infiltrate, such as rheumatoid arthritis and systemic lupus, other inflammatory diseases, viral and bacterial infections, autoimmune diseases and cancer etc. (4, 27, 28). The present review throws light on the kinase-mediated signaling pathways of chemokines and their cognate GPCRs

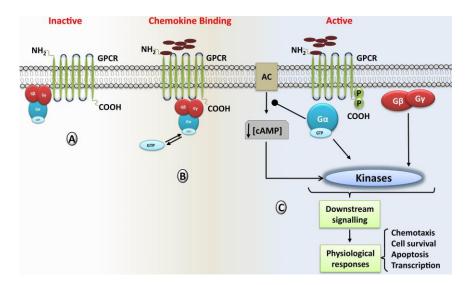


Figure 2. Chemokine induced Signaling cascade mediated by the G protein-coupled receptors (GPCRs); A) inactive stage in which chemokine is not associated with its cognate receptor, and G-protein is intact and inactive state (bound with GDP). B) After binding of chemokine/agonist to its cognate GPCR, it triggers the bound G protein complex leading to GTP-GDP exchange and dissociation of the heterotrimeric G protein complex into Gα and Gβγ subunits. Upon dissociation, GTP remains with the Gα subunit. C) Both the Gα and Gβγ subunit complex activates downstream signaling cascades that ultimately regulate the physiological and pathological response of the cells.

that are crucial in regulating various biological processes.

3. ROLE OF VARIOUS KINASES IN CHEMOKINE/CHEMOKINE RECEPTOR INDUCED SIGNALING EVENTS

3.1. Phosphoinositide 3-kinases (PI3Ks)

Phosphoinositide 3-kinases regulate multiple arrays of cellular responses such as mitogenesis, cell survival, glucose transport, membrane trafficking, superoxide production, membrane ruffling actin reorganization and chemotaxis (29, 30). Kinases of PI3K family phosphorylate the Phosphoinositide lipid (PI) at 3-OH position of the inositol head (31). Depending upon the substrate specificity and structural features, PI3K can be classified into three classes - class I, II and III. Class-I PI3Ks phosphorylate the phosphatidylinositol, phosphatidylinositol (4) phosphate, and phosphatidylinositol (4, 5) bisphosphate. There are four members in this class which can be further subdivided into IA and IB subclasses. The existence of these kinases in multiple isoforms provides an additional advantage

of multiple signal pathway regulation. Subclass IA PI3Ks comprises of three kinases, PI3Kα, PI3Kβ, and PI3Kδ (32). The subclass IB contains the single-member PI3Ky. Class-II PI3Ks utilize the phosphatidylinositol and phosphatidylinositol (4) phosphate as substrates. Class-III PI3Ks employ phosphatidylinositol as the substrate (33, 34). Enzyme phosphate and tensin homolog (PTEN) PI3K terminates the signaling of dephosphorylating the PI (3, 4, 5) P3 (35). Studies using PI3K inhibitors, mouse models with gene knockout experiments, and overexpression of mutated forms of PI3K established that PI3Ks assimilate and transmit signals from diverse surface molecules such as chemokine receptors, B cell receptors, and adhesion molecules. Binding ligands/signaling molecules (such chemokines) to their cognate GPCRs activates the disintegration of receptor associated G protein into Gα and Gβγ subunit. Both these subunits can activate PI3Ks independently. This results in generation of many secondary messengers and effector molecules like phosphatidylinositol (3,4,5) trisphosphate (PIP3) that in turn can activate other protein kinases such as serine/threonine kinases (AKT/PKB and PDK1) to facilitate the flow of signal (31, 36) (Figure 3).

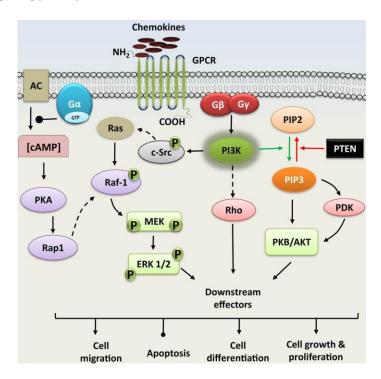


Figure 3. Phosphoinositide 3-kinase (PI3K) and MAPK signaling; PI3K pathway is activated upon agonist binding to receptor G protein coupled receptors (GPCRs). GPCRs can activate PI3Ks via G proteins, such as Gβγ. PI3K phosphorylates the phosphatidylinositol (3, 4)-bisphosphate (PIP2), generating phosphatidylinositol (3, 4, 5)-trisphosphate (PIP3) which recruits other kinases like serine/threonine kinase (PDK/AKT). MAPK signaling cascade is also induced by the Gβγ subunit of G protein that activates the other successive component proteins (cytosolic Src, MEK, and ERK, etc.). Binding of chemokines to G-protein coupled receptors (GPCRs) also activates Ga subunit. This activation hampers the activity of adenylyl cyclase (AC), which lowers the concentration of cAMP, that subsequently regulates the proteins PKA, Rap1, MEK1/2, and ERK1/2. Both these signaling pathways modulate cellular functions, including proliferation, gene expression, cytoskeletal rearrangement, anti-apoptosis, and degranulation.

PI3Ks play a crucial role in regulating the signaling cascade induced by the chemokines (33). PI3K catalyzed signaling pathways induced by chemokines play a consequential role in signal activation of innate and adaptive immunity (35, 37). Activation of PI3K is a rugged signaling event of leukocyte migration and is applicable to the majority of homeostatic and inflammatory chemokine receptors present on the leukocytes (38, 39). The primary evidence for the involvement of PI3Ks in the chemokine mediated signaling cascade was reported using cellular migration experiments (40). It was demonstrated that the CCL5 induced chemotaxis and T cell polarization can be modulated by inhibiting the PI3K. In addition to this, various research groups have shown that chemokines CCL20, CCL3, CXCL1, and CXCL12, etc., stimulate cellular migration via PI3Ks, thus proving the fact that amplification and degradation of PI3K are significant steps in regulating

the chemotactic pathways (41-43). Allergy and inflammatory diseases involve the recruitment of macrophages, dendritic cell, granulocytes, and mast cells. PI3Ks are crucial for the chemoattractant-mediated recruitment of these cells. In the murine model, it has been shown that mice devoid of the PI3Kγ have impeded migration towards chemokines at the site of inflammation (44). Impaired PI3K signaling causes severe defects in both innate and adaptive immunity that lead to different diseases like asthma, rheumatoid arthritis, and cancer (2, 37, 45). Many research groups have also demonstrated that, methods targeting PI3K and its different isoforms using inhibitors as a beneficial therapeutic strategy (Table 1).

Asthma is a common clinical condition that has the characteristics of airway hyperresponsiveness (AHR), lungs infiltration with

Table 1. Summary of phosphoinositide 3-kinases (PI3Ks) inhibitors that are currently in clinical trials (FDA approved)

Inhibitor Compound	Target PI3K	Disease Target	Status	Reference
GSK2269557	ΡΙ3Κδ	Asthma	Trial phase I	(46)
RV-1729	ΡΙ3Κδ/ΡΙ3Κγ	Asthma/COPD	Trial phase I	(47)
IPI-549	РΙЗКγ	Advanced solid tumors	Trial phase I	(48)
Dezapelisib	ΡΙ3Κδ	B-cell malignancies	Trial phase I	(49)
Umbralisib	ΡΙ3Κδ	CLL	Trial phase I	(50)
PI-3065	ΡΙ3Κδ	Advanced solid tumors	Trial phase I	(51)
SF1126	pan PI3K	Neuroblastoma	Trial phase I	(52)
Pilaralisib	pan PI3K	Breast cancer	Trial phase I	(53)
AZD8186	РІЗКβ	Breast cancer	Trial phase I	(54)
SAR260301	РІЗКβ	Solid tumor	Trial phase I	(55)
Acalisib	ΡΙ3Κβ/ ΡΙ3Κγ	Lymphoid Malignancies	Trial phase I	(56)
CL27c	pan PI3K	Asthma/ pulmonary fibrosis	Trial phase I	(57)
AS-604850	РΙЗКγ	Multiple sclerosis	Trial phase I	(58)
IPI-145	ΡΙ3Κδ/ ΡΙ3Κγ	Asthma	Trial phase II	(59)
TG100-115	ΡΙ3Κδ/ΡΙ3Κγ	Asthma/COPD	Trial phase II	(60)
Pictilisib	pan-PI3K	Advanced solid tumors	Trial phase II	(61)
PX-866	pan-PI3K	Prostate cancer	Trial phase II	(62)
GSK2636771	РІЗКβ	Breast cancer	Trial phase II	(63)
Copanlisib	pan-PI3K	Breast cancer	Trial phase II	(64)
Alpelisib	ΡΙ3Κα	Advanced solid tumors	Trial phase III	(65)
Taselisib	ΡΙ3Κα/ ΡΙ3Κδ/ ΡΙ3Κγ	Advanced solid tumors	Trial phase III	(66)
Buparlisib	pan-PI3K	Advanced solid tumors	Trial phase III	(67)
Duvelisib	ΡΙ3Κδ/ΡΙ3Κγ	CLL	FDA approved	(68)
Idelalisib	ΡΙ3Κδ	CLL	FDA approved	(69)
BEZ235	pan-PI3K	Solid tumors	Discontinued	(70)

inflammatory cells, wheezing, shortness of breath and coughing. The pathological events in the asthma are mediated mainly by the airway smooth muscle (ASM) cells, and epithelial cells (71). PI3Ks play a pivotal role in the development and progression of almost all the above mentioned pathophysiological aspects of asthma. The activation of epithelial cells releases the chemokines and cytokines that attract the immune cells. PI3K activation is crucial for the regulation of ASM, as it upregulates the expression of chemokine CXCL8 that recruits different immune cells (72, 73). Further, it has been shown that T cells also contribute to the development of asthma by secreting the immune complexes that induce inflammation. Chemokine receptors recruit the p110γ

and p110 δ members of class-I PI3K family that contribute to activation, differentiation, and proliferation of the T cells (35, 74). PI3K mediated migration of neutrophils was also reported in some cases of asthma (75). The significance of PI3Ks in asthma makes it a potential therapeutic target for treatment of asthma (35). Indeed, various PI3K inhibitors have favorable pharmacokinetics, and are currently considered as promising drug candidates for the treatment of asthma (Table 1).

Mounting evidence reported by different research groups established the involvement of chemokine and chemokine receptors in various stages (development and progression) of

tumorigenesis (76, 77). Chemokine receptors are majorly expressed by the tumor cells, along with generation of respective chemokines within the tumor microenvironment (45, 78, 79). The presence of large number of CCR10 chemokine receptors has been reported on the melanoma cells. Upon binding to its cognate ligand CCL27, the PI3K pathway along with other intermediate pathways are initiated, and antiapoptotic signals are drawn to protect the cells from dying (80). Moreover, the expression of the CCR7 receptor along with its ligands CCL19 and CCL21 has been observed to be involved in the growth and advancement of many types of cancer (58). In the case of non-Hodgkin's lymphoma and pancreatic ductal adenocarcinoma, metastatic and lymphatic spreading is mediated through the PI3K/AKT signaling pathway induced by the high expression of CCR7 (81, 82). The PI3K pathway has been reported to be crucial in the growth and progression of prostate cancer. Binding of chemokine CXCL12 to its receptor CXCR4 induces PI3K pathway, which helps in promoting the proliferation of prostate cancer stem cells (CSCs). The CSCs help in re-emergence of tumor even after the primary treatment (79, 83). Furthermore, chronic lymphocytic leukemia (CLL) is a disease in which B cells are accumulated in the blood and tissue compartment. Many supporting cells like MSCs and NLCs secrete chemokines (CXCL12/ CXCL13) that play critical roles in the progression and retention of this disease (84, 85). According to a recent study the signaling pathway mediated by the PI3Kδ plays a significant role in the development and progression of chronic lymphocytic leukemia (CLL) (29). By using CAL-101, a potent inhibitor of PI3Kδ that inhibits the chemokine mediated signaling, the researchers observed a restriction in the B-cell receptor signaling and its reduced accumulation during CLL. Considering the different isoforms of PI3K, inhibitors have been developed for various isoforms that are currently in trial phase (Table 1).

3.2. G protein coupled receptor kinases (GRKs)

G protein coupled receptor kinases are serine/threonine protein kinases that specifically identify and phosphorylate the GPCRs activated by the agonist. Receptor phosphorylation by GRK is one of the well-studied mechanisms of GPCR

desensitization (86, 87). Binding of ligands to the GPCRs leads to the concurrent activation of G protein and β-arrestin-dependent signal transduction through a series of widely conserved biochemical steps (88). G protein dependent signaling is rapid and is dependent on secondary messengers, while arrestin signaling has slower onset and is signalosome dependent (89). Agonist activated GPCRs are phosphorylated by GRKs, which in turn regulates the binding of β-arrestin (90) (Figure 4). βarrestin binding can initiate the desensitization of G protein signaling at various stages such as recruitment of enzyme that degrades secondary messengers (91, 92), steric hindrance of the receptor (93), and receptor internalization that prevents further binding of the cognate ligands to the receptor (94). βarrestin molecule not only hinders the receptor associated G protein signaling, but also increases the desensitization and internalization of the GPCRs by shifting proteins such as PDEs and cytosolic Src (c-SRC) to the receptors (89). These two proteins turn off the signaling pathway by degrading the cAMP once they reach the membrane. Protein kinase A (PKA), protein kinase B (PKB), protein kinase C (PKC) along with c-SRC also help in enhancing the activity of GRK by phosphorylating it (89, 95) (Figure 4). Due to the propensity of GRKs to interact with numerous proteins involved in the signal transduction process, along with their canonical role of GPCR internalization and desensitization, β-arrestin also initiates additional signaling processes regulated by other kinases including MAPK signaling (89, 95, 96), and receptor tyrosine kinase transactivation (97). GRKs are also involved in the regulation of cellular responses through phosphorylation independent pathways along with phosphorylation mediated processes (98).

There are seven isoforms of GRKs. All the GRKs are involved in the desensitization of GPCRs. The only little differences that have been observed are in the structure and function of GRKs. On the basis of the observed differences, they are categorized into three subfamilies (99). The first family, designated as GRK1 subfamily, comprises GRK1/7, second subfamily known as GRK2 subfamily constitutes GRK2/3, and third subfamily called GRK4 subfamily contains GRK4/5/6. GRK1 and GRK7 are called as visual GRK and are present

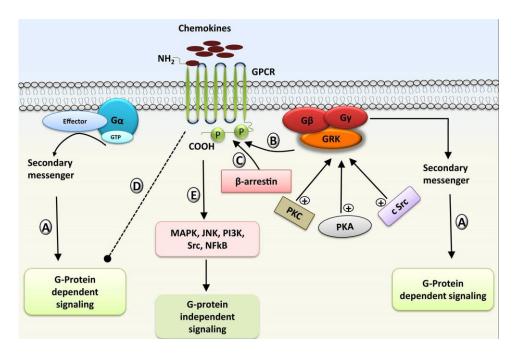


Figure 4. Regulation and desensitization of GPCR by GRKs (G-protein receptor kinases): A) Activation of GPCR and G-protein dependent reactions. B) Recruitment of GRK for GPCR phosphorylation. C) Binding of β-arrestin to phosphorylated GPCR, and formation of signalosome complex leading to desensitization of receptor. (D-E) Inhibition of G protein dependent signaling and activation of G protein independent signaling cascade; Activity of GRK can be enhanced by other protein kinases including protein kinase A (PKA), protein kinase C (PKC), and c-Src.

in retina (100, 101), GRK4 is expressed in testis (102). In contrast, other isoforms like GRK2, GRK3, GRK5, and GRK6 are widely found in various cells across the body (103-105). GRKs are multidomain proteins, having a 25 residue N-terminal region that is unique to GRKs. It also comprises of a regulator of G-protein signaling homology domain (RH) followed by a catalytic (serine threonine-protein kinase) domain that is accountable for phosphorylating the substrates (106).

As GPCR signaling is essential for the proper functioning of the immune cells and also for maintaining homeostasis, dysregulation of the GRK can be deleterious to the cells like leukocytes and neutrophils (107). Such processes often lead to various pathological conditions includina hypertension, chronic inflammatory disease, cardiac failure and rheumatoid arthritis (108).Inflammatory mediators that regulate the expression of GRKs on immune cells via degradation and transcription might also alter the expression of GRKs in different disease conditions (109, 110). As novel interactions of GRKs with numerous proteins are being discovered, GRKs can be the potential target for many diseases (111, 112).

Chemokine mediated signaling is regulated chemokine by **GRKs** through receptor phosphorylation and desensitization (113, 114). GRK mediated desensitization is crucial in sensing the changes in the chemokine gradient, and in turn to control the cellular migration. Moreover, GRKs may also interfere in chemokine signaling by direct interaction with molecules such as MEK. It has been evidenced that CCR2 receptor mediated signaling of ERK1/2 was enhanced in splenocytes of GRK2 knockout mice (115). On a similar note, it was also reported in the GRK2 knockout mice model that CCR5 directly induced the signaling to protein kinase B (PKB), ERK 1/2 and calcium mobilization (116). Neutrophils play a major role in the different infectious and inflammatory diseases. Mobilization of neutrophils is highly controlled by the CXCR4 receptor mediated signaling, which is essentially regulated by GRK6 (116).

The concept of "biased agonism" is also well established in the case of chemokine mediated GRK signaling pathways. This is the phenomenon in which the effector pathways are selectively activated on the basis of the activity induced by a given ligand (117, 118). Some of the agonists may act as "G-Protein biased" i.e., specifically inducing G protein and not the arrestins, while the others may act as "βarrestin biased" by selectively inducing the βarrestins. Chemokine ligands CCL19 and CCL21 of the CCR7 receptor are observed to be biased; both the ligands have equal potency towards the receptor, but only CCL19 induced pathway selectively leads to receptor desensitization through β-arrestin activation (103, 119, 120). Based on this biased agonism phenomenon, many therapeutic agents were proposed for diseases such as Warts Hypogammaglobulinemia Infections Myelokathexis (WHIM) syndrome and HIV infection (121). Principally, the signaling pathway induced by CXCL12 that mediates chemotaxis and receptor internalization through chemokine receptor CXCR4 is regulated by β-arrestin2 and GRK3. In case of WHIM syndrome, a mutation is present at COOH tail of the receptor which leads to the defective CXCR4 signaling. diseased condition, β-arrestin2 In regulates the chemotactic migration of the defective/mutated leukocytes but does not induce receptor internalization (122, 123).

It is well established that the GRKs, specifically GRK2 is involved during various stages of tumorigenesis as it creates a suitable environment that supports tumor progression (124). As supported by cumulative shreds of evidence, GRK2 level has been reported to be upregulated and to alter the other pathways in case of breast cancer (124). This protein is considered to be the key hallmark of cancer as it acts as a crucial oncomodifier in different tissues (125). Deficiency of GRK6 promotes the CXCR2 moderated tumor metastasis and progression in lung carcinoma model (126).

GPCR mediates the most important signal flow for the functioning of cardiovascular system. The involvement of GRKs is also observed in the smooth and regulated flow of the signal in the andergenic system. Hyperactivation of the andergenic system is responsible for maintaining equanimity and

progression of cardiovascular diseases hypertension and heart failure (25, 114). GRK2, GRK3, and GRK5 are seen to be widely involved in most of the cardiovascular diseases (127, 128). A recent study suggested that development of promising compounds that inhibit the activity of GRK5 are handy in preventing and reversing the progression of heart failure (129). Atherosclerosis is a chronic, medically compromised condition that is responsible for the heart related transgression and stroke. It is well evidenced that chemokines of CC and CXC families are entangled in the atherosclerotic plaque in humans as well as in mice (130, 131). GRK5 is seen to impair atherosclerosis, as it inhibits the migration of monocytes by desensitizing the receptor CCR2 (132).

It is well established that the level of GRKs decreases during inflammatory diseases such as multiple sclerosis, sepsis and rheumatoid arthritis (133). In a sepsis patient, clearance of infectious bacteria is initiated by the migration of leukocytes (neutrophils) through CXCL8/IL8 family chemokine. However, due to the signaling by GRK5 and GRK2, the cognate receptor CXCR2 is down-tuned and the sepsis is pursued. Reports suggested that the hindrance in the expression of GRK2 in neutrophil activates the CXCR2 receptor signaling and in turn induces the migration of neutrophil to the site of infection in sepsis (134, 135).

3.3. Janus kinases (JAKs)

Janus kinases are a small family of cytoplasmic kinases that are crucial in providing transmission signals from receptor to nucleus (136, 137). It consists of four members JAK1, JAK2, JAK3, and TYK2. Isoforms JAK1 and JAK2 of this family are crucial in administering a wide range of functions including growth, neural development and host defense (138, 139). JAK3 is mostly seen on the hematopoietic cells that are entangled in cellular signaling leading to immune responses. TYK2 is significant for the immune function and defense mechanism of the host (140). All the isoforms of JAKs possess a unique conserved region called JAKs homology domain (JH1-JH7). Along with the functional domain (JH1) at the C-terminal, they also possess a nonfunctional pseudokinase (JH2) (141).

Mechanistically, signaling through the JAK pathway is relatively simple, as limited molecular partners are involved (142). Binding of ligands with GPCRs induces the multimerization of receptor subunits. Due to this multimerization of receptor, two JAKs are brought into its close vicinity leading to the phosphorylation and activation of JAKs. Activated JAKs phosphorylate the C-terminal intracellular tail of both the receptors, thereby creating a docking site for signal transducer and activator of transcription (STATs). STATs are latent cytoplasmic transcription factors required by all the isoforms of JAK to mediate the signaling pathways (143). JAKs subsequently phosphorylate the tyrosine residue of STATs, which in turn initiates the dimerization of STATs. The activated STATs are then transported to the nucleus. where they bind to several places on the genome to modulate the gene expression (138, 144, 145). Currently, seven mammalian STATs have been recognized (STAT1, STAT2, STAT3, STAT4, STAT5a, STAT5b, STAT6). All these STATs are activated by a specific ligand (141, 144). A small protein family called suppressor of cytokine signaling (SOCS) is observed in the regulation of signaling through JAK/STAT pathway. It has been reported that the activation of the JAK-STAT pathway is regulated by SOCS protein, that interferes with the receptors required for relaying signal to the nucleus (138, 146, 147).

Dysregulation of the JAK/STAT-mediated signaling pathway was reported to be involved in different diseases including autoimmune diseases, inflammatory diseases, cancer, etc. (143, 148, 149). JAK/STAT pathways have also been identified as the major signaling pathway that activates the mechanism of wound healing (150, 151). Although JAK/STAT pathway is mostly implicated in cytokine signaling, its involvement in chemokine and its cognate receptor-mediated signaling has been well documented (152-154).

Among all the signaling pathways activated by chemokines, the involvement of phosphorylated JAK/STAT pathway is triggered soon after chemokines stimulate the GPCR (155). The very first evidence for the involvement of JAKs in chemokine mediated signaling was the manifestation that chemokine CCL2 (MCP-1) binding to CCR2 receptor

induced the activation of JAK2/STAT3 leading to rapid phosphorylation of CCR2 receptor (156). Additionally, by using differentially tagged CCR2 recombinant receptors, it was also established that CCL2 induces the multimerization of CCR2. This finding suggests that the chemokine receptor has the propensity of multimerization upon ligand binding (157). Furthermore, the involvement of JAK/STAT pathways in numerous chemokine receptors signaling including CCR1 (158), CCR2 (156), CCR5 (159, 160), CCR7 (153), CCR9 (152), CXCR4 (155, 161, 162), CXCR3 (163) has been elucidated.

The association of JAK/STAT with malignancies was first identified in 1990 (168). Chemokines and their receptors participate in growth and progression of various cancers through JAK/STAT pathways (169).CXCR4/CXCL12 signaling mediated by JAKs is demonstrated to be crucial for the integrin affinity moderation and T lymphocyte homing. The alliance of this signaling is displayed in case of B-cell chronic lymphocytic leukemia (B-CLL). The obtained data corroborate with the theory that JAK2 is a significant regulator of B-cell adhesion by CXCL12 (154). As JAK/STAT pathways are involved in different types of cancers including colitis-associated cancer, solid-organ malignancies (143, 145), etc., cancer therapies comprising of pan-JAK inhibitors against various isoform of JAKs are under clinical trials (22, 145). Considering the importance of JAK mediated chemokine signaling, researchers have also been investigating them as potential therapeutic agents against a variety of other diseases (140). For example, Osteoarthritis is a chronic medical condition characterized by the degeneration of articular cartilages and synovial membrane inflammation (170). An elevated concentration of chemokines (CXCL8 and CXCL11) is observed in the synovial fluid collected from the Osteoarthritis victim. These chemokines help in disease progression through the JAK/STAT pathway and are also involved in the apoptosis of chondrocytes (171). Similarly, CXCL10 evidenced as the significant biomarker in the case of rheumatoid arthritis (RA), involves JAK/STAT signaling in the disease (172). Tofacitinib, a potent inhibitor of JAK mediated signaling pathway is demonstrated as a potential therapeutic agent against RA (144). The expression of CXCL10 was

Table 2. Summary of Janus kinases, their associated chemokine/chemokine receptors and the JAK inhibitors involved in the treatment of different diseases

Inhibitors	Disease	Chemokine/ chemokine receptor	JAK/STAT involved	References
Maraviroc	Acute Lymphoblastic Leukemia	CXCL12 CXCL13 CCR5	JAK1/STAT3 JAK2/STAT3	(160)
Tofacitinib	Rheumatoid arthritis	CXCL2 CXCL10 CXCL 3	JAK1/STAT1 JAK1/STAT3	(144, 164)
Cepharanthin	Primary Sjögren's syndrome	CXCL10	JAK2/STAT1	(165)
ds-echinoside A	Cancer	CXCR4	JAK2/3-STAT	(166)
Berberine	Cardiovascular Disease	CXCR4	JAK2/STAT	(167)

also observed in the case of primary Sjögren's syndrome (pSS), a common chronic autoimmune disease. The CXCL9/CXCR3 and CXCL10/CXCR3 chemokine-receptor pairs induce JAK/STAT signaling to mediate the amassing of T cell infiltrate in the salivary gland of pSS victim (173). Cepharanthine controls pSS by inhibiting CXCL10 expression in human salivary gland ductal cells by obstructing the JAK2/STAT1 mediated signal flow (165). Several compounds have been synthesized and marketed to inhibit various diseases, where JAK/STAT is a regulatory signaling pathway (Table 2).

3.4. Mitogen-activated protein kinases (MAPKs)

Mitogen-activated protein kinases are serine/threonine protein kinases that play crucial roles in the cellular processes including cell proliferation, differentiation, apoptosis and motility (174, 175). A number of studies have evidenced that the uncontrolled activation of MAPK pathways are involved in the initiation and advancement of different cardiovascular abnormalities cancers. inflammatory diseases such as rheumatoid arthritis (176-178). MAPK pathways constitute a three tiered kinase cascade such as RAF, MEK and ERK family of kinases. Activation of this signal flow is initiated upon binding of ligands to the cell surface cognate receptor, which triggers the activation of RAS through the GTP-GDP exchange. Activated RAS-GTP engages and facilitates the activation of RAF that subsequently phosphorylates and activates the dual specific downstream loop of kinase MEK1 and

MEK2. Phosphorylated MEK1/2, in turn. phosphorylate ERK1 and ERK2. Activated ERK1/2 act on its substrates in the cytosol and nucleus to regulate a broad range of cellular functions including cell growth, survival, differentiation, and cytoskeletal rearrangements, etc. (175, 179, 180) (Figure 3). Until now, three prime MAPK signaling pathways are identified in mammalian cells: They include signal-regulated extracellular protein kinases (ERK1/2), p38 MAPK, and c-Jun NH2-terminal protein kinases (JNKs) (181, 182). The extracellular response kinases (ERKs) are implicated in normal cellular development including proliferation, differentiation, cell death, etc. (183, 184). The activation of c-Jun N-terminal kinase (JNK) is majorly demonstrated to have involvement in response to stress signals including inflammatory cytokines (185, 186). The p38 MAPK pathway is firmly activated by environmental stressors such as lipopolysaccharide, which in turn increase the expression of cytokines/chemokines and their receptors, thus making it a crucial immune response regulator (187, 188). In addition, it is also well documented that signaling through MAPK is stimulated by arrestin in order to relay the signal through its three modules of signaling cascade (189, 190).

The chemokine mediated signaling pathway specificity determines the fortune of the cells. Chemokine induced signaling cascade through PI3K/Akt activation leads to the survival of the cells, while p38 MAPK pathway activation results in cell death (191). Apart from chemokines, there are other stimuli such as cytokines and growth factors that can also activate the MAPK/ERK signaling cascade for

the migration of cells (192). The activity of the MAPK/ERK pathway is often upregulated in cancer as it plays a decisive role in oncogenesis (193, 194). The digressive activation of this pathway in cancer occurs through several mechanisms including a mutation in RAS/RAF and overexpression of receptor kinase (195). Chemokine CXCL8/IL8 induced MAPK pathway is demonstrated to have significant role in growth and proliferation of lung cancer, as the researchers reported that the proliferation and metastasis of lung cancer cells are fostered by integrin $\alpha \nu \beta 6$, which enhance the expression of CXCL8/IL8 through the MEK pathway (196). There is accumulated evidence for the fact that chemokine CXCL12 and its receptor CXCR4 are significant for colon cancer (197). Comparably it has shown that silencing CXCL12 gene leads to the downregulation of MAPK signaling which in turn has inhibitory effect on the metastatic perspective of colon cancer (198).

Abnormal activity of P38 MAPK has been observed in most of the inflammatory diseases (199). Phosphorylation cascade by p38 MAPK is initiated by pathogens and inflammatory stimulus which ultimately leads to the production of antiinflammatory cytokines and the phosphatase enzyme. This enzyme dephosphorylates and inactivates all three major MAP kinases such as p38MAPK, JNK, and ERK and triggers the initiation of multiple inflammatory diseases (200, 201). The JNK and p38 families of MAPKs have been identified to be involved in the pathogenesis of rheumatoid arthritis (202). The involvement of chemokineinduced MAPK pathway is also documented in RA patients. (203). Chemokine CXCL8 is present in large quantity in the synovial fluid of RA patients, whose expression is dependent on p38 MAPK. Moreover, reports established the involvement of MEK in chemokine induced signaling cascade (CCL4/CCR5/c-Jun and c-Fos/CCL2) in RA patients (204). Chemokines can potentially activate chondrocytes in order to release various inflammatory mediators. The high concentration of CXCL12 causes the death of chondrocytes via p38 MAPK activity (205). Further, signaling cascade induced by angiogenic chemokines and their respective receptors (CCL2/CCR2, CXCL8/CXCR2, and CXCL12/CXCR4) at the site of the wound is mediated by the MAPK/ERK pathway, which

provides the base for formulations of various pharmacological inhibitors against MAPK (179, 181, 199, 206). Ulixertinib, an ERK1/2 inhibitor is having significant therapeutic potential against solid-tumor malignancies (194). Talmapimod (207) and pyrimidopyridazinone (208) are p38 MAPK inhibitors formulated against RA are currently in clinical trials. Dilmapimod, a potent p38 MAPK inhibitor has displayed the inhibition of the MAPK pathway in asthma/COPD patients (209). Ralimetinib, another inhibitor of p38 MAPK has manifested its potential against advanced stages of cancer with acceptable pharmacokinetics (210).

3.5. Tec family kinases (TFKs)

Tec family kinases are non-receptor protein tyrosine kinases (PTKs), which are the critical components for signaling in lymphocytes (211, 212). Tec kinase family kinases are activated by a diverse range of surface receptors such as cytokines, chemokines, GPCRs, TLRs, integrins, as well as antigens (213). Kinases of this family are involved in various downstream signaling cascades related to proliferation, differentiation, actin reorganization, cell migration, Ca²⁺ influx, and apoptosis, etc. (214). Tec family member includes ITK (Interleukin-2 inducible T cell kinase), TEC (Tyrosine kinase expressed in hepatocellular carcinoma), RLK (Resting lymphocyte kinase), BTK (Burton's tyrosine kinase), and BMX (Bone marrow tyrosine kinase on chromosome-X) (214, 215). Members of TFK family hold two differentiating characteristics that are unique to them. Firstly, they have Tec homology (TH) domain that is involved in regulatory interactions. Secondly, they possess a pleckstrin homology (PH) domain which initiates the molecular activation by binding with proteins and phospholipids including the subunits of heterotrimeric G proteins (216, 217).

TFKs are mostly seen on hematopoietic cells, where they regulate their response towards external stimuli. The flow of signal regulated by Tec kinases involves the recruitment of immune cells to the site of infection as they facilitate the activation of immune response (218). It is also evident that members of the Tec kinase family contribute to other signaling cascades such as PI3K and MAPK (219). ITK regulates the chemokine receptor signaling

entangled to the migration processes through the stimulation of integrin and cell adhesion (220) It also regulates cytokine induced cell death in CD4+ T cells (221). Chemokine mediated migration of T cells in the case of allergic asthma is regulated by ITK. It was reported that quashing the activity of ITK leads to the reduced T cell migration, thus results in the prevention of allergic asthma (222).

Phosphorylation by Tec kinase at the tyrosine residue is associated with the stimulation of leucocyte (neutrophils) by chemokines (223, 224). The aberrant activation of this pathway may lead to various deleterious effects and pathophysiological conditions (225). Intestinal epithelial cells upon infection with bacteria or activation with other stimuli guide the production of chemokine CXCL8/IL8. Excessive neutrophils accumulated at the mucosal region facilitate the worsening of infection, and the chronic conditions may lead to life-threatening diseases like Crohn disease and ulcer, etc. (225, 226). Modification in the protein tyrosine kinases (PTKs) that regulate the CXCL8 production has been proposed as a novel therapeutic target (227).

BTK is identified as a significant factor in B-cell receptor signaling pathways and B-cell chemotaxis (228). It is evident that the BTK has a crucial role in the pathogenesis of B-cell chronic lymphocytic leukemia (B-CLL), as it plays a consequential role in the CXCL12 induced activation of integrin in both the normal as well as B-CLL condition. It has been demonstrated that BTK may act as the point of convergence between the B-cell receptor and chemokine induced signaling cascade (229)

3.6. Other kinases involved in chemokine mediated signaling

Apart from the above mentioned kinase families, there are many other kinases that play pivotal roles in regulating the signaling pathways induced by the chemokines. These kinases are the members of serine threonine-protein kinase family, that belong to cAMP-dependent protein kinase which includes protein kinase A, protein kinase B/Akt, protein kinase C and, PDK (230, 231). Binding of chemokine to the receptors triggers the dissociation

of G protein; Gα subunit regulates the production of cAMP from adenylyl cyclase (AC) (Figure 3). Protein kinase A (PKA) gets activated upon binding of cAMP to it; further it enters into the nucleus and subsequently phosphorylates the transcription factors (232, 233). PKB is one of the key mediators of PI3K pathway induced by the chemokines (234). Upon activation of PI3K pathway, a second messenger PIP3 is generated from PIP2, which facilitates the translocation of PKB to plasma membrane from cytoplasm, where it plays crucial role in cellular growth and proliferation (235). Chemokine mediated signaling initiates the generation of molecules such as diacylglycerol (DAG), which triggers calcium ion (Ca2+) mobilization, thus activating the Protein kinase C (PKC) for downstream signaling (236).

Src kinases are a small subgroup of the tyrosine kinase family and they act as the essential link between the effector molecules and the key transcription factors during cytoskeleton rearrangements, cell survival, cell growth, and proliferation (237). The Src family kinase regulation can be either activating or inhibitory. Kinases of this family play a crucial role in ITAM (immunoreceptor tyrosine based activation motif) dependent signaling cascade, utilized by immunoreceptors such as lymphocyte antigen receptors, Fc receptors and NK activating receptors. In addition, it also modulate ITIM (immunoreceptor tyrosine based inhibitory motif) dependent path, which negatively regulate both immunoreceptor function and host signaling activities, such as cytokines and integrin signal and regulate the activation of leukocytes (238).

The biological role of Src kinases is manifested in both leukocytes and hematopoietic cells as Gα subunit of G-protein associated with the chemokine receptor activates Src family kinases (237, 239). After getting stimulated by Gα subunit, Src subsequently phosphorylates multiple downstream substrates including Class IA PI 3-Kinase (PI3K), Interleukin-2 inducible T cell kinase (ITK), focal adhesion kinase (FAK), and proline-rich tyrosine kinase 2 (PYK2) (240-242). It is also evident that, once chemokine CXCR12 binds to its cognate receptor, MAPK signaling cascade activation is facilitated by both Lyn and Lck kinases which are the

members of Src kinase family (243). PKA and PKB kinases along with Src are also reported to be involved in enhancing the activity of GRKs (Figure 4).

4. CONCLUSIONS AND FUTURE PERSPECTIVES

The current review has illustrated the significance of various kinases involved in the chemokine mediated signaling cascade and demonstrated their role in regulatory as well as inhibitory pathways on the basis of contemporary development in the research related to chemokines and kinases. After four to five decades of the persistent contribution of several research groups across the globe, it has been established that the kinase system induced by the chemokine/ chemokine receptor engrosses a central position in regulation of the immune system, thus playing a significant role in progression of different initiation and pathophysiological conditions. Overwhelming pieces of evidence have established that kinases are associated with the complex networks of chemokine mediated signaling in which the consequential crosstalk among the components regulates multiple biological activities. However, it's still a matter of debate regarding how to keep the other associated signaling pathway intact by inhibiting one kinase. A fundamental outline of how the chemokine and kinase systems work has been manifested by using mouse models, and there has been substantial progress in the application of this knowledge in clinical practice. Currently, these pathways are potential drug targets for many pathological conditions either involved in innate or adaptive immune components.

Many drug compounds which target kinases are in different phases of the clinical trials. Despite the fact that considerable success has been obtained by using chemokine and associated kinase systems as potential drug targets, satisfactory treatments for chronic immune mediated diseases remain a major unmet medical need. There is high biomedical demand for the development of novel potent kinase inhibitor with clinical specificity and lesser side effects. It is imperative to discern every meaningful interaction between the chemokine and the kinase system so that it can further aid in

formulating alternative therapeutic compounds targeting chemokine/chemokine receptors and their associated kinases.

Apart from drugs targeting kinases, there exists compounds that show inhibitory effects on chemokine and GPCRs (244, 245). They include; engineered chemokines, synthetic GAGs and small molecule receptor targets, which can modulate and deactivate the kinase signaling pathway. Few noted examples include, chemically modified high affinity CCL5 ligands (246), synthetic monosaccharide, 2,4-O-di-sulfated iduronic acid (Di-S-IdoA) (247), selective inhibitors (LY294002 and MK-2206) of CXCR6 (248) etc. These studies establish that kinases, GPCRs, and chemokines are the crucial molecular partners, and are hot targets of pharmaceutical industry. Understanding combinatorial mechanism of the action inhibitor/drug molecules at molecular level will be useful in terms of system biology perspective, as they will elucidate the other key pathways involved, and aid us to formulate molecules with high specificity and minimal side-affects.

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6. REFERENCES

 Behm, B., P. Babilas, M. Landthaler and S. Schreml: Cytokines, chemokines and growth factors in wound healing. J Eur Acad Dermatol Venereol, 26(7), 812-20 (2012)

> DOI: 10.1111/j.1468-3083.2011.04415.x PMid:22211801

 Griffith, J. W., C. L. Sokol and A. D. Luster: Chemokines and chemokine receptors: positioning cells for host defense and immunity. Annu Rev Immunol, 32, 659-702 (2014)

DOI: 10.1146/annurev-immunol-032713-

120145 PMid:24655300

- Poluri, K.: Chemokines: the holy messengers of leukocyte trafficking. Austin J Biotechnol. Bioeng, 1, 1-3 (2014)
- Raman, D., T. Sobolik-Delmaire and A. Richmond: Chemokines in health and disease. Exp Cell Res, 317(5), 575-89 (2011)

DOI: 10.1016/j.yexcr.2011.01.005 PMid:21223965 PMCid:PMC3063402

 Sokol, C. L. and A. D. Luster: The chemokine system in innate immunity. Cold Spring Harb Perspect Biol, 7(5) (2015)

> DOI: 10.1101/cshperspect.a016303 PMid:25635046 PMCid:PMC4448619

- Gulati, K. and K. M. Poluri: Mechanistic and therapeutic overview of glycosaminoglycans: the unsung heroes of biomolecular signaling. Glycoconj J, 33(1), 1-17 (2016) DOI: 10.1007/s10719-015-9642-2 PMid:26635091
- 7. Turner, M. D., B. Nedjai, T. Hurst and D. J. Pennington: Cytokines and chemokines: At the crossroads of cell signalling and inflammatory disease. Biochim Biophys Acta, 1843(11), 2563-2582 (2014)

DOI: 10.1016/j.bbamcr.2014.05.014 PMid:24892271

- Zlotnik, A. and O. Yoshie: The chemokine superfamily revisited. Immunity, 36(5), 705-16 (2012)
 DOI: 10.1016/j.immuni.2012.05.008
 PMid:22633458 PMCid:PMC3396424
- 9. Kufareva, I., C. L. Salanga and T. M. Handel: Chemokine and chemokine

receptor structure and interactions: implications for therapeutic strategies. Immunol Cell Biol, 93(4), 372-383 (2015) DOI: 10.1038/icb.2015.15 PMid:25708536 PMCid:PMC4406842

- Katritch, V. and V. S. Cherezov, Raymond C Structure-function of the G protein-coupled receptor superfamily. Annual review of pharmacology toxicology, 53, 531-556 (2013) DOI: 10.1146/annurev-pharmtox-032112-135923 PMid:23140243 PMCid:PMC3540149
- Gacasan, S. B., D. L. Baker and A. L. Parrill: G protein-coupled receptors: the evolution of structural insight. AIMS biophys, 4(3), 491 (2017)
 DOI: 10.3934/biophy.2017.3.491
 PMid:29951585 PMCid:PMC6018013
- Erlandson, S. C., C. McMahon and A. C. Kruse: Structural basis for G protein-coupled receptor signaling. Annu Rev Biophys, 47, 1-18 (2018)
 DOI: 10.1146/annurev-biophys-070317-032931
 PMid:29498889
- 13. Salanga, C. and T. Handel: Chemokine oligomerization and interactions with receptors and glycosaminoglycans: the role of structural dynamics in function. Exp Cell Res, 317(5), 590-601 (2011) DOI: 10.1016/j.yexcr.2011.01.004 PMid:21223963 PMCid:PMC3089961
- Baggiolini, M.: Chemokines and leukocyte traffic. Nature, 392(6676), 565 (1998)
 DOI: 10.1038/33340

PMid:9560152

15. Stadtmann, A. and A. Zarbock: CXCR2: From Bench to Bedside. Front Immunol,

Kinase mediated signaling pathways of chemokines

3, 263 (2012)

DOI: 10.3389/fimmu.2012.00263 PMid:22936934 PMCid:PMC3426767

- Hughes, C. E. and R. J. Nibbs: A guide to 16. chemokines and their receptors. FEBS J, 285(16), 2944-2971 (2018) DOI: 10.1111/febs.14466 PMid:29637711 PMCid:PMC6120486
- Legler, D. F. and M. Thelen: New insights 17. in chemokine signaling. F1000Res, 7, 95 (2018)DOI: 10.12688/f1000research.13130.1

PMid:29416853 PMCid:PMC5782407

18. Thelen, M.: Dancing to the tune of chemokines. Nat Immunol, 2(2), 129 (2001)

DOI: 10.1038/84224 PMid:11175805

19. Wu, P., T. E. Nielsen and M. H. Clausen: FDA-approved small-molecule kinase inhibitors. Trends Pharmacol Sci, 36(7), 422-39 (2015) DOI: 10.1016/j.tips.2015.04.005

PMid:25975227

20. P.: The Cohen, role of protein phosphorylation in human health and disease. Eur J Biochem, 268(19), 5001-5010 (2001)

DOI: 10.1046/j.0014-2956.2001.02473.x PMid:11589691

- 21. Barnes, P. J.: Kinases as Novel Therapeutic Targets in Asthma and Chronic Obstructive Pulmonary Disease. Pharmacol Rev, 68(3), 788-815 (2016) DOI: 10.1124/pr.116.012518 PMid:27363440
- 22. Yuan, X., H. Wu, H. Bu, J. Zhou and H. Zhang: Targeting the immunity protein kinases for immuno-oncology. Eur J Med

Chem, 163, 413-427 (2019) DOI: 10.1016/j.ejmech.2018.11.072 PMid:30530193

- Moser, B., M. Wolf, A. Walz and P. 23. Loetscher: Chemokines: multiple levels of leukocyte migration control. Trends Immunol, 25(2), 75-84 (2004) DOI: 10.1016/j.it.2003.12.005 PMid:15102366
- 24. O'Hayre, M., C. L. Salanga, T. M. Handel and S. J. Allen: Chemokines and cancer: migration, intracellular signalling and intercellular communication in microenvironment. Biochem J, 409(3), 635-49 (2008) DOI: 10.1042/BJ20071493

PMid:18177271

25. Patel, J., K. M. Channon and E. McNeill: The downstream regulation of chemokine receptor signalling: implications for atherosclerosis. Mediators Inflamm. 2013, 459520 (2013) DOI: 10.1155/2013/459520

PMid:23690662 PMCid:PMC3649756

26. Thelen, M. and J. V. Stein: How chemokines invite leukocytes to dance. Nat Immunol, 9(9), 953 (2008) DOI: 10.1038/ni.f.207

PMid:18711432

- 27. Garin, A. and A. E. Proudfoot: Chemokines as targets for therapy. Exp Cell Res, 317(5), 602-12 (2011) DOI: 10.1016/j.yexcr.2010.12.021 PMid:21376173
- 28. Koelink, P. J., S. A. Overbeek, S. Braber, P. de Kruijf, G. Folkerts, M. J. Smit and A. D. Kraneveld: Targeting chemokine chronic receptors in inflammatory diseases: an extensive review. Pharmacol Ther, 133(1), 1-18 (2012)

- DOI: 10.1016/j.pharmthera.2011.06.008 PMid:21839114
- Hoellenriegel, J., S. A. Meadows, M. Sivina, W. G. Wierda, H. Kantarjian, M. J. Keating, N. Giese, S. O'Brien, A. Yu, L. L. Miller, B. J. Lannutti and J. A. Burger: The phosphoinositide 3'-kinase delta inhibitor, CAL-101, inhibits B-cell receptor signaling and chemokine networks in chronic lymphocytic leukemia. Blood, 118(13), 3603-12 (2011)
 DOI: 10.1182/blood-2011-05-352492
 PMid:21803855 PMCid:PMC4916562
- Vanhaesebroeck, B., S. J. Leevers, K. Ahmadi, J. Timms, R. Katso, P. C. Driscoll, R. Woscholski, P. J. Parker and M. D. Waterfield: Synthesis and function of 3-phosphorylated inositol lipids. Annu Rev Biochem, 70(1), 535-602 (2001) DOI: 10.1146/annurev.biochem.70.1.535 PMid:11395417
- 31. Vanhaesebroeck, B., J. Guillermet-Guibert, M. Graupera and B. Bilanges: The emerging mechanisms of isoform-specific PI3K signalling. Nat Rev Mol Cell Biol, 11(5), 329-41 (2010)
 DOI: 10.1038/nrm2882
 PMid:20379207
- 32. Jean, S. and A. A. Kiger: Classes of phosphoinositide 3-kinases at a glance. J Cell Sci, 127(Pt 5), 923-8 (2014) DOI: 10.1242/jcs.093773 PMid:24587488 PMCid:PMC3937771
- Curnock, A. P., M. K. Logan and S. G. Ward: Chemokine signalling: pivoting around multiple phosphoinositide 3-kinases. Immunology, 105(2), 125-136 (2002)
 DOI: 10.1046/i.1365-2567.2002.01345.x

DOI: 10.1046/j.1365-2567.2002.01345.> PMid:11872087 PMCid:PMC1782650

- 34. Koyasu, S.: The role of PI3K in immune cells. Nat Immunol, 4(4), 313 (2003) DOI: 10.1038/ni0403-313 PMid:12660731
- Yoo, E. J., C. A. Ojiaku, K. Sunder and R. A. Panettieri, Jr.: Phosphoinositide 3-Kinase in Asthma: Novel Roles and Therapeutic Approaches. Am J Respir Cell Mol Biol, 56(6), 700-707 (2017) DOI: 10.1165/rcmb.2016-0308TR PMid:27977296 PMCid:PMC5516292
- 36. Cushing, T. D., D. P. Metz, D. A. Whittington and L. R. McGee: PI3Kdelta and PI3Kgamma as targets for autoimmune and inflammatory diseases. J Med Chem, 55(20), 8559-81 (2012) DOI: 10.1021/jm300847w PMid:22924688
- 37. Fruman, D. A., H. Chiu, B. D. Hopkins, S. Bagrodia, L. C. Cantley and R. T. Abraham: The PI3K Pathway in Human Disease. Cell, 170(4), 605-635 (2017) DOI: 10.1016/j.cell.2017.07.029 PMid:28802037 PMCid:PMC5726441
- 38. Hawkins, P. T. and L. R. Stephens: PI3K signalling in inflammation. Biochim Biophys Acta, 1851(6), 882-97 (2015) DOI: 10.1016/j.bbalip.2014.12.006 PMid:25514767
- G Ward, S.: Phosphoinositide 3-kinases and leukocyte migration. Curr Immunol Rev, 8(2), 154-160 (2012)
 DOI: 10.2174/157339512800099684
- Turner, L., S. G. Ward and J. Westwick: RANTES-activated human T lymphocytes. A role for phosphoinositide 3-kinase. J Immunol, 155(5), 2437-2444 (1995)
- 41. Ferguson, G. J., L. Milne, S. Kulkarni, T.

Sasaki, S. Walker, S. Andrews, T. Crabbe, P. Finan, G. Jones, S. Jackson, M. Camps, C. Rommel, M. Wymann, E. Hirsch, P. Hawkins and L. Stephens: PI(3)Kgamma has an important context-dependent role in neutrophil chemokinesis. Nat Cell Biol, 9(1), 86-91 (2007)

DOI: 10.1038/ncb1517 PMid:17173040

- 42. Turner, S. J., J. Domin, M. D. Waterfield, S. G. Ward and J. Westwick: The CC chemokine monocyte chemotactic peptide-1 activates both the class I p85/p110 phosphatidylinositol 3-kinase and the class II PI3K-C2α. J Biol Chem, 273(40), 25987-25995 (1998) DOI: 10.1074/jbc.273.40.25987 PMid:9748276
- Sotsios, Y., G. C. Whittaker, J. Westwick and S. G. Ward: The CXC chemokine stromal cell-derived factor activates a Gicoupled phosphoinositide 3-kinase in T lymphocytes. J Immunol, 163(11), 5954-5963 (1999)
- 44. Del Prete, A., W. Vermi, E. Dander, K. Otero, L. Barberis, W. Luini, Bernasconi, M. Sironi, A. Santoro, C. Garlanda, F. Facchetti, M. P. Wymann, A. Vecchi, E. Hirsch, A. Mantovani and S. Sozzani: Defective dendritic cell migration and activation of adaptive immunity in PI3Kgamma-deficient mice. EMBO J, 23(17), 3505-15 (2004) DOI: 10.1038/si.emboi.7600361 PMid:15318168 PMCid:PMC516633
- 45. Chow, M. T. and A. D. Luster: Chemokines in cancer. Cancer Immunol Res, 2(12), 1125-31 (2014) DOI: 10.1158/2326-6066.CIR-14-0160 PMid:25480554 PMCid:PMC4258879

- 46. Wilson, R., A. Cahn, A. Deans, I. McSherry, C. Rambaran, A. Sousa and D. Wilbraham: Safety, tolerability and pharmacokinetics (PK) of single and repeat nebulised doses of a novel phosphoinositide 3-kinase δ inhibitor (PI3Kδ), GSK2269557, administered to healthy male subjects in a phase I study. In: Eur Respiratory Soc, (2013)
- 47. Norman, P.: Evaluation of WO2013136076: two crystalline forms of the phosphatidylinositol 3-kinase-delta inhibitor RV-1729. Expert Opin Ther Pat, 24(4), 471-5 (2014)
 DOI: 10.1517/13543776.2014.865725
 PMid:24283201
- 48. Evans, C. A., T. Liu, A. Lescarbeau, S. J. Nair, L. Grenier, J. A. Pradeilles, Q. Glenadel, T. Tibbitts, A. M. Rowley, J. P. DiNitto, E. E. Brophy, E. L. O'Hearn, J. A. Ali, D. G. Winkler, S. I. Goldstein, P. O'Hearn, C. M. Martin, J. G. Hoyt, J. R. Soglia, C. Cheung, M. M. Pink, J. L. Proctor, V. J. Palombella, M. R. Tremblay and A. C. Castro: Discovery of a Selective Phosphoinositide-3-Kinase (PI3K)-gamma Inhibitor (IPI-549) as an Immuno-Oncology Clinical Candidate. ACS Med Chem Lett, 7(9), 862-7 (2016) DOI: 10.1021/acsmedchemlett.6b00238 PMid:27660692 PMCid:PMC5018865
- 49. Shin, N., Y. L. Li, S. Mei, K. H. Wang, L. Hall, K. Katiyar, Q. Wang, G. Yang, B. Rumberger, L. Leffet, X. He, M. Rupar, K. Bowman, M. Favata, J. Li, M. Liu, Y. Li, M. Covington, H. Koblish, M. Soloviev, D. Shuey, T. Burn, S. Diamond, J. Fridman, A. Combs, W. Yao, S. Yeleswaram, G. Hollis, K. Vaddi, R. Huber, R. Newton and P. Scherle: INCB040093 Is a Novel PI3Kdelta Inhibitor for the Treatment of B Cell Lymphoid Malignancies. J

Pharmacol Exp Ther, 364(1), 120-130 (2018)

DOI: 10.1124/jpet.117.244947 PMid:29127109

- 50. Burris III, H. A., I. W. Flinn, M. R. Patel, T. S. Fenske, C. Deng, D. M. Brander, M. Gutierrez, J. H. Essell, J. G. Kuhn and H. P. Miskin: Umbralisib, a novel PI3Kδ and casein kinase-1ε inhibitor, in relapsed or refractory chronic lymphocytic leukaemia and lymphoma: an open-label, phase 1, dose-escalation, first-in-human study. Lancet Oncol, 19(4), 486-496 (2018) DOI: 10.1016/S1470-2045(18)30082-2
- 51. Ali, K., D. R. Soond, R. Pineiro, T. Hagemann, W. Pearce, E. L. Lim, H. Bouabe, C. L. Scudamore, T. Hancox, H. Maecker, L. Friedman, M. Turner, K. Okkenhaug and B. Vanhaesebroeck: Inactivation of PI(3)K p110delta breaks regulatory T-cell-mediated immune tolerance to cancer. Nature, 510(7505), 407-411 (2014)

 DOI: 10.1038/nature13444

PMid:24919154 PMCid:PMC4501086

52. Erdreich-Epstein, A., A. R. Singh, S. Joshi, F. M. Vega, P. Guo, J. Xu, S. Groshen, W. Ye, M. Millard and M. Campan: Association of high microvessel αvβ3 and low PTEN with in poor outcome stage neuroblastoma: rationale for using first in class dual PI3K/BRD4 inhibitor, SF1126. Oncotarget, 8(32), 52193 (2017)

DOI: 10.18632/oncotarget.13386 PMid:28881723 PMCid:PMC5581022

53. Wheler, J., D. Mutch, J. Lager, C. Castell, L. Liu, J. Jiang and A. M. Traynor: Phase I Dose-Escalation Study of Pilaralisib (SAR245408,

XL147) in Combination with Paclitaxel and Carboplatin in Patients with Solid Tumors. Oncologist, 22(4), 377-e37 (2017)

DOI: 10.1634/theoncologist.2016-0257 PMid:28275119 PMCid:PMC5388374

- 54. Barlaam, B., S. Cosulich, S. Degorce, M. Fitzek, S. Green, U. Hancox, C. Lambert-van Brempt, der J. J. Lohmann, M. Maudet, R. Morgentin, M. J. Pasquet, A. Peru, P. Ple, T. Saleh, M. Vautier, M. Walker, L. Ward and N. Warin: Discovery of (R)-8-(1-(3,5difluorophenylamino)ethyl)-N,Ndimethyl-2-morpholino-4-oxo-4Hchrom ene-6-carboxamide (AZD8186): a potent and selective inhibitor of PI3Kbeta and PI3Kdelta for the treatment of PTEN-deficient cancers. J Med Chem, 58(2), 943-62 (2015) DOI: 10.1021/jm501629p PMid:25514658
- 55. Bedard, P. L., M. A. Davies, S. Kopetz, D. Juric, G. I. Shapiro, J. J. Luke, A. Spreafico, B. Wu, C. Castell, C. Gomez, S. Cartot-Cotton, F. Mazuir, M. Dubar, S. Micallef, B. Demers and K. T. Flaherty: First-in-human trial of the PI3Kbeta-selective inhibitor SAR260301 in patients with advanced solid tumors. Cancer, 124(2), 315-324 (2018)

DOI: 10.1002/cncr.31044 PMid:28976556

56. Shugg, R. P., A. Thomson, N. Tanabe, A. Kashishian, B. H. Steiner, K. D. Puri, A. Pereverzev, B. J. Lannutti, F. R. Jirik, S. J. Dixon and S. M. Sims: Effects of isoform-selective phosphatidylinositol 3-kinase inhibitors on osteoclasts: actions on cytoskeletal organization, survival, and resorption. J Biol Chem, 288(49), 35346-57 (2013) DOI: 10.1074/jbc.M113.507525 PMid:24133210 PMCid:PMC3853283

- Campa, C. C., R. L. Silva, J. P. Margaria, T. Pirali, M. S. Mattos, L. R. Kraemer, D. C. Reis, G. Grosa, F. Copperi, E. M. Dalmarco, R. C. P. Lima-Junior, S. Aprile, V. Sala, F. Dal Bello, D. S. Prado, J. C. Alves-Filho, C. Medana, G. D. Cassali, G. C. Tron, M. M. Teixeira, E. Ciraolo, R. C. Russo and E. Hirsch: Inhalation of the prodrug PI3K inhibitor CL27c improves lung function in asthma and fibrosis. Nat Commun, 9(1), 5232 (2018)
 DOI: 10.1038/s41467-018-07698-6
 PMid:30542075 PMCid:PMC6290777
- 58. Li, H., D. Park, P. M. Abdul-Muneer, B. Xu, H. Wang, B. Xing, D. Wu and S. Li: PI3Kgamma inhibition alleviates symptoms and increases axon number in experimental autoimmune encephalomyelitis mice. Neuroscience, 253, 89-99 (2013)

DOI: 10.1016/j.neuroscience.-2013.08.051 PMid:24012746

59. Winkler, D. G., K. L. Faia, J. P. DiNitto, J. A. Ali, K. F. White, E. E. Brophy, M. M. Pink, J. L. Proctor, J. Lussier, C. M. Martin, J. G. Hoyt, B. Tillotson, E. L. Murphy, A. R. Lim, B. D. Thomas, J. R. Macdougall, P. Ren, Y. Liu, L. S. Li, K. A. Jessen, C. C. Fritz, J. L. Dunbar, J. R. Porter, C. Rommel, V. J. Palombella, P. S. Changelian and J. L. Kutok: PI3K-delta and PI3K-gamma inhibition by IPI-145 abrogates immune responses suppresses activity in autoimmune and inflammatory disease models. Chem Biol, 20(11), 1364-74 (2013)

DOI: 10.1016/j.chembiol.2013.09.017 PMid:24211136 60. Doukas, J., L. Eide, K. Stebbins, A. Racanelli-Layton, L. Dellamary, Martin, E. Dneprovskaia, G. Noronha, R. Soll, W. Wrasidlo, L. M. Acevedo and D. A. Cheresh: Aerosolized phosphoinositide 3-kinase gamma/delta inhibitor TG100-115 (3-(2,4-diamino-6-(3-hydroxyphenyl)pteridin-7-yl)phenol) as a therapeutic candidate for asthma and chronic obstructive pulmonary disease. J Pharmacol Exp Ther, 328(3), 758-65 (2009)

DOI: 10.1124/jpet.108.144311

PMid:19056934

- Krop, I. E., I. A. Mayer, V. Ganju, M. Dickler, S. Johnston, S. Morales, D. A. Yardley, B. Melichar, A. Forero-Torres, S. C. Lee, R. de Boer, K. Petrakova, S. Vallentin, E. A. Perez, M. Piccart, M. Ellis, E. Winer, S. Gendreau, M. Derynck, M. Lackner, G. Levy, J. Qiu, J. He and P. Schmid: Pictilisib for oestrogen receptorpositive, aromatase inhibitor-resistant, advanced or metastatic breast cancer (FERGI): a randomised, double-blind, placebo-controlled, phase 2 trial. Lancet Oncol, 17(6), 811-821 (2016)
 DOI: 10.1016/S1470-2045(16)00106-6
- 62. Jimeno, A., J. E. Bauman, C. Weissman, D. Adkins, I. Schnadig, P. Beauregard, D. W. Bowles, A. Spira, B. Levy, N. Seetharamu, D. Hausman, L. Walker, C. M. Rudin and K. Shirai: A randomized, phase 2 trial of docetaxel with or without PX-866, an irreversible oral phosphatidylinositol 3-kinase inhibitor, in patients with relapsed or metastatic head and neck squamous cell cancer. Oral Oncol, 51(4), 383-8 (2015)

DOI: 10.1016/j.oraloncology.-

2014.12.013

PMid:25593016 PMCid:PMC4857706

- 63. Mateo, J., G. Ganji, C. Lemech, H. A. Burris, S. W. Han, K. Swales, S. Decordova, M. P. DeYoung, D. A. Smith, S. Kalyana-Sundaram, J. Wu, M. Motwani, R. Kumar, J. M. Tolson, S. Y. Rha, H. C. Chung, J. P. Eder, S. Sharma, Y. J. Bang, J. R. Infante, L. Yan, J. S. de Bono and H. T. Arkenau: A First-Time-in-Human Study of GSK2636771, a Phosphoinositide 3 Kinase Selective Inhibitor, in Patients with Advanced Solid Tumors. Clin Cancer Res, 23(19), 5981-5992 (2017) DOI: 10.1158/1078-0432.CCR-17-0725 PMid:28645941
- 64. Liu, N., B. R. Rowley, C. O. Bull, C. Schneider, A. Haegebarth, C. A. Schatz, P. R. Fracasso, D. P. Wilkie, M. Hentemann, S. M. Wilhelm, W. J. Scott, D. Mumberg and K. Ziegelbauer: BAY 80-6946 is a highly selective intravenous PI3K inhibitor with potent p110alpha and p110delta activities in tumor cell lines and xenograft models. Mol Cancer Ther, 12(11), 2319-30 (2013)
 DOI: 10.1158/1535-7163.MCT-12-0993-T PMid:24170767
- 65. Ando, Y., S. Iwasa, S. Takahashi, H. Saka, T. Kakizume, K. Natsume, N. Suenaga, C. Quadt and Y. Yamada: Phase I study of alpelisib (BYL719), an alpha-specific PI3K inhibitor, in Japanese patients with advanced solid tumors. Cancer Sci, 110(3), 1021-1031 (2019) DOI: 10.1111/cas.13923 PMid:30588709 PMCid:PMC6398875
- 66. Juric, D., I. Krop, R. K. Ramanathan, T. R. Wilson, J. A. Ware, S. M. Sanabria Bohorquez, H. M. Savage, D. Sampath, L. Salphati, R. S. Lin, H. Jin, H. Parmar, J. Y. Hsu, D. D. Von Hoff and J. Baselga: Phase I Dose-Escalation Study of

- Taselisib, an Oral PI3K Inhibitor, in Patients with Advanced Solid Tumors. Cancer Discov, 7(7), 704-715 (2017) DOI: 10.1158/2159-8290.CD-16-1080 PMid:28331003 PMCid:PMC5501742
- Maira, S. M., S. Pecchi, A. Huang, M. 67. Burger, M. Knapp, D. Sterker, C. Schnell, D. Guthy, T. Nagel, M. Wiesmann, S. Brachmann, C. Fritsch, M. Dorsch, P. Chene, K. Shoemaker, A. De Pover, D. Menezes, G. Martiny-Baron, D. Fabbro, C. J. Wilson, R. Schlegel, F. Hofmann, C. Garcia-Echeverria, W. R. Sellers and C. F. Voliva: Identification and characterization of NVP-BKM120, an orally available pan-class I PI3-kinase inhibitor. Mol Cancer Ther, 11(2), 317-28 (2012)

DOI: 10.1158/1535-7163.MCT-11-0474 PMid:22188813

- 68. Flinn, I. W., S. O'Brien, B. Kahl, M. Patel, Y. Oki, F. F. Foss, P. Porcu, J. Jones, J. A. Burger and N. Jain: Duvelisib, a novel oral dual inhibitor of PI3K-δ, γ, is clinically active in advanced hematologic malignancies. Blood, 131(8), 877-887 (2018)
 - DOI: 10.1182/blood-2017-05-786566 PMid:29191916 PMCid:PMC6033052
- Brown, J. R., J. C. Byrd, S. E. Coutre, D. M. Benson, I. W. Flinn, N. D. Wagner-Johnston, S. E. Spurgeon, B. S. Kahl, C. Bello, H. K. Webb, D. M. Johnson, S. Peterman, D. Li, T. M. Jahn, B. J. Lannutti, R. G. Ulrich, A. S. Yu, L. L. Miller and R. R. Furman: Idelalisib, an inhibitor of phosphatidylinositol 3-kinase p110delta, for relapsed/refractory chronic lymphocytic leukemia. Blood, 123(22), 3390-7 (2014)

DOI: 10.1182/blood-2013-11-535047 PMid:24615777 PMCid:PMC4123414

- 70. Bendell, J. C., C. Kurkjian, J. R. Infante, T. M. Bauer, H. A. Burris, 3rd, F. A. Greco, K. C. Shih, D. S. Thompson, C. M. Lane, L. H. Finney and S. F. Jones: A phase 1 study of the sachet formulation of the oral dual PI3K/mTOR inhibitor BEZ235 given twice daily (BID) in patients with advanced solid tumors. Invest New Drugs, 33(2), 463-71 (2015) DOI: 10.1007/s10637-015-0218-6 PMid:25707361
- 71. Lemanske, R. F., Jr. and W. W. Busse: Asthma: clinical expression and molecular mechanisms. J Allergy Clin Immunol, 125(2 Suppl 2), S95-102 (2010) DOI: 10.1016/j.jaci.2009.10.047 PMid:20176271 PMCid:PMC2853245
- 72. Dragon, S., M. S. Rahman, J. Yang, H. Unruh, A. J. Halayko and A. S. Gounni: IL-17 enhances IL-1beta-mediated CXCL-8 release from human airway smooth muscle cells. Am J Physiol Lung Cell Mol Physiol, 292(4), L1023-9 (2007) DOI: 10.1152/ajplung.00306.2006 PMid:17189320
- 73. Mohamed, J. S. and A. M. Boriek: Stretch augments TGF-beta1 expression through RhoA/ROCK1/2, PTK, and Pl3K in airway smooth muscle cells. Am J Physiol Lung Cell Mol Physiol, 299(3), L413-24 (2010) DOI: 10.1152/ajplung.90628.2008 PMid:20511342 PMCid:PMC2951069
- 74. Jarmin, S. J., R. David, L. Ma, J. G. Chai, H. Dewchand, A. Takesono, A. J. Ridley, K. Okkenhaug and F. M. Marelli-Berg: T cell receptor-induced phosphoinositide-3-kinase p110delta activity is required for T cell localization to antigenic tissue in mice. J Clin Invest, 118(3), 1154-64 (2008)
 DOI: 10.1172/JCI33267

PMid:18259608 PMCid:PMC2230659

- 75. Kampe, M., M. Lampinen, I. Stolt, C. Janson, G. Stalenheim and M. Carlson: PI3-kinase regulates eosinophil and neutrophil degranulation in patients with allergic rhinitis and allergic asthma irrespective of allergen challenge model. Inflammation, 35(1), 230-9 (2012) DOI: 10.1007/s10753-011-9309-5 PMid:21384093
- 76. Nagarsheth, N., M. S. Wicha and W. Zou: Chemokines in the cancer microenvironment and their relevance in cancer immunotherapy. Nat Rev Immunol, 17(9), 559-572 (2017) DOI: 10.1038/nri.2017.49 PMid:28555670 PMCid:PMC5731833
- 77. Vilgelm, A. E. and A. Richmond: Chemokines Modulate Immune Surveillance in Tumorigenesis, Metastasis, and Response to Immunotherapy. Front Immunol, 10, 333 (2019)
 DOI: 10.3389/fimmu.2019.00333
 PMid:30873179 PMCid:PMC6400988
- 78. Massara, M., O. Bonavita, A. Mantovani, M. Locati and R. Bonecchi: Atypical chemokine receptors in cancer: friends or foes? J Leukoc Biol, 99(6), 927-33 (2016) DOI: 10.1189/jlb.3MR0915-431RR PMid:26908826
- Sarvaiya, P. J., D. Guo, I. Ulasov, P. Gabikian and M. S. Lesniak: Chemokines in tumor progression and metastasis. Oncotarget, 4(12), 2171 (2013)
 DOI: 10.18632/oncotarget.1426
 PMid:24259307 PMCid:PMC3926818
- 80. Murakami, T., A. R. Cardones, S. E. Finkelstein, N. P. Restifo, B. A. Klaunberg, F. O. Nestle, S. S. Castillo, P.

1382

- A. Dennis and S. T. Hwang: Immune evasion by murine melanoma mediated through CC chemokine receptor-10. J Exp Med, 198(9), 1337-47 (2003) DOI: 10.1084/jem.20030593 PMid:14581607 PMCid:PMC2194242
- Sperveslage, J., S. Frank, C. Heneweer, J. Egberts, B. Schniewind, M. Buchholz, F. Bergmann, N. Giese, J. Munding, S. A. Hahn, H. Kalthoff, G. Kloppel and B. Sipos: Lack of CCR7 expression is rate limiting for lymphatic spread of pancreatic ductal adenocarcinoma. Int J Cancer, 131(4), E371-81 (2012) DOI: 10.1002/ijc.26502 PMid:22020953
- 82. Yang, J., S. Wang, G. Zhao and B. Sun: Effect of chemokine receptors CCR7 on disseminated behavior of human T cell lymphoma: clinical and experimental study. J Exp Clin Cancer Res., 30(1), 51 (2011)

DOI: 10.1186/1756-9966-30-51 PMid:21548969 PMCid:PMC3113745

- 83. Dubrovska, A., J. Elliott, R. J. Salamone, G. D. Telegeev, A. E. Stakhovsky, I. B. Schepotin, F. Yan, Y. Wang, L. C. Bouchez, S. A. Kularatne, J. Watson, C. Trussell, V. A. Reddy, C. Y. Cho and P. G. Schultz: CXCR4 expression in prostate cancer progenitor cells. PLoS One, 7(2), e31226 (2012)
 DOI: 10.1371/journal.pone.0031226
 PMid:22359577 PMCid:PMC3281066
- 84. Ali, A. Y., X. Wu, N. Eissa, S. Hou, J.-E. Ghia, T. T. Murooka, V. Banerji, J. B. Johnston, F. Lin, S. B. Gibson and A. J. Marshall: Distinct roles for phosphoinositide 3-kinases γ and δ in malignant B cell migration. Leukemia, 32(9), 1958-1969 (2018)

- DOI: 10.1038/s41375-018-0012-5 PMid:29479062 PMCid:PMC6127087
- 85. Burkle, A., M. Niedermeier, A. Schmitt-Graff, W. G. Wierda, M. J. Keating and J. A. Burger: Overexpression of the CXCR5 chemokine receptor, and its ligand, CXCL13 in B-cell chronic lymphocytic leukemia. Blood, 110(9), 3316-25 (2007) DOI: 10.1182/blood-2007-05-089409 PMid:17652619
- 86. Ribas, C., P. Penela, C. Murga, A. Salcedo, C. Garcia-Hoz, M. Jurado-Pueyo, I. Aymerich and F. Mayor, Jr.: The G protein-coupled receptor kinase (GRK) interactome: role of GRKs in GPCR regulation and signaling. Biochim Biophys Acta, 1768(4), 913-22 (2007) DOI: 10.1016/j.bbamem.2006.09.019 PMid:17084806
- 87. Gurevich, E. V., J. J. Tesmer, A. Mushegian and V. V. Gurevich: G protein-coupled receptor kinases: more than just kinases and not only for GPCRs. Pharmacol Ther, 133(1), 40-69 (2012) DOI: 10.1016/j.pharmthera.2011.08.001 PMid:21903131 PMCid:PMC3241883
- Lefkowitz, R. J. and E. J. Whalen: betaarrestins: traffic cops of cell signaling. Curr Opin Cell Biol, 16(2), 162-8 (2004) DOI: 10.1016/j.ceb.2004.01.001 PMid:15196559
- 89. Luttrell, L. M. and D. Gesty-Palmer: Beyond desensitization: physiological relevance of arrestin-dependent signaling. Pharmacol Rev, 62(2), 305-30 (2010)

DOI: 10.1124/pr.109.002436 PMid:20427692 PMCid:PMC2879915

90. DeWire, S. M., S. Ahn, R. J. Lefkowitz and S. K. Shenoy: β-Arrestins and Cell

Signaling. Annu. Rev. Physiol, 69(1), 483-510 (2007)

DOI: 10.1146/annurev.physiol.69.-

022405.154749 PMid:17305471

 Nelson, C. D., S. J. Perry, D. S. Regier, S. M. Prescott, M. K. Topham and R. J. Lefkowitz: Targeting of diacylglycerol degradation to M1 muscarinic receptors by ß-arrestins. Science, 315(5812), 663-666 (2007)

DOI: 10.1126/science.1134562

PMid:17272726

92. Perry, S. J., G. S. Baillie, T. A. Kohout, I. McPhee, M. M. Magiera, K. L. Ang, W. E. Miller, A. J. McLean, M. Conti and M. D. Houslay: Targeting of cyclic AMP degradation to β2-adrenergic receptors by β-arrestins. Science, 298(5594), 834-836 (2002)

DOI: 10.1126/science.1074683

PMid:12399592

- 93. Lohse, M., S. Andexinger, J. Pitcher, S. Trukawinski, J. Codina, J. Faure, M. G. Caron and R. J. J. o. B. C. Lefkowitz: Receptor-specific desensitization with purified proteins. Kinase dependence and receptor specificity of beta-arrestin and arrestin in the beta 2-adrenergic receptor and rhodopsin systems. J Biol Chem, 267(12), 8558-8564 (1992)
- 94. Ferguson, S. S. J. P. r.: Evolving concepts in G protein-coupled receptor endocytosis: the role in receptor desensitization and signaling. Pharmacol Rev, 53(1), 1-24 (2001)
- 95. Kohout, T. A. and R. J. Lefkowitz: Regulation of G protein-coupled receptor kinases and arrestins during receptor desensitization. Mol Pharmacol, 63(1), 9-18 (2003)

DOI: 10.1124/mol.63.1.9 PMid:12488531

- 96. Schmid, C. L. and L. M. Bohn: Physiological and pharmacological implications of beta-arrestin regulation. Pharmacol Ther, 121(3), 285-93 (2009) DOI: 10.1016/j.pharmthera.2008.11.005 PMid:19100766 PMCid:PMC2656564
- 97. Noma, T., A. Lemaire, S. V. Naga Prasad, L. Barki-Harrington, D. G. Tilley, J. Chen, P. Le Corvoisier, J. D. Violin, H. Wei, R. J. Lefkowitz and H. A. Rockman: Beta-arrestin-mediated beta1-adrenergic receptor transactivation of the EGFR confers cardioprotection. J Clin Invest, 117(9), 2445-58 (2007) DOI: 10.1172/JCI31901

PMid:17786238 PMCid:PMC1952636

98. Watari, K., M. Nakaya and H. Kurose: Multiple functions of G protein-coupled receptor kinases. J Mol Signal, 9(1), 1 (2014)

> DOI: 10.1186/1750-2187-9-1 PMid:24597858 PMCid:PMC3973964

- Komolov, K. E. and J. L. Benovic: G protein-coupled receptor kinases: Past, present and future. Cell Signal, 41, 17-24 (2018)
 DOI: 10.1016/j.cellsig.2017.07.004
 - PMid:28711719 PMCid:PMC5722692
- 100. Vogalis, F., T. Shiraki, D. Kojima, Y. Wada, Y. Nishiwaki, J. L. Jarvinen, J. Sugiyama, K. Kawakami, I. Masai, S. Kawamura, Y. Fukada and T. D. Lamb: Ectopic expression of cone-specific G-protein-coupled receptor kinase GRK7 in zebrafish rods leads to lower photosensitivity and altered responses. J Physiol, 589(Pt 9), 2321-48 (2011) DOI: 10.1113/jphysiol.2010.204156 PMid:21486791 PMCid:PMC3098706

Abbreviations: GPCRs, G protein coupled receptors; PI3Ks, Phosphoinositide 3-kinases; GRKs, G protein coupled receptors kinase; MAPKs, Mitogen-activated protein kinases; JAKs, Janus Kinases; STAT, signal transducer and activator of transcription; ERKs, Extracellular signal-regulated kinases; PKA, Protein kinase A; PKB, Protein kinase B; PKC, Protein kinase C; TFKs, Tec family kinases; ITK, Interleukin-2 inducible T cell kinase; RLK, Resting lymphocyte kinase; BTK, Burton's tyrosine kinase; TEC, Tyrosine kinase expressed in hepatocellular carcinoma; BMX, Bone marrow tyrosine kinase on chromosome-X; TLRs, Toll like receptor; TH, Tec Homology; PH, Pleckstrin Homology; PTK, Protein tyrosine kinase; B-CLL, B-cell chronic lymphocytic leukemia: GTP, Guanosine triphosphate; GDP, Guanosine diphosphate; ATP, Adenosine triphosphate; JNKs, c-Jun NH2terminal protein kinases; EGF, Epidermal derived growth factor; RA, Rheumatoid arthritis; pSS, Primary Sjögren's syndrome; SOCS, Suppressor of cytokine signaling; JH, JAKs homology; WHIM, Warts Hypogammaglobulinemia Infections Myelokathexis; c-SRC, cytosolic Src; CLL, Chronic lymphocytic leukemia; COPD, Chronic obstructive pulmonary diseases; AC, Adenylyl cyclase; DAG, Diacylglycerol; PLC, Phospholipase C; IP3, inositol (1,4,5) triphosphate; FDA, Food and drug associations; MS, Multiple sclerosis; PIP2, phosphatidylinositol-4-5-bisphosphate; PIP3, Phosphatidylinositol-3-4-5-trisphosphate: PTEN, phosphate and tensin homolog: CAMP, Cyclic adenosine mono phosphate; ITIM, Immunoreceptor tyrosine-based inhibitory motif: ITAM immunoreceptor tyrosine-based Activation motif

Key Words: Chemokines, Chemokine receptors, Kinases, G protein coupled receptors, Molecular signaling, Leukocyte trafficking, Review

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