

Expression and role of long non-coding RNA H19 in carcinogenesis

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1. ABSTRACT

With the recent advent of whole genome and transcriptome sequencing technologies, long non-coding RNAs have been brought into the spotlight in molecular biology. H19 was one of the first reported long non-coding RNAs; its expression is high in embryonic organs and absent or greatly reduced in most adult tissues. Accumulating evidence suggests that H19 plays crucial roles in embryogenesis. However, its levels are increased in different cancers, including breast, hepato-gastrointestinal, urological, respiratory, and brain tumors. Although there have been several controversial reports as to whether H19 is oncogenic or tumor-suppressive, most studies have indicated that H19 is associated with growth, migration, invasion, and/or metastasis in many cancers; however, its reported functional mechanisms vary among cancer types. Furthermore, serum H19 levels in patients with certain cancers have been suggested to be useful for diagnosis and prognosis. Thus, H19 long non-coding

RNA might be a candidate for development of promising therapeutic and diagnostic modalities for several cancers. The purpose of this review is to provide an inclusive report on the functional role of H19 in different cancers.

2. INTRODUCTION

Genome-wide transcriptome studies, based on recently developed microarray and next-generation sequencing technologies, have determined that at least 70% of the mammalian genome is actively transcribed and that most transcripts are derived from non-protein-coding genes (1–4). In the classical central dogma of biology, RNA is only an intermediate between DNA and proteins. Therefore, non-coding RNAs were previously considered useless. However, they have recently gained much importance in molecular biology. Increasing evidence suggests the importance of non-coding RNAs in embryogenesis, several physiological

functions, and various diseases (1, 3, 4). Non-coding RNAs are broadly divided into small and long molecules based on the number of ribonucleotides. The former group includes small interfering RNA (siRNA), microRNA (miRNA), piwi-interacting RNA (piRNA), and small nuclear RNA (snRNA), which consists of a ribonucleotide smaller than 200 nt. In contrast, long non-coding RNAs (lncRNAs) range in length from several hundreds to tens of thousands of ribonucleotides. Many of these are mRNA-like transcripts that are transcribed by RNA polymerase II, but fail to encode an open reading frame.

One of the first reported lncRNAs was H19 (1). Human H19 is a 2.3-kb RNA molecule encoded by the *H19* gene located on chromosome 11p15.5. It is known as an imprinted gene expressed only from the maternal allele. This gene was originally discovered by different research groups (5). Pachnes *et al.* identified it by screening a murine fetal liver cDNA library for clones containing RNA sequences whose amounts decrease after birth (6). It was also isolated by Davis *et al.* in a screen to identify myogenic differentiation genes in myocytes, and was initially called MyoH (7). Poirier *et al.* also isolated it as a gene activated during embryonic stem cell differentiation (8). H19 is highly expressed during embryonic development and is repressed shortly after birth (9). Therefore, it has been considered that H19 plays a key role in embryogenesis (5). Furthermore, H19 was found to be re-expressed in a wide variety of tumor types (Figure 1A) and was suggested to have oncogenic or tumor suppressor abilities (1, 9). There are many varying reports in the literature discussing the functional mechanism of H19 lncRNA in the development and metastasis of different cancer types. One such function is as a microRNA precursor (10), as exon 1 of H19 encodes two conserved miRNAs: miR-675-3p and miR-675-5p (Figure 1B). Another contention is that H19 functions as a competing endogenous RNA (ceRNA), acting as a microRNA ‘sponge’ through its miRNA binding sites (Figure 1C) (11). Another mechanism involves its interaction with epigenetic regulatory factors such as polycomb-group proteins (Figure 1D). It is thought that certain lncRNAs might facilitate the recruitment of regulatory factors to the promoter of target genes (12). In the following sections, we describe the profiles of H19 expression in fetal and adult organs and the reported roles and functional mechanisms of H19 based on tumor types.

3. H19 IN FETAL AND ADULT ORGANS

H19 is expressed in most organs during early stages of embryogenesis in humans, mice, cattle, and sheep (5, 13, 14). In human fetal organs, H19 is maximally expressed in the adrenal tissue, muscles, and liver (15). H19 has also been found to be highly expressed in the muscles and kidney in bovine

fetuses, and in the liver, skeletal muscle, and heart in sheep fetuses (13, 14). In contrast, no or significantly decreased expression of H19 has been observed in the brain of human and bovine fetuses (9, 13). Prominent expression of H19 occurs in the placenta, including the amnion, chorion, and allantois, at a much higher level than that in fetal organs (9, 13, 15). In particular, H19 expression is most abundant in intermediate trophoblasts and villous cytotrophoblasts in placental tissues (9, 15). H19 has been reported to regulate trophoblast cell differentiation and proliferation via imprinting linkage with *Igf2* or miR-675 embedded within H19 (16).

Dramatically decreased expression of H19 is detected in most adult tissues (9). Skeletal muscle is the organ with the highest level of H19 expression in adult humans, mice, and cattle (5, 13). Day *et al.* reported that H19 promoted skeletal muscle differentiation and regeneration by giving rise to miR-675-3p and miR-675-5p (17). A fairly high level of H19 expression is maintained in normal adult adrenal tissue (18). In contrast to neoplasms in most other organs, H19 is also highly expressed in adrenocortical adenomas but reduced in carcinomas, suggesting that loss of H19 expression may be associated with malignancy in adrenocortical neoplasms (18).

4. H19 IN CANCER

4.1. H19 in head and neck cancer

Frequent loss of imprinting (LOI) of the *H19* gene has long been identified in head and neck squamous cell carcinoma (HNSCC) (19). A significant correlation has also been demonstrated between H19 expression in HNSCC and recurrence in patients (20). However, in a prediction analysis of microarray data with leave-one-out cross validation, H19 expression was positively correlated with a low risk of recurrence in larynx squamous cell carcinoma (LSCC) (21). Nevertheless, another study indicated that H19 expression in LSCC is inversely correlated with survival in patients (22). The same study also demonstrated that H19 suppressed the activity of miR-148a-3p, thereby enhancing the expression of its target gene, the DNA methyltransferase enzyme DNMT1, which promoted proliferation, migration, and invasion of LSCC cells. In nasopharyngeal carcinoma, H19 lncRNA promoted invasive properties via inhibition of miR-630 and the subsequent regulation of enhancer of zeste homolog 2 (EZH2), which is a target of the miRNA (23). In these cancers, H19 lncRNA acts as an endogenous ‘sponge’ for microRNA.

4.2. H19 in esophageal cancer

LOI with H19 overexpression was observed frequently in esophageal squamous cell carcinoma

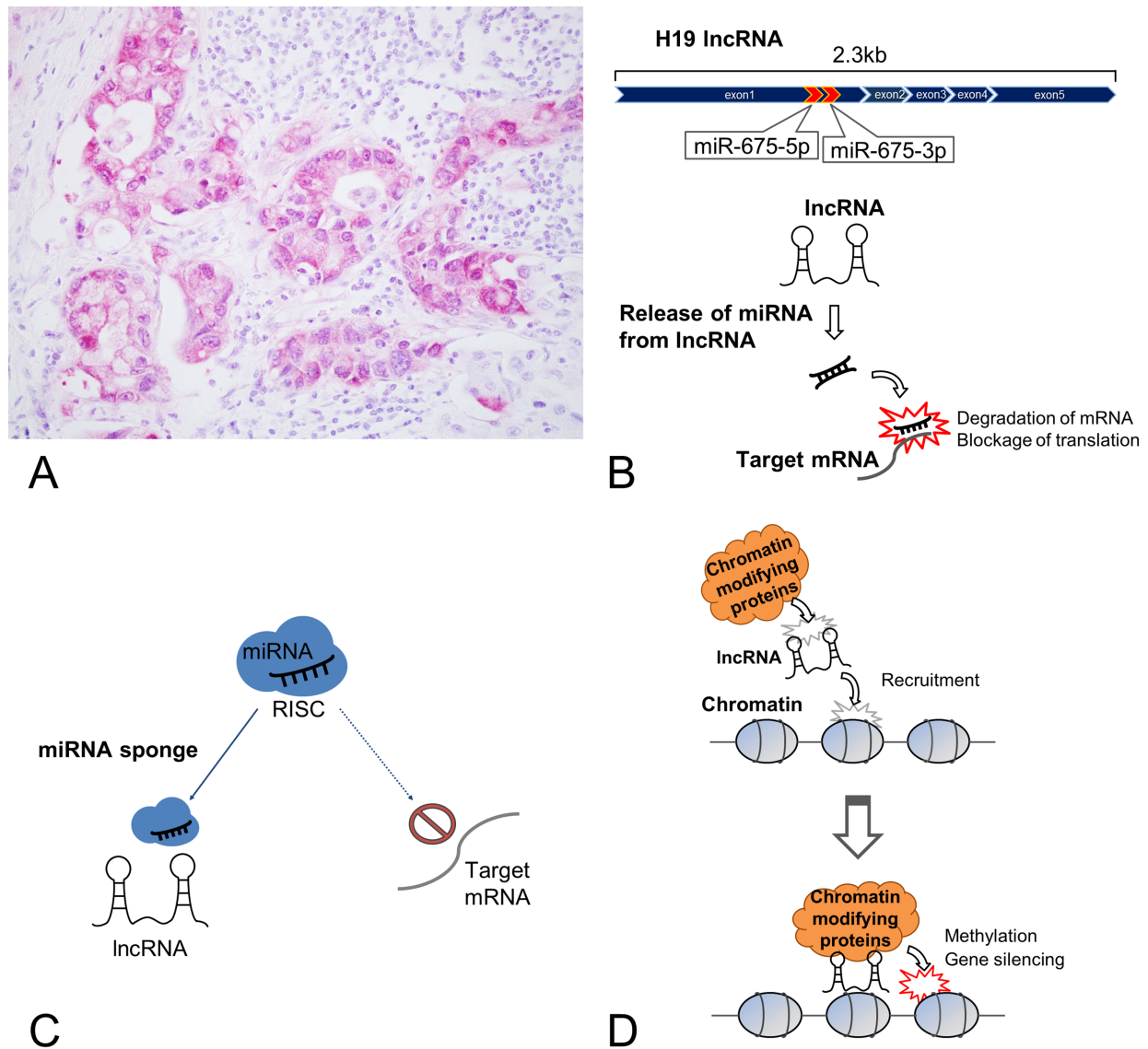


Figure 1. A. H19 lncRNA expressed in human pancreatic carcinoma cells. *In situ* hybridization. B. Some lncRNAs give rise to miRNAs that degrade target mRNAs or block translation. H19 exon1 also encodes two conserved microRNAs, miR-675-3p and miR-675-5p. C. Some lncRNAs share the same miRNA binding sequence with mRNAs. These lncRNAs are considered to act as competing endogenous RNAs (ceRNAs). D. Some lncRNAs can interact with chromatin-modifying proteins such as Polycomb-Repressive Complex 2 (PRC2) that are involved in gene silencing. These lncRNAs are considered to facilitate the recruitment of chromatin modifying proteins to promoter of target genes.

(ESCC) (24). A marked correlation was evident between the expression of H19 and tumor invasion depth, tumor stage, and metastasis (25, 26). H19 lncRNA promoted cell proliferation, migration, invasion, and G0/G1 phase arrest, and induced the epithelial-to-mesenchymal transition (EMT), as indicated by downregulation of the epithelial marker E-cadherin with upregulation of the mesenchymal markers vimentin and fibronectin and metastasis-associated protein MMP-9 in ESCC cells (25, 26). Regarding the functional mechanism of H19 in ESCC tumorigenesis, Zhou *et al.* demonstrated that miR-675-5p encoded within the *H19* gene plays a key role by targeting REPS2 via the RalBP1/RAC1/CDC42 signaling pathway (27). In addition, it was reported that

H19 DMR methylation might play crucial roles in ESCC progression via IGF2 imprinting (28).

4.3. H19 in gastric cancer

Increased levels of H19 in gastric carcinoma (GC) tissues and cells have been reported (29–31). Among the 135 studied lncRNAs, H19 was the most highly upregulated in GC tissue compared to in paired non-tumorous tissues (32). Enhanced H19 expression in GC tissues correlated significantly with poor overall patient survival (29, 30). Surprisingly, H19 levels in gastric juice from GC patients were significantly higher than those from normal subjects (30). Furthermore,

when the levels of lncRNAs were examined in the plasma of GC patients, H19 was found to be significantly upregulated in patients compared to in healthy controls and significantly downregulated in post-operative samples (33–35). Thus, H19 lncRNA might be useful as a diagnostic biomarker for GC.

The role of H19 lncRNA in gastric carcinogenesis has been addressed in several studies. Yang *et al.* reported that H19 was associated with p53 inactivation, which was followed by suppression of apoptosis and increased cell proliferation (31). Zhang *et al.* reported that H19 lncRNA was induced by the oncogene c-Myc and regulated proliferation in GC cells (29). Zhuang *et al.* demonstrated that H19 modulates GC cell proliferation via its mature product miR-675 by targeting the tumor suppressor runt domain transcription factor 1 (RUNX1) (36). Liu *et al.* also confirmed that RUNX1 is a downstream molecule of the H19/miR-675 axis and that inhibition of RUNX1 stimulates activation of the Akt/mTOR pathway to enhance GC cell proliferation and invasion (37). In contrast, Li *et al.* found that H19 promoted proliferation, migration, invasion, and metastasis in GC cells and identified CALN1 as a target gene of H19-derived miR-675. In addition, H19 was suggested to likely play an additional role through direct binding to ISM1 (38). Furthermore, Zhou *et al.* reported that miR-141 could bind to a sequence in H19 and suppress H19 expression in GC cells (39).

4.4. H19 in colorectal cancer

Enhanced expression of H19 has long been known to occur in colorectal carcinomas (CRCs) and has been suggested to result from LOI (40). High H19 expression levels in CRC tissues were significantly correlated with worse overall survival and disease-free survival in patients (11). A recent evaluation of single nucleotide polymorphisms of *H19* provided evidence that *H19* rs2839689 is positively associated with susceptibility to CRC in the Chinese population (41).

Some researchers have proposed a mechanism for the effect of H19 in the progression of CRC. Tsang *et al.* confirmed an inverse expression pattern between H19/miR-675 and the tumor suppressor Rb in human CRC tissues and cell lines. Furthermore, they identified that H19-derived miR-675 targets Rb to increase cell growth and soft-agar colony formation in colon carcinoma cells (10). Liang *et al.* found that overexpression of H19 promoted EMT and tumor growth in colon carcinoma cells (42), and they focused on the function of H19 as a ceRNA in this process. H19 was found to serve as a molecular 'sponge' for miR-138 and miR-200a, targeting mesenchymal marker genes including vimentin, ZEB1, and ZEB2. In contrast, Han *et al.* confirmed that an RNA-binding protein, eukaryotic translation

initiation factor 4A3 (eIF4A3), could bind to H19 lncRNA (11). Combining eIF4A3 with H19 prevented the recruitment of eIF4A3 to cell-cycle-associated mRNA and consequently led to aberrant proliferation of CRC cells.

4.5. H19 in liver cancer

There is contradictory evidence indicating that *H19* can act as an oncogene or a tumor suppressor in hepatocellular carcinoma (HCC). In support of its role as an oncogene, Matouk *et al.* reported that in the Hep3B HCC cell line, H19 expression was elicited in response to hypoxic stress and that knockdown of H19 inhibited tumorigenicity after the cells were subcutaneously injected into nude mice (43). In addition, microarray analysis of H19 knockdown in Hep3B cells after hypoxic stress revealed modulation of the expression of genes involved in angiogenesis, survival, and tumorigenesis. Furthermore, Yang *et al.* demonstrated that high H19 levels in resected HCC tissues were associated with shorter disease-free survival in patients (44).

In contrast, a tumor suppressor role has also been proposed for H19; the results of Zhang *et al.* appeared to directly contrast with the results of the reports described above (45). In particular, Zhang *et al.* showed that H19 expression was lower in intratumoral tissues (T) than in peritumoral tissues (L) and that a low T/L ratio for H19 was an independent predictor of poor outcome in HCC patients. In addition, suppression of H19 expression increased *in vitro* invasion in HCC cells; in addition, intrahepatic metastasis in orthotopic xenograft tumors with reduced H19 expression was greater compared to that in controls. H19 was found to bind to the protein complex hnRNP U/PCAF/RNAPol II to activate the miR-200 family, which consists of EMT suppressive miRNAs, by promoting histone acetylation. Therefore, H19 lncRNA could upregulate the miR-200 family and suppress EMT in HCC cells.

Hernandez *et al.* attempted to solve this paradox (46). They demonstrated that α -fetoprotein-secreting HCCs that are often associated with poor prognosis displayed elevated expression of H19 and its product miR-675. The increased expression of miR-675 led to reduced expression of the tumor suppressor Rb, and therefore stimulated proliferation in HCC cell lines. Furthermore, miR-675 dramatically increased anchorage-independent growth. miR-675 also directly inhibited the expression of a well-known EMT mediator, Twist1, and consequently induced downregulation of the mesenchymal phenotype. This included reduced expression of the mesenchymal cytoskeleton protein vimentin, enhanced expression of the adhesion protein E-cadherin, transformation of cell morphology from a spindle to an epithelioid shape, and reduction of invasive capacity in HCC cells. Although these results were apparently contradictory, the authors concluded

that the epithelial phenotype provided an advantage for cell proliferation and that metastasizing carcinoma cells implanted in secondary organs might induce a mesenchymal-to-epithelial transition (MET) program, required for secondary tumor formation, through the H19/miR-675 pathway.

Recently, Conigliaro *et al.* found that HCC stem-cell-like cells (indicated by CD90 positivity) released exosomes that had the ability to promote tube formation and cell-cell adhesion in endothelial cells (47). LncRNA profiling revealed that the exosomes derived from CD90+ HCC cells were enriched in H19 lncRNA. After upregulation or downregulation of H19 in endothelial cells, it was clarified that H19 plays a critical role in angiogenesis. Thus, H19 lncRNA can be released by cancer stem-like cells via exosomes to potentially affect the cancer microenvironment.

4.6. H19 in bladder cancer

H19 expression in bladder carcinoma (BC) has long been suggested to be useful as a prognostic marker for early recurrence (48–50). SNP polymorphisms in the *H19* gene have been reported to be associated with an increased or decreased risk of suffering BC (51, 52).

Matouk *et al.* showed that overexpression of H19 in BC cells enhanced tumor growth after the cells were subcutaneously injected into mice (43). Additionally, H19 silencing using siRNA decreased tumor volumes *in vivo*. Luo *et al.* reported that H19 lncRNA increased cell migration through its association with EZH2, which led to activation of the Wnt/ β -catenin pathway and subsequent inhibition of E-cadherin expression in BC cells (12). They also demonstrated that H19 increased BC cell proliferation through enhanced expression of the DNA-binding protein inhibitor ID2 (53). Liu *et al.* confirmed that miR-675 derived from H19 lncRNA promoted BC cell proliferation through induction of G1 arrest and suppression of cell apoptosis, which was likely dependent on its negative regulation of the tumor suppressor p53 (54). The Yes-associated protein 1 (YAP1) oncogene was identified by Li *et al.* as an upstream gene that induces H19 lncRNA expression (55).

4.7. H19 in renal cancer

It has been shown that epigenetic H19 silencing occurs as an early event in the tumorigenesis of Wilms tumors (56). Therefore, H19 was originally thought to be a tumor suppressor in renal neoplasms (57). However, Wang *et al.* showed that clear cell renal carcinoma (ccRC) tissues had significantly higher levels of H19 compared to adjacent normal tissues and that silencing of H19 reduced cell proliferation, migration, and invasion of RC cells (58). Furthermore,

patients bearing ccRC with higher H19 expression had poorer overall survival.

4.8. H19 in lung cancer

LOI of H19 leading to its overexpression has been described as a frequent event during the development of lung carcinoma (LC) (59). Higher expression of H19 is associated with advanced tumor-node-metastasis stage and poorer survival in LC patients (60, 61). H19 knockdown experiments revealed that H19 is involved in cell proliferation, clonogenicity, and anchorage-independent growth in LC (60, 62). Furthermore, H19 was shown to promote cell cycle progression by downregulating miR-107 (63).

As in BC and GC cells, H19 is known to be directly regulated by c-Myc in LC cells (60, 62, 63). Moreover, mineral dust-induced gene (*mdig*) was reported to demethylate H3K9me3 on the histone H3 peptide, a key regulator and epigenetic marker of heterochromatin, which was followed by the de-repression of H19 lncRNA (61). In a statistical study of lncRNA genetic polymorphisms, H19 rs2107425 was strongly associated with LC susceptibility in individuals under 50 years of age and H19 rs2839698 showed a relationship with platinum-based chemotherapy response in small-cell LC (64).

4.9. H19 in breast cancer

In human breast carcinomas (BCs), the expression of H19 lncRNA is evident in cancer cells or stromal cells (65). Adriaenssens *et al.* performed *in situ* hybridization (ISH) with 102 BCs and found that the *H19* gene was obviously expressed at a higher level in 74 cases (72.5%) when compared to expression in normal breast tissue. H19 upregulation in BCs was significantly correlated with T values (TNM classification). However, of these carcinomas, only eight cases (7.8%) exhibited overexpression of H19 in the carcinoma cells themselves, whereas in 99 cases (97.1%), expression occurred in stromal cells (66). In BCs, H19 lncRNA might contribute to carcinogenesis through the formation of cancer-associated fibroblasts.

Although H19 overexpression in BC cells was detected at a lower frequency than expected, this phenomenon seemed to be important for cancer progression. Multidrug-resistant MCF-7/AdrVp BC cells display abundant expression of H19 relative to parental MCF-7 cells (67). Moreover, in an *in vitro* study using MDA-MB-231 BC cells, a clone stably transfected with the genomic sequence of the human H19 gene was shown to form more and larger colonies in soft agar during anchorage-independent growth assays and formed more and larger subcutaneous tumors after injection into *scid* mice (68). A follow-up study from the same research group showed that H19

promotes the G1-S transition, as the H19 promoter is activated by E2F1 in BC cells (69). Furthermore, the same research group identified that miR-675, derived from H19 lncRNA, enhanced proliferation and migration in BC cells by downregulating genes of the ubiquitin ligase E3 family (70). Barsyte-Lovejoy *et al.* demonstrated a strong association between c-Myc and H19 expression in samples from BC patients (62). The product of the *MYC* oncogene directly binds to conserved E-boxes at the *H19* promoter and enhances the transcription of the maternal H19 allele. In addition, knockdown of H19 expression significantly decreased clonogenicity and anchorage-independent growth in BC cells.

A relationship between the expression of H19 and hormone receptors has been suggested in BC. H19 overexpression in BCs was significantly correlated with the presence of both estrogen and progesterone receptors (66). H19 expression was shown to depend on the estrogen receptor, and H19 was suggested to mediate estrogen-induced cell proliferation in BC cells (71). H19 was also reported to be associated with luminal progenitor cell differentiation that is regulated by estrogen (72).

Recently, Zhang *et al.* evaluated whether circulating H19 RNA in the plasma could serve as a novel biomarker for the diagnosis and monitoring of BC (73). H19 levels were significantly increased in plasma from BC patients compared to that from healthy volunteers and the value of H19 as a biomarker was higher than that of carcinoembryonic antigen and carbohydrate antigen 153.

4.10. H19 in cervical cancer

According to a previous report, LOI of the *H19* gene occurred in 36% and 100% of informative cervical and endometrial carcinoma cases, respectively (74). When H19 expression was assessed in cervical intraepithelial neoplasia 3 (CIN3) samples by *in situ* hybridization, the signals were present exclusively in areas of CIN3 that were within the cervical epithelium (75). In cervical carcinoma cells, H19 lncRNA was overexpressed and promoted cell proliferation and sphere formation, but did not affect apoptosis and migration (76). H19 expression was regulated by TGF- β 1 treatment or the hypoxia inducer CoCl₂ in a cell line-specific manner. In addition, extracellular vesicles released from cervical carcinoma cells into the culture medium contained H19 lncRNA.

4.11. H19 in brain neoplasms

Imprinting status varies in different types of brain tumors (77–79). Shi *et al.* discovered high expression of H19 lncRNA in high-grade gliomas and

found that H19 stimulated glioblastoma cell invasion by modulating the expression of Cadherin13, which was a direct target of miR-675 derived from H19 (80). Jiang *et al.* reported that H19 expression levels in glioblastoma tissues were associated with patient survival (81). H19 was shown to promote cell invasion in Matrigel assays, angiogenesis in endothelial tube formation assays, and tumorigenicity in a xenograft mouse model. In addition, H19 was suggested to be correlated with stemness based on the enrichment of H19 lncRNA in stem cell marker CD133+ glioblastoma cells and based on the high degree of sphere formation in H19-overexpressing cells. Li *et al.* later observed low cell proliferation and high rates of apoptosis, in addition to downregulation of the stem cell markers CD133, NANOG, Oct4, and Sox2, in H19-deficient glioblastoma cells (82). Jiang *et al.* also demonstrated decreased stemness in H19-deficient glioblastoma cells. In addition, the observed enhancement of the ability of tumor suppressor let-7 miRNA to inhibit the expression of its target HMGA2 oncogene was related to the self-renewal of cancer stem cells (83). The function of H19 as a let-7 ‘sponge’ was responsible for this phenomenon.

5. CONCLUSION

Although the oncogene or tumor suppressor status of H19 remains controversial in some cancers, H19 is highly expressed and correlated with growth, migration, invasion, and/or metastasis in most types of cancer. Restricted expression of H19 has been shown in adult normal tissues of several organs. These findings indicate that this lncRNA might serve as a novel candidate for molecular targeted therapy, using nucleic acid-based medicine, for several cancers. Furthermore, recent studies have suggested that H19 lncRNA is released from a wide variety of cancers and is detectable in the sera of patients as a stable form that is protected from endogenous RNases. Thus, circulating H19 lncRNA could be a promising new biomarker for the early detection or prognosis of cancers.

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7. REFERENCES

1. E. A. Gibb, C. J. Brown and W. L. Lam: The functional role of long non-coding RNA in human carcinomas. *Mol Cancer* 10, 38 (2011)
DOI: 10.1186/1476-4598-10-38

2. L. D. Stein: Human genome: end of the beginning. *Nature* 431, 915–916 (2004)
DOI: 10.1038/431915a
3. X. Shi, M. Sun, H. Liu, Y. Yao and Y. Song: Long non-coding RNAs: a new frontier in the study of human diseases. *Cancer Lett* 339, 159–166 (2013)
DOI: 10.1016/j.canlet.2013.06.013
4. J. Sana, P. Faltejskova, M. Svoboda and O. Slaby: Novel classes of non-coding RNAs and cancer. *J Transl Med* 10, 103 (2012)
DOI: 10.1186/1479-5876-10-103
5. A. Gabory, H. Jammes and L. Dandolo: The H19 locus: role of an imprinted non-coding RNA in growth and development. *Bioessays* 32, 473–480 (2010)
DOI: 10.1002/bies.200900170
6. V. Pachnis, A. Belayew and S. M. Tilghman: Locus unlinked to alpha-fetoprotein under the control of the murine raf and Rif genes. *Proc Natl Acad Sci U S A* 81, 5523–5527 (1984)
7. R. L. Davis, H. Weintraub and A. B. Lassar: Expression of a single transfected cDNA converts fibroblasts to myoblasts. *Cell* 51, 987–1000 (1987)
DOI: 10.1016/0092-8674(87)90585-X
8. F. Poirier, C. T. Chan, P. M. Timmons, E. J. Robertson, M. J. Evans and P. W. Rigby: The murine H19 gene is activated during embryonic stem cell differentiation *in vitro* and at the time of implantation in the developing embryo. *Development* 113, 1105–1114 (1991)
9. I. Ariel, S. Ayesh, E. J. Perlman, G. Pizov, V. Tanos, T. Schneider, V. A. Erdmann, D. Podeh, D. Komitowski, A. S. Quasem, N. de Groot and A. Hochberg: The product of the imprinted H19 gene is an oncofetal RNA. *Mol Pathol* 50, 34–44 (1997)
DOI: 10.1136/mp.50.1.34
10. W. P. Tsang, E. K. Ng, S. S. Ng, H. Jin, J. Yu, J. J. Sung and T. T. Kwok: Oncofetal H19-derived miR-675 regulates tumor suppressor RB in human colorectal cancer. *Carcinogenesis* 31, 350–358 (2010)
DOI: 10.1093/carcin/bgp181
11. D. Han, X. Gao, M. Wang, Y. Qiao, Y. Xu, J. Yang, N. Dong, J. He, Q. Sun, G. Lv, C. Xu, J. Tao and N. Ma: Long noncoding RNA H19 indicates a poor prognosis of colorectal cancer and promotes tumor growth by recruiting and binding to eIF4A3. *Oncotarget* 7, 22159–22173 (2016)
DOI: 10.18632/oncotarget.8063
12. M. Luo, Z. Li, W. Wang, Y. Zeng, Z. Liu and J. Qiu: Long non-coding RNA H19 increases bladder cancer metastasis by associating with EZH2 and inhibiting E-cadherin expression. *Cancer Lett* 333, 213–221 (2013)
DOI: 10.1016/j.canlet.2013.01.033
13. H. Khatib and V. Schutzkus: The expression profile of the H19 gene in cattle. *Mamm Genome* 17, 991–996 (2006)
DOI: 10.1007/s00335-006-0038-2
14. L. G. Naimeh, B. C. Schutte, W. S. Hamilton and E. Tsalikian: Ontogeny of the H19 gene in sheep and effect of maternal fasting on its expression in the fetus. *Endocr Res* 27, 417–431 (2001)
15. R. Goshen, J. Rachmilewitz, T. Schneider, N. de-Groot, I. Ariel, Z. Palti and A. A. Hochberg: The expression of the H-19 and IGF-2 genes during human embryogenesis and placental development. *Mol Reprod Dev* 34, 374–379 (1993)
DOI: 10.1002/mrd.1080340405
16. W. L. Gao, M. Liu, Y. Yang, H. Yang, Q. Liao, Y. Bai, Y. X. Li, D. Li, C. Peng and Y. L. Wang: The imprinted H19 gene regulates human placental trophoblast cell proliferation via encoding miR-675 that targets Nodal Modulator 1 (NOMO1). *RNA Biol* 9, 1002–1010 (2012)
DOI: 10.4161/rna.20807
17. B. K. Dey, K. Pfeifer and A. Dutta: The H19 long noncoding RNA gives rise to microRNAs miR-675–3p and miR-675–5p to promote skeletal muscle differentiation and regeneration. *Genes Dev* 28, 491–501 (2014)
DOI: 10.1101/gad.234419.113
18. J. Liu, A. I. Kahri, P. Heikkila, V. Ilvesmaki and R. Voutilainen: H19 and insulin-like growth factor-II gene expression in adrenal tumors and cultured adrenal cells. *J Clin Endocrinol Metab* 80, 492–496 (1995)
DOI: 10.1210/jcem.80.2.7531713

19. A. K. el-Naggar, S. Lai, S. A. Tucker, G. L. Clayman, H. Goepfert, W. K. Hong and V. Huff: Frequent loss of imprinting at the IGF2 and H19 genes in head and neck squamous carcinoma. *Oncogene* 18, 7063–7069 (1999)
DOI: 10.1038/sj.onc.1203192
20. L. I. Esteves, A. C. Javaroni, I. N. Nishimoto, J. Magrin, J. A. Squire, L. P. Kowalski, C. A. Rainho and S. R. Rogatto: DNA methylation in the CTCF-binding site I and the expression pattern of the H19 gene: does positive expression predict poor prognosis in early stage head and neck carcinomas? *Mol Carcinog* 44, 102–110 (2005)
DOI: 10.1002/mc.20126
21. V. Mirisola, R. Mora, A. I. Esposito, L. Guastini, F. Tabacchiera, L. Paleari, A. Amaro, G. Angelini, M. Dellepiane, U. Pfeffer and A. Salami: A prognostic multigene classifier for squamous cell carcinomas of the larynx. *Cancer Lett* 307, 37–46 (2011)
DOI: 10.1016/j.canlet.2011.03.013
22. T. Wu, L. Qu, G. He, L. Tian, L. Li, H. Zhou, Q. Jin, J. Ren, Y. Wang, J. Wang, X. Kan, M. Liu, J. Shen, M. Guo and Y. Sun: Regulation of laryngeal squamous cell cancer progression by the lncRNA H19/miR-148a-3p/DNMT1 axis. *Oncotarget* 7, 11553–11566 (2016)
DOI: 10.18632/oncotarget.7270
23. X. Li, Y. Lin, X. Yang, X. Wu and X. He: Long noncoding RNA H19 regulates EZH2 expression by interacting with miR-630 and promotes cell invasion in nasopharyngeal carcinoma. *Biochem Biophys Res Commun* 473, 913–919 (2016)
DOI: 10.1016/j.bbrc.2016.03.150
24. K. Hibi, H. Nakamura, A. Hirai, Y. Fujikake, Y. Kasai, S. Akiyama, K. Ito and H. Takagi: Loss of H19 imprinting in esophageal cancer. *Cancer Res* 56, 480–482 (1996)
25. D. Tan, Y. Wu, L. Hu, P. He, G. Xiong, Y. Bai and K. Yang: Long noncoding RNA H19 is up-regulated in esophageal squamous cell carcinoma and promotes cell proliferation and metastasis. *Dis Esophagus* (2016)
DOI: 10.1111/dote.12481
26. C. Huang, L. Cao, L. Qiu, X. Dai, L. Ma, Y. Zhou, H. Li, M. Gao, W. Li, Q. Zhang, K. Han and H. Lv: Upregulation of H19 promotes invasion and induces epithelial-to-mesenchymal transition in esophageal cancer. *Oncol Lett* 10, 291–296 (2015)
DOI: 10.3892/ol.2015.3165
27. Y. W. Zhou, H. Zhang, C. J. Duan, Y. Gao, Y. D. Cheng, D. He, R. Li and C. F. Zhang: miR-675-5p enhances tumorigenesis and metastasis of esophageal squamous cell carcinoma by targeting REPS2. *Oncotarget* 7, 30730–30747 (2016)
DOI: 10.18632/oncotarget.8950
28. T. Gao, B. He, Y. Pan, L. Gu, L. Chen, Z. Nie, Y. Xu, R. Li and S. Wang: H19 DMR methylation correlates to the progression of esophageal squamous cell carcinoma through IGF2 imprinting pathway. *Clin Transl Oncol* 16, 410–417 (2014)
DOI: 10.1007/s12094-013-1098-x
29. E. B. Zhang, L. Han, D. D. Yin, R. Kong, W. De and J. Chen: c-Myc-induced, long, noncoding H19 affects cell proliferation and predicts a poor prognosis in patients with gastric cancer. *Med Oncol* 31, 914 (2014)
DOI: 10.1007/s12032-014-0914-7
30. J. S. Chen, Y. F. Wang, X. Q. Zhang, J. M. Lv, Y. Li, X. X. Liu and T. P. Xu: H19 serves as a diagnostic biomarker and up-regulation of H19 expression contributes to poor prognosis in patients with gastric cancer. *Neoplasia* 63, 223–230 (2016)
DOI: 10.4149/207_150821N454
31. F. Yang, J. Bi, X. Xue, L. Zheng, K. Zhi, J. Hua and G. Fang: Up-regulated long non-coding RNA H19 contributes to proliferation of gastric cancer cells. *Febs J* 279, 3159–3165 (2012)
DOI: 10.1111/j.1742-4658.2.012.0.8694.x
32. H. Song, W. Sun, G. Ye, X. Ding, Z. Liu, S. Zhang, T. Xia, B. Xiao, Y. Xi and J. Guo: Long non-coding RNA expression profile in human gastric cancer and its clinical significances. *J Transl Med* 11, 225 (2013)
DOI: 10.1186/1479-5876-11-225
33. D. Hashad, A. Elbanna, A. Ibrahim and G. Khedr: Evaluation of the Role of Circulating Long Non-Coding RNA H19 as a Promising Novel Biomarker in Plasma of Patients with Gastric Cancer. *J Clin Lab Anal* (2016)
DOI: 10.1002/jcla.21987
34. X. Zhou, C. Yin, Y. Dang, F. Ye and G. Zhang: Identification of the long non-coding

- RNA H19 in plasma as a novel biomarker for diagnosis of gastric cancer. *Sci Rep* 5, 11516 (2015)
DOI: 10.1038/srep11516
35. T. Arita, D. Ichikawa, H. Konishi, S. Komatsu, A. Shiozaki, K. Shoda, T. Kawaguchi, S. Hirajima, H. Nagata, T. Kubota, H. Fujiwara, K. Okamoto and E. Otsuji: Circulating long non-coding RNAs in plasma of patients with gastric cancer. *Anticancer Res* 33, 3185–3193 (2013)
36. M. Zhuang, W. Gao, J. Xu, P. Wang and Y. Shu: The long non-coding RNA H19-derived miR-675 modulates human gastric cancer cell proliferation by targeting tumor suppressor RUNX1. *Biochem Biophys Res Commun* 448, 315–322 (2014)
DOI: 10.1016/j.bbrc.2013.12.126
37. G. Liu, T. Xiang, Q. F. Wu and W. X. Wang: Long Noncoding RNA H19-Derived miR-675 Enhances Proliferation and Invasion via RUNX1 in Gastric Cancer Cells. *Oncol Res* 23, 99–107 (2016)
DOI: 10.3727/096504015x14496932933575
38. H. Li, B. Yu, J. Li, L. Su, M. Yan, Z. Zhu and B. Liu: Overexpression of lncRNA H19 enhances carcinogenesis and metastasis of gastric cancer. *Oncotarget* 5, 2318–2329 (2014)
DOI: 10.18632/oncotarget.1913
39. X. Zhou, F. Ye, C. Yin, Y. Zhuang, G. Yue and G. Zhang: The Interaction Between MiR-141 and lncRNA-H19 in Regulating Cell Proliferation and Migration in Gastric Cancer. *Cell Physiol Biochem* 36, 1440–1452 (2015)
DOI: 10.1159/000430309
40. H. Cui, P. Onyango, S. Brandenburg, Y. Wu, C. L. Hsieh and A. P. Feinberg: Loss of imprinting in colorectal cancer linked to hypomethylation of H19 and IGF2. *Cancer Res* 62, 6442–6446 (2002)
41. S. Li, Y. Hua, J. Jin, H. Wang, M. Du, L. Zhu, H. Chu, Z. Zhang and M. Wang: Association of genetic variants in lncRNA H19 with risk of colorectal cancer in a Chinese population. *Oncotarget* 7, 25470–25477 (2016)
DOI: 10.18632/oncotarget.8330
42. W. C. Liang, W. M. Fu, C. W. Wong, Y. Wang, W. M. Wang, G. X. Hu, L. Zhang, L. J. Xiao, D. C. Wan, J. F. Zhang and M. M. Waye: The lncRNA H19 promotes epithelial to mesenchymal transition by functioning as miRNA sponges in colorectal cancer. *Oncotarget* 6, 22513–22525 (2015)
DOI: 10.18632/oncotarget.4154
43. I. J. Matouk, N. DeGroot, S. Mezan, S. Ayeshe, R. Abu-lail, A. Hochberg and E. Galun: The H19 non-coding RNA is essential for human tumor growth. *PLoS One* 2, e845 (2007)
DOI: 10.1371/journal.pone.0000845
44. Z. Yang, Y. Lu, Q. Xu, B. Tang, C. K. Park and X. Chen: HULC and H19 Played Different Roles in Overall and Disease-Free Survival from Hepatocellular Carcinoma after Curative Hepatectomy: A Preliminary Analysis from Gene Expression Omnibus. *Dis Markers* 2015, 191029 (2015)
DOI: 10.1155/2015/191029
45. L. Zhang, F. Yang, J. H. Yuan, S. X. Yuan, W. P. Zhou, X. S. Huo, D. Xu, H. S. Bi, F. Wang and S. H. Sun: Epigenetic activation of the MiR-200 family contributes to H19-mediated metastasis suppression in hepatocellular carcinoma. *Carcinogenesis* 34, 577–586 (2013)
DOI: 10.1093/carcin/bgs381
46. J. M. Hernandez, A. Elahi, C. W. Clark, J. Wang, L. A. Humphries, B. Centeno, G. Bloom, B. C. Fuchs, T. Yeatman and D. Shibata: miR-675 mediates downregulation of Twist1 and Rb in AFP-secreting hepatocellular carcinoma. *Ann Surg Oncol* 20 Suppl 3, S625–635 (2013)
DOI: 10.1245/s10434-013-3106-3
47. A. Conigliaro, V. Costa, A. Lo Dico, L. Saieva, S. Buccheri, F. Dieli, M. Manno, S. Raccosta, C. Mancone, M. Tripodi, G. De Leo and R. Alessandro: CD90+ liver cancer cells modulate endothelial cell phenotype through the release of exosomes containing H19 lncRNA. *Mol Cancer* 14, 155 (2015)
DOI: 10.1186/s12943-015-0426-x
48. I. Ariel, M. Sughayer, Y. Fellig, G. Pizov, S. Ayeshe, D. Podeh, B. A. Libdeh, C. Levy, T. Birman, M. L. Tykocinski, N. de Groot and A. Hochberg: The imprinted H19 gene is a marker of early recurrence in human bladder carcinoma. *Mol Pathol* 53, 320–323 (2000)
49. M. J. Cooper, M. Fischer, D. Komitowski, A. Shevelev, E. Schulze, I. Ariel, M. L. Tykocinski, S. Miron, J. Ilan, N. de Groot and

- A. Hochberg: Developmentally imprinted genes as markers for bladder tumor progression. *J Urol* 155, 2120–2127 (1996)
DOI: 10.1016/S0022-5347(01)66120-2
50. I. Ariel, O. Lustig, T. Schneider, G. Pizov, M. Sappir, N. De-Groot and A. Hochberg: The imprinted H19 gene as a tumor marker in bladder carcinoma. *Urology* 45, 335–338 (1995)
DOI: 10.1016/0090-4295(95)80030-1
51. Q. Hua, X. Lv, X. Gu, Y. Chen, H. Chu, M. Du, W. Gong, M. Wang and Z. Zhang: Genetic variants in lncRNA H19 are associated with the risk of bladder cancer in a Chinese population. *Mutagenesis* 31, 531–538 (2016)
DOI: 10.1093/mutage/gew018
52. G. W. Verhaegh, L. Verkleij, S. H. Vermeulen, M. den Heijer, J. A. Witjes and L. A. Kiemeny: Polymorphisms in the H19 gene and the risk of bladder cancer. *Eur Urol* 54, 1118–1126 (2008)
DOI: 10.1016/j.eururo.2008.01.060
53. M. Luo, Z. Li, W. Wang, Y. Zeng, Z. Liu and J. Qiu: Upregulated H19 contributes to bladder cancer cell proliferation by regulating ID2 expression. *Febs J* 280, 1709–1716 (2013)
DOI: 10.1111/febs.12185
54. C. Liu, Z. Chen, J. Fang, A. Xu, W. Zhang and Z. Wang: H19-derived miR-675 contributes to bladder cancer cell proliferation by regulating p53 activation. *Tumour Biol* 37, 263–270 (2016)
DOI: 10.1007/s13277-015-3779-2
55. S. Li, Z. Yu, S. S. Chen, F. Li, C. Y. Lei, X. X. Chen, J. M. Bao, Y. Luo, G. Z. Lin, S. Y. Pang and W. L. Tan: The YAP1 oncogene contributes to bladder cancer cell proliferation and migration by regulating the H19 long noncoding RNA. *Urol Oncol* 33, 427.e421–410 (2015)
DOI: 10.1016/j.urolonc.2015.06.003
56. M. A. Frevel, S. J. Sowerby, G. B. Petersen and A. E. Reeve: Methylation sequencing analysis refines the region of H19 epimutation in Wilms tumor. *J Biol Chem* 274, 29331–29340 (1999)
DOI: 10.1074/jbc.274.41.29331
57. S. Zhou, J. Wang and Z. Zhang: An emerging understanding of long noncoding RNAs in kidney cancer. *J Cancer Res Clin Oncol* 140, 1989–1995 (2014) doi:10.1007/s00432-014-1699-y
DOI: 10.1007/s00432-014-1699-y
58. L. Wang, Y. Cai, X. Zhao, X. Jia, J. Zhang, J. Liu, H. Zhen, T. Wang, X. Tang, Y. Liu and J. Wang: Down-regulated long non-coding RNA H19 inhibits carcinogenesis of renal cell carcinoma. *Neoplasma* 62, 412–418 (2015)
DOI: 10.4149/neo_2015_049
59. M. Kondo, H. Suzuki, R. Ueda, H. Osada, K. Takagi, T. Takahashi and T. Takahashi: Frequent loss of imprinting of the H19 gene is often associated with its overexpression in human lung cancers. *Oncogene* 10, 1193–1198 (1995)
60. E. Zhang, W. Li, D. Yin, W. De, L. Zhu, S. Sun and L. Han: c-Myc-regulated long non-coding RNA H19 indicates a poor prognosis and affects cell proliferation in non-small-cell lung cancer. *Tumour Biol* 37, 4007–4015 (2016)
DOI: 10.1007/s13277-015-4185-5
61. B. Chen, M. Yu, Q. Chang, Y. Lu, C. Thakur, D. Ma, Z. Yi and F. Chen: Mdig de-represses H19 large intergenic non-coding RNA (lincRNA) by down-regulating H3K9me3 and heterochromatin. *Oncotarget* 4, 1427–1437 (2013)
DOI: 10.18632/oncotarget.1155
62. D. Barsyte-Lovejoy, S. K. Lau, P. C. Boutros, F. Khosravi, I. Jurisica, I. L. Andrusis, M. S. Tsao and L. Z. Penn: The c-Myc oncogene directly induces the H19 noncoding RNA by allele-specific binding to potentiate tumorigenesis. *Cancer Res* 66, 5330–5337 (2006)
DOI: 10.1158/0008-5472.can-06-0037
63. J. Cui, J. Mo, M. Luo, Q. Yu, S. Zhou, T. Li, Y. Zhang and W. Luo: c-Myc-activated long non-coding RNA H19 downregulates miR-107 and promotes cell cycle progression of non-small cell lung cancer. *Int J Clin Exp Pathol* 8, 12400–12409 (2015)
64. W. J. Gong, J. Y. Yin, X. P. Li, C. Fang, D. Xiao, W. Zhang, H. H. Zhou, X. Li and Z. Q. Liu: Association of well-characterized lung cancer lncRNA polymorphisms with lung cancer susceptibility and platinum-based chemotherapy response. *Tumour Biol* 37, 8349–8358 (2016)
DOI: 10.1007/s13277-015-4497-5

65. T. Dugimont, J. J. Curgy, N. Wernert, A. Delobelle, M. B. Raes, A. Joubel, D. Stehelin and J. Coll: The H19 gene is expressed within both epithelial and stromal components of human invasive adenocarcinomas. *Biol Cell* 85, 117–124 (1995)
DOI: 10.1016/0248-4900(96)85272-5
66. E. Adriaenssens, L. Dumont, S. Lottin, D. Bolle, A. Lepretre, A. Delobelle, F. Bouali, T. Dugimont, J. Coll and J. J. Curgy: H19 overexpression in breast adenocarcinoma stromal cells is associated with tumor values and steroid receptor status but independent of p53 and Ki-67 expression. *Am J Pathol* 153, 1597–1607 (1998)
DOI: 10.1016/s0002-9440(10)65748-3
67. L. A. Doyle, W. Yang, A. K. Rishi, Y. Gao and D. D. Ross: H19 gene overexpression in atypical multidrug-resistant cells associated with expression of a 95-kilodalton membrane glycoprotein. *Cancer Res* 56, 2904–2907 (1996)
68. S. Lottin, E. Adriaenssens, T. Dupressoir, N. Berteaux, C. Montpellier, J. Coll, T. Dugimont and J. J. Curgy: Overexpression of an ectopic H19 gene enhances the tumorigenic properties of breast cancer cells. *Carcinogenesis* 23, 1885–1895 (2002)
DOI: 10.1093/carcin/23.11.1885
69. N. Berteaux, S. Lottin, D. Monte, S. Pinte, B. Quatannens, J. Coll, H. Hondermarck, J. J. Curgy, T. Dugimont and E. Adriaenssens: H19 mRNA-like noncoding RNA promotes breast cancer cell proliferation through positive control by E2F1. *J Biol Chem* 280, 29625–29636 (2005)
DOI: 10.1074/jbc.M504033200
70. C. Vennin, N. Spruyt, F. Dahmani, S. Julien, F. Bertucci, P. Finetti, T. Chassat, R. P. Bourette, X. Le Bourhis and E. Adriaenssens: H19 non coding RNA-derived miR-675 enhances tumorigenesis and metastasis of breast cancer cells by downregulating c-Cbl and Cbl-b. *Oncotarget* 6, 29209–29223 (2015)
DOI: 10.18632/oncotarget.4976
71. H. Sun, G. Wang, Y. Peng, Y. Zeng, Q. N. Zhu, T. L. Li, J. Q. Cai, H. H. Zhou and Y. S. Zhu: H19 lncRNA mediates 17 β -estradiol-induced cell proliferation in MCF-7 breast cancer cells. *Oncol Rep* 33, 3045–3052 (2015)
DOI: 10.3892/or.2015.3899
72. P. Basak, S. Chatterjee, S. Weger, M. C. Bruce, L. C. Murphy and A. Raouf: Estrogen regulates luminal progenitor cell differentiation through H19 gene expression. *Endocr Relat Cancer* 22, 505–517 (2015)
DOI: 10.1530/ERC-15-0105
73. K. Zhang, Z. Luo, Y. Zhang, L. Zhang, L. Wu, L. Liu, J. Yang, X. Song and J. Liu: Circulating lncRNA H19 in plasma as a novel biomarker for breast cancer. *Cancer Biomark* 17, 187–194 (2016)
DOI: 10.3233/cbm-160630
74. C. Lee, S. J. Kim, J. Y. Na, C. S. Park, S. Y. Lee, I. H. Kim and Y. K. Oh: Alterations in Promoter Usage and Expression Levels of Insulin-like Growth Factor-II and H19 Genes in Cervical and Endometrial Cancer. *Cancer Res Treat* 35, 314–322 (2003)
DOI: 10.4143/crt.2003.35.4.314
75. T. Feigenberg, O. N. Gofrit, G. Pizov, A. Hochberg and A. Benshushan: Expression of the h19 oncofetal gene in premalignant lesions of cervical cancer: a potential targeting approach for development of nonsurgical treatment of high-risk lesions. *ISRN Obstet Gynecol* 2013, 137509 (2013)
DOI: 10.1155/2013/137509
76. T. Iempridee: Long non-coding RNA H19 enhances cell proliferation and anchorage-independent growth of cervical cancer cell lines. *Exp Biol Med (Maywood)* (2016)
DOI: 10.1177/1535370216670542
77. S. Muller, D. Zirkel, M. Westphal and W. Zumkeller: Genomic imprinting of IGF2 and H19 in human meningiomas. *Eur J Cancer* 36, 651–655 (2000)
DOI: 10.1016/S0959-8049(99)00328-7
78. S. Uyeno, Y. Aoki, M. Nata, K. Sagisaka, T. Kayama, T. Yoshimoto and T. Ono: IGF2 but not H19 shows loss of imprinting in human glioma. *Cancer Res* 56, 5356–5359 (1996)
79. S. Albrecht, A. Waha, A. Koch, J. A. Kraus, C. G. Goodyer and T. Pietsch: Variable imprinting of H19 and IGF2 in fetal cerebellum and medulloblastoma. *J Neuropathol Exp Neurol* 55, 1270–1276 (1996)
80. Y. Shi, Y. Wang, W. Luan, P. Wang, T. Tao, J. Zhang, J. Qian, N. Liu and Y. You: Long non-coding RNA H19 promotes glioma cell

invasion by deriving miR-675. *PLoS One* 9, e86295 (2014)
DOI: 10.1371/journal.pone.0086295

81. X. Jiang, Y. Yan, M. Hu, X. Chen, Y. Wang, Y. Dai, D. Wu, Y. Wang, Z. Zhuang and H. Xia: Increased level of H19 long noncoding RNA promotes invasion, angiogenesis, and stemness of glioblastoma cells. *J Neurosurg* 124, 129–136 (2016)
DOI: 10.3171/2014.12.jns1426
82. W. Li, P. Jiang, X. Sun, S. Xu, X. Ma and R. Zhan: Suppressing H19 Modulates Tumorigenicity and Stemness in U251 and U87MG Glioma Cells. *Cell Mol Neurobiol* (2016)
DOI: 10.1007/s10571-015-0320-5
83. W. Jiang, S. Finniss, S. Cazacu, C. Xiang, Z. Brodie, T. Mikkelsen, L. Poisson, D. B. Shackelford and C. Brodie: Repurposing phenformin for the targeting of glioma stem cells and the treatment of glioblastoma. *Oncotarget* (2016)
DOI: 10.18632/oncotarget.10919

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