

Myeloid heme oxygenase-1: a new therapeutic target in anti-inflammation

Mingyi Zhao¹, Minghua Yang², Weitao Que³, Lin Zhong³, Masayuki Fujino^{4, 5}, Xiao-Kang Li⁴

¹Department of Pediatrics, The Third Xiangya Hospital, Central South University, Changsha, 410006, China;

²Department of Pediatrics, Xiangya Hospital, Central South University, Changsha, 410000, China;

³Department of Surgery, Shanghai General Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China; ⁴Research Institute, National Center for Child Health and Development, 2-10-1 Okura,

Setagaya-ku, Tokyo 157-8535, Japan; ⁵National Institute of Infectious Diseases, 1-23-1 Toyama, Shinjuku-ku, Tokyo 162-8640, Japan

TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Myeloid cells and inflammation
 - 3.1. Functional of granulocytes in inflammation
 - 3.2. Functional of macrophages in inflammation
 - 3.3. Functional of monocytes in inflammation
 - 3.4. Function of DCs in inflammation
4. Role of myeloid HO-1 in the pathological process of inflammation
 - 4.1. Role of myeloid HO-1 effects on NKT and NK cells
 - 4.2. Role of myeloid HO-1 effects on T cells
 - 4.3. Role of myeloid HO-1 effects on B cells
 - 4.4. Molecular mechanisms of myeloid HO-1 gene regulation in inflammation
 - 4.5. Protective role of myeloid HO-1 in inflammation diseases
 - 4.6. Myeloid HO-1 bridges a crosstalk between autophagy, apoptosis, oxidative stress, and inflammation
5. Compounds as myeloid HO-1 expression inducer
 - 5.1. Aminolevulinic acid (5-ALA)
 - 5.2. Dihydroquercetin (DHQ)
6. Summary and conclusion
7. Acknowledgments
8. References

1. ABSTRACT

An increasing amount of evidence reveals that an orchestrated interplay between myeloid subpopulations in the hematopoietic system plays a significant role in supporting normal functions of the immune system and facilitating homeostatic restoration upon exogenous or endogenous insults. Heme oxygenase-1 (HO-1), a microsomal enzyme discovered decades ago, can metabolize pro-oxidant heme into biliverdin, free iron, and carbon monoxide. This enzymatic reaction produces biological materials, contributing to major immunomodulatory effects. Specifically, HO-1 expression in myeloid cells has been generally acknowledged to drive potent anti-inflammatory and immunosuppressive responses. In this review, the authors focused on elucidating

the potential mechanisms underlying myeloid HO-mediated immunomodulation phenotypes, and discussed the potential application of myeloid-specific HO-1 induction as an anti-inflammation therapeutic strategy.

2. INTRODUCTION

When the immune system is activated after a detrimental attack, an organism must be able to balance the stimulated proinflammatory and corresponding anti-inflammatory processes to achieve homeostasis for maintaining normal physiologic status. This is significant as the production of proinflammatory factors is crucial for successfully eliminating the infective

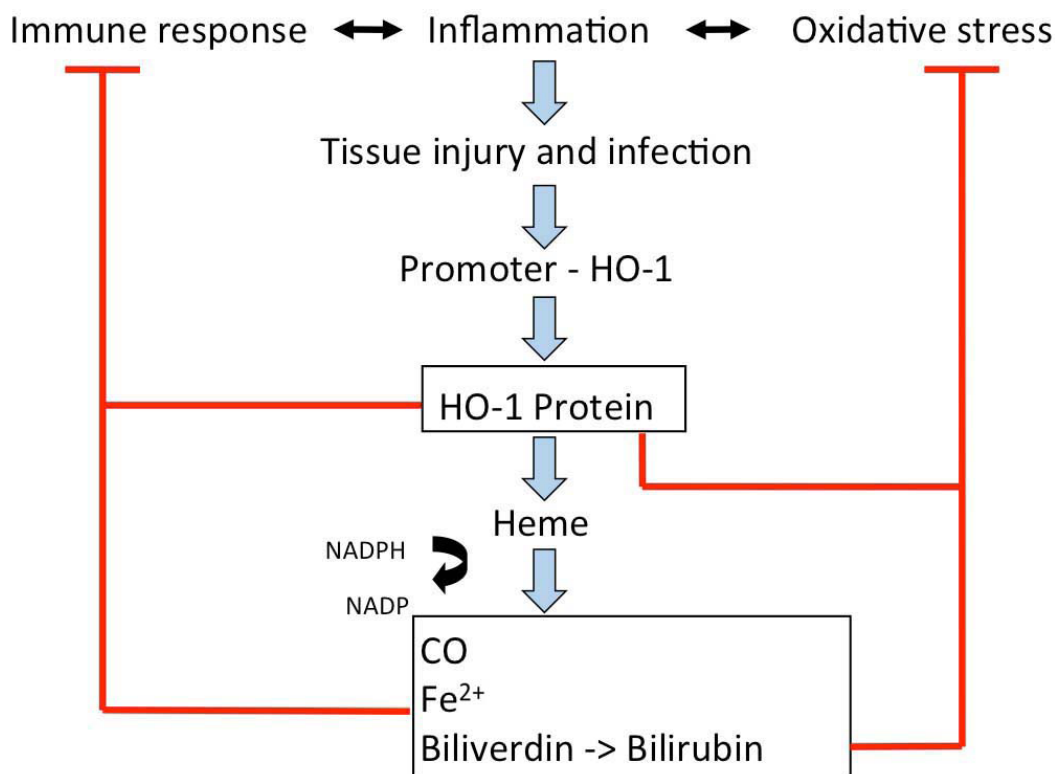


Figure 1. HO-1 metabolizes heme. The HO-1 promoter has elements for many transcription factors and responds to heme. These induce HO-1 protein, which then metabolizes carbon monoxide, iron, and biliverdin (which is converted into bilirubin in a coupled reaction). HO-1 and the three by-products appear to have anti-inflammatory and anti-oxidant properties. NADP, nicotinamide adenine dinucleotide phosphate; NADPH, β -nicotinamide adenine dinucleotide 2'-phosphate reduced tetrasodium salt.

agents, whereas the restoration of homeostasis is necessary for minimizing immune system disorders associated with pathology (1). Accumulating studies have demonstrated that the coordinated interplay between myeloid cells (e.g. macrophages, neutrophils, monocytes, and dendritic cells) is required for maintaining this delicate yet important physiologic balance (2, 3).

Heme oxygenase-1 (HO-1) is a microsomal enzyme catabolizing pro-oxidant heme into biliverdin, free iron, and carbon monoxide (CO) (4, 5). These by-products appear to have potent cytoprotective properties (6) as shown in Figure 1. HO-1 expression is ubiquitously induced as a response to diverse stimuli related to oxidative stress and inflammation, including cytokines, hypoxia, hyperoxia, heavy metals, nitric oxide (NO), heat shock, thiol-reactive substances, hydrogen peroxide, and shear stress, where this expression activates or suppresses cell-intrinsic pathways (7, 8). Moreover, increasing evidence indicates that HO-1 derived from myeloid cells contributes to the pathogenesis of numerous virus infections and alters responses of infectious and organ-specific autoimmune diseases (5, 9). This review highlights the current progress on: i) the association between myeloid cells and inflammation; ii) the

contribution of myeloid HO-1 to immune responses; iii) the mechanisms underlying myeloid HO-1 regulation and this enzyme's capability for treating immune disorders.

3. MYELOID CELLS AND INFLAMMATION

Myeloid cells include four subgroups, namely, granulocytes, monocytes, macrophages, and dendritic cells (DCs) (10). They can be promptly recruited to the infected or damaged sites through peripheral or lymphatic trafficking, with the facilitation of a number of different chemokine receptors (11-14). The infiltrated myeloid cells are activated for phagocytosis and secrete a diverse variety of inflammatory mediators to realize functional plasticity in immunity (15-17).

3.1. Functional of granulocytes in inflammation

Granulocytes, a differentiated lineage derived from hematopoietic stem cells in bone marrow (BM), are the most abundant component of the myeloid population. They include four subcategories, namely, neutrophils, eosinophils, basophils, and mast cells. Neutrophils are generated in high number in BM and can be triggered for activation while in circulation through communication with signals from the resident

macrophages at the infected or injured site (17). Eosinophils, in addition to localizing in BM, are also found in various organs (e.g., gastrointestinal tract, heart, and lung). They are important in forming an immune barrier against parasites and allergens, and therefore support physiologic homeostasis (18). Basophils and mast cells can contribute to the development and progression of Type 2 T helper (Th2) cytokine-mediated inflammation (19). They are able to release potent inflammatory mediators, such as proteases, histamine, heparin, and other molecules, which can initiate and modulate an inflammatory response by acting on various tissues and cells (20).

3.2. Functional of macrophages in inflammation

Macrophages perform diverse roles in homeostasis and disease progression that can involve developmental integrity, tissue repair machinery, and immune modulations. These functions of macrophages could be executed in a well-controlled manner, and could involve multilevel cellular responses. For instance, recognition of pathogen-associated molecular patterns (PAMPs), phagocytic clearance, and cytokine secretion are significant components of macrophage-mediated responses. The accumulative results could further lead to recruitment of other types of cells, triggering a systematic immune response. In general, macrophages can elicit inflammation, scavenge tissues, and actively regulate immune responses. Notably, the primary function of macrophages is to initiate adaptive immune responses by presenting pathogen-derived antigens to T cells (21). Moreover, macrophages are one major cell type responsible for efferocytosis, the process of engulfing and destroying dying cells, thus releasing harmful intracellular biomaterials to trigger inflammation that could lead to autoimmunity (22, 23).

3.3. Functional of monocytes in inflammation

Monocytes, an important component of the innate immune system, are among the first batches of cell types to sense and respond to the local insults by migrating from the peripheral blood to the infected site and differentiating into tissue macrophages and dendritic cells to take part in the immune response. Endowed with dualism due to their heterogeneity and functional plasticity, monocytes are able to exert versatile immune functions during the different stages of immunity (24). They can produce proinflammatory factors (e.g., TNF- α , interleukin (IL)-6, IL-1 β), reactive oxygen species (ROS), proteolytic enzymes, and other active molecules to actively dictate an early cascade of immune responses. On the other hand, by secreting anti-inflammatory factors (e.g., IL-10), monocytes can also actively engage in the inflammation suppression process which is essential for maintaining tissue homeostasis (25, 26).

3.4. Function of DCs in inflammation

As discussed in previous studies, the immune system can be efficiently activated by anatomical, immunological, genetic, and environmental cues in the early stages of life. In addition to conventional DCs which arise from hematopoietic progenitor cells in the BM, a heterogeneous sub-population called inflammatory DCs can be derived from monocytes which are recruited to the local sites where inflammation occurs. It has also been observed that the nuclear factor kappa B (NF- κ B) pathway engaged in TLR-4 signaling is well preserved in neonatal monocyte-derived DCs (MoDCs), which are in charge of maintaining the generation of pro-inflammatory factors (e.g., TNF- α , IL-6, and IL-8). This helps scholars to better understand the molecular mechanisms underlying the diminished ability of circulating MoDCs to generate inflammatory cytokines such as TNF- α or IL-1 β , contributing to the vulnerability of newborns to bacterial sepsis (27).

4. ROLE OF MYELOID HO-1 IN THE PATHOLOGICAL PROCESS OF INFLAMMATION

Inflammation is a dynamic process engaging diverse cellular and subcellular elements that undergo a complex interplay. Those elements operate in a collaborative fashion to defend the host against deleterious stimulation (28). Current views state that the inflammation process can involve both the innate immune system and adaptive immune system for orchestrated regulation of immunity. Essentially, modulating the myeloid HO-1 may cross-regulate these two processes.

4.1. Role of myeloid HO-1 effects on NKT and NK cells

Natural-killer-T (NKT) cells, harboring cytotoxicity and cytokine-producing functions, are significant components of the innate immune system. They can quickly respond to virally infected cells and tumor cells and can generate diverse cytokines correspondingly. In addition to their long-term acknowledged effector lymphocyte function, recent discoveries underscore their regulatory role in communicating with other types of lymphocytes as well. For instance, antigen-presenting DCs can be activated by NKTs (29), and T cells can be induced to form differential Th1 cells after direct interaction with NKTs (30). Genetically, deletion of HO-1 in multi-drug resistance protein 2 (Mdr2) knockout mice leads to a more severe inflammation phenotype compared with Mdr2-only knockout littermates. This exacerbation could be explained by the corresponding elevated frequencies of T cells, NKT cells, and mature dendritic cells (DCs) compared to Mdr2-only knockout littermates (31). NK cells can interfere with the delicate balance between the regulatory and effector input part

of the immune response through eliminating regulatory T cells (T-reg) (32). Induced HO-1 expression in mice reveals a severe suppression of T cell and NK-cell-mediated spleen cell toxicity (32).

4.2. Role of myeloid HO-1 effects on T cells

T-lymphocytes are essential players and a core component for adaptive immunity, governing the host's immune response toward different pathogens (33). T-lymphocytes are mainly divided into two subpopulations based on the marker expression pattern on their cell surface, namely, CD4+ and CD8+. In addition to carrying out multiple layers of functions in immunity, CD4+ helper T cells promote activation of cells engaged in the innate immune system, B-lymphocytes, and other nonimmune cell types. On the other hand, they contribute to dampening the active immune response based on their regulatory property. CD8+ cytotoxic T cells can specifically destroy their target (i.e., cancer cells, infected cells, or damaged cells from other sources) through releasing their cytotoxins to induce apoptosis. Releasing soluble factors and cytokines is an important means for T cells to communicate with other cells as well as defining the local immune environment. The distinct patterns of released cytokines could be proinflammatory or anti-inflammatory. Additionally, T cells release soluble factors that can activate and recruit immune cells or modulate their functions (34). Several studies have indicated that myeloid-derived HO-1 affects the function and development of human and murine CD4+ and CD8+ regulatory T cells (T-reg). Myeloid dendritic cells expressing HO-1 have been shown to potentially drive the enhancement of Foxp3 regulatory T cells through unknown mechanisms (35). However, the contribution of HO-1 activity in the inhibition of inflammatory molecules produced by T cells has not yet been identified. Findings support the notion that myeloid HO-1 expression can actively impact many aspects of T cell biology potentially through a dissected cell-cell communication network. Brandsma (36) reported that HO-1 induction, particularly in lung alveolar macrophages, was correlated with the inhibition of smoking-induced B-cell infiltrates in lung. This phenotype was accompanied with elevated CD4+CD25+ regulatory T cells in lung (36). These studies provided strong evidence about the myeloid HO-1 function in regulating inflammation, and further enabled scholars to infer that manipulation of myeloid HO-1 level could serve as a novel means to regulate immune cell (i.e., T-regs) responses toward an anti-inflammation outcome. Moreover, our previous findings revealed that the application of the precursor of heme anabolism, 5-aminolevulinic acid (5-ALA), together with the byproduct of heme catabolism, ferrous iron, interestingly upregulated HO-1 expression in myeloid cells and led to T cell proliferation repression in a murine cardiac allograft model (37). However, in this

model, regulatory T cells and myeloid cells received significant expansion, leading to alleviation of the organ allotransplantation rejection phenotype and eventually to successful allograft acceptance. Collectively, all these findings support the idea that myeloid-derived HO-1 plays a critical role in immune tolerance, and manipulation of its expression level adds a novel layer of regulation beneficial for clinical treatment of inflammation.

4.3. Role of myeloid HO-1 effects on B cells

B cells, a critical component of adaptive immunity, carry out the humoral immune response through secreting antibody molecules. Furthermore, B cells can also release cytokines and actively engage in regulating the function of other immune cells (e.g., T cells and DCs) to realize its regulatory role in immunity (38). Kapturczak *et al.* discovered that HO-1 gene deletion in mice led to diminishing the number of B220+ cells in the lymph nodes (39). In these animals, levels of serum immunoglobulin M (IgM) were dramatically increased and irregular immunoglobulin profiling was demonstrated (40). These data strongly support the concept that HO-1 gene expression can impact B cell functions (41). We can infer that myeloid-derived HO-1 might contribute to the above-described phenotype, and that this enzyme is vital for B cell inflammation.

4.4. Molecular mechanisms of myeloid HO-1 gene regulation

HO-1 gene expression primarily responds to numerous biological stress stimuli including oxidative stresses and their inductions, which are regulated through diverse signaling pathways primarily at the transcriptional level (42). Among the identified HO-1 gene regulatory DNA motifs in mouse models, stress-responsive elements or antioxidant response elements (ARE) have been shown to be a dominant cis-regulatory element which also exists in the human HO-1 gene promoter region. Transcriptional regulation of the HO-1 gene can be carried out through dynamic interactions between transcriptional activators or repressors and the regulatory DNA motifs. Transcriptional activators such as nuclear factor erythroid 2-related factor-2 (Nrf2) or transcriptional repressors (e.g., Bach1) can competitively occupy the ARE domain of the HO-1 gene promoter, which can lead to modulation of gene expression. Furthermore, evidence supports the notion that inactivation of Bach1 was indeed a prerequisite for HO-1 gene induction. To add an extra regulatory layer to this dynamic process, it was shown that HO-1 gene expression can also be regulated by its substrate heme, which is able to form a complex with the transcriptional repressor Bach1 to prevent its binding with the HO-1 gene enhancer and promote its nuclear export as well. Previous results, obtained by the authors, showed that application of

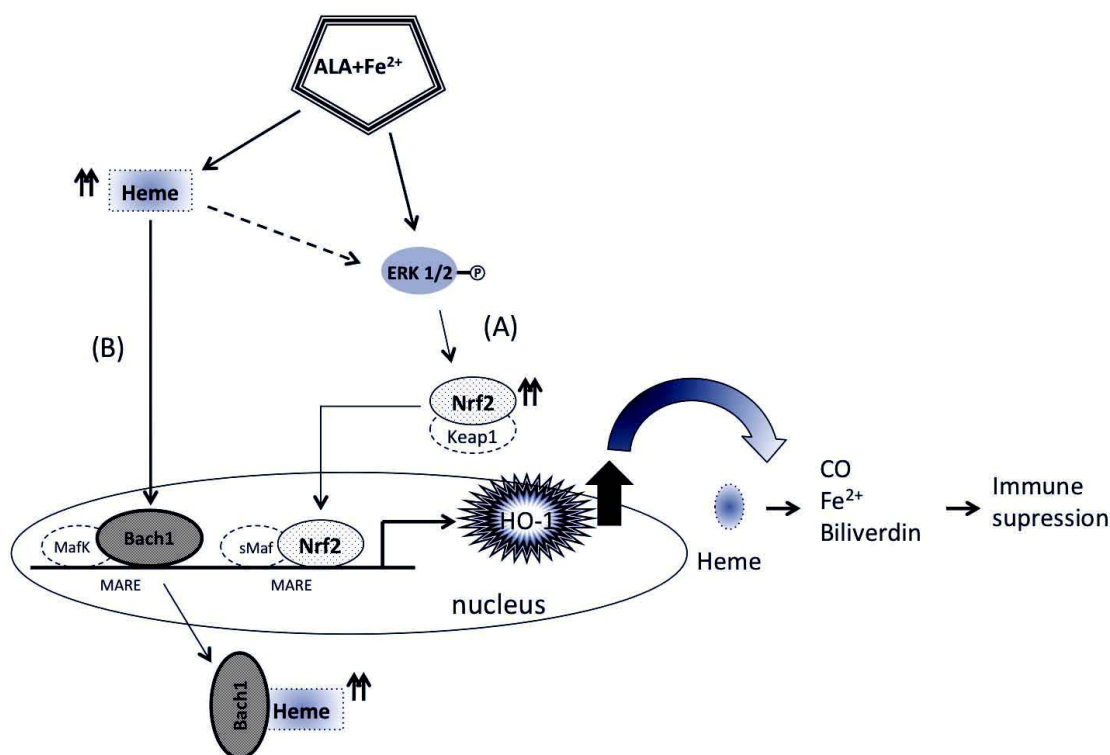


Figure 2. A model of HO-1 induction based on our observations. 5-ALA metabolism induced HO-1 expression through (A) and (B) pathways in myeloid cells (RAW264.7.). Treatment with 5-ALA combined with Fe^{2+} induces the phosphorylation of ERK and p38 MAPK. These activated MAPKs lead to HO-1 expression through their effects on post-transcriptional factors, such as Nrf2. On the other hand, exposure to 5-ALA/ Fe^{2+} increases the intracellular levels of heme. Under conditions with a higher concentration of heme, the HO-1 repressor, Bach1, is inactivated by direct binding of heme to Bach1, which allows for increased expression of HO-1. The induction of HO-1 via at least two pathways leads to reduced levels of inflammatory cytokines.

the precursor of heme anabolism (5-ALA) together with the byproduct of heme catabolism (ferrous iron) in the macrophage cell line RAW264.7. up-regulated the HO-1 gene expression for an anti-inflammation effect, which could be suppressed by mitogen-activated protein kinase (MAPK) pathway inhibitors (43). NFE2L2 (Nrf2) activator, and intracellular increment at the heme level, were observed in the treated RAW264.7. cells, suggesting a Bach1 inactivation mechanism to facilitate HO-1 gene upregulation, as shown in Figure 2) (43).

A number of other proximal promoter components, such as upstream stimulatory factor, basic helix loop helix proteins, NF- κ B, signal transducer and activator of transcription 3 (STAT3), activator protein-2, hypoxia-inducible factor (HIF)-1, activating transcription factor 2 (ATF2), and heat shock factor 1 (HSF1) are all among the growing inventory list contributing to HO-1 gene regulation. The multiplicity of this complex gene fine-tuning network establishes a platform for developing pharmaceutical intervention methods targeting HO-1 gene production for therapeutic application. Pharmaceutical activation or inhibition of some promoter elements can induce expression of HO-1 proteins that efficiently rescue cells from inflammatory stress (44).

MAPK signaling is well acknowledged for its significant role to elicit a variety of biological changes in response to extracellular cues. Our lab and other groups revealed that MAPK signaling cascade is a crucial player involved in Nrf2 transcription factor activation, which is required for inducing HO-1 expression to exert its anti-oxidative stress function (43, 45-48). In addition, other signaling molecules including tyrosine kinase, protein kinase C, NF- κ B, and phosphatidylinositol 3-kinase/Akt are all included in the critical list for HO-1 expression regulation (see Figure 3) (49-51).

4.5. Protective role of myeloid HO-1 in inflammation diseases

In mammals, HO-1, as a stress-responsive enzyme, catabolizes heme into CO, free iron, and biliverdin in the presence of molecular oxygen and electron source for nicotinamide adenine dinucleotide phosphate (NADPH) cytochrome p450 reductase. The generated biliverdin is further reduced to bilirubin by biliverdin reductase. Both biliverdin and bilirubin showed direct anti-oxidative properties by eliminating excessive ROS whose elevated level is believed to significantly contribute to chronic inflammation and pathogenesis (52-

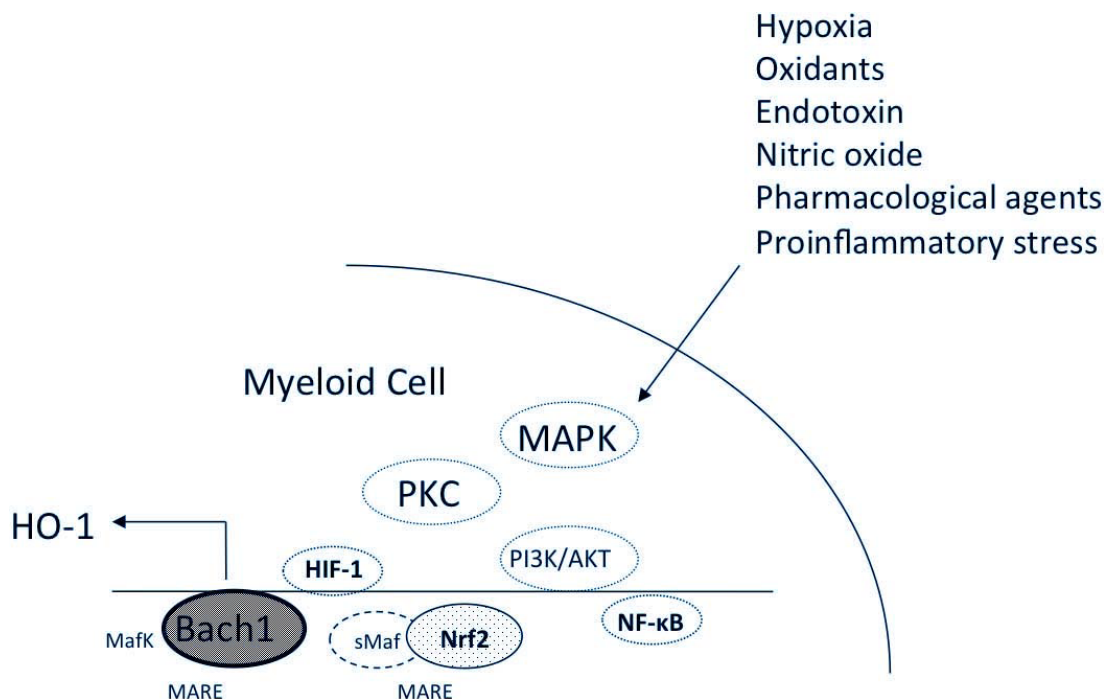


Figure 3. A diagram of the HO-1 signaling pathways. External and internal stimulus factors, such as hypoxia, oxidants, endotoxin, nitric oxide, pharmacological agents, or proinflammatory stress, up-regulate HO-1 expression via several kinase cascades and transcription factors.

54). In addition, CO, through directly regulating diverse transcription regulation programs and inflammation-associated molecules, significantly conveys the beneficial effect of HO-1 protein in combating inflammation (55). Recently, it was discovered that HO-1's anti-inflammation and immunomodulation function can be tightly linked to myeloid or endothelial origin-derived expression (56). In myeloid cells, HO-1 is believed to induce downregulation of tumor necrosis factor (TNF- α) and upregulation of IL10; these HO-1 mediated events at the molecular level are further associated with cell differencing commitment to tolerogenic antigen presenting cells, which are promising mediators for treating inflammatory diseases. Importantly, in multiple elegantly established *in vivo* organ-system inflammatory models, evidence indicated that HO-1 could positively execute its protective potential for homeostasis recovery through modulation of tissue responses to injury, as shown in Table 1.

4.6. Myeloid HO-1 bridges a crosstalk between autophagy, apoptosis, oxidative stress, and inflammation

A growing body of evidence supports the concept that autophagy, an adaptive cell catabolic program that eliminates harmful intracellular components, can play diverse roles in response to ROS-triggered oxidative stress signals. Through preserving mitochondrial integrity and degrading nonfunctional proteins, autophagy provides a survival

mechanism under stress which is also significant for modulating the inflammation process and immune functions (57). HO-1 or exogenous application of its reaction product CO, has been shown to confer a cytoprotective role through the modulation of apoptotic-autophagy and/or inflammatory signaling in models of inflammatory stress, as depicted in Figure 4 (58-60). Indeed, the HO-1/CO axis has also been shown to protect stress-insulted cells from incurred apoptosis, therefore shifting cell fate toward survival.

Using pharmacological intervention or genetic manipulation methods, *in vivo* evidence highlights the importance of HO-1 and its end products in affording protective effect in tissue injury models through induction of autophagy (61). Furthermore, in macrophages stimulated by lipopolysaccharide (LPS), HO-1 was shown to mediate autophagy which further leads to anti-inflammation and cytokine production repression phenotype (60-63). A previous study also suggested that HO-1 induction by 5-ALA together with ferrous iron (SFC) inhibited hypoxia-caused cellular injury in cardiomyocytes through autophagy (45).

5. COMPOUNDS AS MYELOID HO-1 EXPRESSION INDUCER

Various inducers for HO-1 expression are known to have potent cytoprotective properties, such

Table 1. Protective role of HO-1 expression in inflammation diseases

Organ	Injury/Disease Model	Mechanism	Reference
Brain	<ul style="list-style-type: none"> HIV infection Diabetes Parkinson's disease Hypoxia-induced brain injury Lipopolysaccharide-induced acute inflammation 	<ul style="list-style-type: none"> Endogenous antioxidant and immune modulatory NF-κB p65 inhibition Nrf2 pathway PI3K/Akt/Nrf-2 signal pathways Oxidative reaction 	(66) (67) (68) (69) (70)
Eyes	<ul style="list-style-type: none"> Endotoxin-induced uveitis 	<ul style="list-style-type: none"> Increased cytokine production 	(71)
Skin	<ul style="list-style-type: none"> UVB irradiation TPA-induced inflammation 	<ul style="list-style-type: none"> Nrf2 pathway IL-1 beta and TNF-alpha 	(53, 72) (73)
Lung	<ul style="list-style-type: none"> LPS-induced inflammation Acute pulmonary inflammation 	<ul style="list-style-type: none"> p38MAPK-dependent Inhibiting the release of segmented PMNs from the BM 	(74) (75)
Heart	<ul style="list-style-type: none"> Cardiac dysfunction 	<ul style="list-style-type: none"> Inhibition of oxidative stress, inflammation, and apoptosis 	(76)
Gastrointestinal tract	<ul style="list-style-type: none"> Inflammatory diseases 	<ul style="list-style-type: none"> Anti-inflammatory, anti-proliferative, antioxidant, and anti-apoptotic 	(77)
Liver	<ul style="list-style-type: none"> Liver fibrosis Ischemia and reperfusion 	<ul style="list-style-type: none"> Nrf2/ARE signaling pathway Nrf-2/Akt-p70S6k signaling pathway Notch1/Hes1/Stat3 signaling 	(78) (79) (80)
Bone marrow	<ul style="list-style-type: none"> Transplantation 	<ul style="list-style-type: none"> Reduced T-cell activation ex vivo 	(81)
Kidney	<ul style="list-style-type: none"> Ischemia/reperfusion Polycystic kidney disease 	<ul style="list-style-type: none"> Produce the potent cellular antioxidant bilirubin Reduced antioxidant enzyme 	(52, 82) (83, 84)

GI: Gastrointestinal, BM: Bone marrow,

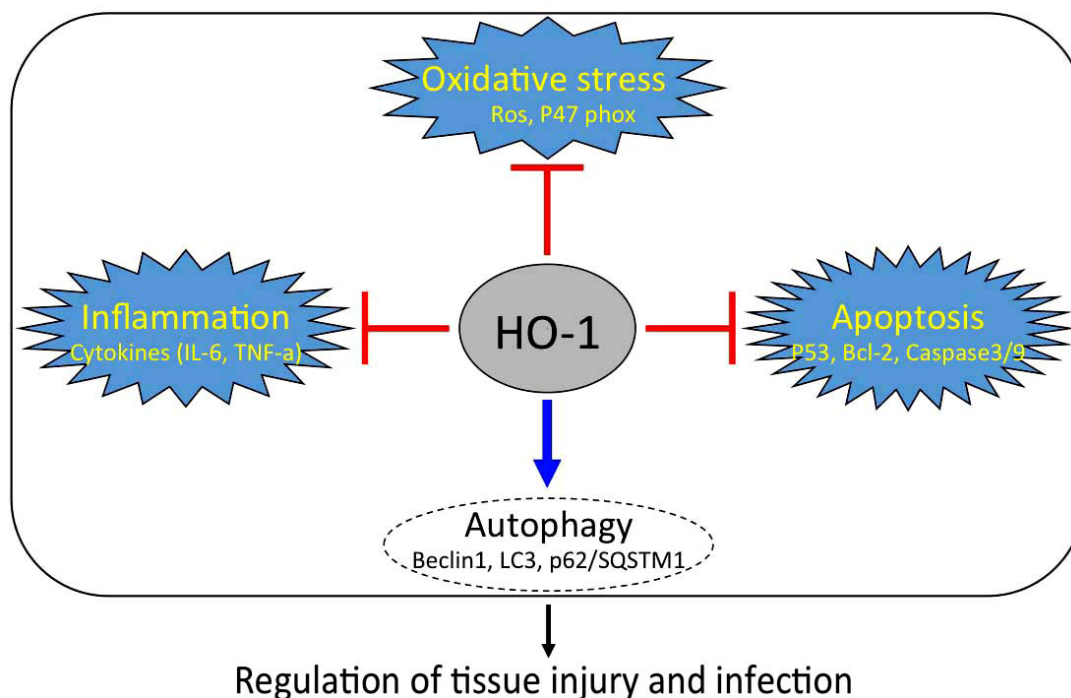


Figure 4. Crosstalk between autophagy, apoptosis, oxidative stress and inflammation by HO-1 overexpression.

as hemin, epigallocatechin-3-gallate, rosiglitazone, gastrodin, methotrexate (MTX), and curcumin. This review details the effects of two compounds, 5-ALA and dihydroquercetin (DHQ), which have been recently studied by our research group.

5.1. Aminolevulinic acid (5-ALA)

5-ALA, a naturally occurring important metabolic intermediate, is a necessary precursor of heme. A previous study discovered that in a renal

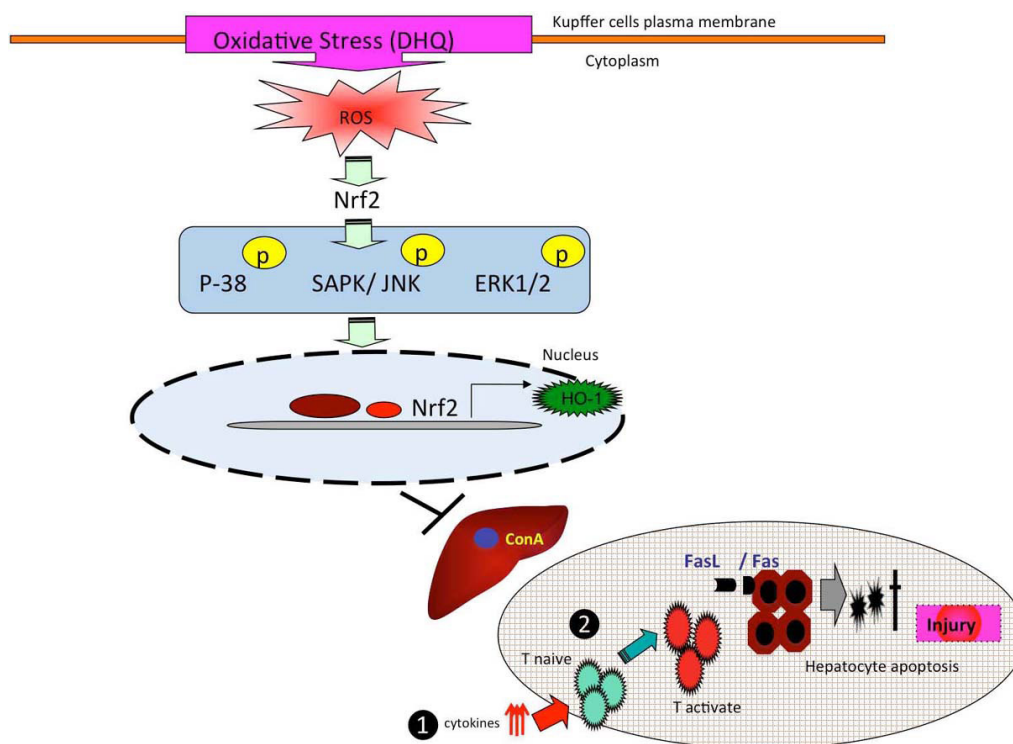


Figure 5. Dihydroquercetin (DHQ) ameliorated concanavalin A-induced experimental fulminant hepatitis in mice and enhanced HO-1 expression through the MAPK/Nrf2 antioxidant pathway in RAW cells.

ischemia reperfusion injury (IRI) model, a 5-ALA/SFC combination application before the surgery dramatically attenuated the severity of the IRI with significantly alleviated tubular damage and apoptosis inhibition. Furthermore, the detected amount of renal thiobarbituric acid-reactive substance was considerably lower in the 5-ALA/SFC combination application group, which could be attributed to the elevated HO-1 regulation followed by the corresponding high production of the end product CO in initiating the protective effect through downregulating TNF- α and interferon gamma (IFN- γ) in the renal parenchymal tissue. Interestingly, macrophage infiltration was found to be diminished in the 5-ALA/SFC combination-treated renal group, suggesting a potential mechanism of 5-ALA/SFC—HO-1—CO axis-induced immunomodulatory effect in protecting kidney from detrimental stimuli, IRI in this case (64).

5.2. Dihydroquercetin (DHQ)

DHQ, a natural compound found in plants, has been long perceived as a potent anti-oxidant agent that holds promise for fighting a variety of diseases, including liver disease. Using our established *in vivo* experimental platform (65), we were able to show that DHQ application efficiently protected mouse livers from concanavalin A (ConA)-induced damage, evidenced by reduced liver function impairment index serum alanine transaminase (ALT) and aspartate transaminase (AST) levels, improved histopathological outcome,

and boosted animal survival rate. We further found that various proinflammatory cytokine mRNA levels underwent remarkable reduction in the liver tissues from DHQ-treated animals, suggesting involvement of an immunomodulation process as a potential mechanism explaining DHQ's beneficial effect in liver protection. Based on the belief that liver macrophages/Kupffer cells carry essential distinct functions in response to oxidative stress and inflammatory stimuli for governing liver homeostasis and pathogenesis, the authors were prompted to explore their response by applying ConA to a mouse RAW264 macrophage cell line. The results showed that DHQ treatment can obviously block the transcription level of IFN- γ and TNF- α , and further diminish their secretion in culture. To dissect the potential underlying mechanisms, the authors focused on HO-1 and demonstrated that its expression was markedly raised in a dose- and time-dependent fashion by DHQ, probably through the signaling from the upregulated transcriptional activation factor Nrf2. Furthermore, DHQ stimulated phosphorylation of three MAPK family members, and application of the inhibitors of MEK/ERK (PD98059), p38 (SB203580), and JNK (SP600125) blocked DHQ-mediated HO-1 upregulation (65) (see Figure 5).

6. SUMMARY AND CONCLUSION

In summary, the presence of myeloid HO-1 serves a protective role at the site of tissue

injury. Given this positive role, perhaps laboratory approaches, in order to increase myeloid HO-1 levels for anti-inflammation therapeutic purposes, should be considered for translation from benchtop to clinical applications. However, further understanding the mechanisms of myeloid HO-1 in the pathogenesis of inflammation diseases may be required to safely create and efficiently design treatments for use in clinical practice.

7. ACKNOWLEDGMENTS

Mingyi Zhao and Minghua Yang contributed equally to this work. The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest. This work was partly supported by research grants from the Ministry of Education, Culture, Sports, Science and Technology of Japan (Grants-in-Aid 24/02741, 15F15756, 15K10043 and 17H04277), the National Center for Child Health and Development (26-6), JSPS under JSPS Postdoctoral Fellowship For Foreign Researchers (P12741), National Natural Sciences Foundation of China (81500231 and 81570154), and Natural Sciences Foundation of Hunan Province (2015JJ6118).

8. REFERENCES

1. T. S. Lai, Z. H. Wang and S. X. Cai: Mesenchymal Stem Cell Attenuates Neutrophil-predominant Inflammation and Acute Lung Injury in an *In vivo* Rat Model of Ventilator-induced Lung Injury. *Chin Med J (Engl)*, 128(3), 361-7 (2015)
DOI: 10.4103/0366-6999.150106
2. K. S. Burrack and T. E. Morrison: The role of myeloid cell activation and arginine metabolism in the pathogenesis of virus-induced diseases. *Front Immunol*, 5, 428 (2014)
DOI: 10.3389/fimmu.2014.00428
3. P. J. Murray and T. A. Wynn: Protective and pathogenic functions of macrophage subsets. *Nat Rev Immunol*, 11(11), 723-37 (2011)
DOI: 10.1038/nri3073
4. A. Paine, B. Eiz-Vesper, R. Blasczyk and S. Immenschuh: Signaling to heme oxygenase-1 and its anti-inflammatory therapeutic potential. *Biochem Pharmacol*, 80(12), 1895-903 (2010)
DOI: 10.1016/j.bcp.2010.07.014
5. V. Koliarakis and G. Kollias: A new role for myeloid HO-1 in the innate to adaptive crosstalk and immune homeostasis. *Adv Exp Med Biol*, 780, 101-11 (2011)
DOI: 10.1007/978-1-4419-5632-3_9
6. S. Immenschuh, E. Baumgart-Vogt and S. Mueller: Heme Oxygenase-1 and Iron in Liver Inflammation: A Complex Alliance. *Current Drug Targets*, 11(12), 1541-1550 (2010)
DOI: 10.2174/1389450111009011541
7. E. P. Carter, C. Garat and M. Imamura: Continual emerging roles of HO-1: protection against airway inflammation. *Am J Physiol Lung Cell Mol Physiol*, 287(1), L24-5 (2004)
DOI: 10.1152/ajplung.00097.2004
8. C. Chauveau, S. Remy, P. J. Royer, M. Hill, S. Tanguy-Royer, F. X. Hubert, L. Tesson, R. Brion, G. Beriou, M. Gregoire, R. Josien, M. C. Cuturi and I. Anegon: Heme oxygenase-1 expression inhibits dendritic cell maturation and proinflammatory function but conserves IL-10 expression. *Blood*, 106(5), 1694-702 (2005)
DOI: 10.1182/blood-2005-02-0494
9. P. Mandal, M. T. Pritchard and L. E. Nagy: Anti-inflammatory pathways and alcoholic liver disease: role of an adiponectin/interleukin-10/heme oxygenase-1 pathway. *World J Gastroenterol*, 16(11), 1330-6 (2010)
DOI: 10.3748/wjg.v16.i11.1330
10. I. De Kleer, F. Willems, B. Lambrecht and S. Goriely: Ontogeny of myeloid cells. *Front Immunol*, 5, 423 (2014)
DOI: 10.3389/fimmu.2014.00423
11. E. G. Perdiguerro, K. Klapproth, C. Schulz, K. Busch, E. Azzoni, L. Crozet, H. Garner, C. Trouillet, M. F. de Bruijn, F. Geissmann and H. R. Rodewald: Tissue-resident macrophages originate from yolk-sac-derived erythro-myeloid progenitors. *Nature* (2014)
DOI: 10.1038/nature13989
12. Y. Miyazaki, A. Vieira-de-Abreu, E. S. Harris, A. M. Shah, A. S. Weyrich, H. C. Castro-Faria-Neto and G. A. Zimmerman: Integrin α (D) β (2) (CD11d/CD18) Is Expressed by Human Circulating and Tissue Myeloid Leukocytes and Mediates Inflammatory Signaling. *Plos One*, 9(11) (2014)
DOI: 10.1371/journal.pone.0112770
13. J. Polz, A. Remke, S. Weber, D. Schmidt, D. Weber-Steffens, A. Pietryga-Krieger, N. Muller, U. Ritter, S. Mostbock and D. N. Mannel: Myeloid suppressor cells require membrane TNFR2 expression for

- suppressive activity. *Immun Inflamm Dis*, 2(2), 121-30 (2014)
DOI: 10.1002/iid3.19
14. T. T. Rohn: The Triggering Receptor Expressed on Myeloid Cells 2: "TREM-ming" the Inflammatory Component Associated with Alzheimer's Disease. *Oxidative Medicine and Cellular Longevity* (2013)
DOI: 10.1155/2013/860959
15. C. Auffray, D. K. Fogg, E. Narni-Mancinelli, B. Senechal, C. Trouillet, N. Saederup, J. Leemput, K. Bigot, L. Campisi, M. Abitbol, T. Molina, I. Charo, D. A. Hume, A. Cumano, G. Lauvau and F. Geissmann: CX3CR1+ CD115+ CD135+ common macrophage/DC precursors and the role of CX3CR1 in their response to inflammation. *J Exp Med*, 206(3), 595-606 (2009)
DOI: 10.1084/jem.20081385
16. A. S. Dighe, D. Campbell, C. S. Hsieh, S. Clarke, D. R. Greaves, S. Gordon, K. M. Murphy and R. D. Schreiber: Tissue-specific targeting of cytokine unresponsiveness in transgenic mice. *Immunity*, 3(5), 657-66 (1995)
DOI: 10.1016/1074-7613(95)90136-1
17. S. H. Lee, J. A. Carrero, R. Uppaluri, J. M. White, J. M. Archambault, K. S. Lai, S. R. Chan, K. C. F. Sheehan, E. R. Unanue and R. D. Schreiber: Identifying the Initiating Events of Anti-Listeria Responses Using Mice with Conditional Loss of IFN-gamma Receptor Subunit 1 (IFNGR1). *Journal of Immunology*, 191(8), 4223-4234 (2013)
DOI: 10.4049/jimmunol.1300910
18. M. R. Konikoff, C. Blanchard, C. Kirby, B. K. Buckmeier, M. B. Cohen, J. E. Heubi, P. E. Putnam and M. E. Rothenberg: Potential of blood eosinophils, eosinophil-derived neurotoxin, and eotaxin-3 as biomarkers of eosinophilic esophagitis. *Clin Gastroenterol Hepatol*, 4(11), 1328-36 (2006)
DOI: 10.1016/j.cgh.2006.08.013
19. M. C. Siracusa, B. S. Kim, J. M. Spergel and D. Artis: Basophils and allergic inflammation. *J Allergy Clin Immunol*, 132(4), 789-801; quiz 788 (2013)
DOI: 10.1016/j.jaci.2013.07.046
20. E. L. Gibson, Y. Vaucher and J. J. Corrigan, Jr.: Eosinophilia in premature infants: relationship to weight gain. *J Pediatr*, 95(1), 99-101 (1979)
DOI: 10.1016/S0022-3476(79)80097-9
21. J. Neefjes, M. L. Jongsma, P. Paul and O. Bakke: Towards a systems understanding of MHC class I and MHC class II antigen presentation. *Nat Rev Immunol*, 11(12), 823-36 (2011)
DOI: 10.1038/nri3084
22. R. C. Taylor, S. P. Cullen and S. J. Martin: Apoptosis: controlled demolition at the cellular level. *Nature Reviews Molecular Cell Biology*, 9(3), 231-241 (2008)
DOI: 10.1038/nrm2312
23. R. Rodriguez-Manzanet, M. A. Sanjuan, H. Y. Wu, F. J. Quintana, S. Xiao, A. C. Anderson, H. L. Weiner, D. R. Green and V. K. Kuchroo: T and B cell hyperactivity and autoimmunity associated with niche-specific defects in apoptotic body clearance in TIM-4-deficient mice. *Proceedings of the National Academy of Sciences of the United States of America*, 107(19), 8706-8711 (2010)
DOI: 10.1073/pnas.0910359107
24. T. Strunk, P. Temming, U. Gembruch, I. Reiss, P. Bucskey and C. Schultz: Differential maturation of the innate immune response in human fetuses. *Pediatr Res*, 56(2), 219-26 (2004)
DOI: 10.1203/01.PDR.0000132664.66975.79
25. T. R. Kollmann, J. Crabtree, A. Rein-Weston, D. Blimkie, F. Thommai, X. Y. Wang, P. M. Lavoie, J. Furlong, E. S. Fortuno, A. M. Hajjar, N. R. Hawkins, S. G. Self and C. B. Wilson: Neonatal Innate TLR-Mediated Responses Are Distinct from Those of Adults. *Journal of Immunology*, 183(11), 7150-7160 (2009)
DOI: 10.4049/jimmunol.0901481
26. G. Schuler and N. Romani: Generation of mature dendritic cells from human blood - An improved method with special regard to clinical applicability. *Dendritic Cells in Fundamental and Clinical Immunology*, Vol 3, 417, 7-13 (1997)
DOI: 10.1007/978-1-4757-9966-8_2
27. J. H. Peters, J. Ruppert, R. K. H. Gieseler, H. M. Najar and H. Xu: Differentiation of Human Monocytes into Cd14 Negative Accessory Cells - Do Dendritic Cells Derive from the Monocytic Lineage. *Pathobiology*, 59(3), 122-126 (1991)
DOI: 10.1159/000163628
28. N. Sugimoto, T. Rui, M. Yang, S. Bharwani, O. Handa, N. Yoshida, T. Yoshikawa and P. R. Kvietys: Points of control exerted along the macrophage-endothelial cell-

- polymorphonuclear neutrophil axis by PECAM-1 in the innate immune response of acute colonic inflammation. *Journal of Immunology*, 181(3), 2145-2154 (2008)
DOI: 10.4049/jimmunol.181.3.2145
29. F. C. Robertson, J. A. Berzofsky and M. Terabe: NKT cell networks in the regulation of tumor immunity. *Front Immunol*, 5, 543 (2014)
DOI: 10.3389/fimmu.2014.00543
30. A. Martin-Fontecha, L. L. Thomsen, S. Brett, C. Gerard, M. Lipp, A. Lanzavecchia and F. Sallusto: Induced recruitment of NK cells to lymph nodes provides IFN-gamma for T(H)1 priming. *Nat Immunol*, 5(12), 1260-5 (2004)
DOI: 10.1038/ni1138
31. R. Barikbin, M. Sandmann, A. Quaas, K. Karimi, G. Sass and G. Tiegs: P0454 : An additional heme oxygenase-1 knockout increases maturation of dendritic cells and liver inflammation in Mdr2 knockout mice. *Journal of Hepatology*, 62, Supplement 2, S482-S483 (2015)
DOI: 10.1016/S0168-8278(15)30662-0
32. S. Roy, P. F. Barnes, A. Garg, S. Wu, D. Cosman and R. Vankayalapati: NK cells lyse T regulatory cells that expand in response to an intracellular pathogen. *Journal of Immunology*, 180(3), 1729-1736 (2008)
DOI: 10.4049/jimmunol.180.3.1729
33. A. Schumacher and A. C. Zenclussen: Effects of heme oxygenase-1 on innate and adaptive immune responses promoting pregnancy success and allograft tolerance. *Front Pharmacol*, 5, 288 (2014)
DOI: 10.3389/fphar.2014.00288
34. I. Hudic and Z. Fatusic: Progesterone-induced blocking factor (PIBF) and Th-1/Th-2 cytokine in women with threatened spontaneous abortion. *Journal of Perinatal Medicine*, 37(4), 338-342 (2009)
DOI: 10.1515/JPM.2009.061
35. K. Karimi, N. Kandiah, J. Chau, J. Bienenstock and P. Forsythe: A Lactobacillus rhamnosus Strain Induces a Heme Oxygenase Dependent Increase in Foxp3+Regulatory T Cells. *Plos One*, 7(10) (2012)
DOI: 10.1371/journal.pone.0047556
36. C. A. Brandsma, M. N. Hylkema, B. W. van der Strate, D. J. Slebos, M. A. Luinge, M. Geerlings, W. Timens, D. S. Postma and H. A. Kerstjens: Heme oxygenase-1 prevents smoke induced B-cell infiltrates: a role for regulatory T cells? *Respir Res*, 9, 17 (2008)
DOI: 10.1186/1465-9921-9-17
37. J. Hou, S. Cai, Y. Kitajima, M. Fujino, H. Ito, K. Takahashi, F. Abe, T. Tanaka, Q. Ding and X. K. Li: 5-Aminolevulinic acid combined with ferrous iron induces carbon monoxide generation in mouse kidneys and protects from renal ischemia-reperfusion injury. *Am J Physiol Renal Physiol*, 305(8), F1149-57 (2013)
DOI: 10.1152/ajprenal.00275.2013
38. P. Youinou, C. Jamin, J. O. Pers, C. Berthou, A. Saraux and Y. Renaudineau: B lymphocytes are required for development and treatment of autoimmune diseases. *Ann N Y Acad Sci*, 1050, 19-33 (2005)
DOI: 10.1196/annals.1313.003
39. S. Bancos, C. J. Baglole, I. Rahman and R. P. Phipps: Induction of heme oxygenase-1 in normal and malignant B lymphocytes by 15-deoxy-Delta(12,14)-prostaglandin J(2) requires Nrf2. *Cell Immunol*, 262(1), 18-27 (2010)
DOI: 10.1016/j.cellimm.2009.12.003
40. M. H. Kapturczak, C. Wasserfall, T. Brusko, M. Campbell-Thompson, T. M. Ellis, M. A. Atkinson and A. Agarwal: Heme oxygenase-1 modulates early inflammatory responses: evidence from the heme oxygenase-1-deficient mouse. *Am J Pathol*, 165(3), 1045-53 (2004)
DOI: 10.1016/S0002-9440(10)63365-2
41. M. Watanabe-Matsui, A. Muto, T. Matsui, A. Itoh-Nakadai, O. Nakajima, K. Murayama, M. Yamamoto, M. Ikeda-Saito and K. Igarashi: Heme regulates B-cell differentiation, antibody class switch, and heme oxygenase-1 expression in B cells as a ligand of Bach2. *Blood*, 117(20), 5438-48 (2011)
DOI: 10.1182/blood-2010-07-296483
42. S. W. Ryter, J. Alam and A. M. K. Choi: Heme oxygenase-1/carbon monoxide: From basic science to therapeutic applications. *Physiological Reviews*, 86(2), 583-650 (2006)
DOI: 10.1152/physrev.00011.2005
43. Y. Nishio, M. Fujino, M. Zhao, T. Ishii, M. Ishizuka, H. Ito, K. Takahashi, F. Abe, M. Nakajima, T. Tanaka, S. Taketani, Y. Nagahara and X. K. Li: 5-Aminolevulinic acid combined

- with ferrous iron enhances the expression of heme oxygenase-1. *Int Immunopharmacol*, 19(2), 300-7 (2014)
DOI: 10.1016/j.intimp.2014.02.003
44. N. A. Kaidery, R. Banerjee, L. Yang, N. A. Smirnova, D. M. Hushpalian, K. T. Liby, C. R. Williams, M. Yamamoto, T. W. Kensler, R. R. Ratan, M. B. Sporn, M. F. Beal, I. G. Gazaryan and B. Thomas: Targeting Nrf2-mediated gene transcription by extremely potent synthetic triterpenoids attenuate dopaminergic neurotoxicity in the MPTP mouse model of Parkinson's disease. *Antioxid Redox Signal*, 18(2), 139-57 (2013)
DOI: 10.1089/ars.2011.4491
45. M. Zhao, H. Guo, J. Chen, M. Fujino, H. Ito, K. Takahashi, F. Abe, M. Nakajima, T. Tanaka, J. Wang, H. Huang, S. Zheng, M. Hei, J. Li, S. Huang, J. Li, X. Ma, Y. Chen, L. Zhao, J. Zhuang, P. Zhu and X. K. Li: 5-aminolevulinic acid combined with sodium ferrous citrate ameliorates H₂O₂-induced cardiomyocyte hypertrophy via activation of the MAPK/Nrf2/HO-1 pathway. *Am J Physiol Cell Physiol*, 308(8), C665-72 (2015)
DOI: 10.1152/ajpcell.00369.2014
46. M. Zhao, P. Zhu, M. Fujino, Y. Nishio, J. Chen, H. Ito, K. Takahashi, M. Nakajima, T. Tanaka, L. Zhao, J. Zhuang and X. K. Li: 5-Aminolevulinic acid with sodium ferrous citrate induces autophagy and protects cardiomyocytes from hypoxia-induced cellular injury through MAPK-Nrf-2-HO-1 signaling cascade. *Biochem Biophys Res Commun*, 479(4), 663-669 (2016)
DOI: 10.1016/j.bbrc.2016.09.156
47. J. C. Chen, K. C. Huang and W. W. Lin: HMG-CoA reductase inhibitors upregulate heme oxygenase-1 expression in murine RAW264.7. macrophages via ERK, p38 MAPK and protein kinase G pathways. *Cell Signal*, 18(1), 32-9 (2006)
DOI: 10.1016/j.cellsig.2005.03.016
48. G. Jiang, Y. Hu, L. Liu, J. Cai, C. Peng and Q. Li: Gastrodin protects against MPP(+)-induced oxidative stress by up regulates heme oxygenase-1 expression through p38 MAPK/Nrf2 pathway in human dopaminergic cells. *Neurochem Int*, 75, 79-88 (2014)
DOI: 10.1016/j.neuint.2014.06.003
49. E. M. Harrison, S. J. McNally, L. Devey, O. J. Garden, J. A. Ross and S. J. Wigmore: Insulin induces heme oxygenase-1 through the phosphatidylinositol 3-kinase/Akt pathway and the Nrf2 transcription factor in renal cells. *Febs Journal*, 273(11), 2345-2356 (2006)
DOI: 10.1111/j.1742-4658.2006.05224.x
50. R. M. Ogborne, S. A. Rushworth and M. A. O'Connell: Epigallocatechin activates haem oxygenase-1 expression via protein kinase C delta and Nrf2. *Biochemical and Biophysical Research Communications*, 373(4), 584-588 (2008)
DOI: 10.1016/j.bbrc.2008.06.068
51. A. I. Rojo, M. Salinas, M. Salazar, S. Takahashi, G. Suske, V. Calvo, M. R. de Sagarra and A. Cuadrado: Regulation of heme oxygenase-1 gene expression through the phosphatidylinositol 3-kinase/PKC-zeta pathway and Sp1. *Free Radical Biology and Medicine*, 41(2), 247-261 (2006)
DOI: 10.1016/j.freeradbiomed.2006.04.002
52. M. D. Maines, V. S. Raju and N. Panahian: Spin trap (N-t-butyl-alpha-phenylnitron)-mediated suprainduction of heme oxygenase-1 in kidney ischemia/reperfusion model: role of the oxygenase in protection against oxidative injury. *J Pharmacol Exp Ther*, 291(2), 911-9 (1999)
53. Y. C. Xiang, G. Liu, L. Yang and J. L. Zhong: UVA-induced protection of skin through the induction of heme oxygenase-1. *Bioscience Trends*, 5(6), 239-244 (2011)
DOI: 10.5582/bst.2011.v5.6.239
54. P. E. M. Gibbs and M. D. Maines: Biliverdin inhibits activation of NF-kappa B: Reversal of inhibition by human biliverdin reductase. *International Journal of Cancer*, 121(11), 2567-2574 (2007)
DOI: 10.1002/ijc.22978
55. S. W. Ryter and A. M. K. Choi: Heme Oxygenase-1/Carbon Monoxide From Metabolism to Molecular Therapy. *American Journal of Respiratory Cell and Molecular Biology*, 41(3), 251-260 (2009)
DOI: 10.1165/rcmb.2009-0170TR
56. P. Mandal, P. H. Park, M. R. McMullen, B. T. Pratt and L. E. Nagy: The Anti-Inflammatory Effects of Adiponectin Are Mediated via a Heme Oxygenase-1-Dependent Pathway in Rat Kupffer Cells. *Hepatology*, 51(4), 1420-1429 (2010)
DOI: 10.1002/hep.23427

57. S. W. Ryter and A. M. K. Choi: Regulation of Autophagy in Oxygen-Dependent Cellular Stress. *Current Pharmaceutical Design*, 19(15), 2747-2756 (2013)
DOI: 10.2174/1381612811319150010
58. I. Petrache, L. E. Otterbein, J. Alam, G. W. Wiegand and A. M. Choi: Heme oxygenase-1 inhibits TNF-alpha-induced apoptosis in cultured fibroblasts. *Am J Physiol Lung Cell Mol Physiol*, 278(2), L312-9 (2000)
DOI: 10.1152/ajplung.2000.278.2.L312
59. E. H. Carchman, J. Rao, P. A. Loughran, M. R. Rosengart and B. S. Zuckerbraun: Heme oxygenase-1-mediated autophagy protects against hepatocyte cell death and hepatic injury from infection/sepsis in mice. *Hepatology*, 53(6), 2053-62 (2011)
DOI: 10.1002/hep.24324
60. P. Waltz, E. H. Carchman, A. C. Young, J. Rao, M. R. Rosengart, D. Kaczorowski and B. S. Zuckerbraun: Lipopolysaccharide induces autophagic signaling in macrophages via a TLR4, heme oxygenase-1 dependent pathway. *Autophagy*, 7(3), 315-20 (2011)
DOI: 10.4161/auto.7.3.14044
61. D. Morse, L. Lin, A. M. Choi and S. W. Ryter: Heme oxygenase-1, a critical arbitrator of cell death pathways in lung injury and disease. *Free Radic Biol Med*, 47(1), 1-12 (2009)
DOI: 10.1016/j.freeradbiomed.2009.04.007
62. Y. K. Wang, R. Z. Chen, M. H. Li, Y. Q. Xie, Y. Z. Zou, J. B. Ge and H. Z. Chen: The Role of Triggering Receptor Expressed on Myeloid Cells-1 in the Pathogenesis of Acute Viral Myocarditis. *Circulation*, 122(2), E327-E327 (2010)
63. Y. Zhao, L. Zhang, Y. Qiao, X. Zhou, G. Wu, L. Wang, Y. Peng, X. Dong, H. Huang, L. Si, X. Zhang, L. Zhang, J. Li, W. Wang, L. Zhou and X. Gao: Heme oxygenase-1 prevents cardiac dysfunction in streptozotocin-diabetic mice by reducing inflammation, oxidative stress, apoptosis and enhancing autophagy. *PLoS One*, 8(9), e75927 (2013)
DOI: 10.1371/journal.pone.0075927
64. J. Hou, Q. Zhang, M. Fujino, S. Cai, H. Ito, K. Takahashi, F. Abe, M. Nakajima, T. Tanaka, J. Xu, H. Zou, Q. Ding and X. K. Li: 5-Aminolevulinic acid with ferrous iron induces permanent cardiac allograft acceptance in mice via induction of regulatory cells. *J Heart Lung Transplant*, 34(2), 254-63 (2015)
DOI: 10.1016/j.healun.2014.09.037
65. M. Zhao, J. Chen, P. Zhu, M. Fujino, T. Takahara, S. Toyama, A. Tomita, L. Zhao, Z. Yang, M. Hei, L. Zhong, J. Zhuang, S. Kimura and X. K. Li: Dihydroquercetin (DHQ) ameliorated concanavalin A-induced mouse experimental fulminant hepatitis and enhanced HO-1 expression through MAPK/Nrf2 antioxidant pathway in RAW cells. *Int Immunopharmacol*, 28(2), 938-44 (2015)
DOI: 10.1016/j.intimp.2015.04.032
66. S. S. Ambegaokar and D. L. Kolson: Heme oxygenase-1 dysregulation in the brain: implications for HIV-associated neurocognitive disorders. *Curr HIV Res*, 12(3), 174-88 (2014)
DOI: 10.2174/1570162X12666140526122709
67. K. Sahin, M. Tuzcu, C. Orhan, H. Gencoglu, M. Ulas, M. Atalay, N. Sahin, A. Hayirli and J. R. Komorowski: The effects of chromium picolinate and chromium histidinate administration on NF-kappaB and Nrf2/HO-1 pathway in the brain of diabetic rats. *Biol Trace Elem Res*, 150(1-3), 291-6 (2012)
DOI: 10.1007/s12011-012-9475-9
68. I. Lastres-Becker, A. Ulusoy, N. G. Innamorato, G. Sahin, A. Rabano, D. Kirik and A. Cuadrado: alpha-Synuclein expression and Nrf2 deficiency cooperate to aggravate protein aggregation, neuronal death and inflammation in early-stage Parkinson's disease. *Hum Mol Genet*, 21(14), 3173-92 (2012)
DOI: 10.1093/hmg/dds143
69. Y. M. Ha, M. Y. Kim, M. K. Park, Y. S. Lee, Y. M. Kim, H. J. Kim, J. H. Lee and K. C. Chang: Higenamine reduces HMGB1 during hypoxia-induced brain injury by induction of heme oxygenase-1 through PI3K/Akt/Nrf-2 signal pathways. *Apoptosis*, 17(5), 463-74 (2012)
DOI: 10.1007/s10495-011-0688-8
70. S. Maeda, I. Nakatsuka, Y. Hayashi, H. Higuchi, M. Shimada and T. Miyawaki: Heme oxygenase-1 induction in the brain during lipopolysaccharide-induced acute inflammation. *Neuropsychiatr Dis Treat*, 4(3), 663-7 (2008)
DOI: 10.2147/NDT.S3063

71. S. Rossi, M. D'Amico, A. Capuano, M. Romano, P. Petronella and C. Di Filippo: Hyperglycemia in streptozotocin-induced diabetes leads to persistent inflammation and tissue damage following uveitis due to reduced levels of ciliary body heme oxygenase-1. *Mediators Inflamm*, 2006(4), 60285 (2006)
DOI: 10.1155/MI/2006/60285
72. C. L. L. Saw, A. Y. Yang, M. T. Huang, Y. Liu, J. H. Lee, T. O. Khor, Z. Y. Su, L. M. Shu, Y. P. Lu, A. H. Conney and A. N. T. Kong: Nrf2 null enhances UVB-induced skin inflammation and extracellular matrix damages. *Cell and Bioscience*, 4 (2014)
DOI: 10.1186/2045-3701-4-39
73. Y. Iba, K. Watanabe, K. Ozaki, O. Aozasa, K. Ishizawa, T. Matsuura, H. Oyama and T. Masukawa: Altered gene expression profiles associated with enhanced skin inflammation induced by 12-O-tetradecanoylphorbol-13-acetate in streptozotocin-diabetic mice. *International Immunopharmacology*, 15(3), 614-619 (2013)
DOI: 10.1016/j.intimp.2013.01.007
74. E. J. Park, S. W. Park, H. J. Kim, J. H. Kwak, D. U. Lee and K. C. Chang: Dehydrocostuslactone inhibits LPS-induced inflammation by p38MAPK-dependent induction of hemeoxygenase-1 *in vitro* and improves survival of mice in CLP-induced sepsis *in vivo*. *International Immunopharmacology*, 22(2), 332-340 (2014)
DOI: 10.1016/j.intimp.2014.07.012
75. F. M. Konrad, S. Braun, K. C. Ngamsri, I. Vollmer and J. Reutershan: Heme oxygenase-1 attenuates acute pulmonary inflammation by decreasing the release of segmented neutrophils from the bone marrow. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 307(9), L707-L717 (2014)
DOI: 10.1152/ajplung.00145.2014
76. Y. Zhang, G. Jiang, M. Sauler and P. J. Lee: Lung endothelial HO-1 targeting *in vivo* using lentiviral miRNA regulates apoptosis and autophagy during oxidant injury. *Faseb Journal*, 27(10), 4041-4058 (2013)
DOI: 10.1096/fj.13-231225
77. M. Chang, J. Xue, V. Sharma and A. Habtezion: Protective role of hemeoxygenase-1 in gastrointestinal diseases. *Cell Mol Life Sci* (2014)
DOI: 10.1007/s00018-014-1790-1
78. J. Q. Ma, J. Ding, L. Zhang and C. M. Liu: Protective effects of ursolic acid in an experimental model of liver fibrosis through Nrf2/ARE pathway. *Clin Res Hepatol Gastroenterol* (2014)
DOI: 10.1016/j.clinre.2014.09.007
79. S. Shimada, M. Fukai, K. Wakayama, T. Ishikawa, N. Kobayashi, T. Kimura, K. Yamashita, T. Kamiyama, T. Shimamura, A. Taketomi and S. Todo: Hydrogen sulfide augments survival signals in warm ischemia and reperfusion of the mouse liver. *Surg Today* (2014)
DOI: 10.1007/s00595-014-1064-4
80. J. Huang, X. D. Shen, S. Yue, J. Zhu, F. Gao, Y. Zhai, R. W. Busuttill, B. Ke and J. W. Kupiec-Weglinski: Adoptive transfer of heme oxygenase-1 (HO-1)-modified macrophages rescues the nuclear factor erythroid 2-related factor (Nrf2) antiinflammatory phenotype in liver ischemia/reperfusion injury. *Mol Med*, 20, 448-55 (2014)
DOI: 10.2119/molmed.2014.00103
81. A. Gerbitz, P. Ewing, A. Wilke, T. Schubert, G. Eissner, B. Dietl, R. Andreesen, K. R. Cooke and E. Holler: Induction of heme oxygenase-1 before conditioning results in improved survival and reduced graft-versus-host disease after experimental allogeneic bone marrow transplantation. *Biol Blood Marrow Transplant*, 10(7), 461-72 (2004)
DOI: 10.1016/j.bbmt.2004.04.001
82. H. Shimizu, T. Takahashi, T. Suzuki, A. Yamasaki, T. Fujiwara, Y. Odaka, M. Hirakawa, H. Fujita and R. Akagi: Protective effect of heme oxygenase induction in ischemic acute renal failure. *Crit Care Med*, 28(3), 809-17 (2000)
DOI: 10.1097/00003246-200003000-00033
83. R. L. Maser, D. Vassmer, B. S. Magenheimer and J. P. Calvet: Oxidant stress and reduced antioxidant enzyme protection in polycystic kidney disease. *J Am Soc Nephrol*, 13(4), 991-9 (2002)
84. J. Zhou, X. Ouyang, T. R. Schoeb, S. Bolisetty, X. Cui, S. Mrug, B. K. Yoder, M. R. Johnson, A. J. Szalai and M. Mrug: Kidney injury accelerates cystogenesis via

pathways modulated by heme oxygenase and complement. *J Am Soc Nephrol*, 23(7), 1161-71 (2012)
DOI: 10.1681/ASN.2011050442

Key Words: Myeloid, Heme Oxygenase-1, Anti-inflammation, Review

Send correspondence to: Masayuki Fujino, National Institute of Infectious Diseases, 1-23-1, Toyama, Shinjuku-ku, Tokyo 162-8640, Japan, Tel: 81-3-5258-1111 Fax+81-3-5258-1150 E-mail: mfujino-kkr@umin.ac.jp