The zinc sensing receptor, ZnR/GPR39, in health and disease

Laxmi Sunuwar¹, David Gilad¹, Michal Hershfinkel¹

¹Department of Physiology and Cell Biology and The Zlotowski Center for Neuroscience, Faculty of Health Sciences, Ben-Gurion university of the Negev, Beer Sheva, Israel

TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. ZnR/GPR39 in health and disease
 - 3.1. A distinct Zn²⁺-sensing Gq-protein coupled Receptor
 - 3.2. Crosstalk between Zn²⁺, pH and ZnR/GPR39
 - 3.3. Zn²⁺ acts as a neurotransmitter via ZnR/GPR39
 - 3.3.1. The Zn²⁺-containing neuron
 - 3.3.2. ZnR/GPR39 in neurons
 - 3.3.3. ZnR/GPR39 plays a homeostatic role during seizure activity
 - 3.4. ZnR/GPR39 signaling in Epithelial Cells
 - 3.4.1. The digestive system

3.4.2. Skin

- 3.5. A paracrine role for ZnR/GPR39
- 3.6. ZnR/GPR39 in cancer
- 4. Conclusions and future prospects
- 5. References

1. ABSTRACT

While zinc has had a well-established structural role for many years, it is only during the last two decades that its role as a signaling molecule has been recognized. Ionic zinc, Zn2+, that is endogenously released during physiological activity acts as a first messenger, triggering the activity of a distinct Zn²⁺-sensing-receptor, ZnR. The ZnR is a member of the Gq-coupled receptor family, and the molecular moiety mediating its activity is GPR39. In this review, we will discuss the role of the ZnR/GPR39 in mediating Zn²⁺-dependent signaling in epithelial tissues and in neurons, where Zn²⁺ homeostasis plays physiological as well as pathological roles. Importantly, ZnR/GPR39 activates signaling that regulates a remarkably wide range of cell functions, including proliferation, differentiation and survival, as well as modulation of ion transport, and thereby, regulation of Na⁺, H⁺ and Cl homeostasis. Moreover, signaling activated by ZnR/GPR39 plays a key role in mediating effects of Zn^{2+} in health and disease. Thus, ZnR/GPR39 provides a unique target for therapeutically modifying the actions of zinc in a specific and selective manner.

2. INTRODUCTION

Ionic zinc, Zn²⁺, plays an essential role in the development and function of every system in the body. Its involvement is particularly prominent in the digestive, immune, nervous, secretory and integumentary systems (1-6). The utility of zinc in treatment of airway spasms or skin wounds, was described already in the 19th century (7, 8). A link between Zn²⁺ and carbonic anhydrase was shown 70 years ago (9, 10), and this was followed by the identification of Zn2+ as a cofactor of numerous enzymes. Zinc was then found to play an important structural role in various proteins, particularly transcription factors (11-13). Based on its importance in numerous physiological and cellular processes, it is hardly surprising that changes in extracellular as well as intracellular Zn²⁺ content trigger cellular signaling pathways. Moreover, Zn2+ transport into or out of organelles or its release from buffering proteins into the cytoplasm trigger subcellular changes in Zn²⁺ that is now considered a second messenger (14, 15). In this way, Zn²⁺ is now recognized as a metal ion that acts as a first or second messenger in physiological processes (16, 17). While most cellular Zn^{2+} is bound to proteins, transient changes in its extracellular or intracellular

concentration occur under both physiological and pathological conditions. Its functional significance is shown definitively by the potentially lethal effects of Zn^{2+} deficiency or increases of cellular Zn^{2+} (18, 19).

The strict maintenance of intracellular Zn²⁺ homeostasis is accomplished by means of a plethora of discrete Zn2+ transporters and buffer proteins (16, 20-22). These are largely represented by three families of proteins: 1) the 10 member zinc transporter family (ZnTs, SLC30), which transports this ion out of the cell or into cytoplasmic organelles and vesicles, 2) the 14 member Zip (SLC39) family that moves Zn²⁺ into the cytoplasm, and 3) the Zn²⁺ chelating metallothioneins, loosely binding this ion within the cytoplasm (20, 21, 23-25). Cellular Zn2+- containing vesicles are particularly abundant in the pancreas, brain, mammary gland, salivary gland and digestive system, and exocytosis of these vesicles triggers transient changes in extracellular Zn²⁺ levels (26-31). Extracellular Zn²⁺ levels were also increased following injury of keratinocytic cells that resulted in release of endogenous Zn2+ (32). Activity of the released Zn2+ as a signaling molecule requires that changes in Zn²⁺ concentration are transient. Several mechanisms could operate to guickly induce reuptake of Zn²⁺ into the cells via the extensive transporter system consisting of Zip and ZnT proteins mentioned above. Buffering of intracellular Zn²⁺ by metallothioneins or extracellular Zn²⁺ by albumin or citrate may also serve to lower the extracellular free Zn²⁺ concentration (13, 33).

Extracellular Zn²⁺ acts as a signaling molecule via the ZnR/GPR39 (34, 35), which has been functionally identified as a Zn2+-dependent, G-protein coupled receptor that senses changes in extracellular Zn²⁺ and, in response, activates downstream signaling pathways. Importantly, ZnR/GPR39 can be activated by endogenous Zn²⁺ released from vesicles or following injury (32, 36-38). Extracellular Zn2+ also indirectly activates cell signaling via allosteric modulation of ZnR/GPR39 signaling , and hence, dissection of the pathways activated by this ion may reveal specific therapeutic targets. For example, while Zn²⁺ interacts with major neuronal membrane transporters, e.g., the dopamine transporter, NMDA, glycine and GABA and modulates their activity (39-43), the distinct downstream signaling pathways triggered by the ZnR/GPR39 may be essential for regulating neuronal inhibition (37, 38, 44). In epithelial cells, Zn2+ regulates the activity of purinergic receptors and the store-operated channel (SOC), representing an important link between Zn2+ and intracellular Ca2+ (45-47). In fibroblasts, extracellular Zn²⁺ upregulates the PI3 kinase pathway. leading to activation of AKT and increased cell survival (48). Similar effects have also been shown to be mediated by Zn²⁺-dependent activation of the mitogen-activated protein kinase (MAPK) (49). Although intracellular Zn2+ may affect

these pathways via regulation protein phosphatases (50, 51), ZnR/GPR39 is the major link between Zn²⁺ and both, the PI3 and MAP kinase pathways (52, 53).

3. ZnR/GPR39 IN HEALTH AND DISEASE

3.1. A distinct Zn²⁺-sensing Gq-protein coupled Receptor

The ZnR/GPR39 is activated by physiological concentrations of extracellular Zn²⁺, inducing release of Ca²⁺ from thapsigargin-sensitive intracellular stores via the IP3 pathway (35, 54). Inhibitors of Gag (55, 56), inositol 1,4,5-trisphosphate (IP3) receptor and the phospholipase C (PLC) attenuate this Zn2+-dependent Ca²⁺ rise, indicating that the Ca²⁺ release is mediated by activation of a Gag-coupled receptor (35, 52). ZnR/ GPR39-dependent Ca²⁺ release has thus far been observed in numerous epithelia, including colonocytes. keratinocytes, pancreatic cells, prostate cancer cells and salivary gland cells (52, 57-59). Interestingly, Zn2+ has a well-established role in the normal function of these tissues, and Zn²⁺ dyshomeostasis is associated with diarrhea, growth retardation, skin lesions, impaired salivary secretion and taste disorders (60, 61). The ZnR/GPR39-dependent Ca2+ rise induced by Zn2+, enhances activation of the mitogen activated protein kinase, MAPK, and PI3 kinase pathways that are closely linked to enhanced cell survival and proliferation (62). Thus, ZnR/GPR39 may be the mediator of many of the well-established, health-promoting functions of Zn²⁺ (63). Finally, Zn²⁺ dyshomeostasis is also associated with neurological disorders, including Alzheimer's disease, ischemia and epilepsy (64-67). The ZnR/ GPR39 has been identified in neurons postsynaptic to vesicular Zn²⁺-containing synaptic boutons (68). The physiological role of ZnR/GPR39 in its diverse contexts will be discussed further in this review.

It should be emphasized that ZnR/GPR39 is highly specific to Zn²⁺, as other biologically relevant heavy metal ions (e.g. Mn²⁺, Cu²⁺ and Fe²⁺) do not produce a Ca2+ response (35). In addition, ZnR/ GPR39 is sufficient and necessary to trigger Zn2+dependent signaling. Nevertheless, it can interact with another, well-described cation receptor, the Ca2+ sensing receptor (CaSR (69, 70)). Heterodimerization of GPCRs diversifies the physiological response of these receptors to their ligands and may play an important role in their regulation (71, 72). The ZnR/ GPR39 and CaSR exhibit similarities in their signaling pathways, for instance, both are activated via a Gqdependent mechanism. Although Zn²⁺-dependent activity does not require the presence of extracellular Ca²⁺, this ion alters the apparent cooperativity and affinity of ZnR/GPR39 to Zn²⁺ (57). Similarly, spermine, a CaSR ligand, synergistically increases the cellular response when applied with Zn²⁺, though it does not activate the ZnR/GPR39 itself (53). Finally, silencing

of the CaSR downregulates ZnR/GPR39 response, and direct interaction between the CaSR and ZnR/ GPR39 is monitored using co-immunoprecipitation (53). Previous studies have shown that changes in the surface expression of the CaSR occur following exposure to its ligand (73). We posit that CaSR localization, following spermine or Ca²⁺ application, also affects the surface expression of ZnR/GPR39, thereby enhancing the Zn²⁺-dependent response.

Regulation of GPCRs activity is also achieved by desensitization, occurring after a brief exposure to their ligands. In this process, the desensitized receptor is internalized and may undergo degradation (74, 75). Differences in the degree of functional desensitization among GPCRs reflect the ratio between the recycling versus degradation of the receptor (76). Following its release, decrease of extracellular Zn²⁺ level may be facilitated by its reuptake via the Zip transporters. found in most cells, or chelation by Zn²⁺-binding proteins. However extracellular Zn²⁺, in contrast to most GPCRs ligands, is not rapidly degraded. Protection of cells from excessive Ca2+ signals triggered by continued activation of ZnR/GPR39 is achieved via desensitization of the ZnR/GPR39. Exposure to subtoxic concentrations of Zn2+ leads to profound and prolonged desensitization likely involving ZnR/GPR39 degradation (52, 58, 68). In the normal prostate Zn²⁺ is found in the presence of citrate, an extracellular Zn²⁺ binding protein that is especially abundant in the prostate. While the complex of Zn²⁺ with citrate does not activate ZnR/GPR39 signaling, it does induce desensitization of the ZnR/GPR39 (58). This may suggest a mechanism by which the ZnR/GPR39 is quiescent in the normal prostate when Zn²⁺ is largely complexed with citrate, but nevertheless desensitizes the receptor. However, during carcinogenesis when citrate and Zn²⁺ levels decrease (77-78). ZnR/GPR39 will be re-sensitized and can function to enhance cell proliferation in the prostate (58).

3.2. Crosstalk between Zn²⁺, pH and ZnR/GPR39

Analysis of the structural basis for constitutive activity of GPR39, known then as an orphan receptor, revealed that application of Zn²⁺ increased the activity of this receptor (79, 80). Though constitutive activity of this receptor was less than half compared to other neurotensin- and ahrelin-receptor family members. Zn²⁺ was considered of little physiological significance and not suggested as an endogenous ligand of GPR39. Instead, obestatin, a short peptide linked to obesity, was suggested to activate GPR39 (81), but activation of GPR39 by this peptide was not reproduced in further studies (82, 83). In contrast, an unbiased study identified Zn²⁺ as the endogenous agonist of GPR39 (84). The Zn²⁺ binding site on GPR39 was found to consist of two histidine residues: His17, His19 (85), and an aspartate residue: Asp313. This Asp313

was suggested to act as a tethered inverse agonist that upon binding of Zn2+ is diverted to enable Zn2+ binding to the histidines. Binding of Zn²⁺ to histidine occurs via an imidazole group and is pH sensitive, being most efficient between pH 7-8. Hence, when the extracellular pH drops to 6.5, the Zn2+ activated, ZnR/ GPR39-dependent Ca²⁺ response and subsequent phosphorylation of MAP or PI3 kinase is completely abolished (32, 86, 87). Attenuation of Zn²⁺-dependent signaling at pH 6.5 is short-lived and reversible (86, 87). By overexpressing GPR39 mutated at what was expected to be the pH sensitive residues, i.e., His17 and His19, we showed that ZnR/GPR39 maintains its pH-dependence and Zn²⁺ signaling is still abolished at pH 6.5. Similarly, other extracellular-facing histidines fail to reverse the pH sensitivity. Eventually, Asp313 was identified as the pH sensing component for ZnR/ GPR39. The replacement of this residue with the pH insensitive alanine results in Zn2+-dependent Ca2+ responses that are similar at pH 7.4. or 6.5. while its substitution by His or Glu pH-sensitive residues restores the pH sensitivity of the receptor (86). Reducing the activity of the ZnR/GPR39 following changes in extracellular, but not intracellular, pH may result from different protonation states of the involved residues (88). Yet another possible mechanism involves local conformational changes of the binding site as was shown for pH sensitivity of the Ca2+-sensing receptor and the mGluR4 glutamate receptor (89, 90).

Altogether. ZnR/GPR39 mediates Zn²⁺-dependent signaling while tuned to sense physiologically-relevant changes in extracellular pH (86). Importantly, changes in this range of pH commonly occur under physiological conditions within the digestive system lumen, the epidermis and the brain (91-95). Hence, the pH sensitivity of ZnR/GPR39 may serve as an important regulator of physiological and pathological responses to Zn²⁺. For example, inflammatory bowel disease may induce local pH changes that render ZnR/GPR39 signaling inefficient for Zn²⁺ enhancement of proliferation. Such mechanism could underlie the erosion of the epithelial layer, occurring in this disease (96).

Interestingly, ZnR/GPR39 itself regulates the pH of the intracellular and extracellular microenvironments via upregulation of Na⁺/H⁺ exchange (NHE) activity. The activity of NHE exchangers is upregulated following a drop in intracellular pH, serving as an important factor in the recovery from intracellular acid loads in many cell types (97). Activation of ZnR/ GPR39 signaling, and its downstream phosphorylation of the MAPK pathway, results in upregulation of NHE activity in colonocytes, keratinocytes and neurons (32, 36, 52, 87, 98). In all of these cells, the effect of Zn²⁺ on regulation of NHE is completely lost in the absence of ZnR/GPR39. Such upregulation of NHE activity may have important effects on the function of neurons or epithelial cells. In neurons, intracellular acid load, accumulated during repetitive firing, is largely the result of metabolic H⁺ generation (99). The intracellular pH changes can affect neuronal excitability by modulating the activity of ion channels, transporters and receptors (100-102). Thus, ZnR/GPR39-dependent regulation of intracellular pH may play a role in neuronal excitability.

By mediating Na⁺-dependent H⁺ export, NHE exchangers may induce changes in the extracellular pH while accelerating recovery of the intracellular environment. In neurons, acidifying the cell surfaces that abut the synaptic cleft can regulate various functions, including GABA signaling or dendritic spine growth (103, 104). On the other hand, sustained NHE activity contributes to tissue acidosis during ischemic neuronal injury (105). Under these conditions, inhibition of NHE was claimed to be neuroprotective (106, 107). The pH sensitivity of neuronal ZnR/GPR39. which is inactive at acidic pH, suggests a homeostatic mechanism by which NHE-mediated decrease of the extracellular pH following Zn²⁺ activation of ZnR/ GPR39, serves to prevent excessive tissue acidification (87). In keratinocytes, ZnR/GPR39 also upregulates NHE activity (32), which may result in an acidic apical surface. In the skin, such acidification is required for formation of an effective permeability barrier (108) and NHE1 is an important regulator of this function (109). Interestingly, Zn²⁺ deficiency is commonly associated with inflammation, possibly reflecting breakdown of this barrier. Elegant in vivo experiments have shown an important role for NHE2 activation in gastric epithelial repair (110), interestingly NHE2 activity in this study did not induce changes in surface pH because they may have been masked by HCO₂ buffering. It is not clear if ZnR/GPR39 upregulation of NHE activity in the presence of physiological HCO₃ affects extracellular pH in various tissues, hence further studies comparing the role of ZnR/GPR39 and NHE in the presence of physiological concentrations of HCO₂ are required.

3.3. Zn²⁺ acts as a neurotransmitter via ZnR/ GPR39

3.3.1. The Zn²⁺-containing neuron

Deficiency of Zn^{2+} is associated with developmental malformations and impaired cognitive performance (66, 111). There is an increasing body of evidence suggesting that disturbances of Zn^{2+} homeostasis play an important role in the etiology of various neurological disorders, including Alzheimer's disease (112, 113), amyotrophic lateral sclerosis (114, 115), ischemia (116-118), spreading depression (119-121) and autistic spectrum disorders (122, 123). Zinc deficiency has also been closely associated to epilepsy, with lack of dietary Zn^{2+} leading to enhanced susceptibility to epileptic seizures in mouse models (124, 125) as well as in humans (126-133). Furthermore, Zn²⁺ administrated to mice, in studies using a kindling model of epilepsy, reduced seizure activity (134, 135). In contrast, excessive rise of extracellular as well as intracellular Zn²⁺, correlates with neurotoxicity and cell death (51, 118, 136-138). Rapid increases in the concentration of intracellular Zn²⁺ may occur following an episode of oxidative or nitrosative stress, inducing liberation of Zn²⁺ bound to intracellular proteins and frequently resulting in cell death (18, 51, 118, 139). For this reason, Zn²⁺ chelation is increasingly contemplated as a potentially viable therapeutic strategy in some neurological conditions such as ischemia/stroke (138, 140-142), though such an approach could be detrimental in others (143). In contrast, in Alzheimer's disease, metal ionophores that may restore intracellular Zn²⁺ levels have been considered for use as therapeutic agents (144).

A unique pool of Zn²⁺, representing a mere 10% of total brain zinc. is concentrated in synaptic vesicles within a subclass of excitatory cortical neurons, it is often referred to as 'synaptic Zn^{2+'}. It is this Zn2+ that is demonstrated by the so-called 'Timm's' staining method (29, 145, 146). Loading of Zn²⁺ into synaptic vesicles is mediated by a specific transporter. ZnT3, which is expressed in discrete regions of the brain, including hippocampus, amygdala, neocortex and auditory brainstem among other regions. Knockout of the ZnT3 gene results in mice lacking synaptic Zn2+ (125, 147). Early studies did not show a clear phenotype of these mice except for enhanced susceptibility to seizure, yet later studies indicate that ZnT3 knockout mice may also have impaired learning, memory and fear-conditioning processes and autistic-like behavior (148-154). Synaptic Zn²⁺ is stored together with glutamate, the principle excitatory neurotransmitter of the mammalian CNS, and is co-released with it into the synaptic cleft (136, 155) in a Ca2+- and activitydependent manner (156-159). Synaptic Zn2+ has been suggested to modulate membrane excitability via direct interaction with post synaptic targets, e.g., GABA, NMDA and glycine receptors (39, 66, 160-165). Binding of synaptic Zn²⁺ to ZnR/GPR39 provides a pathway underlying, for example, the effects of Zn²⁺ during seizure (44).

3.3.2. ZnR/GPR39 in neurons

Metabotropic pathways, triggering slow intracellular second messenger systems, play a critical role in neurotransmission (166). Indeed, metabotropic receptors activate intracellular signaling leading to delayed modulation of ion channels and membrane transporters. Glutamate receptors (mGluRs), for example, mediate changes in synaptic plasticity by activating the mitogen-activated protein kinase, MAPK, pathway (167, 168). Hence, we hypothesized that the metabotropic Zn²⁺ sensing receptor ZnR/GPR39 is a distinct target of synaptic Zn²⁺ that may underlie the

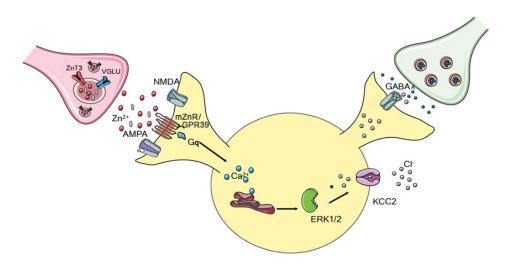


Figure 1. Schematic representations of ZnR/GPR39 signaling in CA3 neurons. Zn²⁺ is transported into synaptic vesicles by ZnT3 and is stored and coreleased together with glutamate. Synaptically released Zn²⁺ may then bind ZnR/GPR39, inducing post-synaptic intracellular Ca²⁺ rises and activation of the MAPK pathway. ZnR/GPR39 signaling enhances KCC2 activity and the Cl⁻ gradient, affecting GABA_A inhibitory responses. Image was created using Servier Medical Art. ZnT3 - Zn²⁺ transporter 3, VGLUT – Vesicular glutamate transporter, ZnR/GPR39 – metabotropic Zn²⁺-sensing receptor, mGluR – metabotropic glutamate receptor, AMPA and NMDA are ionotropic glutamate receptors, MAPK – mitogen activated kinase.

effects of this ion in the brain. Initial efforts focused on the hippocampus, as this region demonstrates intense Timm's staining, particularly of mossy fiber boutons aligned with their postsynaptic, CA3 neurons (28). Application of exogenous Zn²⁺ resulted in intracellular Ca²⁺ release from the endoplasmic reticulum stores in these neurons, but not in adjacent astroglia (38, 68). Pharmacological dissection of the signaling pathway triggered by Zn²⁺ revealed that intracellular Ca²⁺ rise is PLC and Gq dependent (see Figure 1), in accordance with ZnR/GPR39 activity seen in epithelial cells. Furthermore, activation of MAP kinase by Zn²⁺ in neurons requires ZnR/GPR39 activation and the resulting Ca²⁺ release (68, 87). Importantly, physiological stimulation of the mossy fibers was shown to trigger endogenous synaptic Zn²⁺ release (157, 158) and ZnR/GPR39-dependent Ca²⁺ rises in the post synaptic CA3 neurons (38). The presence of a non-membrane permeable, extracellular, Zn²⁺ chelator reduced the stimulus-induced Ca2+ rise suggesting that the endogenous synaptic Zn²⁺ is sufficient for activating ZnR/GPR39. ZnR/GPR39-dependent Ca2+signaling was monitored when exogenous Zn2+ was applied to hippocampal slices from ZnT3 knockout mice, which lack synaptic Zn²⁺, but stimulation of the mossy fibers in these slices yielded a significantly decreased Ca2+ signal in the CA3 neurons that was similar to the signal triggered in WT slices in the presence of the Zn²⁺ chelator (38). Thus, synaptic Zn²⁺ release is essential for a postsynaptic metabotropic response, indicating the physiological relevance of this receptor (68). Importantly, the Zn²⁺-dependent Ca²⁺ signaling is absent in GPR39 knockout mice supporting the conclusion that the molecular moiety mediating neuronal ZnR responses is GPR39 (38, 169). Similar ZnR/GPR39 activity, triggered by

physiologically relevant electrostimulation of synaptic Zn²⁺-containing fibers, has been described in the dorsal cochlear nucleus, a region of the auditory brainstem (37). In these neurons, activation of ZnR/GPR39 inhibits glutamate release by inducing synthesis of the endocannabinoid, 2-arachidonoylglycerol (2-AG), which reduces synaptic strength (37). A role for ZnR/GPR39 in regulation of the CREB/BDNF/TrkB pathway, and thereby in depression, has also been postulated, though it is not clear at present how Gq signaling activates this pathway or whether these effects are lost in ZnR/GPR39 knockout mice (170, 171).

In the hippocampus, ZnR/GPR39 activation by synaptic Zn²⁺ results in enhancement of inhibitory tone, apparently by means of an increase in the neuronal Cl⁻ efflux pathway (38, 169). The major neuronal outward transporter of CI- is the K+/CIcotransporter, KCC2, which is necessary and sufficient to create a CI⁻ equilibrium potential that is negative with respect to the resting membrane voltage (172, 173). The activity of KCC2 is therefore crucial for rendering Cl⁻ channels, i.e. GABA, and glycine receptors, activity as inhibitory (174, 175). This important co-transporter is highly regulated via its phosphorylation, and by changes in its expression during neuronal activity, thereby modulating the inhibitory effect of GABA and glycine (176-178). Interestingly, ZnR/GPR39 activation in neurons and downstream phosphorylation of MAPK results in enhanced K⁺-dependent Cl⁻ transport that is mediated by KCC2. This activity is not reproduced in ZnR/GPR39 knockout mice (38, 169). It has been shown that Gq-dependent signaling enhances KCC2 surface expression and thereby upregulates KCC2dependent transport (38). These results were the first demonstration of a direct and distinct target for synaptic Zn^{2+} and suggest that this ion acts as a neurotransmitter (Figure 1). Moreover, in this way, neuronal inhibitory tone is enhanced by Zn^{2+} binding to ZnR/GPR39.

3.3.3. ZnR/GPR39 plays a homeostatic role during seizure activity

Importantly, decreased KCC2 function renders the, normally inhibitory, GABA post-synaptic potentials excitatory thereby increasing seizure susceptibility (175, 176, 179). On the other hand, a study in neonatal rats revealed that a single seizure episode in vivo or a brief treatment of acute brain slices with the excitotoxin, kainite, result in activation of KCC2 in the hippocampus. This correlated with an increase in the plasmalemal fraction of KCC2, and not with an increase in its overall concentration (180). As suggested above, loss of synaptic Zn²⁺, thought to act as an inhibitory neuromodulator, is associated with epileptogenesis (181, 182). Moreover, administration of a Zn²⁺-deficient diet is sufficient to reduce synaptic Zn²⁺ levels, and results in greater susceptibility to kainate-induced seizures (183, 184). Similarly, ZnT3 knockout mice show enhanced sensitivity to seizureinducing pharmaceuticals, including the glutamatergic excitotoxin, kainite, which induces limbic seizures (124). More recent studies show that ZnT3 knockout mice are also more prone to febrile hyperthermia induced seizures (133). Interestingly, Zn²⁺ supplementation may decrease febrile seizure recurrence in children (185).

It was demonstrated that Zn²⁺, acting via ZnR/GPR39, upregulates KCC2 activity and may be sufficient to decrease seizure severity. Indeed, ZnR/GPR39 knockout animals exhibit an enhanced susceptibility to kainate-induced seizures, exhibiting significantly higher behavioral seizure severity scores and more seizures over longer periods of time, compared to wildtype controls (44). Treating hippocampal slices with kainate is sufficient to upregulate KCC2 activity in an extracellular Zn2+dependent manner. Kainate-induced upregulation of KCC2 is also dependent on ZnR/GPR39 signaling. as it is abolished by treatment with Gg. PLC or MAPK inhibitors. Finally, kainate-induced KCC2 upregulation is absent in ZnR/GPR39 knockout mice, suggesting that the receptor itself is necessary for this process. Taken together, these findings support a homeostatic role for ZnR/GPR39 triggered by seizure-induced synaptic Zn²⁺ release (44).

3.4. ZnR/GPR39 signaling in epithelial cells

3.4.1. The digestive system

The intestinal epithelial layer separates the body from the luminal contents, which includes metabolites but also toxins, bacteria and pathogens. Hence, it is hardly surprising that this laver undergoes continuous renewal of its cell population every 3-4 days, a process requiring continuous cell proliferation and differentiation. This layer of epithelium, on one hand, is responsible for the selective uptake of digested metabolites from the lumen, which necessitates multiple transporter proteins. On the other hand, this epithelium requires tight anatomical and physiological barrier to prevent invasion by foreign organisms and substances. Numerous studies link Zn²⁺ to proper function of the digestive system, including absorption and barrier functions (186-189). At the cellular level, Zn²⁺ promotes proliferation, differentiation, survival (17, 98) and barrier formation (190-193) in colon epithelial cell (colonocyte) cultures. The presence of a Zn2+triggered, Gq-dependent mechanism for activation of Ca²⁺ cellular signaling in colonocytes enabled the demonstration of a functional ZnR/GPR39 in these cells (35).

Initial studies indicated that GPR39 is widelyexpressed throughout the digestive system (194, 195). We subsequently showed that luminal application of Zn²⁺ is sufficient to activate ZnR/GPR39, suggesting that the receptor is present on the apical side of colon epithelial cells, facing the colonic lumen (98). Sources of luminal Zn2+ that could interact with ZnR/ GPR39 include exogenous or dietary Zn²⁺, as well as endogenous sources, e.g., Zn2+ released from pancreatic digestive enzymes, Zn²⁺ from salivary gland vesicles or from Paneth cells in the intestinal epithelium (26-31, 196). In addition, Zn²⁺ is released from all mammalian cells following injury or death (32), and more selectively via a process mediated by Zn²⁺ transporters such as ZnT6 (196). Finally, endogenous Zn²⁺ released into the gastrointestinal lumen via epithelial shedding alone may account for its physiological concentration (98), sufficient to activate the ZnR/GPR39.

As mentioned above, Zn2+- activated, ZnR/ GPR39-dependent Ca²⁺ signaling plays a key role in enhancing proliferation of epithelial cells (35, 98). The downstream pathways activated by ZnR/ GPR39 include MAPK and AKT, both hallmarks of cell proliferation and survival (36, 52). Both are of major importance to the constant renewal of the epithelium, required to replace those cells constantly shed into the intestinal lumen. It seems noteworthy as well that ZnR/GPR39 has been implicated in accelerating proliferation and differentiation of preadipocytes (198). The short chain fatty acid, butyrate, present at high concentrations in the colon (199). imposes an acidic stress and exerts a pro-apoptotic effect on colonocytes (199-202). The acidic load imposed on colonocytes by the presence of butyrate (203) is reduced by activation of ZnR/GPR39 and its downstream activation of NHE (52, 98). Colonocyte cell death, induced by prolonged exposure to

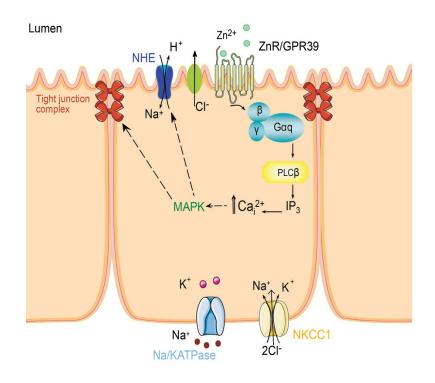


Figure 2. Schematic representations of ZnR/GPR39 signaling in colonocytes. Colonocytes form a permeability barrier via expression of tight junction complexes on the apical side and express ion transporters regulating Na⁺, Cl⁻ and K⁺ absorption. Zn²⁺ on the luminal side may activate the ZnR/GPR39 and trigger downstream Ca²⁺ rises and ERK1/2 pathway activation. This results in upregulation of the Na⁺/H⁺ exchanger and the formation of tight junctions. Image was created using Servier Medical Art. NHE - Na⁺/H⁺ exchanger, ZnR/GPR39 – metabotropic Zn²⁺-sensing receptor, MAPK – mitogen activated kinase, Na/KATPase – Na⁺/K⁺ ATPase, NKCC1 – Na⁺/K⁺/Cl- cotransporter 1.

butyrate, is attenuated by activation of ZnR/GPR39dependent MAP and PI3 kinase pathways (98). The rescue of cells from butyrate-induced cell death, however, requires further activation of the pro-survival glycoprotein, clusterin, also known as apolipoprotein J (204, 205). A role for ZnR/GPR39 in enhancing proliferation and cell survival (see Figure 2) may also underlie the effects of Zn²⁺ in therapies accelerating healing of gastric ulcers (206).

It has been suggested that Zn2+ reduces the severity and extends the times between bouts of inflammatory ulcerative disease, such as occur in Crohn's disease and colitis (186, 187, 207-209)). Amajor factor in these diseases is breakdown of the epithelial barrier that precedes the inflammatory response (210, 211). Under conditions of Zn²⁺ deficiency, occurring due to insufficient dietary Zn²⁺ or genetically-induced by loss of transporters responsible for Zn²⁺ absorption. barrier function is compromised, leading to increased permeability (190, 212, 213). On the other hand, Zn²⁺-dependent activation of ZnR/GPR39 enhances expression of junctional complex proteins ZO-1 and occludin, (36), critical elements of the specialized junctions between adjacent colonocytes that compose the intestinal barrier (214, 215). Moreover, the role of ZnR/GPR39 in regulating expression of these proteins was elucidated by their significant reduction in the colon epithelium of ZnR/GPR39 knockout mice (98).

Interestingly, redistribution of ZO-1 takes place early on during epithelial cell shedding, as it is essential for maintaining the barrier function during the ongoing epithelial renewal in the intestines (216). It would be of interest, then, to examine the effect of Zn^{2+} , released from redundant colon epithelial cells, on the expression and reorganization of ZO-1 during incorporation of new cells into the epithelial layer.

A crucial function of the intestinal epithelium is regulation of ion and solute transport, by which osmotic gradients for water movement are maintained (217). Interestingly, under normal conditions or during experimentally-induced diarrhea, Zn^{2+} was suggested to modulate ion transport in the colon (218, 219). In colon cell lines and native colonocytes, Zn^{2+} , acting via ZnR/GPR39, promotes upregulation of Na⁺/H⁺ exchanger activity (52, 98). While this reduces the intracellular acid load, it also induces rapid uptake of Na⁺ from the lumen. Because of the key role played by the colonocytic apical NHE3 in attenuating diarrhea (217, 220, 221), its upregulation by ZnR/GPR39 in the presence of Zn²⁺ is likely to enhance this protective effect.

3.4.2. Skin

One of the first physiological functions demonstrated for extracellular zinc involves its

enhancement of wound healing. Zinc deficiency has long been linked to skin lesions (222, 223). Topical application of zinc-containing ointments, moreover, stimulates wound healing and re-epithelialization processes (8, 223-225). Importantly, severe skin lesions manifest in both dietary and genetic Zn²⁺ deficiencies and can be reversed by dietary Zn²⁺ supplementation (226, 227). Concenital Zn²⁺ deficiency, accompanied by severe skin lesions, is also characteristic to disorders linked to dysfunction of Zn²⁺ transporters, such as Acrodermatitis Enteropathica (AE), a genetic mutation in the intestinal Zn²⁺ uptake transporter ZIP4, or the Transient Neonatal Zinc Deficiency (TNZD). a symptomatic disorder in breastfed babies that is associated with mutations in the mammary gland Zn²⁺ transporter ZnT2 (60, 228-230). Extracellular Zn2+, at concentrations found in the epidermis (222, 223, 231) triggers Ca²⁺ release from thapsigargin-sensitive stores that is largely mediated by ZnR/GPR39 (32). Moreover, injury of keratinocytes releases Zn²⁺ at concentrations that are sufficient to trigger ZnR/GPR39 response in neighboring keratinocytes (32). Surprisingly, this Zn²⁺dependent Ca²⁺ response in a keratinocyte epithelial skin cell-line possesses a dramatically higher affinity to Zn²⁺ compared to other cells (32). This may be explained by the tendency of G-protein coupled receptors to form dimers with other members, thereby affecting their affinity to their endogenous ligands (71, 72). Indeed. ZnR/GPR39 is able to interact with CaSR (53), suggesting interaction between GPCRs may underlie the high affinity of the keratinocytic ZnR/ GPR39 to Zn²⁺. In keratinocytes. Zn²⁺-dependent upregulation of MAPK activity and NHE ion transport are both mediated via activation of ZnR/GPR39 (32). The importance of ZnR/GPR39 upregulation of NHE activity in keratinocytes is underlined by the role of this transport in induction and maintenance of an acidic local extracellular microenvironment essential to the efficacy of the permeability barrier in the skin (108, 203, 232). It appears likely, therefore, that ZnR/ GPR39 activation may regulate the formation of the permeability barrier and enhance anti-inflammatory effects, associated with topical Zn²⁺ application during wound healing. Moreover, accelerated wound closure in the presence of Zn²⁺ indicates that activation of ZnR/ GPR39 also directly promotes healing, while silencing the receptor or pharmacologically inhibiting its signaling pathway reverses the effect (32). Interestingly, although in colonocytes ZnR/GPR39 upregulation of NHE does not affect cell survival (98), this pathway clearly enhances keratinocytes proliferation (32). Since the keratinocytic-ZnR/GPR39 undergoes rapid and profound desensitization following exposure to high levels of extracellular Zn²⁺ (32), topical application of Zn²⁺ for extended periods (for example in bandages) may actually hinder wound healing. As such, the discovery of agonists that activate, but that do not desensitize, ZnR/GPR39 could provide better wound healing solutions.

3.5. A paracrine role for ZnR/GPR39

Signaling pathways activated by ZnR/GPR39 are also involved in secretion of first messengers that may trigger paracrine signaling in neighboring cells. Indeed, activation of ZnR/GPR39 in a salivary gland duct epithelial cell line. HSY, induces release of ATP (57). Moreover, application of Zn²⁺ to co-cultures of HSY cells, expressing a functional ZnR/GPR39, and vascular smooth muscle cells (VSMCs) that do not express a functional ZnR (57), induces a Ca²⁺ rise in both cell types. To determine a paracrine effect, we showed that treatment with the non-permeable ATP scavenger apyrase inhibited the Zn²⁺-dependent Ca²⁺ rise in the VSMC but not in the HSY cells (57). Altogether these experiments suggested that ZnR/ GPR39 activity in HSY cells triggers ATP release that activated metabotropic signaling in the neighboring VSMC.

Such paracrine effects influence and could amplify the increased proliferation and migration processes elicited by Zn²⁺- activated ZnR/GPR39 to neighboring cells, not expressing the receptor. Hence, Zn²⁺-activated ZnR/GPR39 may enhance the wound healing ability by promoting proliferation and migration of both keratinocytes and fibroblasts (233), although fibroblasts lack a functional ZnR (35). Taken together, ZnR/GPR39 may be capable of augmenting growth of neighboring metastatic cells following increased secretion of ATP or S100A4 (234).

3.6. ZnR/GPR39 in cancer

Since ZnR/GPR39 enhances cell proliferation and migration a role in carcinogenesis must be considered. A role for Zn²⁺ in the development of breast cancer has been explored, and changes in the expression or function of transporters from the ZIP and ZnT families have been linked to the development and progression of the malignant process (235-237). Similarly, butyrate has been shown to induce apoptosis of colon cancer cells. The finding that ZnR/GPR39 rescues colonocytes from butyrate-induced apoptosis (98), suggests a potential role for this receptor in the etiology of colon cancer that has yet to be addressed. In contrast, Zn²⁺ via ZnR/GPR39 signaling-induces release of intracellular Ca2+ in androgen-independent, but not in androgen-dependent prostate cancer cells (58). Changes in intracellular Ca2+ may also regulate S100A calcium binding proteins that enable cell migration and invasion of cancer cells, and are specifically linked to enhanced prostate cancer growth (238-240). Specifically, S100A4 is present in tumor interstitial fluid and is thought to enhance metastatic cell proliferation and angiogenesis (241). Proliferation and invasiveness of a prostate cancer cell line (PC3) is enhanced by S100A4 via induction of the metalloprotease, MMP-9 (242). Importantly, S100A4

expression is triggered in PC-3 cells by extracellular Zn^{2+} at concentrations that activate ZnR/GPR39 (53). In contrast, S100A4 expression is reduced following silencing of ZnR/GPR39 with siGPR39 (53). It will be interesting to assess the effect of ZnR/GPR39 expression on the level of extracellular S100A4 in prostate tumors and its relation to metastasis.

Robust changes in extracellular Zn²⁺ during prostate tumorigenesis have been suggested to serve as a marker for prostate cancer (243, 244). In PC-3 cells, ZnR/GPR39 activation induces PI3K pathway upregulation, measured via phosphorylation of AKT (53). Surprisingly. ZnR/GPR39 expression is itself associated with higher levels of total AKT expression, which are associated with a more malignant phenotype of adrenal carcinomas (245). In addition, constitutive activation of the PI3K signaling pathway is correlated with severity of prostate tumors (246). Importantly, zinc itself and zinc transporters, most prominent ZIP1, ZIP2, ZIP3 and ZnT7, have been implicated in playing an important role in prostate cancer (5, 77, 243, 247-250), but whether there is a causative or synergistic link between these changes and ZnR/ GPR39 activity is not understood. Finally, changes in GPR39 expression levels and patterns were observed in human esophageal squamous cell carcinoma (251) and in gastric adenocarcinomas (252), but a role for Zn²⁺ in activation of ZnR/GPR39 was not determined.

4. CONCLUSIONS AND FUTURE PROSPECTS

It is now established that ZnR/GPR39 is a distinct target for free ionic Zn²⁺ released in response to physiological or pathological activity. The receptor is a functional GPCR that mediates Zn²⁺-dependent signaling in epithelial cells and in neurons, providing for the first time, a molecular target to explain many of the physiological actions of Zn²⁺. Described for many years as a ubiquitous structural element of virtually all cells, Zn²⁺ is now identified, in addition, as a signaling molecule in a wide variety of contexts. The ZnR/GPR39 is an important regulator of Zn²⁺-dependent signaling, and may serve as a handle to modulate physiological processes.

Although Zn²⁺ plays a role in many pathological conditions, Zn²⁺ itself, because of its lack of specificity and the often conflicting intracellular versus extracellular effects, offers a far less attractive address for therapeutic interventions than a distinct GPCR. Hence, ZnR/GPR39 is a potentially promising candidate for therapeutic intervention in diseases as diverse as epilepsy and colitis, which in general are poorly controlled by existing therapies. By focusing on ZnR/GPR39, the molecular pathways responsible for the symptoms of diseases may be directly and effectively targeted. That the Gq protein-coupled receptor family (GPCR) is currently a major focus of the pharmaceutical industry (253, 254), provides reason for optimism that specific and effective agonists and antagonists for ZnR/GPR39 will be forthcoming.

5. ACKNOWLEDGEMENT

Laxmi Sunuwar and David Gilad contributed equally to this paper. We thank Dr. WF Silverman for helpful suggestions and critical reading of the manuscript.

6. REFERENCES

- N Roohani, R Hurrell, R Kelishadi, and R Schulin: Zinc and its importance for human health: An integrative review. *J Res Med Sci.* 18(2) 144-57 (2013)
- 2. AS Prasad: Zinc: an overview. *Nutrition. 11(1 Suppl)* 93-9 (1995)
- AS Prasad: Zinc in human health: effect of zinc on immune cells. *Mol Med.* 14(5-6) 353-7 (2008) DOI: 10.2119/2008-00033.Prasad
- HH Sandstead, CJ Frederickson, and JG Penland: History of zinc as related to brain function. *J Nutr.* 130(2S Suppl) 496S-502S (2000)
- SL Kelleher, NH McCormick, V Velasquez, and V Lopez: Zinc in specialized secretory tissues: roles in the pancreas, prostate, and mammary gland. *Adv Nutr.* 2(2) 101-11 (2011) DOI: 10.3945/an.110.000232
- T Kambe, BP Weaver, and GK Andrews: The genetics of essential metal homeostasis during development. *Genesis.* 46(4) 214-28 (2008) DOI: 10.1002/dvg.20382
- J Waters: On the Efficacy of Oxide of Zinc in Laryngismus Stridulus. *Prov Med Surg J* 2(33) 125-7 (1841) DOI: 10.1136/bmj.s1-2.33.125
- DG Barceloux: Zinc. J. Toxicol. Clin. Toxicol. 37(2) 279-92 (1999) DOI: 10.1081/CLT-100102426
- 9. BL Vallee: Zinc and carbonic anhydrase content of red cells in normals and in pernicious anemia. *J Clin Invest.* 27(4) 559 (1948)
- 10. BL Vallee and MD Altschule: Zinc in the mammalian organism, with particular

reference to carbonic anhydrase. *Physiol Rev.* 29(4) 370-88 (1949)

- 11. W Maret: Zinc biochemistry, physiology, and homeostasis – recent insights and current trends. *BioMetals*. *14(3-4)* 187-190 (2001) DOI: 10.1023/A:1012945110820
- BL Vallee and KH Falchuk: The biochemical basis of zinc physiology. *Physiol-Rev.* 73(1) 79-118 (1993)
- W Maret: Zinc biochemistry: from a single zinc enzyme to a key element of life. *Adv Nutr.* 4(1) 82-91 (2013) DOI: 10.3945/an.112.003038
- 14. S Yamasaki, K Sakata-Sogawa, A Hasegawa, T Suzuki, K Kabu, E Sato, T Kurosaki, S Yamashita, M Tokunaga, K Nishida, and T Hirano: Zinc is a novel intracellular second messenger. J. Cell Biol. 177(4) 637-45 (2007) DOI: 10.1083/jcb.200702081
- 15. W Maret: Zinc in the biosciences. *Metallomics.* 6(7) 1174 (2014) DOI: 10.1039/C4MT90021A
- T Fukada, S Yamasaki, K Nishida, M Murakami, and T Hirano: Zinc homeostasis and signaling in health and diseases: Zinc signaling. *J Biol Inorg Chem.* 16(7) 1123-34 (2011) DOI: 10.1007/s00775-011-0797-4
- 17. M Hershfinkel. Zinc, a Dynamic Signaling Molecule, In: Molecular Biology of Metal Homeostasis and Detoxification, Eds: M Tamas and E Martinoia, Berlin Heidelberg (2006)
- Y Zhang, H Wang, J Li, DA Jimenez, ES Levitan, E Aizenman, and PA Rosenberg: Peroxynitrite-induced neuronal apoptosis is mediated by intracellular zinc release and 12-lipoxygenase activation. *J Neurosci.* 24(47) 10616-27 (2004) DOI: 10.1523/JNEUROSCI.2469-04.2004
- PD Zalewski, IJ Forbes, and WH Betts: Correlation of apoptosis with change in intracellular labile Zn(II) using zinquin [(2-methyl-8-p-toluenesulphonamido-6quinolyloxy)acetic acid], a new specific fluorescent probe for Zn(II). *Biochem. J.* 296(Pt 2) 403-8 (1993) DOI: 10.1042/bj2960403

- I Sekler, SL Sensi, M Hershfinkel, and WF Silverman: Mechanism and regulation of cellular zinc transport. *Mol. Med.* 13(7-8) 337-43 (2007) DOI: 10.2119/2007-00037.Sekler
- 21. A Krezel and W Maret: Zinc-buffering capacity of a eukaryotic cell at physiological pZn. J Biol Inorg Chem. 11(8) 1049-62 (2006) DOI: 10.1007/s00775-006-0150-5
- M Vasak: Advances in metallothionein structure and functions. *J Trace Elem Med Biol.* 19(1) 13-7 (2005) DOI: 10.1016/j.jtemb.2005.03.003
- JP Liuzzi and RJ Cousins: Mammalian zinc transporters. Annu Rev Nutr. 24 151-72 (2004) DOI: 10.1146/annurev.nutr.24.012003.132402
- 24. DJ Eide: Zinc transporters and the cellular trafficking of zinc. *Biochim Biophys Acta. 1763(7)* 711-22 (2006) DOI: 10.1016/j.bbamcr.2006.03.005
- 25. RA Colvin, WR Holmes, CP Fontaine, and W Maret: Cytosolic zinc buffering and muffling: their role in intracellular zinc homeostasis. *Metallomics.* 2(5) 306-17 (2010) DOI: 10.1039/b926662c
- CJ Frederickson, BA Rampy, S Reamy Rampy, and GA Howell: Distribution of histochemically reactive zinc in the forebrain of the rat. *J. Chem. Neuroanat.* 5(6) 521-30 (1992) DOI: 10.1016/0891-0618(92)90007-D
- CJ Frederickson, J Perez-Clausell, and G Danscher: Zinc-containing 7S-NGF complex. Evidence from zinc histochemistry for localization in salivary secretory granules. J Histochem Cytochem. 35(5) 579-83 (1987) DOI: 10.1177/35.5.2435783
- CJ Frederickson and G Danscher: Zinccontaining neurons in hippocampus and related CNS structures. *Prog Brain Res.* 83 71-84 (1990) DOI: 10.1016/S0079-6123(08)61242-X
- 29. G Danscher and M Stoltenberg: Zincenriched neurons. *J Neurochem.* 85(Suppl 2) 10. (2003) DOI: 10.1046/j.1471-4159.85.s2.10_2.x

- K Ishii, M Sato, M Akita, and H Tomita: Localization of zinc in the rat submandibular gland and the effect of its deficiency on salivary secretion. *Ann Otol Rhinol Laryngol. 108(3)* 300-8 (1999) DOI: 10.1177/000348949910800315
- N McCormick, V Velasquez, L Finney, S Vogt, and SL Kelleher: X-ray fluorescence microscopy reveals accumulation and secretion of discrete intracellular zinc pools in the lactating mouse mammary gland. *PLoS One. 5(6)* e11078 (2010) DOI: 10.1371/journal.pone.0011078
- H Sharir, A Zinger, A Nevo, I Sekler, and M Hershfinkel: Zinc released from injured cells is acting via the Zn2+-sensing receptor, ZnR, to trigger signaling leading to epithelial repair. *J Biol Chem. 285(34)* 26097-106 (2010) DOI: 10.1074/jbc.M110.107490
- W Maret: Zinc coordination environments in proteins determine zinc functions. *J. Trace Elem. Med. Biol.* 19(1) 7-12 (2005) DOI: 10.1016/j.jtemb.2005.02.003
- W Maret: From the Cover: Crosstalk of the group IIa and IIb metals calcium and zinc in cellular signaling. *Proc Nat Acad Sci USA*. 98(22) 12325-7 (2001)
 DOI: 10.1073/pnas.231481398
- M Hershfinkel, A Moran, N Grossman, and I Sekler: A zinc-sensing receptor triggers the release of intracellular Ca2+ and regulates ion transport. *Proc Nat Acad Sci USA. 98(20)* 11749-54 (2001) DOI: 10.1073/pnas.201193398
- 36. L Cohen, I Sekler, and M Hershfinkel: The zinc sensing receptor, ZnR/GPR39, controls proliferation and differentiation of colonocytes and thereby tight junction formation in the colon. *Cell Death Dis.* 5 e1307 (2014) DOI: 10.1038/cddis.2014.262
- T Perez-Rosello, CT Anderson, FJ Schopfer, Y Zhao, D Gilad, SR Salvatore, BA Freeman, M Hershfinkel, E Aizenman, and T Tzounopoulos: Synaptic Zn2+ inhibits neurotransmitter release by promoting endocannabinoid synthesis. J Neurosci. 33(22) 9259-72 (2013) DOI: 10.1523/JNEUROSCI.0237-13.2013
- 38. E Chorin, O Vinograd, I Fleidervish, D Gilad, S Herrmann, I Sekler, E Aizenman, and M

Hershfinkel: Upregulation of KCC2 activity by zinc-mediated neurotransmission via the mZnR/GPR39 receptor. *J Neurosci.* 31(36) 12916-26 (2011) DOI: 10.1523/JNEUROSCI.2205-11.2011

- AM Hosie, EL Dunne, RJ Harvey, and TG Smart: Zinc-mediated inhibition of GABA(A) receptors: discrete binding sites underlie subtype specificity. *Nat Neurosci.* 6(4) 362-9 (2003) DOI: 10.1038/nn1030
- 40. Y Han and SM Wu: Modulation of glycine receptors in retinal ganglion cells by zinc. *Proc. Natl. Acad. Sci. USA.* 96(6) 3234-8 (1999) DOI: 10.1073/pnas.96.6.3234
- 41. JW Lynch, P Jacques, KD Pierce, and PR Schofield: Zinc potentiation of the glycine receptor chloride channel is mediated by allosteric pathways. *J Neurochem.* 71(5) 2159-68 (1998) DOI: 10.1046/j.1471-4159.1998.71052159.x
- 42. P Paoletti, P Ascher, and J Neyton: Highaffinity zinc inhibition of NMDA NR1-NR2A receptors. *J Neurosci.* 17(15) 5711-25. (1997)
- GA Herin and E Aizenman: Amino terminal domain regulation of NMDA receptor function. *Eur J Pharmacol.* 500(1-3) 101-11 (2004) DOI: 10.1016/j.ejphar.2004.07.015
- 44. D Gilad, S Shorer, M Ketzef, A Friedman, I Sekler, E Aizenman, and M Hershfinkel: Homeostatic regulation of KCC2 activity by the zinc receptor mZnR/GPR39 during seizures. *Neurobiol Dis, 81* 4-13 (2015) DOI: 10.1016/j.nbd.2014.12.020
- 45. A Gore, A Moran, M Hershfinkel, and I Sekler: Inhibitory mechanism of storeoperated Ca2+ channels by zinc. *J Biol Chem.* 279(12) 11106-11 (2004) DOI: 10.1074/jbc.M400005200
- 46. SS Wildman, BF King, and G Burnstock: Modulatory activity of extracellular H+ and Zn2+ on ATP-responses at rP2X1 and rP2X3 receptors. *Br. J. Pharmacol.* 128(2) 486-92 (1999) DOI: 10.1038/sj.bjp.0702802
- 47. C Acuna-Castillo, B Morales, and JP Huidobro-Toro: Zinc and copper modulate

differentially the P2X4 receptor. J Neurochem. 74(4) 1529-37 (2000) DOI: 10.1046/j.1471-4159.2000.0741529.x

- 48. S Kim, Y Jung, D Kim, H Koh, and J Chung; Extracellular zinc activates p70 S6 kinase through the phosphatidylinositol 3-kinase signaling pathway. J Biol Chem. 275(34) 25979-84 (2000) DOI: 10.1074/jbc.M001975200
- 49. SY Oh, KS Park, JA Kim, and KY Choi: Differential modulation of zinc-stimulated p21(Cip/WAF1) and cyclin D1 induction by inhibition of PI3 kinase in HT-29 colorectal cancer cells. Exp Mol Med. 34(1) 27-31. (2002)

DOI: 10.1038/emm.2002.4

- 50. Y Ho, R Samarasinghe, ME Knoch, M Lewis, E Aizenman, and DB DeFranco: Selective inhibition of mitogen-activated protein kinase phosphatases by zinc accounts for extracellular signal-regulated kinase 1/2-dependent oxidative neuronal cell death. Mol. Pharmacol. 74(4) 1141-51 (2008)DOI: 10.1124/mol.108.049064
- 51. Y Zhang, E Aizenman, DB DeFranco, and PA Rosenberg: Intracellular zinc release, 12-lipoxygenase activation and MAPK dependent neuronal and oligodendroglial death. Mol Med. 13(7-8) 350-5 (2007) DOI: 10.2119/2007-00042.Zhang
- 52. H Azriel-Tamir, H Sharir, B Schwartz, and M Hershfinkel: Extracellular zinc triggers ERKdependent activation of Na+/H+ exchange in colonocytes mediated by the zinc-sensing receptor. J Biol Chem. 279(50) 51804-16 (2004)DOI: 10.1074/jbc.M406581200
- 53. H Asraf, S Salomon, A Nevo, I Sekler, D Mayer, and M Hershfinkel: The ZnR/ GPR39 Interacts with the CaSR to Enhance Signaling in Prostate and Salivary Epithelia. J Cell Physiol, 229(7) 868-77 (2013) DOI: 10.1002/jcp.24514
- 54. W Maret: Crosstalk of the group IIa and IIb metals calcium and zinc in cellular signaling. Proc Nat Acad Sci USA. 98(22) 12325-7 (2001)DOI: 10.1073/pnas.231481398
- 55. J Takasaki, T Saito, M Taniguchi, T Kawasaki, Y Moritani, K Hayashi, and M Kobori: A

novel Galphaq/11-selective inhibitor. J. Biol. Chem. 279(46) 47438-45 (2004) DOI: 10.1074/jbc.M408846200

- 56. M Taniguchi, K Suzumura, K Nagai, T Kawasaki, J Takasaki, M Sekiguchi, Y Moritani, T Saito, K Hayashi, S Fujita, S Tsukamoto, and K Suzuki: YM-254890 analogues, novel cyclic depsipeptides with Galpha(q/11) inhibitory activity from Chromobacterium sp. QS3666. Bioorg. Med. Chem. 12(12) 3125-33 (2004)
- 57. H Sharir and M Hershfinkel: The extracellular zinc-sensing receptor mediates intercellular communication by inducing ATP release. Biochem Biophys Res Commun. 332(3) 845-52 (2005) DOI: 10.1016/j.bbrc.2005.05.036
- 58. N Dubi, L Gheber, D Fishman, I Sekler, and M Hershfinkel: Extracellular zinc and zinccitrate, acting through a putative zinc-sensing receptor, regulate growth and survival of prostate cancer cells. Carcinogenesis. 29(9) 1692-700 (2008) DOI: 10.1093/carcin/bgn027
- 59. B Holst, KL Egerod, C Jin, PS Petersen, MV Ostergaard, J Hald, AM Sprinkel, J Storling, T Mandrup-Poulsen, JJ Holst, P Thams, C Orskov, N Wierup, F Sundler, OD Madsen, and TW Schwartz: G protein-coupled receptor 39 deficiency is associated with pancreatic islet dysfunction. Endocrinology. 150(6) 2577-85 (2009) DOI: 10.1210/en.2008-1250
- 60. TKambe, KFukue, R Ishida, and S Miyazaki: Overview of Inherited Zinc Deficiency in Infants and Children. J Nutr Sci Vitaminol (Tokyo). 61 Suppl S44-6 (2015)
- 61. M Komai, T Goto, H Suzuki, T Takeda, and Y Furukawa: Zinc deficiency and taste dysfunction; contribution of carbonic anhydrase, a zinc-metalloenzyme, to normal taste sensation. Biofactors. 12(1-4) 65-70 (2000)DOI: 10.1002/biof.5520120111
- 62. WH Chappell, LS Steelman, JM Long, RC Kempf, SL Abrams, RA Franklin, J Basecke, F Stivala, M Donia, P Fagone, G Malaponte, MC Mazzarino, F Nicoletti, M Libra, D Maksimovic-Ivanic, S Mijatovic, G Montalto, M Cervello, P Laidler, M Milella, A Tafuri, A Bonati, C Evangelisti, L Cocco, AM Martelli, and JA McCubrey: Ras/Raf/MEK/ERK and

PI3K/PTEN/Akt/mTOR inhibitors: rationale and importance to inhibiting these pathways in human health. *Oncotarget. 2(3)* 135-64 (2011) DOI: 10.18632/oncotarget.240

- 63. RS MacDonald: The role of zinc in growth and cell proliferation. *J. Nutr.* 130(5S Suppl) 1500S-8S (2000)
- 64. DW Choi and JY Koh: Zinc and brain injury. *Annu. Rev. Neurosci.* 21 347-75 (1998) DOI: 10.1146/annurev.neuro.21.1.347
- JH Weiss, SL Sensi, and JY Koh: Zn(2+): a novel ionic mediator of neural injury in brain disease. *Trends Pharmacol. Sci.* 21(10) 395-401 (2000) DOI: 10.1016/S0165-6147(00)01541-8
- 66. SL Sensi, P Paoletti, JY Koh, E Aizenman, Al Bush, and M Hershfinkel: The neurophysiology and pathology of brain zinc. *J Neurosci.* 31(45) 16076-85 (2011) DOI: 10.1523/JNEUROSCI.3454-11.2011
- 67. Al Bush: Copper, zinc, and the metallobiology of Alzheimer disease. *Alzheimer Dis. Assoc. Disord. 17(3)* 147-50. (2003). DOI: 10.1097/00002093-200307000-00005
- L Besser, E Chorin, I Sekler, WF Silverman, S Atkin, JT Russell, and M Hershfinkel: Synaptically released zinc triggers metabotropic signaling via a zinc-sensing receptor in the hippocampus. *J Neurosci.* 29(9) 2890-901 (2009) DOI: 10.1523/JNEUROSCI.5093-08.2009
- 69. SC Brennan, U Thiem, S Roth, A Aggarwal, IS Fetahu, S Tennakoon, AR Gomes, ML Brandi, F Bruggeman, R Mentaverri, D Riccardi, and E Kallay: Calcium sensing receptor signalling in physiology and cancer. *Biochim Biophys Acta*, (2012)
- EM Brown: The extracellular C2+ sensing receptor.: central mediator of systemic calcium homeostasis [In Process Citation]. *Annu Rev Nutr. 20* 507-33 (2000) DOI: 10.1146/annurev.nutr.20.1.507
- 71. I Gomes, A Gupta, J Filipovska, HH Szeto, JE Pintar, and LA Devi: A role for heterodimerization of mu and delta opiate receptors in enhancing morphine analgesia. *Proc. Natl. Acad. Sci. USA. 101(14)* 5135-9 (2004) DOI: 10.1073/pnas.0307601101

- 72. L Albizu, MN Balestre, C Breton, JP Pin, M Manning, B Mouillac, C Barberis, and T Durroux: Probing the existence of G proteincoupled receptor dimers by positive and negative ligand-dependent cooperative binding. *Mol. Pharmacol.* 70(5) 1783-91 (2006) DOI: 10.1124/mol.106.025684
- 73. MP Grant, A Stepanchick, A Cavanaugh, and GE Breitwieser: Agonist-driven maturation and plasma membrane insertion of calcium-sensing receptors dynamically control signal amplitude. *Sci Signal. 4*(200) ra78 (2011) DOI: 10.1126/scisignal.2002208
- 74. PH McDonald and RJ Lefkowitz: Beta-Arrestins: new roles in regulating heptahelical receptors' functions. *Cell Signal. 13(10)* 683-9 (2001) DOI: 10.1016/S0898-6568(01)00203-0
- TA Kohout and RJ Lefkowitz: Regulation of G protein-coupled receptor kinases and arrestins during receptor desensitization. *Mol. Pharmacol.* 63(1) 9-18 (2003) DOI: 10.1124/mol.63.1.9
- 76. ML Mohan, NT Vasudevan, MK Gupta, EE Martelli, and SV Naga Prasad: G-protein coupled receptor resensitizationappreciating the balancing act of receptor function. *Curr Mol Pharmacol, May* (2012)
- LC Costello, P Feng, B Milon, M Tan, and RB Franklin: Role of zinc in the pathogenesis and treatment of prostate cancer: critical issues to resolve. *Prostate Cancer Prostatic Dis.* 7(2) 111-7 (2004) DOI: 10.1038/sj.pcan.4500712
- LC Costello and RB Franklin: The clinical relevance of the metabolism of prostate cancer; zinc and tumor suppression: connecting the dots. *Mol Cancer.* 5(1) 17 (2006) DOI: 10.1186/1476-4598-5-17
- B Holst, ND Holliday, A Bach, CE Elling, HM Cox, and TW Schwartz: Common structural basis for constitutive activity of the ghrelin receptor family. *J. Biol. Chem.* 279(51) 53806-17 (2004) DOI: 10.1074/jbc.M407676200
- 80. CE Elling, TM Frimurer, LO Gerlach, R Jorgensen, B Holst, and TW Schwartz: Metal ion site engineering indicates a global toggle switch model for seven-transmembrane

receptor activation. *J Biol Chem.* 281(25) 17337-46 (2006) DOI: 10.1074/jbc.M512510200

- 81. JV Zhang, PG Ren, O Avsian-Kretchmer, CW Luo, R Rauch, C Klein, and AJ Hsueh: Obestatin, a peptide encoded by the ghrelin gene, opposes ghrelin's effects on food intake. *Science.* 310(5750) 996-9 (2005) DOI: 10.1126/science.1117255
- 82. B Holst, KL Egerod, E Schild, SP Vickers, S Cheetham, LO Gerlach, L Storjohann, CE Stidsen, R Jones, AG Beck-Sickinger, and TW Schwartz: GPR39 signaling is stimulated by zinc ions but not by obestatin. *Endocrinology.* 148(1) 13-20 (2007) DOI: 10.1210/en.2006-0933
- E Lauwers, B Landuyt, LArckens, L Schoofs, and W Luyten: Obestatin does not activate orphan G protein-coupled receptor GPR39. *Biochem Biophys Res Commun.* 351(1) 21-5 (2006) DOI: 10.1016/j.bbrc.2006.09.141
- 84. S Yasuda, T Miyazaki, K Munechika, M Yamashita, Y Ikeda, and A Kamizono: Isolation of Zn2+ as an endogenous agonist of GPR39 from fetal bovine serum. J. Recept. Signal Transduct. Res. 27(4) 235-46 (2007) DOI: 10.1080/10799890701506147
- L Storjohann, B Holst, and TW Schwartz: Molecular mechanism of Zn2+ agonism in the extracellular domain of GPR39. *FEBS Lett.* 582(17) 2583-8 (2008) DOI: 10.1016/j.febslet.2008.06.030
- L Cohen, HAsraf, I Sekler, and M Hershfinkel: Extracellular pH regulates zinc signaling via an Asp residue of the zinc-sensing receptor (ZnR/GPR39). *J Biol Chem. 287(40)* 33339-50 (2012) DOI: 10.1074/jbc.M112.372441
- 87. T Ganay, H Asraf, E Aizenman, M Bogdanovic, I Sekler, and M Hershfinkel: Regulation of Neuronal pH by the Metabotropic Zinc Receptor mZnR/GPR39. *J Neurochem*, 135(5) 897-907 (2015) DOI: 10.1111/jnc.13367
- J Srivastava, DL Barber, and MP Jacobson: Intracellular pH sensors: design principles and functional significance. *Physiology* (*Bethesda*). 22 30-9 (2007) DOI: 10.1152/physiol.00035.2006

- SJ Quinn, M Bai, and EM Brown: pH Sensing by the calcium-sensing receptor. J Biol Chem. 279(36) 37241-9 (2004) DOI: 10.1074/jbc.M404520200
- 90. C Levinthal, L Barkdull, P Jacobson, L Storjohann, BC Van Wagenen, TM Stormann, and LG Hammerland: Modulation of group III metabotropic glutamate receptors by hydrogen ions. *Pharmacology. 83(2)* 88-94 (2009) DOI: 10.1159/000180124
- M Sharma, K Sahu, A Dube, and PK Gupta: Extracellular pH influences the mode of cell death in human colon adenocarcinoma cells subjected to photodynamic treatment with chlorin p6. *J Photochem Photobiol B. 81(2)* 107-13 (2005). DOI: 10.1016/j.jphotobiol.2005.07.001
- 92. M Sandoval, J Burgos, FV Sepulveda, and LP Cid: Extracellular pH in restricted domains as a gating signal for ion channels involved in transepithelial transport. *Biol Pharm Bull.* 34(6) 803-9 (2011) DOI: 10.1248/bpb.34.803
- 93. DA Perdikis, R Davies, A Zhuravkov, B Brenner, L Etter, and MD Basson: Differential effects of mucosal pH on human (Caco-2) intestinal epithelial cell motility, proliferation, and differentiation. *Dig Dis Sci.* 43(7) 1537-46 (1998) DOI: 10.1023/A:1018871016691
- 94. P Holzer: Acid sensing by visceral afferent neurones. *Acta Physiol (Oxf). 201(1)* 63-75 (2011)
 DOI: 10.1111/j.1748-1716.2010.02143.x
- 95. P Holzer: Acid-sensitive ion channels and receptors. *Handb Exp Pharmacol, (194)* 283-332 (2009)
- 96. SG Nugent, D Kumar, DS Rampton, and DF Evans: Intestinal luminal pH in inflammatory bowel disease: possible determinants and implications for therapy with aminosalicylates and other drugs. *Gut.* 48(4) 571-7 (2001) DOI: 10.1136/gut.48.4.571
- 97. J Orlowski and S Grinstein: Na+/H+ exchangers. *Compr Physiol.* 1(4) 2083-100 (2011) DOI: 10.1002/cphy.c110020
- 98. L Cohen, H Azriel-Tamir, N Arotsker, I Sekler, and M Hershfinkel: Zinc Sensing Receptor

Signaling, Mediated by GPR39, Reduces Butyrate-Induced Cell Death in HT29 Colonocytes via Upregulation of Clusterin. *PLoS One.* 7(4) e35482 (2012) DOI: 10.1371/journal.pone.0035482

- 99. K Kaila, P Panula, T Karhunen, and E Heinonen: Fall in intracellular pH mediated by GABAA receptors in cultured rat astrocytes. *Neurosci Lett.* 126(1) 9-12 (1991) DOI: 10.1016/0304-3940(91)90358-Z
- 100. M Chesler: The regulation and modulation of pH in the nervous system. *Prog Neurobiol. 34(5)* 401-27 (1990) DOI: 10.1016/0301-0082(90)90034-E
- 101. SF Traynelis and SG Cull-Candy: Pharmacological properties and H+ sensitivity of excitatory amino acid receptor channels in rat cerebellar granule neurones. *J Physiol.* 433 727-63 (1991) DOI: 10.1113/jphysiol.1991.sp018453
- 102. J Church, KA Baxter, and JG McLarnon: pH modulation of Ca2+ responses and a Ca2+-dependent K+ channel in cultured rat hippocampal neurones. *J Physiol. 511 (Pt 1)* 119-32 (1998) DOI: 10.1111/j.1469-7793.1998.119bi.x
- 103. CJ Dietrich and M Morad: Synaptic acidification enhances GABAA signaling. *J Neurosci.* 30(47) 16044-52 (2010) DOI: 10.1523/JNEUROSCI.6364-09.2010
- 104. GH Diering, F Mills, SX Bamji, and M Numata: Regulation of dendritic spine growth through activity-dependent recruitment of the brainenriched Na(+)/H(+) exchanger NHE5. *Mol Biol Cell.* 22(13) 2246-57 (2011) DOI: 10.1091/mbc.E11-01-0066
- 105. N Manhas, Y Shi, J Taunton, and D Sun: p90 activation contributes to cerebral ischemic damage via phosphorylation of Na+/H+ exchanger isoform 1. *J Neurochem.* 114(5) 1476-86 (2010)
- 106. Y Wang, J Luo, X Chen, H Chen, SW Cramer, and D Sun: Gene inactivation of Na+/H+ exchanger isoform 1 attenuates apoptosis and mitochondrial damage following transient focal cerebral ischemia. *Eur J Neurosci.* 28(1) 51-61 (2008) DOI: 10.1111/j.1460-9568.2008.06304.x
- 107.TI Lam, AM Brennan-Minnella, SJ Won, Y Shen, C Hefner, Y Shi, D Sun, and RA

Swanson: Intracellular pH reduction prevents excitotoxic and ischemic neuronal death by inhibiting NADPH oxidase. *Proc Natl Acad Sci U S A. 110(46)* E4362-8 (2013) DOI: 10.1073/pnas.1313029110

- 108. JP Hachem, M Behne, I Aronchik, M Demerjian, KR Feingold, PM Elias, and TM Mauro: Extracellular pH Controls NHE1 expression in epidermis and keratinocytes: implications for barrier repair. *J. Invest. Dermatol.* 125(4) 790-7 (2005) DOI: 10.1111/j.0022-202X.2005.23836.x
- 109. MJ Behne, JW Meyer, KM Hanson, NP Barry, S Murata, D Crumrine, RW Clegg, E Gratton, WM Holleran, PM Elias, and TM Mauro: NHE1 regulates the stratum corneum permeability barrier homeostasis. Microenvironment acidification assessed with fluorescence lifetime imaging. *J Biol Chem.* 277(49) 47399-406 (2002) DOI: 10.1074/jbc.M204759200
- 110. L Xue, E Aihara, TC Wang, and MH Montrose: Trefoil factor 2 requires Na/H exchanger 2 activity to enhance mouse gastric epithelial repair. *J Biol Chem.* 286(44) 38375-82 (2011) DOI: 10.1074/jbc.M111.268219
- 111. CJ Frederickson, SW Suh, D Silva, and RB Thompson: Importance of zinc in the central nervous system: the zinc-containing neuron. *J Nutr. 130(5S Suppl)* 1471S-83S (2000)
- 112. NT Watt, IJ Whitehouse, and NM Hooper: The role of zinc in Alzheimer's disease. *Int J Alzheimers Dis. 2011* 971021 (2010)
- 113. S Ayton, P Lei, and Al Bush: Metallostasis in Alzheimer's disease. *Free Radic Biol Med.* 62 76-89 (2013) DOI: 10.1016/j.freeradbiomed.2012.10.558
- 114. JB Hilton, AR White, and PJ Crouch: Metaldeficient SOD1 in amyotrophic lateral sclerosis. *J Mol Med (Berl).* 93(5) 481-7 (2015) DOI: 10.1007/s00109-015-1273-3
- 115. J Hennig, C Andresen, AK Museth, P Lundstrom, LA Tibell, and BH Jonsson: Local destabilization of the metal-binding region in human copper-zinc superoxide dismutase by remote mutations is a possible determinant for progression of ALS. *Biochemistry.* 54(2) 323-33 (2015) DOI: 10.1021/bi500606j

- 116. BK Bitanihirwe and MG Cunningham: Zinc: the brain's dark horse. *Synapse.* 63(11) 1029-49 (2009) DOI: 10.1002/syn.20683
- 117. MA Aras, RA Saadi, and E Aizenman: Zn2+ regulates Kv2.1 voltage-dependent gating and localization following ischemia. *Eur J Neurosci.* 30(12) 2250-7 (2009) DOI: 10.1111/j.1460-9568.2009.07026.x
- 118. MA Aras and E Aizenman: Redox regulation of intracellular zinc: molecular signaling in the life and death of neurons. *Antioxid Redox Signal. 15(8)* 2249-63 (2011) DOI: 10.1089/ars.2010.3607
- 119. RE Carter, I Aiba, RM Dietz, CT Sheline, and CW Shuttleworth: Spreading depression and related events are significant sources of neuronal Zn2+ release and accumulation. *J Cereb Blood Flow Metab.* 31(4) 1073-84 (2011) DOI: 10.1038/jcbfm.2010.183
- 120. RE Carter, I Aiba, RM Dietz, CT Sheline, and CW Shuttleworth: Spreading depression and related events are significant sources of neuronal Zn(2+) release and accumulation. *J Cereb Blood Flow Metab.* 31(4) 1073-84 (2010) DOI: 10.1038/jcbfm.2010.183
- 121. RM Dietz, JH Weiss, and CW Shuttleworth: Zn2+ influx is critical for some forms of spreading depression in brain slices. *J Neurosci.* 28(32) 8014-24 (2008) DOI: 10.1523/JNEUROSCI.0765-08.2008
- 122. AJ Russo and R Devito: Analysis of Copper and Zinc Plasma Concentration and the Efficacy of Zinc Therapy in Individuals with Asperger's Syndrome, Pervasive Developmental Disorder Not Otherwise Specified (PDD-NOS) and Autism. *Biomark Insights.* 6 127-33 (2011) DOI: 10.4137/BMI.S7286
- 123. G Vela, P Stark, M Socha, AK Sauer, S Hagmeyer, and AM Grabrucker: Zinc in gutbrain interaction in autism and neurological disorders. *Neural Plast.* 2015 972791 (2015) DOI: 10.1155/2015/972791
- 124. TB Cole, CA Robbins, HJ Wenzel, PA Schwartzkroin, and RD Palmiter: Seizures and neuronal damage in mice lacking vesicular zinc. *Epilepsy Res. 39(2)* 153-69 (2000) DOI: 10.1016/S0920-1211(99)00121-7

- 125. TB Cole, HJ Wenzel, KE Kafer, PA Schwartzkroin, and RD Palmiter: Elimination of zinc from synaptic vesicles in the intact mouse brain by disruption of the ZnT3 gene. *Proc. Natl. Acad. Sci. USA. 96(4)* 1716-21 (1999) DOI: 10.1073/pnas.96.4.1716
- 126. K Saad, E Hammad, AF Hassan, and R Badry: Trace element, oxidant, and antioxidant enzyme values in blood of children with refractory epilepsy. *Int J Neurosci.* 124(3) 181-6 (2014) DOI: 10.3109/00207454.2013.831851
- 127. HN Farahani, AR Ashthiani, and MS Masihi: Study on serum zinc and selenium levels in epileptic patients. *Neurosciences (Riyadh)*. *18(2)* 138-42 (2013)
- 128. RW Wojciak, E Mojs, M Stanislawska-Kubiak, and W Samborski: The serum zinc, copper, iron, and chromium concentrations in epileptic children. *Epilepsy Res.* 104(1-2) 40-4 (2013) DOI: 10.1016/j.eplepsyres.2012.09.009
- 129. M Seven, SY Basaran, M Cengiz, S Unal, and A Yuksel: Deficiency of selenium and zinc as a causative factor for idiopathic intractable epilepsy. *Epilepsy Res.* 104(1-2) 35-9 (2013) DOI: 10.1016/j.eplepsyres.2012.09.013
- 130. JM Blasco-Ibanez, J Poza-Aznar, C Crespo, Al Marques-Mari, FJ Gracia-Llanes, and FJ Martinez-Guijarro: Chelation of synaptic zinc induces overexcitation in the hilar mossy cells of the rat hippocampus. *Neurosci Lett.* 355(1-2) 101-4. (2004) DOI: 10.1016/j.neulet.2003.10.053
- 131. R Ganesh and L Janakiraman: Serum zinc levels in children with simple febrile seizure. *Clin Pediatr (Phila).* 47(2) 164-6 (2008) DOI: 10.1177/0009922807306165
- 132. HJ Goldberg and EM Sheehy: Fifth day fits: an acute zinc deficiency syndrome? *Arch Dis Child.* 57(8) 633-5 (1982) DOI: 10.1136/adc.57.8.633
- 133. Hildebrand MS, Phillips AM, Mullen SA, Adlard PA, Hardies K, Damiano JA, Wimmer V, Bellows ST, McMahon JM, Burgess R, Hendrickx R, Weckhuysen S, Suls A, De Jonghe P, Scheffer IE, Petrou S, Berkovic SF, and R CA: Loss of synaptic Zn(2+) transporter function increases risk of febrile seizures. *Sci Rep.* 5 17816 (2015) DOI: 10.1038/srep17816

- 134. SM Elsas, S Hazany, WL Gregory, and I Mody: Hippocampal zinc infusion delays the development of afterdischarges and seizures in a kindling model of epilepsy. *Epilepsia. 50(4)* 870-9 (2009) DOI: 10.1111/j.1528-1167.2008.01913.x
- 135. AM Baraka, W Hassab El Nabi, and S El Ghotni: Investigating the role of zinc in a rat model of epilepsy. *CNS Neurosci Ther. 18(4)* 327-33 (2012) DOI: 10.1111/j.1755-5949.2011.00252.x
- 136. SL Sensi, P Paoletti, Al Bush, and I Sekler: Zinc in the physiology and pathology of the CNS. *Nat Rev Neurosci.* 10(11) 780-91 (2009) DOI: 10.1038/nrn2734
- 137. S Pal, KA Hartnett, JM Nerbonne, ES Levitan, and E Aizenman: Mediation of Neuronal Apoptosis by Kv2.1-Encoded Potassium Channels. *J. Neurosci.* 23(12) 4798-4802 (2003)
- 138. MC McCord and E Aizenman: The role of intracellular zinc release in aging, oxidative stress, and Alzheimer's disease. *Front Aging Neurosci.* 6 77 (2014) DOI: 10.3389/fnagi.2014.00077
- 139. E Aizenman, AK Stout, KA Hartnett, KE Dineley, B McLaughlin, and IJ Reynolds: Induction of neuronal apoptosis by thiol oxidation: putative role of intracellular zinc release. *J. Neurochem.* 75(5) 1878-88 (2000) DOI: 10.1046/j.1471-4159.2000.0751878.x
- 140. SR Bareggi and U Cornelli: Clioquinol: review of its mechanisms of action and clinical uses in neurodegenerative disorders. *CNS Neurosci Ther. 18(1)* 41-6 (2012) DOI: 10.1111/j.1755-5949.2010.00231.x
- 141. MH Park, SJ Lee, HR Byun, Y Kim, YJ Oh, JY Koh, and JJ Hwang: Clioquinol induces autophagy in cultured astrocytes and neurons by acting as a zinc ionophore. *Neurobiol Dis.* 42(3) 242-51 (2011) DOI: 10.1016/j.nbd.2011.01.009
- 142. MC McCord and E Aizenman: Convergent Ca2+ and Zn2+ signaling regulates apoptotic Kv2.1 K+ currents. *Proc Natl Acad Sci U S A. 110(34)* 13988-93 (2013) DOI: 10.1073/pnas.1306238110
- 143. MI Dominguez, JM Blasco-Ibanez, C Crespo, Al Marques-Mari, and FJ Martinez-

Guijarro: Zinc chelation during non-lesioning overexcitation results in neuronal death in the mouse hippocampus. *Neuroscience*. *116(3)* 791-806 (2003) DOI: 10.1016/S0306-4522(02)00731-5

- 144. MA Greenough, J Camakaris, and Al Bush: Metal dyshomeostasis and oxidative stress in Alzheimer's disease. *Neurochem Int. 62(5)* 540-55 (2013) DOI: 10.1016/j.neuint.2012.08.014
- 145. G Danscher: Exogenous selenium in the brain. A histochemical technique for light and electron microscopical localization of catalytic selenium bonds. *Histochemistry*. *76(3)* 281-93 (1982) DOI: 10.1007/BF00543951
- 146. G Danscher: The autometallographic zincsulphide method. A new approach involving in vivo creation of nanometer-sized zinc sulphide crystal lattices in zinc-enriched synaptic and secretory vesicles. *Histochem J. 28(5)* 361-73 (1996) DOI: 10.1007/BF02331399
- 147. DH Linkous, JM Flinn, JY Koh, A Lanzirotti, PM Bertsch, BF Jones, LJ Giblin, and CJ Frederickson: Evidence that the ZNT3 protein controls the total amount of elemental zinc in synaptic vesicles. *J Histochem Cytochem. 56*(*1*) 3-6 (2008) DOI: 10.1369/jhc.6A7035.2007
- 148. MH Yoo, TY Kim, YH Yoon, and JY Koh: Autism phenotypes in ZnT3 null mice: Involvement of zinc dyshomeostasis, MMP-9 activation and BDNF upregulation. *Sci Rep.* 6 28548 (2016) DOI: 10.1038/srep28548
- 149. PA Adlard, JM Parncutt, DI Finkelstein, and AI Bush: Cognitive loss in zinc transporter-3 knock-out mice: a phenocopy for the synaptic and memory deficits of Alzheimer's disease? *J Neurosci.* 30(5) 1631-6 (2010) DOI: 10.1523/JNEUROSCI.5255-09.2010
- 150. G Martel, C Hevi, O Friebely, T Baybutt, and GP Shumyatsky: Zinc transporter 3 is involved in learned fear and extinction, but not in innate fear. *Learn Mem.* 17(11) 582-90 (2010) DOI: 10.1101/lm.1962010
- 151. G Martel, C Hevi, N Kane-Goldsmith, and GP Shumyatsky: Zinc transporter ZnT3 is involved in memory dependent on the

hippocampus and perirhinal cortex. *Behav Brain Res. 223(1)* 233-8 (2011) DOI: 10.1016/j.bbr.2011.04.020

- 152. SA Kodirov, S Takizawa, J Joseph, ER Kandel, GP Shumyatsky, and VY Bolshakov: Synaptically released zinc gates long-term potentiation in fear conditioning pathways. *Proc Natl Acad Sci U S A. 103(41)* 15218-23. (2006) DOI: 10.1073/pnas.0607131103
- 153. PA Adlard, J Parncutt, V Lal, S James, D Hare, P Doble, DI Finkelstein, and Al Bush: Metal chaperones prevent zinc-mediated cognitive decline. *Neurobiol Dis.* 81 196-202 (2015) DOI: 10.1016/i.pbd.2011.12.012

DOI: 10.1016/j.nbd.2014.12.012

- 154. C Sindreu, RD Palmiter, and DR Storm: Zinc transporter ZnT-3 regulates presynaptic Erk1/2 signaling and hippocampusdependent memory. *Proc Natl Acad Sci U S A. 108(8)* 3366-70 (2011) DOI: 10.1073/pnas.1019166108
- 155. CJ Frederickson, JY Koh, and Al Bush: The neurobiology of zinc in health and disease. *Nat. Rev. Neurosci.* 6(6) 449-62 (2005) DOI: 10.1038/nrn1671
- 156. CJ Frederickson, LJ Giblin, 3rd, RV Balaji, R Masalha, CJ Frederickson, Y Zeng, EV Lopez, JY Koh, U Chorin, L Besser, M Hershfinkel, Y Li, RB Thompson, and A Krezel: Synaptic release of zinc from brain slices: factors governing release, imaging, and accurate calculation of concentration. *J. Neurosci. Methods. 154*(*1-2*) 19-29 (2006) DOI: 10.1016/j.jneumeth.2005.11.014
- 157. J Qian and JL Noebels: Visualization of transmitter release with zinc fluorescence detection at the mouse hippocampal mossy fibre synapse. *J Physiol.* 566(Pt 3) 747-58. (2005)
 DOI: 10.1113/jphysiol.2005.089276
- 158. J Qian and JL Noebels: Exocytosis of vesicular zinc reveals persistent depression of neurotransmitter release during metabotropic glutamate receptor long-term depression at the hippocampal CA3-CA1 synapse. *J Neurosci.* 26(22) 6089-95 (2006) DOI: 10.1523/JNEUROSCI.0475-06.2006
- 159. Y Li, CJ Hough, CJ Frederickson, and JM Sarvey: Induction of mossy fiber --> Ca3 long-term potentiation requires translocation

of synaptically released Zn2+. J Neurosci. 21(20) 8015-25 (2001)

- 160. M Gielen, B Siegler Retchless, L Mony, JW Johnson, and P Paoletti: Mechanism of differential control of NMDA receptor activity by NR2 subunits. *Nature.* 459(7247) 703-7 (2009) DOI: 10.1038/nature07993
- 161. T Perez-Rosello, CT Anderson, C Ling, SJ Lippard, and T Tzounopoulos: Tonic zinc inhibits spontaneous firing in dorsal cochlear nucleus principal neurons by enhancing glycinergic neurotransmission. *Neurobiol Dis. 81* 14-9 (2015) DOI: 10.1016/j.nbd.2015.03.012
- 162. TG Smart, AM Hosie, and PS Miller: Zn2+ ions: modulators of excitatory and inhibitory synaptic activity. *Neuroscientist. 10(5)* 432-42. (2004) DOI: 10.1177/1073858404263463
- 163. K Vogt, J Mellor, G Tong, and R Nicoll: The actions of synaptically released zinc at hippocampal mossy fiber synapses. *Neuron.* 26(1) 187-96 (2000) DOI: 10.1016/S0896-6273(00)81149-6
- 164. BI Kalappa, CT Anderson, JM Goldberg, SJ Lippard, and T Tzounopoulos: AMPA receptor inhibition by synaptically released zinc. *Proc Natl Acad Sci U S A. 112(51)* 15749-54 (2015) DOI: 10.1073/pnas.1512296112
- 165. CT Anderson, RJ Radford, ML Zastrow, DY Zhang, UP Apfel, SJ Lippard, and T Tzounopoulos: Modulation of extrasynaptic NMDA receptors by synaptic and tonic zinc. *Proc Natl Acad Sci U S A. 112(20)* E2705-14 (2015) DOI: 10.1073/pnas.1503348112
- 166. RE Nicholls, XL Zhang, CP Bailey, BR Conklin, ER Kandel, and PK Stanton: mGluR2 acts through inhibitory Galpha subunits to regulate transmission and longterm plasticity at hippocampal mossy fiber-CA3 synapses. *Proc Natl Acad Sci U S A. 103(16)* 6380-5 (2006) DOI: 10.1073/pnas.0601267103
- 167. LJ Volk, CA Daly, and KM Huber: Differential roles for group 1 mGluR subtypes in induction and expression of chemically induced hippocampal long-term depression. *J Neurophysiol.* 95(4) 2427-38 (2006) DOI: 10.1152/jn.00383.2005

- 168. JQ Wang, EE Fibuch, and L Mao: Regulation of mitogen-activated protein kinases by glutamate receptors. *J Neurochem.* 100(1) 1-11 (2007) DOI: 10.1111/j.1471-4159.2006.04208.x
- 169. RA Saadi, K He, KA Hartnett, K Kandler, M Hershfinkel, and E Aizenman: SNAREdependent upregulation of potassium chloride co-transporter 2 activity after metabotropic zinc receptor activation in rat cortical neurons in vitro. *Neuroscience. 210* 38-46 (2012) DOI: 10.1016/j.neuroscience.2012.03.001
- 170. A Cichy, M Sowa-Kucma, B Legutko, L Pomierny-Chamiolo, A Siwek, A Piotrowska, B Szewczyk, E Poleszak, A Pilc, and G Nowak: Zinc-induced adaptive changes in NMDA/glutamatergic and serotonergic receptors. *Pharmacol Rep.* 61(6) 1184-91 (2009) DOI: 10.1016/S1734.1140(00)70182.3

DOI: 10.1016/S1734-1140(09)70182-3

- 171. K Mlyniec and G Nowak: Up-regulation of the GPR39 Zn(2+)-sensing receptor and CREB/BDNF/TrkB pathway after chronic but not acute antidepressant treatment in the frontal cortex of zinc-deficient mice. *Pharmacol Rep.* 67(6) 1135-40 (2015) DOI: 10.1016/j.pharep.2015.04.003
- 172. H Lee, CX Chen, YJ Liu, E Aizenman, and K Kandler: KCC2 expression in immature rat cortical neurons is sufficient to switch the polarity of GABA responses. *Eur J Neurosci. 21(9)* 2593-9 (2005) DOI: 10.1111/j.1460-9568.2005.04084.x
- 173. J Lu, M Karadsheh, and E Delpire: Developmental regulation of the neuronalspecific isoform of K-Cl cotransporter KCC2 in postnatal rat brains. *J Neurobiol.* 39(4) 558-68 (1999) DOI: 10.1002/(SICI)1097-4695(19990615)39: 4<558::AID-NEU9>3.0.CO;2-5
- 174. M Farrant and K Kaila: The cellular, molecular and ionic basis of GABA(A) receptor signalling. *Prog Brain Res.* 160 59-87 (2007) DOI: 10.1016/S0079-6123(06)60005-8
- 175. T Viitanen, E Ruusuvuori, K Kaila, and J Voipio: The K+-Cl cotransporter KCC2 promotes GABAergic excitation in the mature rat hippocampus. *J Physiol. 588(Pt 9)* 1527-40 (2010) DOI: 10.1113/jphysiol.2009.181826

- 176. L Zhu, D Lovinger, and E Delpire: Cortical neurons lacking KCC2 expression show impaired regulation of intracellular chloride. *J Neurophysiol.* 93(3) 1557-68 (2005) DOI: 10.1152/jn.00616.2004
- 177. L Zhu, N Polley, GC Mathews, and E Delpire: NKCC1 and KCC2 prevent hyperexcitability in the mouse hippocampus. *Epilepsy Res.* 79(2-3) 201-12 (2008) DOI: 10.1016/j.eplepsyres.2008.02.005
- 178. G Huberfeld, L Wittner, S Clemenceau, M Baulac, K Kaila, R Miles, and C Rivera: Perturbed chloride homeostasis and GABAergic signaling in human temporal lobe epilepsy. *J Neurosci.* 27(37) 9866-73 (2007) DOI: 10.1523/JNEUROSCI.2761-07.2007
- 179. NS Woo, J Lu, R England, R McClellan, S Dufour, DB Mount, AY Deutch, DM Lovinger, and E Delpire: Hyperexcitability and epilepsy associated with disruption of the mouse neuronal-specific K-Cl cotransporter gene. *Hippocampus. 12(2)* 258-68 (2002) DOI: 10.1002/hipo.10014
- 180. S Khirug, F Ahmad, M Puskarjov, R Afzalov, K Kaila, and P Blaesse: A single seizure episode leads to rapid functional activation of KCC2 in the neonatal rat hippocampus. J Neurosci. 30(36) 12028-35 (2010) DOI: 10.1523/JNEUROSCI.3154-10.2010
- 181. K Mitsuya, N Nitta, and F Suzuki: Persistent zinc depletion in the mossy fiber terminals in the intrahippocampal kainate mouse model of mesial temporal lobe epilepsy. *Epilepsia.* 50(8) 1979-90 (2009) DOI: 10.1111/j.1528-1167.2009.02055.x
- 182. J Qian, K Xu, J Yoo, TT Chen, G Andrews, and JL Noebels: Knockout of Zn transporters Zip-1 and Zip-3 attenuates seizure-induced CA1 neurodegeneration. J Neurosci. 31(1) 97-104 (2011) DOI: 10.1523/JNEUROSCI.5162-10.2011
- 183. A Takeda, H Itoh, H Tamano, and N Oku: Responsiveness to kainate in young rats after 2-week zinc deprivation. *Biometals. 19(5)* 565-72 (2006) DOI: 10.1007/s10534-005-6145-9
- 184. A Takeda, H Itoh, M Hirate, and N Oku: Region-specific loss of zinc in the brain in pentylentetrazole-induced seizures and seizure susceptibility in zinc deficiency.

Epilepsy Res. 70(1) 41-8 (2006) DOI: 10.1016/j.eplepsyres.2006.03.002

- 185. R Fallah, S Sabbaghzadegan, SA Karbasi, and F Binesh: Efficacy of zinc sulfate supplement on febrile seizure recurrence prevention in children with normal serum zinc level: A randomised clinical trial. Nutrition. 31(11-12) 1358-61 (2015) DOI: 10.1016/j.nut.2015.05.024
- 186. DS Alam, M Yunus, S El Arifeen, HR Chowdury, CP Larson, DA Sack, AH Baqui, and RE Black: Zinc treatment for 5 or 10 days is equally efficacious in preventing diarrhea in the subsequent 3 months among Bangladeshi children. J Nutr. 141(2) 312-5 (2011)

DOI: 10.3945/jn.110.120857

187. CL Walker and RE Black: Zinc for the treatment of diarrhoea: effect on diarrhoea morbidity, mortality and incidence of future episodes. Int J Epidemiol. 39 Suppl 1 i63-9 (2010)

DOI: 10.1093/ije/dyg023

- 188. S Sazawal, RE Black, MK Bhan, N Bhandari, A Sinha, and S Jalla: Zinc supplementation in young children with acute diarrhea in India. N Engl J Med. 333(13) 839-44 (1995) DOI: 10.1056/NEJM199509283331304
- 189. GW Lindenmayer, RJ Stoltzfus, and AJ Prendergast: Interactions between zinc deficiency and environmental enteropathy in developing countries. Adv Nutr. 5(1) 1-6 (2014) DOI: 10.3945/an.113.004838
- 190. A Finamore, M Massimi, L Conti Devirgiliis, and E Mengheri: Zinc deficiency induces membrane barrier damage and increases neutrophil transmigration in Caco-2 cells. J Nutr. 138(9) 1664-70 (2008)
- 191. J Geiser, KJ Venken, RC De Lisle, and GK Andrews: A mouse model of acrodermatitis enteropathica: loss of intestine zinc transporter ZIP4 (SIc39a4) disrupts the stem cell niche and intestine integrity. PLoS Genet. 8(6) e1002766 (2012) DOI: 10.1371/journal.pgen.1002766
- 192. CN Glover, NR Bury, and C Hogstrand: Intestinal zinc uptake in freshwater rainbow trout: evidence for apical pathways associated with potassium efflux and modified by calcium. Biochim Biophys Acta. 1663(1-2) 214-21 (2004) DOI: 10.1016/j.bbamem.2004.03.008

- 193. CN Glover, NR Bury, and C Hogstrand: Zinc uptake across the apical membrane of freshwater rainbow trout intestine is mediated by high affinity, low affinity, and histidine-facilitated pathwavs. Biochim Biophys Acta. 1614(2) 211-9 (2003) DOI: 10.1016/S0005-2736(03)00178-0
- 194. D Moechars, I Depoortere, B Moreaux, B de Smet, I Goris, L Hoskens, G Daneels, S Kass, L Ver Donck, T Peeters, and B Coulie: Altered gastrointestinal and metabolic function in the GPR39-obestatin receptorknockout mouse. Gastroenterology. 131(4) 1131-41 (2006) DOI: 10.1053/j.gastro.2006.07.009
- 195. I Depoortere: GI functions of GPR39: novel biology. Curr Opin Pharmacol. 12(6) 647-52 (2012)DOI: 10.1016/j.coph.2012.07.019
- 196. GL Gopalsamy, DH Alpers, HJ Binder, CD Tran, BS Ramakrishna, I Brown, M Manary, E Mortimer, and GP Young: The relevance of the colon to zinc nutrition. Nutrients. 7(1) 572-83 (2015) DOI: 10.3390/nu7010572
- 197. YY Yu, CP Kirschke, and L Huang: Immunohistochemical analysis of ZnT1, 4, 5, 6, and 7 in the mouse gastrointestinal tract. J Histochem Cytochem. 55(3) 223-34 (2007)DOI: 10.1369/jhc.6A7032.2006
- 198. X Dong, S Tang, W Zhang, W Gao, and Y Chen: GPR39 activates proliferation and differentiation of porcine intramuscular preadipocytes through targeting the PI3K/ AKT cell signaling pathway. J Recept Signal Transduct Res, 1-9 (2015)
- 199. D Scharlau, A Borowicki, N Habermann, T Hofmann, S Klenow, C Miene, U Munjal, K Stein, and M Glei: Mechanisms of primary cancer prevention by butyrate and other products formed during gut flora-mediated fermentation of dietary fibre. Mutat Res. 682(1) 39-53 (2009) DOI: 10.1016/j.mrrev.2009.04.001
- 200. Y Zhang, L Zhou, YL Bao, Y Wu, CL Yu, YX Huang, Y Sun, LH Zheng, and YX Li: Butyrate induces cell apoptosis through activation of JNK MAP kinase pathway in human colon cancer RKO cells. Chem Biol Interact. 185(3) 174-81 (2010) DOI: 10.1016/j.cbi.2010.03.035

- 201. DC Yu, JS Waby, H Chirakkal, CA Staton, and BM Corfe: Butyrate suppresses expression of neuropilin I in colorectal cell lines through inhibition of Sp1 transactivation. *Mol Cancer. 9*(276) 276 (2010) DOI: 10.1186/1476-4598-9-276
- 202. M Bordonaro, DL Lazarova, and AC Sartorelli: Butyrate and Wnt signaling: a possible solution to the puzzle of dietary fiber and colon cancer risk? *Cell Cycle.* 7(9) 1178-83 (2008) DOI: 10.4161/cc.7.9.5818
- 203. C Stock, RA Cardone, G Busco, H Krahling, A Schwab, and SJ Reshkin: Protons extruded by NHE1: digestive or glue? *Eur. J. Cell. Biol.* 87(8-9) 591-9 (2008) DOI: 10.1016/j.ejcb.2008.01.007
- 204. B Pajak and A Orzechowski: Clusterin: the missing link in the calcium-dependent resistance of cancer cells to apoptogenic stimuli. *Postepy Hig Med Dosw (Online).* 60 45-51 (2006)
- 205. P Mazzarelli, S Pucci, and LG Spagnoli: CLU and colon cancer. The dual face of CLU: from normal to malignant phenotype. *Adv Cancer Res. 105* 45-61 (2009) DOI: 10.1016/S0065-230X(09)05003-9
- 206. W Opoka, D Adamek, M Plonka, W Reczynski, B Bas, D Drozdowicz, P Jagielski, Z Sliwowski, P Adamski, and T Brzozowski: Importance of luminal and mucosal zinc in the mechanism of experimental gastric ulcer healing. *J Physiol Pharmacol.* 61(5) 581-91 (2010)
- 207. M Krasovec and E Frenk: Acrodermatitis enteropathica secondary to Crohn's disease. *Dermatology. 193(4)* 361-3 (1996). DOI: 10.1159/000246296
- 208. GC Sturniolo, W Fries, E Mazzon, V Di Leo, M Barollo, and R D'Inca: Effect of zinc supplementation on intestinal permeability in experimental colitis. *J Lab Clin Med.* 139(5) 311-5 (2002) DOI: 10.1067/mlc.2002.123624
- 209. HH Luk, JK Ko, HS Fung, and CH Cho: Delineation of the protective action of zinc sulfate on ulcerative colitis in rats. *Eur J Pharmacol.* 443(1-3) 197-204. (2002) DOI: 10.1016/S0014-2999(02)01592-3
- 210. A Nusrat, JR Turner, and JL Madara: Molecular physiology and pathophysiology

of tight junctions. IV. Regulation of tight junctions by extracellular stimuli: nutrients, cytokines, and immune cells. *Am J Physiol Gastrointest Liver Physiol.* 279(5) G851-7 (2000)

- 211. KL Edelblum and JR Turner: The Tight Junction in Inflammatory Disease: Communication Breakdown. *Curr Opin Pharmacol* 9715-720 (2009) DOI: 10.1016/j.coph.2009.06.022
- 212. KM Hoque and HJ Binder: Zinc in the treatment of acute diarrhea: current status and assessment. *Gastroenterology.* 130(7) 2201-5 (2006) DOI: 10.1053/j.gastro.2006.02.062
- 213. J Geiser, RC De Lisle, D Finkelstein, PA Adlard, AI Bush, and GK Andrews: Clioquinol synergistically augments rescue by zinc supplementation in a mouse model of acrodermatitis enteropathica. *PLoS One.* 8(8) e72543 (2013) DOI: 10.1371/journal.pone.0072543
- 214. M Furuse, M Itoh, T Hirase, A Nagafuchi, S Yonemura, and S Tsukita: Direct association of occludin with ZO-1 and its possible involvement in the localization of occludin at tight junctions. *J Cell Biol.* 127(6 Pt 1) 1617-26 (1994) DOI: 10.1083/jcb.127.6.1617
- 215. H Chiba, M Osanai, M Murata, T Kojima, and N Sawada: Transmembrane proteins of tight junctions. *Biochim Biophys Acta. 1778(3)* 588-600 (2008) DOI: 10.1016/j.bbamem.2007.08.017
- 216. Y Guan, AJ Watson, AM Marchiando, E Bradford, L Shen, JR Turner, and MH Montrose: Redistribution of the tight junction protein ZO-1 during physiological shedding of mouse intestinal epithelial cells. *Am J Physiol Cell Physiol.* 300(6) C1404-14 (2011) DOI: 10.1152/ajpcell.00270.2010
- 217.V Singh, J Yang, TE Chen, NC Zachos, O Kovbasnjuk, AS Verkman, and M Donowitz: Translating molecular physiology of intestinal transport into pharmacologic treatment of diarrhea: stimulation of Na+ absorption. *Clin Gastroenterol Hepatol. 12(1)* 27-31 (2014) DOI: 10.1016/j.cgh.2013.10.020
- 218. M Medani, VA Bzik, A Rogers, D Collins, R Kennelly, DC Winter, DJ Brayden, and AW

Baird: Zinc sulphate attenuates chloride secretion in human colonic mucosae in vitro. *Eur J Pharmacol.* 696(1-3) 166-71 (2012) DOI: 10.1016/j.ejphar.2012.09.017

- 219. RB Canani, P Cirillo, V Buccigrossi, S Ruotolo, A Passariello, P De Luca, F Porcaro, G De Marco, and A Guarino: Zinc inhibits cholera toxin-induced, but not Escherichia coli heat-stable enterotoxin-induced, ion secretion in human enterocytes. *J Infect Dis. 191(7)* 1072-7 (2005) DOI: 10.1086/428504
- 220. AC Girardi and F Di Sole: Deciphering the mechanisms of the Na+/H+ exchanger-3 regulation in organ dysfunction. *Am J Physiol Cell Physiol.* 302(11) C1569-87 (2012) DOI: 10.1152/ajpcell.00017.2012
- 221. JR Thiagarajah, EA Ko, L Tradtrantip, M Donowitz, and AS Verkman: Discovery and development of antisecretory drugs for treating diarrheal diseases. *Clin Gastroenterol Hepatol. 12(2)* 204-9 (2014) DOI: 10.1016/j.cgh.2013.12.001
- 222. M Andrews and C Gallagher-Allred: The role of zinc in wound healing. *Adv. Wound Care. 12(3)* 137-8 (1999)
- 223. AB Lansdown, U Mirastschijski, N Stubbs, E Scanlon, and MS Agren: Zinc in wound healing: Theoretical, experimental, and clinical aspects. *Wound Repair Regen. 15(1)* 2-16 (2007) DOI: 10.1111/j.1524-475X.2006.00179.x
- 224. AB Lansdown: Zinc in the healing wound. *Lancet.* 347(9003) 706-7 (1996) DOI: 10.1016/S0140-6736(96)90072-0
- 225. JR Schwartz, RG Marsh, and ZD Draelos: Zinc and skin health: overview of physiology and pharmacology. *Dermatol. Surg. 31(7 Pt 2)* 837-47; discussion 847 (2005) DOI: 10.1111/j.1524-4725.2005.31729
- 226. SL Jensen, C McCuaig, A Zembowicz, and MA Hurt: Bullous lesions in acrodermatitis enteropathica delaying diagnosis of zinc deficiency: a report of two cases and review of the literature. *J. Cutan. Pathol. 35 Suppl 1* 1-13 (2008) DOI: 10.1111/j.1600-0560.2008.00981.x
- 227. H Takahashi, M Nakazawa, K Takahashi, M Aihara, M Minami, T Hirasawa, and Z Ikezawa: Effects of zinc deficient diet

on development of atopic dermatitis-like eruptions in DS-Nh mice. *J. Dermatol. Sci. 50(1)* 31-9 (2008) DOI: 10.1016/j.jdermsci.2007.11.002

- 228. GK Andrews: Regulation and function of Zip4, the acrodermatitis enteropathica gene. *Biochem. Soc. Trans.* 36(*Pt 6*) 1242-6 (2008) DOI: 10.1042/BST0361242
- 229. I Lasry, YA Seo, H Ityel, N Shalva, B Pode-Shakked, F Glaser, B Berman, I Berezovsky, A Goncearenco, A Klar, J Levy, Y Anikster, SL Kelleher, and YG Assaraf: A dominant negative heterozygous G87R mutation in the zinc transporter, ZnT-2 (SLC30A2), results in transient neonatal zinc deficiency. *J Biol Chem.* 287(35) 29348-61 (2012) DOI: 10.1074/jbc.M112.368159
- 230. W Chowanadisai, B Lonnerdal, and SL Kelleher: Identification of a mutation in SLC30A2 (ZnT-2) in women with low milk zinc concentration that results in transient neonatal zinc deficiency. *J Biol Chem.* 281(51) 39699-707 (2006) DOI: 10.1074/jbc.M605821200
- 231. YB Nitzan, I Sekler, and WF Silverman: Histochemical and histofluorescence tracing of chelatable zinc in the developing mouse. *J Histochem Cytochem*. *52*(*4*) 529-39 (2004) DOI: 10.1177/002215540405200411
- 232. L Stuwe, M Muller, A Fabian, J Waning, S Mally, J Noel, A Schwab, and C Stock: pH dependence of melanoma cell migration: protons extruded by NHE1 dominate protons of the bulk solution. *J. Physiol.* 585(Pt 2) 351-60 (2007) DOI: 10.1113/jphysiol.2007.145185
- 233. JS Huang, JJ Mukherjee, T Chung, KS Crilly, and Z Kiss: Extracellular calcium stimulates DNA synthesis in synergism with zinc, insulin and insulin-like growth factor I in fibroblasts. *Eur. J. Biochem.* 266(3) 943-51 (1999) DOI: 10.1046/j.1432-1327.1999.00932.x
- 234. B Schmidt-Hansen, J Klingelhofer, B Grum-Schwensen, A Christensen, S Andresen, C Kruse, T Hansen, N Ambartsumian, E Lukanidin, and M Grigorian: Functional significance of metastasis-inducing S100A4(Mts1) in tumor-stroma interplay. J Biol Chem. 279(23) 24498-504 (2004) DOI: 10.1074/jbc.M400441200

- 235. C Hogstrand, P Kille, RI Nicholson, and KM Taylor: Zinc transporters and cancer: a potential role for ZIP7 as a hub for tyrosine kinase activation. *Trends Mol. Med.* 15(3) 101-11 (2009) DOI: 10.1016/j.molmed.2009.01.004
- 236. V Lopez and SL Kelleher: Zip6-attenuation promotes epithelial-to-mesenchymal transition in ductal breast tumor (T47D) cells. *Exp Cell Res. 316(3)* 366-75 (2010) DOI: 10.1016/j.yexcr.2009.10.011
- 237. SAlam and SL Kelleher: Cellular mechanisms of zinc dysregulation: a perspective on zinc homeostasis as an etiological factor in the development and progression of breast cancer. *Nutrients. 4*(*8*) 875-903 (2012) DOI: 10.3390/nu4080875
- 238. A Hermani, B De Servi, S Medunjanin, PA Tessier, and D Mayer: S100A8 and S100A9 activate MAP kinase and NF-kappaB signaling pathways and trigger translocation of RAGE in human prostate cancer cells. *Exp Cell Res.* 312(2) 184-97 (2006) DOI: 10.1016/j.yexcr.2005.10.013
- 239. A Hermani, J Hess, B De Servi, S Medunjanin, R Grobholz, L Trojan, P Angel, and D Mayer: Calcium-binding proteins S100A8 and S100A9 as novel diagnostic markers in human prostate cancer. *Clin Cancer Res. 11(14)* 5146-52 (2005) DOI: 10.1158/1078-0432.CCR-05-0352
- 240. S Grebhardt, K Muller-Decker, F Bestvater, M Hershfinkel, and D Mayer: Impact of S100A8/A9 expression on prostate cancer progression in vitro and in vivo. *J Cell Physiol.* 229(5) 661-71 (2014) DOI: 10.1002/jcp.24489
- 241.K Boye and GM Maelandsmo: S100A4 and metastasis: a small actor playing many roles. *Am J Pathol. 176(2)* 528-35 (2010) DOI: 10.2353/ajpath.2010.090526
- 242. ML Joiner, OM Koval, J Li, BJ He, C Allamargot, Z Gao, ED Luczak, DD Hall, BD Fink, B Chen, J Yang, SA Moore, TD Scholz, S Strack, PJ Mohler, WI Sivitz, LS Song, and ME Anderson: CaMKII determines mitochondrial stress responses in heart. *Nature.* 491(7423) 269-73 (2012) DOI: 10.1038/nature11444
- 243.L Huang, CP Kirschke, and Y Zhang: Decreased intracellular zinc in human

tumorigenic prostate epithelial cells: a possible role in prostate cancer progression. *Cancer Cell Int.* 6 10 (2006) DOI: 10.1186/1475-2867-6-10

- 244. LC Costello, Y Liu, J Zou, and RB Franklin: Evidence for a zinc uptake transporter in human prostate cancer cells which is regulated by prolactin and testosterone. *J Biol Chem.* 274(25) 17499-504 (1999) DOI: 10.1074/jbc.274.25.17499
- 245. M Fassnacht, D Weismann, S Ebert, P Adam, M Zink, F Beuschlein, S Hahner, and B Allolio: AKT is highly phosphorylated in pheochromocytomas but not in benign adrenocortical tumors. *J Clin Endocrinol Metab.* 90(7) 4366-70 (2005) DOI: 10.1210/jc.2004-2198
- 246. A Arcaro and AS Guerreiro: The phosphoinositide 3-kinase pathway in human cancer: genetic alterations and therapeutic implications. *Curr Genomics. 8(5)* 271-306 (2007) DOI: 10.2174/138920207782446160
- 247. J Zou, BC Milon, MM Desouki, LC Costello, and RB Franklin: hZIP1 zinc transporter down-regulation in prostate cancer involves the overexpression of ras responsive element binding protein-1 (RREB-1). *Prostate.* 71(14) 1518-24 (2011) DOI: 10.1002/pros.21368
- 248. RB Franklin and LC Costello: Zinc as an anti-tumor agent in prostate cancer and in other cancers. *Arch Biochem Biophys. 463(2)* 211-7 (2007) DOI: 10.1016/j.abb.2007.02.033
- 249. RB Franklin, B Milon, P Feng, and LC Costello: Zinc and zinc transporters in normal prostate and the pathogenesis of prostate cancer. *Front Biosci.* 10 2230-9 (2005) DOI: 10.2741/1692
- 250. S Tepaamorndech, L Huang, and CP Kirschke: A null-mutation in the Znt7 gene accelerates prostate tumor formation in a transgenic adenocarcinoma mouse prostate model. *Cancer Lett.* 308(1) 33-42 (2011) DOI: 10.1016/j.canlet.2011.04.011
- 251. F Xie, H Liu, YH Zhu, YR Qin, Y Dai, T Zeng, L Chen, C Nie, H Tang, Y Li, L Fu, and XY Guan: Overexpression of GPR39 contributes to malignant development

of human esophageal squamous cell carcinoma. *BMC Cancer. 11* 86 (2011) DOI: 10.1186/1471-2407-11-86

- 252. BO Alen, S Leal-Lopez, MO Alen, P Viano, V Garcia-Castro, CS Mosteiro, A Beiras, FF Casanueva, R Gallego, T Garcia-Caballero, JP Camina, and Y Pazos: The role of the obestatin/GPR39 system in human gastric adenocarcinomas. *Oncotarget*, 7(5) 5957-71 (2015)
- 253. D Wootten, A Christopoulos, and PM Sexton: Emerging paradigms in GPCR allostery: implications for drug discovery. *Nat Rev Drug Discov. 12(8)* 630-44 (2013) DOI: 10.1038/nrd4052
- 254. C Custodi, R Nuti, TI Oprea, and A Macchiarulo: Fitting the complexity of GPCRs modulation into simple hypotheses of ligand design. *J Mol Graph Model.* 38 70-81 (2012). DOI: 10.1016/j.jmgm.2012.07.002

Key Words: Zinc, GPR39, Zinc Signaling, Neuron, Keratinocyte, Epithelium, Intestine, Colon, Review

Send correspondence to: Michal Hershfinkel, Department of Physiology and Cell Biology and The Zlotowski Center for Neuroscience, Faculty of Health Sciences, POB 653, Ben-Gurion Ave. Ben-Gurion university of the Negev, Beer Sheva, 84105, Israel, Tel: 972-8-6477318, E-mail: hmichal@bgu.ac.il