Bone marrow mesenchymal stem cells in hepatocellular carcinoma

Peng Gong¹, Yingxin Wang¹, Jing Zhang¹, Zhongyu Wang¹

¹Department of Hepatobiliary Surgery, the First Affiliated Hospital of Dalian Medical University, Dalian, China

TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. BMSCs and promotion of tumor growth
 - 3.1. BMSC role in HCC tumor initiation and growth
 - 3.2. BMSCs home to sites with tumor cells
- 4. BMSCs and antitumor activity
 - 4.1. Animal study findings of BMSCs' therapeutic potential
 - 4.2. Clinical applications of BMSCs
- 5. Conclusions
- 6. Acknowledgments
- 9. References

1. ABSTRACT

Bone marrow mesenchymal stem cells (BMSCs) are non-hematopoietic multipotent stem cells capable of differentiating into mature cells. Studies in animal models have indicated that hepatocellular carcinoma (HCC) may originate from genetically mutated BMSCs. Moreover, it has been shown that BMSCs are influenced by and can modulate their micro-environment via secreted cytokines that promote tumor initiation, growth, and homing to tumor sites. Based on these features, BMSCs have been recognized as a putative target of molecular therapies to treat and prevent HCC. In this review we discuss the role of human BMSCs in HCC pathogenesis and their therapeutic potential.

2. INTRODUCTION

Human bone marrow mesenchymal stem cells (hBMSCs) are non-hematopoietic multipotent stem cells capable of differentiating into both mesenchymal and non-mesenchymal cell types, including osteoblasts, adipocytes, and chondrocytes (1-5). Many studies have shown that hBMSCs can stimulate tumor growth and metastasis *in vivo* by facilitating the expansion of tumor-associated fibroblasts, stimulating angiogenesis, suppressing the cytotoxic function of immune cells, and secreting chemokines (6-9). Interestingly, opposing effects of BMSCs on tumor cell growth have been observed according to the use of *in vitro* and *in vivo* experimental

systems; for example, exposure to BMSCs in vitro led to transient arrest of tumor cells in the G(1) phase of the cell cycle, while in vivo exposure led to increased tumor growth (10). This discrepancy between experimental systems may reflect the ability of BMSCs to interact with their environment and other endogenous factors to form a cancer stem cell niche in which tumor cells can preserve their potential to proliferate and sustain the malignant process. Such a dynamic mechanism may reveal several molecules and pathways that hold promise for therapeutic manipulation. To this end, hBMSCs are generally considered well suited for clinical application because they are easily obtained from patients, with their procurement posing no ethical concerns, and can be used in autologous transplantation (11.12). Moreover, the tropism of hBMSCs and tumors implies that such cells could potentially serve as gene delivery vectors in cancer gene therapy (13). Before the full clinical benefit of hBMSC-based therapies may be fully realized, however, a detailed understanding of the effects of unmodified hBMSCs on tumor progression must be achieved.

Hepatocellular carcinoma (HCC) is the fifth most prevalent cancer worldwide, and the third in terms of cancer-related deaths (14). In China, the mortality rate of HCC has steadily increased since the 1990s, emerging as the second leading cause of cancer deaths (15). While high incidences of recurrence and metastasis have been implicated as the primary causes of HCC's poor prognosis (16), the pathogenic underlying mechanisms have yet to be elucidated. Considering the collective findings indicating hBMSCs as an effective delivery vehicle, researchers have begun to investigate the possibility of using hBMSCsmediated gene therapy to treat HCC. A recent study in nude mice demonstrated that hBMSCs may play a role in the pathogenic mechanisms of HCC, further suggesting their potential as effective therapeutic agents of HCC (17). In this review, we will discuss the dual roles of hBMSCs in HCC and the implications of these roles for developing more effective HCC prevention and treatment strategies.

3. BMSCs AND PROMOTION OF TUMOR GROWTH

The effects of BMSCs on the initiation, progression, and metastasis of certain tumor types have been demonstrated in various experimental and model systems (9,18,19), as has the involvement of their secreted paracrine factors (20-22). These studies have indicated that BMSCs participate in liver regeneration by migrating to the affected site and differentiating into hepatic precursor cells (oval cells) and hepatocytes. However, chronic conditions of liver injury, such as hepatitis virus infection, alcoholism and congenital fibrosis, overcome the normal regenerative mechanisms and cause extracellular matrix (ECM) remodeling, cirrhosis, and liver failure. Alterations in the ECM components of the liver tissues triggers a signaling cascade that inhibits the transactivation potential of liverspecific transcription factors, leading to the arrest of BMSC differentiation and poorly differentiated liver tissues that are characteristic of HCC (23-36).

3.1. BMSC role in HCC tumor initiation and growth

The established association between cancer and chronic tissue injury, primarily related to the inflammatory response, suggests that cancer growth may represent the continuous operation of an unregulated state of tissue repair (37). Taken together, the well-known roles of the Hedgehog and Wnt signaling pathways in tissue regeneration, stem cell renewal, and cancer growth suggest that carcinogenesis proceeds by misappropriation of the homeostatic mechanisms that govern tissue repair and stem cell self-renewal (38). Malignant transformation of hepatocytes, therefore, may occur in the context of chronic inflammation and regeneration, which is in line with the potential role for BMSCs contributing to the eventual development of HCC (39).

In vitro experiments have demonstrated that HCC can be derived from genetically mutated rat BMSCs (40). When a rat hepatoma cell line was treated with the medium of cultured rat BMSCs, containing the full complement of secreted factors, cellular proliferation and cell division were markedly enhanced in a dose-dependent manner (41). Similarly, isolated hBMSCs were shown to be able to differentiate into hepatocyte-like cells that resembled poorly differentiated human hepatoma cell lines (42). Furthermore, when the hepatocyte nuclear factoralpha (HNF-4□ transcription factor was overexpressed in the isolated hBMSCs, the expression levels of other hepatocyte-specific genes, liver-enriched transcription factor genes, and cytochrome P450 genes became markedly up-regulated, indicating that HNF-4□ plays a significant role in promoting the process of hBMSC differentiation towards the hepatocyte phenotype (42). In addition, primary BMSCs were shown to exhibit immunosuppressive properties following injection into mice (43). Taken together, these findings suggest that BMSCs promote tumor growth.

3.2. BMSCs home to sites with tumor cells

BMSCs' involvement in tumor invasion and angiogenesis (44-47), immunosuppression (48,49), and inhibition of apoptosis (10) has been demonstrated in multiple studies and various experimental systems. In addition, hBMSCs have been shown to selectively localize to xenotransplanted human gliomas following intravascular administration, where they successfully delivered antiglioma agents (50,51). This capacity to localize to gliomas may reflect the intrinsic ability of BMSCs to home to solid tumors, regardless of the underlying cell type (52-55). However, recent studies of the BMSCs tumor homing mechanism indicated that this process may actually generate a microenvironment that is suitable for tumor cells, thereby promoting tumor growth and negating any potential therapeutic benefit (56). Focused studies of the tumor microenvironment revealed that BMSCs can induce formation of this cancer-related system and promote its maintenance by functionally interacting with cancer cells (57). Such a tumor microenvironment may contribute to HCC initiation by providing a concentrated locale of soluble factors, such as cytokines, that would stimulate BMSCs to differentiate into fibroblasts or vascular endothelial cells (58,59). Likewise, the BMSC secreted

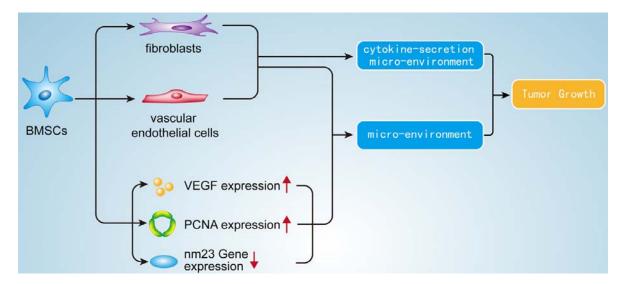


Figure 1. Bone marrow mesenchymal stem cells (BMSCs) promote tumor growth. BMSCs potentially participate in the formation of a tumor microenvironment and a secreted cytokine microenvironment via differentiation into fibroblasts or vascular endothelial cells, which are involved in the initiation of HCC. In addition, BMSC transplantation could increase the expression of vascular endothelial cell growth factor (VEGF) and proliferating cell nuclear antigen (PCNA), while decreasing the expression of the tumor metastasis inhibiting gene nm23 in hepatoma cells, resulting in the formation of the tumor microenvironment and promoting tumor growth.

factors in the microenvironment may induce HCC or promote its growth. Shao *et al.* reported that BMSC transplantation in a rat hepatoma model resulted in increased expression of vascular endothelial cell growth factor (VEGF) and of proliferating cell nuclear antigen (PCNA), but decreased expression of the tumor metastasis inhibiting gene nm23, and concluded that the consequent microenvironment supported the observed increase in tumor growth (60) (Figure 1). Although several studies have demonstrated that BMSCs can migrate to tumor and injury sites and to incorporate into the tumor stroma, the effects of the interactions between BMSCs and tumor cells and the mechanisms underlying these effects have yet to be elucidated.

On the other hand, hBMSCs and human hepatoma cells exhibit different responses to environmental stimuli, likely reflecting their unique cellular behavior and surface characteristics. The newly developed hydrophobic poly(3biodegradable, polyester hydroxybutyrate-co-3-hydroxyhexnoate) (PHBHHx) has attracted the attention of tissue engineering researchers hoping to improve the distribution of hBMSCs in vivo. It has been proposed that the orientation of scars on the hBMSC surface would guide the intracellular growth direction of the actin cytoskeleton. In contrast, it has been demonstrated that the surface characteristics of the human hepatoma cell line C3A/HepG2 are obviously associated with their metabolic activity but not with their morphology. C3A/HepG2 cells exhibit unique cellular The characteristics when compared to the hMSCs/HepG2 cells which likely contribute to their differential responses to environmental stimuli (61). Therefore, it is possible that the microenvironment may exert different influence on BMSCs and hepatoma cells.

Recent studies have shown that BMSCs that home to tumors not only generate a suitable microenvironment for tumor cells, but also promote tumor metastasis (24,62,63). However, only a few reports to date have addressed the ability of BMSCs to promote HCC metastasis in this manner.

4. BMSCs AND ANTITUMOR ACTIVITY

MSCs can be expanded in culture for long periods of time without a loss of differentiation capacity. Furthermore, since MSCs are particularly amenable to genetic modification/correction, they can be harvested from a patient's own bone marrow even if the patient's liver disease were the result of an underlying genetic defect. Genetically corrected autologous MSCs could thus be propagated to generate a sufficient number of cells to achieve a meaningful level of engraftment following transplantation. In vitro studies have provided definitive evidence that BMSCs can, under appropriate conditions, differentiate into cells with all of the characteristics of functional hepatocytes (64-67). In addition, BMSCexosomes have been proposed as another potentially manipulable regulatory mechanism of the paracrine action of BMSCs. Bruno et al. reported that BMSC-derived microvesicles can protect against acute tubular injury via horizontal transfer of mRNA (68). Thus, BMSCs appear to be able to exert beneficial effects in a wide range of injuries and disease states within the liver, including HCC. Du and colleagues demonstrated that IFN- $\Box\text{-stimulated}$ hBMSCs were able to induce tumor cell apoptosis in vitro via tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) (69). Thus, these beneficial therapeutic effects coupled with the ability of BMSCs to home to sites of injury and tumors

further support the therapeutic potential of these cells for HCC (70,71).

4.1. Animal study findings of BMSCs' therapeutic potential

Studies based on HCC animal models have revealed the antitumor activity of BMSCs. However, the effects of unmodified BMSCs on tumor progression remain unclear. Some studies have suggested that BMSCs can induce tumor cell necrosis or suppress tumor growth. For example, Jiang et al. found that BMSCs not only engraft within the livers of carcinoma-bearing BALB/c mice but also differentiate to hepatocyte-like cells (74). And at the same time, BMSCs might induce tumor cells necrosis. In addition, administration of MSCs in chemically-induced HCC rats suppressed the tumor growth, as evidenced by marked down-regulation of Wnt signaling target genes that are associated with antiapoptosis, mitogenesis, cell proliferation and cell cycle regulation, as well as amelioration of liver histopathology and function (75). The BMSC-inhibited tumor growth was also shown to be correlated with increased overall survival of HCC rats (76). When rat BMSCs labeled with superparamagnetic iron oxide (SPIO) were transplanted into HCC rats, the tumor volume at post-transplantation weeks 1 and 2 was found to be significantly smaller than that in the control rats, as determined by magnetic resonance imaging (MRI).

BMSCs potentially control the growth of tumor cells in metabolism as one of the important factors to inhibit the proliferation of malignant cells. Qiao and colleagues found that the clonality and proliferation of hepatoma cells were inhibited upon culture in medium harvested from BMSC cultures; moreover, the inhibited hepatoma cells expressed significantly lower levels of nuclear transcription factor P8, the member of RhoGTP family CDC42EP and NK-kappaB2, but higher levels of metallothionein (MT) than the controls (77). All of these genes and cytokines are known to be involved in tumor cell metabolism. Recently. Lu and colleagues demonstrated that BMSCs exhibit potential inhibitory effects on tumor cell growth in vitro and in vivo, without inducing host immunosuppression, by triggering apoptotic cell death and arrest in the G(0)/G(1) phase (78). It is possible that these findings reflect BMSC-induced up-regulation of the cell cycle negative regulator p21 and/or the apoptosisassociated protease caspase 3 in tumor cells.

The tropism of hBMSCs toward tumors suggests that these cells may prove useful as gene delivery vectors for cancer gene therapy (79,80). Due to its dual role as a reporter and therapy gene, the sodium iodide symporter (NIS) allows non-invasive imaging of functional NIS expression by (123)I-scintigraphy or (124)I-PET imaging to be carried out prior to the application of a therapeutic dose of (131)I. As such, NIS expression monitoring has provided a novel approach by which to evaluate mesenchymal stem cells (MSCs) as gene delivery vehicles for tumor therapy. Knoop *et al.* stably transfected bone marrow-derived CD34⁻MSCs with NIS cDNA, and three cycles of systemic BMSC-mediated NIS gene delivery followed by (131)I application resulted in a significant delay in tumor growth

(81). These results demonstrated the tumor-specific accumulation and therapeutic efficacy of radioiodine after BMSC-mediated NIS gene delivery in HCC tumors, and suggest that NIS-mediated radionuclide therapy of metastatic cancer using BMSCs as gene delivery vehicles may prove efficacious (81). In addition, the use of engineered BMSCs as therapeutic vehicles has been reported for the HCC xenografted mouse model. When isolated MSCs from bone marrow of C57/Bl6 p53^{-/-} mice were injected, exogenous BMSCs were recruited to the growing HCC xenografts and this process was accompanied by activation of the CCL5 or Tie2 promoters within the injected BMSCs. Furthermore, stem cellmediated introduction of suicide genes into the HCC xenografted tumor followed by administration of a routine drug regimen effectively resolved the HCC (82).

4.2. Clinical applications of BMSCs

The discovery of pluripotent stem cells made the prospect of cell therapy and tissue regeneration a clinical reality, particularly following the evidenced contribution of bone marrow-derived stem cells in hepatic regeneration. Since then, several research groups have aimed at developing effective and convenient clinical applications of BMSCs to treat HCC. Fürst et al. treated patients with malignant liver lesions using a combination of portal vein embolization (PVE) with BMSCs administration, and found that the method produced substantially more robust hepatic regeneration than PVE alone (83). In HCC patients who are otherwise unsuitable for resection, transplantation, ablation therapy or arterial chemoembolization, administration of stem cell differentiation stage factors have been shown to be effective (84). This finding suggests that cytokines secreted by BMSCs could potentially inhibit the growth of HCC by impacting the tumor microenvironment in patients.

Infusion of BMSCs prior to trans-arterial chemoembolization may help to promote liver regeneration, consequently increasing liver volume and the hepatic reserve, in patients with HCC. In a long-term follow-up clinical trial, 527 patients with hepatitis B virus (HBV)-related decompensated liver cirrhosis were found to experience improved hepatic function in the early period of treatment and lower tendency of HCC development in the long-term. Moreover, the long-term observation of these patients revealed no change in the incidence of HCC following the administration of BMSCs, suggesting the possibility of an improved survival rate associated with the BMSC treatment (85). The safety and efficacy of BMSCs in HCC have been reported by Ismail et al. (86). In that study, Child-Pugh class B patients with unresectable HCC were treated by transarterial chemoembolization and injected, during the same session, with autologous bone marrow mononuclear layer containing stem cells into the hepatic artery feeding the contralateral lobe of the liver. Results obtained at the 3 month follow-up indicated that BMSC infusion into the hepatic artery synchronized with transcatheter arterial chemoembolization (TACE) was safe and feasible for patients with chronic liver disease complicated with HCC, as evidenced by remarkable improvements in both biological and radiological

volumetric parameters (86). Although this study was carried out with only four patients, its findings indicate the promise of BMSCs in the therapy of HCC. Furthermore, a case report demonstrated that the combined treatment using autologous BMSC transplantation and TACE was an appropriate and sufficient alternative treatment for HCC patients who are unable to tolerate TACE due to hepatic dysfunction (87). Therefore, BMSCs may also benefit patients with advanced HCC, who can no longer tolerate invasive therapies due to the severe hepatic dysfunction. In the future, clinical studies involving a greater number of samples are needed to further clarify the efficacy of BMSCs in the therapy of HCC.

5. CONCLUSIONS

BMSCs play dual roles in HCC, promoting the initiation and progression of tumors and inhibiting tumor growth. As such, these cells may represent a useful target of HCC therapy or a delivery vehicle for antitumor agents. In animal models, HCC has been demonstrated to be derived from genetically mutated BMSCs. Moreover, BMSCs are potentially involved in the formation of microenvironments, such as those composed of secreted cytokines, that promote tumor growth and homing to tumor sites, whereas the BMSCs, in turn, may be influenced by the microenvironment itself to further support tumor development and growth. The initial efforts to develop BMSC-based therapies for HCC have shown promise. However, additional studies using larger sample size are needed to further clarify the efficacy and safety of BMSCs in clinical applications.

6. ACKNOWLEDGMENTS

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7. REFERENCES

- 1.Bianco P, Riminucci M, Gronthos S, Robey PG. Bone marrow stromal stem cells: nature, biology, and potential applications. *Stem Cells* 19, 180–192. (2001)
- 2.Friedenstein AJ, Chailakhyan RK, Latsinik NV, Panasyuk AF, Keiliss-Borok IV. Stromal cells responsible for transferring the microenvironment of the hemopoietic tissues. *Cloning in vitro and retransplantation in vivo. Transplantation* 17, 331–340. (1974)
- 3.Owen M, Friedenstein AJ. Stromal stem cells: marrow-derived osteogenic precursors. *Ciba Found Symp* 136, 42–60. (1988)
- 4.Pittenger MF, *et al.* Multilineage potential of adult human mesenchymal stem cells. *Science* 284, 143–147. (1999)
- 5.Prockop DJ. Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* 276, 71–74. (1997)

- 6.Spaeth EL, Dembinski JL, Sasser AK, Watson K, Klopp A, Hall B, *et al.* Mesenchymal stem cell transition to tumor-associated fibroblasts contributes to fibrovascular network expansion and tumor progression. *PloS One* 4, e4992. (2009)
- 7.Sun B, Zhang S, Ni C, Zhang D, Liu Y, Zhang W, et al. Correlation between melanoma \$angiogenesis and the mesenchymal stem cells and endothelial progenitor cells derived from bone marrow. Stem Cells Development 14, 292–8. (2005)
- 8.Djouad F, Plence P, Bony C, Tropel P, Apparailly F, Sany J, *et al.* Immunosuppressive effect of mesenchymal stem cells favors tumor growth in allogeneic animals. *Blood* 102, 3837–44. (2003)
- 9.Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, *et al.* Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature* 449, 557–63. (2007)
- 10.Ramasamy R, Lam EW-F, Soeiro I, Tisato V, Bonnet D, Dazzi F. Mesenchymal stem cells inhibit proliferation and apoptosis of tumor cells: impact on *in vivo* tumor growth. *Leukemia* 21, 304-10. (2007)
- 11.Digirolamo CM, Stokes D, Colter D, Phinney DG, Class R, Prockop DJ. Propagation and senescence of human marrow stromal cells in culture: a simple colony-forming assay identifies samples with the greatest potential to propagate and differentiate. *Br J Haematol* 107, 275–81. (1999)
- 12. Colter DC, Class R, DiGirolamo CM, Prockop DJ. Rapid expansion of recycling stem cells in cultures of plastic-adherent cells from human bone marrow. *Proc Natl Acad Sci USA* 97, 3213–8. (2000)
- 13.Kidd S, Spaeth E, Dembinski JL, Dietrich M, Watson K, Klopp A, *et al.* Direct evidence of mesenchymal stem cells tropism for tumor and wounding microenvironments using *in vivo* bioluminescent imaging. *Stem Cells* 27, 2614-23. (2009)
- 14.Bruix J, Sherman M, Llovet JM, Beaugrand M, Lencioni R, Burroughs AK, Christensen E, Pagliaro L, Colombo M, Rodés J; EASL Panel of Experts on HCC. Clinical management of hepatocellular carcinoma. Conclusions of the Barcelona-2000 EASL conference. European Association for the Study of the Liver. *J Hepatol* 35(3), 421-30. (2001)
- 15.He J, Gu D, Wu X, Reynolds K, Duan X, Yao C, Wang J, Chen CS, Chen J, Wildman RP, Klag MJ, Whelton PK. Major causes of death among men and women in China. *N Engl J Med* 353(11), 1124-34. (2005)
- 16.Nishida N, Goel A. Genetic and epigenetic signatures in human hepatocellular carcinoma: a systematic review. *Curr Genomics* 12(2), 130-7. (2011)

- 17.Gao Y, Yao A, Zhang W, Lu S, Yu Y, Deng L, Yin A, Xia Y, Sun B, Wang X. Human mesenchymal stem cells overexpressing pigment epithelium-derived factor inhibit hepatocellular carcinoma in nude mice. *Oncogene* 29(19), 2784-94. (2010)
- 18.Zhu W, Xu W, Jiang R, Qian H, Chen M, Hu J, Cao W, Han C, Chen Y. Mesenchymal stem cells derived from bone marrow favor tumor cell growth *in vivo*. *Exp Mol Pathol* 80(3), 267-74. (2006)
- 19.Shinagawa K, Kitadai Y, Tanaka M, Sumida T, Kodama M, Higashi Y, Tanaka S, Yasui W, Chayama K. Mesenchymal stem cells enhance growth and metastasis of colon cancer. *Int J Cancer* 127(10), 2323-33. (2010)
- 20.Bouffi C, Djouad F, Mathieu M, Noël D, Jorgensen C. Multipotent mesenchymal stromal cells, rheumatoid arthritis: risk or benefit? *Rheumatology (Oxford)* 48(10), 1185-9. (2009)
- 21.Beckermann BM, Kallifatidis G, Groth A, Frommhold D, Apel A, Mattern J, Salnikov AV, Moldenhauer G, Wagner W, Diehlmann A, Saffrich R, Schubert M, Ho AD, Giese N, Büchler MW, Friess H, Büchler P, Herr I. EGF expression by mesenchymal stem cells contributes to angiogenesis in pancreatic carcinoma. *Br J Cancer* 99(4), 622-31. (2008)
- 22.Roorda BD, Elst A, Boer TG, Kamps WA, de Bont ES. Mesenchymal stem cells contribute to tumor cell proliferation by direct cell–cell contact interactions. *Cancer Invest* 28(5), 526-34. (2010)
- 23. Nagaki M, Shidoji Y, Yamada Y, Sugiyama A, Tanaka M, Akaike T, Ohnishi H, Moriwaki H, Muto Y. Regulation of hepatic genes and liver transcription factors in rat hepatocytes by extracellular matrix. *Biochem Biophys Res Commun* 210(1), 38-43. (1995)
- 24.Brill S, Zvibel I, Halpern Z, Oren R. The role of fetal and adult hepatocyte extracellular matrix in the regulation of tissue-specific gene expression in fetal and adult hepatocytes. *Eur J Cell Biol* 81(1), 43–50. (2002)
- 25.Sidhu JS, Liu F, Omiecinski CJ. Phenobarbital responsiveness as a uniquely sensitive indicator of hepatocyte differentiation status: requirement of dexamethasone and extracellular matrix in establishing the functional integrity of cultured primary rat hepatocytes. *Exp Cell Res* 292(2), 252–64. (2004)
- 26.Rana B, Mischoulon D, Xie Y, Bucher NL, Farmer SR. Cellextracellular matrix interactions can regulate the switch between growth and differentiation in rat hepatocytes: reciprocal expression of C/EBP alpha and immediate-early growth response transcription factors. *Mol Cell Biol* 14(9), 5858–69. (1994)
- 27.Wu XZ, Chen D, Xie GR. Extracellular matrix remodeling in hepatocellular carcinoma: effects of soil on seed? *Med Hypotheses* 66(6), 1115–20. (2006)

- 28. Fujii M, Yamashita Y, Shirabe K, Ijima H, Nakazawa K, Funatsu K, Sugimachi K. Basic study about the development of the hybrid-artificial liver support system using human hepatoma cell lines (Hep G2, Huh 7): effects on liver functions by extracellular matrix (type I collagen) in monolayer culture. Fukuoka Igaku Zasshi 92(8), 299-305. (2001)
- 29.Zhao M, Laissue JA, Zimmermann A. Tenascin and type IV collagen expression in liver cell dysplasia and in hepatocellular carcinoma. *Histol Histopathol* 11(20), 323–33. (1996)
- 30.Kudriavtseva EI, Morozova OV, Rudinskaia TD, Engel'gard NV. Disruption of intercellular contacts and cell-extracellular matrix interaction in rapidly growing murine hepatocarcinoma. *Arkh Patol* 63(4), 33–7. (2001)
- 31.McCaughan GW, Siah CL, Abbott C, Wickson J, Ballesteros M, Bishop GA. Dipeptidyl peptidase IV is down-regulated in rat hepatoma cells at the mRNA level. *J Gastroenterol Hepatol* 8(2), 142–5. (1993)
- 32.Caron JM. Induction of albumin gene transcription in hepatocytes by extracellular matrix proteins. *Mol Cell Biol* 10(3), 1239–43. (1990)
- 33. Jaskiewicz K, Chasen MR, Robson SC. Differential expression of extracellular matrix proteins and integrins in hepatocellular carcinoma and chronic liver disease. *Anticancer Res* 13(6A), 2229–37. (1993)
- 34.Seebacher T, Medina JL, Bade EG. Laminin alpha 5, a major transcript of normal and malignant rat liver epithelial cells, is differentially expressed in developing and adult liver. Exp Cell Res 237(1), 70–6. (1997)
- 35.Torimura T, Ueno T, Inuzuka S, Kin M, Ohira H, Kimura Y, Majima Y, Sata M, Abe H, Tanikawa K. The extracellular matrix in hepatocellular carcinoma shows different localization patterns depending on the differentiation and the histological pattern of tumors: immunohistochemical analysis. *J Hepatol* 21(1), 37–46. (1994)
- 36.Yao M, Zhou XD, Zha XL, Shi DR, Fu J, He JY, Lu HF, Tang ZY. Expression of the integrin alpha5 subunit and its mediated cell adhesion in hepatocellular carcinoma. *J Cancer Res Clin Oncol* 123(8), 435–40. (1997)
- 37. Coussens LM, Werb Z. Inflammation and cancer. *Nature* 420(6917), 860–7. (2002)
- 38.Beachy PA, Karhadkar SS, Berman DM. Tissue repair and stem cell renewal in carcinogenesis. *Nature* 432(7015), 324–31. (2004)
- 39.Wu XZ, Chen D. Helicobacter pylori and hepatocellular carcinoma: correlated or uncorrelated? *J Gastrointerol Hepatol* 21(2), 345–7. (2006)
- 40.Zhang GQ, Fang CH, Gao P, Yan Z, Zheng Q, Chen GH. Study of mesenchymal stem cells transfected with

- oncogenes differentiate into hepatocellular carcinoma of rats. Zhonghua Wai Ke Za Zhi 45(9), 605-8. (2007)
- 41.Zhou J, Xiang H, Zhu Z, Lo H, Luo Y, Wang P, Li Y, Yin C. Efect of paracrine substance of rat bone marrow mesenchymal stem cells on the proliferation Of CBRH-7919 hepatoma cells. *Journal of Tianjin Medical University* 17(4), 455-458. (2011)
- 42.Chen ML, Lee KD, Huang HC, Tsai YL, Wu YC, Kuo TM, Hu CP, Chang C. HNF- 4α determines hepatic differentiation of human mesenchymal stem cells from bone marrow. World J *Gastroenterol* 16(40), 5092-103. (2010)
- 43.Djouad F, Plence P, Bony C, Tropel P, Apparailly F, Sany J, Noël D, Jorgensen C. Immunosuppressive effect of mesenchymal stem cells favors tumor growth in allogeneic animals. *Blood*.102(10), 3837-44. (2003)
- 44.Sun B, Zhang S, Ni C, Zhang D, Liu Y, Zhang W, Zhao X, Zhao C, Shi M. Correlation between melanoma angiogenesis and the mesenchymal stem cells and endothelial progenitor cells derived from bone marrow. *Stem Cells Dev* 14(3), 292-8. (2005)
- 45.Zhu W, Xu W, Jiang R, Qian H, Chen M, Hu J, Cao W, Han C, Chen Y. Mesenchymal stem cells derived from bone marrow favor tumor cell growth *in vivo*. *Exp Mol Pathol* 80(3), 267-74. (2006)
- 46.Annabi B, Naud E, Lee YT, Eliopoulos N, Galipeau J. Vascular progenitors derived from murine bone marrow stromal cells are regulated by fibroblast growth factor and are avidly recruited by vascularizing tumors. *J Cell Biochem* 91(6), 1146-58. (2004)
- 47.Hung SC, Deng WP, Yang WK, Liu RS, Lee CC, Su TC, Lin RJ, Yang DM, Chang CW, Chen WH, Wei HJ, Gelovani JG. Mesenchymal stem cell targeting of microscopic tumors and tumor stroma development monitored by noninvasive *in vivo* positron emission tomography imaging. *Clin Cancer Res* 11(21), 7749-56. (2005)
- 48.Djouad F, Bony C, Apparailly F, Louis-Plence P, Jorgensen C, Noel D. Earlier onset of syngeneic tumors in the presence of mesenchymal stem cells. *Transplantation* 82(8), 1060–6. (2006)
- 49.Djouad F, Plence P, Bony C, Tropel P, Apparailly F, Sany J, Noel D, Jorgensen C. Immunosuppressive effect of mesenchymal stem cells favors tumor growth in allogeneic animals. *Blood* 102(10), 3837–44. (2003)
- 50.Nakamizo A, Marini F, Amano T, Khan A, Studeny M, Gumin J, Chen J, Hentschel S, Vecil G, Dembinski J, Andreeff M, Lang FF. Human bone marrow-derived mesenchymal stem cells in the treatment of gliomas. *Cancer Res* 65(8), 3307–18. (2005)
- 51.Nakamura K, Ito Y, Kawano Y, Kurozumi K, Kobune M, Tsuda H, Bizen A, Honmou O, Niitsu Y, Hamada H. Antitumor effect of genetically engineered mesenchymal

- stem cells in a rat glioma model. Gene Ther 11(14) 1155-64. (2004)
- 52.Hall B, Dembinski J, Sasser AK, Studeny M, Andreeff M, Marini F. Mesenchymal stem cells in cancer: tumorassociated fibroblasts and cell-based delivery vehicles. *Int J Hematol* 86(1), 8–16. (2007)
- 53.Studeny M, Marini FC, Champlin RE, Zompetta C, Fidler IJ, Andreeff M. Bone marrow-derived mesenchymal stem cells as vehicles for interferon-beta delivery into tumors. *Cancer Res* 62(13), 3603–8. (2002)
- 54.Studeny M, Marini FC, Dembinski JL, Zompetta C, Cabreira-Hansen M, Bekele BN, Champlin RE, Andreeff M. Mesenchymal stem cells: potential precursors for tumor stroma and targeted-delivery vehicles for anticancer agents. *J Natl Cancer Inst* 96(21), 1593–603. (2004)
- 55.Klopp AH, Spaeth EL, Dembinski JL, Woodward WA, Munshi A, Meyn RE, Cox JD, Andreeff M, Marini FC. Tumor irradiation increases the recruitment of circulating mesenchymal stem cells into the tumor microenvironment. *Cancer Res* 67(24), 11687–95. (2007)
- 56.Zhu W, Xu W, Jiang R, Qian H, Chen M, Hu J, Cao W, Han C, Chen Y. Mesenchymal stem cells derived from bone marrow favor tumor cell growth *in vivo*. *Exp Mol Pathol* 80(3), 267-74. (2006)
- 57.Roorda BD, ter Elst A, Kamps WA, de Bont ES. Bone marrowderived cells and tumor growth: contribution of bone marrowderived cells to tumor micro-environments with special focus on mesenchymal stem cells. *Crit Rev Oncol Hematol* 69(3), 187-98. (2009)
- 58.Leek RD, Harris AL, Lewis CE. Cytokine networks in solid human tumors--regulation of angiogenesis. *J Leukoc Biol* 56(4), 423-35. (1994)
- 59.Budhu A, Wang XW. The role of cytokines in hepatocellular carcinoma. *J Leukoc Biol* 80(6), 1197-213. (2006)
- 60.Shao ZH, Wang PJ, Li MH, Zhang W, Zheng SQ, Zhao XH, Wang GL, Shang MY, Mao XQ. Effects of tmsenehymal stem cell transplantation on growth of liver cancer:experiment with rats. *Zhonghua Yi Xue Za Zhi* 89(7), 491-6. (2009)
- 61.Yu BY, Hu SW, Sun YM, Lee YT, Young TH. Modulating the activities of human mesenchymal stem cells (hMSCs) and C3A/HepG2 hepatoma cells by modifying the surface characteristics of poly(3-hydroxybutyrate-co-3-hydroxyhexnoate) (PHBHHx). *J Biomater Sci Polym Ed* 2009;20(9):1275-93.
- 62. Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, Richardson AL, Polyak K, Tubo R, Weinberg RA. Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature* 449(7162), 557-63. (2007)

- 63.Shinagawa K, Kitadai Y, Tanaka M, Sumida T, Kodama M, Higashi Y, Tanaka S, Yasui W, Chayama K. Mesenchymal stem cells enhance growth and metastasis of colon cancer. *Int J Cancer* 127(10), 2323-33. (2010)
- 64.Aurich H, Sgodda M, Kaltwasser P, Vetter M, Weise A, Liehr T, Brulport M, Hengstler JG, Dollinger MM, Fleig WE, Christ B. Hepatocyte differentiation of mesenchymal stem cells from human adipose tissue *in vitro* promotes hepatic integration *in vivo*. *Gut* 58(4), 570–581. (2009)
- 65.Banas A, Teratani T, Yamamoto Y, Tokuhara M, Takeshita F, Quinn G, Okochi H, Ochiya T. Adipose tissue-derived mesen- chymal stem cells as a source of human hepatocytes. *Hepatology* 46(1), 219–228. (2007)
- 66.Pan RL, Chen Y, Xiang LX, Shao JZ, Dong XJ, Zhang GR. Fetal liver- conditioned medium induces hepatic specification from mouse bone marrow mesenchymal stromal cells: a novel strategy for hepatic trans- differentiation. *Cytotherapy* 10(7), 668–675. (2008)
- 67.Stock P, Staege MS, Müller LP, Sgodda M, Völker A, Volkmer I, Lützkendorf J, Christ B. Hepatocytes derived from adult stem cells. *Transplant Proc* 40(2), 620–623. (2008)
- 68.Bruno S, Grange C, Deregibus MC, Calogero RA, Saviozzi S, Collino F, Morando L, Busca A, Falda M, Bussolati B, Tetta C, Camussi G. Mesenchymal stem cell-derived microvesicles protect against acute tubular injury. *Am Soc Nephrol* 20(5), 1053-67. (2009)
- 69.Du J, Zhou L, Chen X, Yan S, Ke M, Lu X, Wang Z, Yu W, Xiang AP. IFN-γ-primed human bone marrow mesenchymal stem cells induced tumor cell apoptosis *in vitro* via tumor necrosis factor-related apoptosis-inducing ligand. *Int J Biochem Cell Biol* 44(8), 1305-14. (2012)
- 70.Studeny M, Marini FC, Champlin RE, Zompetta C, Fidler IJ, Andreeff M. Bone marrow-derived mesenchymal stem cells as vehicles for interferon-beta delivery into tumors. *Cancer Res* 62(13), 3603-8. (2002)
- 71.Studeny M, Marini FC, Dembinski JL, Zompetta C, Cabreira-Hansen M, Bekele BN, Champlin RE, Andreeff M. Mesenchymal stem cells: potential precursors for tumor stroma and targeted-delivery vehicles for anticancer agents. *J Natl Cancer Inst* 96(21), 1593-603. (2004)
- 72.Digirolamo CM, Stokes D, Colter D, Phinney DG, Class R, Prockop DJ. Propagation and senescence of human marrow stromal cells in culture: a simple colony-forming assay identifies samples with the greatest potential to propagate and differentiate. *Br J Haematol* 107(2), 275–81. (1999)
- 73.Colter DC, Class R, DiGirolamo CM, Prockop DJ. Rapid expansion of recycling stem cells in cultures of plastic-adherent cells from human bone marrow. *Proc Natl Acad Sci U S A* 97(7), 3213–8. (2000)
- 74.Jiang H, Ren J. Inoculation of murine bone marrow mesenchymal stem cells induces tumor necrosis in mouse

- with orthotopic hepatocellular carcinoma. *Beijing Da Xue Xue Bao* 40(5), 453-8. (2008)
- 75. Abdel aziz MT, El Asmar MF, Atta HM, Mahfouz S, Fouad HH, Roshdy NK, Rashed LA, Sabry D, Hassouna AA, Taha FM. Efficacy of mesenchymal stem cells in suppression of hepatocarcinorigenesis in rats: possible role of Wnt signaling. *J Exp Clin Cancer Res* 30, 49. (2011)
- 76. Cheng S, Wang P, Li M, Zhang W. Magnetic labeling bone mesenchymal stem cells exert potent antitumorigenic effects in a model of hepatocellular carcinoma. *Journal of Clinical Radiology* 28(10), 1454-1458. (2009)
- 77.Qiao L, Zhao TJ, Shan CL, Ye LH, Zhang XD. Gene expression analyses of human mesenchymal stem cell mediated inhibition on proliferation of hepatoma cells. *Chinese Journal Of Biochemistry And Molecular Biology* 23(12), 1037-1044. (2007)
- 78.Lu YR, Yuan Y, Wang XJ, Wei LL, Chen YN, Cong C, Li SF, Long D, Tan WD, Mao YQ, Zhang J, Li YP, Cheng JQ. The growth inhibitory effect of mesenchymal stem cells on tumor cells *in vitro* and *in vivo*. *Cancer Biol Ther* 7(2), 245-51. (2008)
- 79.Kidd S, Spaeth E, Dembinski JL, Dietrich M, Watson K, Klopp A, Battula VL, Weil M, Andreeff M, Marini FC. Direct evidence of mesenchymal stem cells tropism for tumor and wounding microenvironments using *in vivo* bioluminescent imaging. *Stem Cells* 27(10), 2614-23. (2009)
- 80.Kucerova L, Altanerova V, Matuskova M, Tyciakova S, Altaner C. Adipose tissuederived human mesenchymal stem cells mediated prodrug cancer gene therapy. *Cancer Research* 67(13), 6304-13. (2007)
- 81.Knoop K, Kolokythas M, Klutz K, Willhauck MJ, Wunderlich N, Draganovici D, Zach C, Gildehaus FJ, Böning G, Göke B, Wagner E, Nelson PJ, Spitzweg C. Image-guided, tumor stroma-targeted 131I therapy of hepatocellular cancer after systemic mesenchymal stem cell-mediated NIS gene delivery. *Mol Ther* 19(9), 1704-13. (2011)
- 82. Niess H, Bao Q, Conrad C, Zischek C, Notohamiprodjo M, Schwab F, Schwarz B, Huss R, Jauch KW, Nelson PJ, Bruns CJ. Selective targeting of genetically engineered mesenchymal stem cells to tumor stroma microenvironments using tissue-specific suicide gene expression suppresses growth of hepatocellular carcinoma. *Ann Surg* 254(5), 767-74, discussion 774-5. (2011)
- 83.Fürst G, Schulte am Esch J, Poll LW, Hosch SB, Fritz LB, Klein M, Godehardt E, Krieg A, Wecker B, Stoldt V, Stockschläder M, Eisenberger CF, Mödder U, Knoefel WT. Portal Vein Embolization and Autologous CD133+ Bone Marrow Stem Cells for Liver Regeneration: Initial Experience. *Radiology* 243(1), 171-9. (2007)
- 84.Livraghi T, Meloni F, Frosi A, Lazzaroni S, Bizzarri TM, Frati L, Biava PM. Treatment with stem cell differentiation stage factors of intermediate-advanced

Bmscs in hepatocellular carcinoma

hepatocellular carcinoma: an open randomized clinical trial. *Oncol Res* 15(7-8), 399-408. (2005)

85.Peng L, Xie DY, Lin BL, Liu J, Zhu HP, Xie C, Zheng YB, Gao ZL. Autologous bone marrow mesenchymal stem cell transplantation in liver failure patients caused by hepatitis B: short-term and long-term outcomes. *Hepatology* 54(3), 820–8. (2011)

86.Ismail A, AlDorry A, Shaker M, Elwekeel R, Mokbel K, Zakaria D, Meshaal A, Eldeen FZ, Selim A. Simultaneous injection of autologous mononuclear cells with TACE in HCC patients; preliminary study. *J Gastrointest Cancer* 42(1), 11-9. (2011)

87. Huang XB, Wang XW, Li J, Zheng L, Zhao HZ, Liang P, Dai JG. Are autologous bone marrow stem cell transplantation and transcatheter arterial embolization the best choices for patients with hepatocellular carcinoma and hepatic dysfunction? Report of a case. *Surg Today* (Epub ahead of print). (2011)

Abbreviations: BMSCs: Bone marrow mesenchymal stem cells; HCC: hepatocellular carcinoma; ECM: extracellular matrix; VEGF: vascular endothelial cell growth factor; PCNA: proliferating cell nuclear antigen; PHBHHx: poly (3-hydroxybutyrate-co-3-hydroxyhexnoate); MRI: magnetic resonance imaging; MSCs: mesenchymal stem cells; PVE: portal vein embolization; HBV: hepatitis B virus; TACE: transcatheter arterial chemoembolization

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Send correspondence to: Peng Gong, No.222 Zhongshan Road, Department of Hepatobiliary Surgery, the First Affiliated Hospital of Dalian Medical University, Dalian, China,116011, Tel: 86 15541198928, Fax: 86 0411-83622844, E-mail: doctorgongpeng@yeah.net