

Associations between adipokines and obesity-related cancer

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1. ABSTRACT

There is increasing evidence that obesity may have pathophysiological effects that extend beyond its well-known co-morbidities; in particular its role in cancer has received considerable epidemiological support. As adipose tissue becomes strongly established as an endocrine organ, two of its most abundant and most investigated adipokines, leptin and adiponectin, are also taken beyond their traditional roles in energy homeostasis, and are implicated as mediators of the effects of obesity on cancer development. This review examines these adipokines in relation to the prostate, breast, colorectal, thyroid, renal, pancreatic, endometrial and oesophageal cancers, and how they may orchestrate the influence of obesity on the development of these malignancies.

2. INTRODUCTION

Obesity has reached epidemic proportions worldwide. Recent data from different countries show that only one third of the population is normal weight, and approximately one third is obese. Obesity leads to several co-morbidities, such as diabetes, dyslipidaemia, hypertension, sleep apnoea, osteoarthritis, menstrual disorders, infertility, gout, stroke, ischaemic heart disease, congestive heart failure, deep vein thrombosis and pulmonary embolism (1).

Increased risk of cancer is another co-morbidity that is related to obesity. Since 1979, several epidemiological studies and meta-analyses have consistently demonstrated that an increased BMI (body

Table 1. Cancers strongly (relative risk greater than 1.2) associated with an increase in BMI of 5 kg/m²

Cancer	RR (95% CI) for Men	RR (95% CI) for Women
Oesophageal adenocarcinoma	1.52 (1.33–1.74)	1.51 (1.31–1.74)
Thyroid	1.33 (1.04–1.70)	1.14 (1.06–1.23)
Colon	1.24 (1.20–1.28)	1.09 (1.05–1.13) ¹
Renal	1.24 (1.15–1.34)	1.34 (1.25–1.43)
Endometrium	NA	1.59 (1.50–1.68)
Gallbladder	1.09 (0.99–1.21) ¹	1.59 (1.02–2.47)

¹ Cancer with RR less than 1.2. All P less than 0.05, except for gallbladder cancer in men. NA: not applicable. Adapted from (3)

mass index) is associated with several types of cancer, such as prostate cancer in men, breast cancer in women, and oesophageal adenocarcinoma in both sexes.

Given the strong epidemiological evidence linking obesity and cancer, the identification of its pathophysiological basis is made necessary, to allow the development of preventive and even therapeutic strategies. Obesity presents with several hormonal changes, such as increased oestrogens, insulin and insulin-like growth factors. In addition, alterations in the immune response, in the nuclear factor kappa B (NF-kappa B) system, in oxidative stress, and in peroxidation are common in obesity. Finally, hypertension, acid reflux and increased iodine uptake are also considered other alterations that may predispose obese individuals to specific types of cancer (2). It is out of the scope of this manuscript to review all the pathophysiological bases of obesity and cancer. Here, based on the literature identified through searches on the PubMed database, we will address the roles of adipokines – polypeptides produced mainly by the adipose tissue – in the development of cancer types associated with obesity that were listed in a recent meta-analysis (3), plus prostate (the most common cancer in men), breast (the most common cancer in women) and pancreatic (the cancer with the lowest 5-year survival rate) cancers. Since no evidence between adipokines and gallbladder cancer has been described in the literature, this type of cancer will not be discussed in this review.

3. OBESITY AND CANCER: EPIDEMIOLOGICAL EVIDENCE

The first and largest epidemiological study investigating the associations between weight and cancer was published in 1979. This was a long-term prospective study involving 750,000 men and women evaluated between 1959 and 1972, showing that mortality was approximately 90% higher in men and women who were more than 40% heavier than the average. In that group, cancer mortality was also increased by a third among men, due to colon and rectum cancers, and by 55% among women, due to cancers of the gallbladder, biliary passages, breast, cervix, endometrial, and ovary (4).

In 2008, the World Cancer Research Fund issued an Expert Report acknowledging the association of excess body fat with increased risk of oesophageal adenocarcinoma and cancers of the pancreas, colon,

rectum, post-menopausal breast, endometrium and kidney (5). Subsequently, a very large epidemiological study, the Million Women Study, identified associations between high body mass index (BMI) and increased risk of endometrial cancer (2.39), oesophageal adenocarcinoma (2.38), pre-menopausal colorectal cancer (1.61), kidney cancer (1.53), leukaemia (1.50), post-menopausal breast cancer (1.40), multiple myeloma (1.31), pancreatic cancer (1.24), non-Hodgkin's lymphoma (1.17), and ovarian cancer (1.14). The risk for all cancers combined was 1.12 (6).

The first meta-analysis published in 2001 demonstrated that nearly 35,000 and 37,000 new cases of cancer in Europe were related to obesity and to overweight, respectively. The cancers that were most strongly associated with overweight and obesity were endometrium, kidney, gallbladder, colon and breast (7). Several other meta-analyses have been published, associating obesity or high BMI with increased risk for several types of cancer: colon (both sexes) and rectal (men) (8–11), gallbladder (12), liver (13, 14), kidney (15, 16), pancreatic (17, 18), ovarian (19, 20), breast (21) and prostate cancers (22); leukaemia, non-Hodgkin's lymphoma and multiple myeloma (23, 24).

A more recent meta-analysis of 141 articles confirmed strong associations between higher BMI and increased risk for oesophageal adenocarcinoma, and for thyroid, colon, and renal cancers in men; and for endometrial, gallbladder, oesophageal adenocarcinoma and renal cancers in women (Table 1). That meta-analysis found additional weaker, but still positive, associations (relative risk less than 1.20) between increased BMI and rectal cancer and malignant melanoma (men); postmenopausal breast, pancreatic, thyroid, and colon cancers (women); and leukaemia, multiple myeloma, and non-Hodgkin lymphoma (both sexes) (3). Of particular interest, the same meta-analysis found that increased BMI was negatively associated with lung cancer and with oesophageal squamous cancer, both in men and in women. However, most of the lung cancer patients are smokers and tend to have lower BMIs than non-smokers. When non-smokers are analysed separately, that negative association ceases to exist (3, 25). Another meta-analysis found associations between high BMI and all 20 cancer types that were analysed, except oesophageal and prostate cancer (26).

4. ADIPOKINES – AN OVERVIEW

Over the last decade, there has been a paradigm shift from the concept that adipose tissue is a mere storage site for energy (27). It is now known that adipose tissue is a metabolically active organ that plays active roles in energy homeostasis, immunity, endocrine balance and bone remodelling (28). Adipose tissue is responsible for the synthesis and secretion of several polypeptide growth factors and cytokines, known as adipokines or adipocytokines. They are produced exclusively, or substantially, by white adipose tissue preadipocytes and mature adipocytes. The list of adipokines grows by the day,

with more than 50 molecules being listed to date (29). The most abundant and well-known ones are leptin – the first described adipokine, produced mainly by the adipose tissue, and adiponectin – produced exclusively by it.

Adipokines play an important role in the pathophysiology of cancer in obesity. The list of adipocytokines is extensive, and we will discuss the roles of the two most abundant adipokines in cancer: leptin and adiponectin.

5. LEPTIN

Leptin is a 16-kDa adipokine that is produced mainly, but not exclusively, by white adipose tissue. Others sites of production include the placenta, intestine, stomach, ovaries, bone marrow, brain, pituitary, liver, mammary epithelial cells and skeletal muscle. Leptin levels are positively correlated with white adipose tissue mass, and are therefore increased in obesity. Its synthesis is influenced by insulin, tumour necrosis factor alpha (TNF- α), glucocorticoids, sex hormones and prostaglandins. Its expression is also stimulated by hypoxia (commonly found in solid tumours), through the hypoxia-induced factor-1 (HIF-1) (30).

The main role of leptin is to regulate energy homeostasis by controlling energy intake and energy expenditure, through its action on the arcuate nucleus of the hypothalamus. It has additional effects in the endocrine and immune systems, including reproduction, glucose homeostasis, bone formation, tissue remodelling, and inflammation (31, 32). Leptin binds to its receptor and activates different signalling pathways, such as the JAK/STAT (Janus Kinase/Signal Transducer and Activator of Transcription), MAPK (mitogen-activated protein kinase), PI3K/Akt (phosphatidylinositol 3-kinase/protein-kinase B), AMPK (5' AMP-activated protein kinase) and IRS (insulin receptor substrate) pathways, which affect cell proliferation and survival. For a review on the intracellular signalling pathways activated by leptin, see Fruhbeck (33).

Congenital leptin deficiency, which is observed in rare cases of mutations in the leptin gene, is associated with hyperphagia, impaired thermogenesis, immune defects, insulin resistance, dyslipidaemia, lipotoxicity, hypogonadotropic hypogonadism, functional and structural alterations in the brain, and impairment of cognitive development, all reversible by leptin treatment (34-40). Low levels of leptin are also observed in patients with lipodystrophic syndromes and in anorexia nervosa. Leptin is a pleiotropic hormone, being mitogenic, anti-apoptotic, pro-angiogenic, and pro-inflammatory in various cellular systems. Being the most abundant adipokine and increased in obesity, its role in the pathogenesis of cancer has been extensively investigated.

Leptin stimulates growth, migration and invasion in tumour cell models, which are relevant in the pathogenesis of cancer (30, 41). Leptin also increases the production of cytokines by macrophages (such as IL-6, IL-12, and TNF- α), stimulating cancer cells (42). By

reducing tissue sensitivity to insulin, leptin is responsible for hyperinsulinaemia, which also stimulates cell growth. Leptin appears to be involved in angiogenesis as well (43). Leptin also plays a role in hormone-dependent neoplasms, such as in the cancer of the endometrium and in breast cancer, by activating aromatase (44), an enzyme catalysing the transformation of androstenedione to oestrone.

5.1. Prostate cancer

Epidemiological studies found contradictory results associating leptin with prostate cancer. A recent review summarises those studies, with 3 out of 9 studies showing positive associations between blood leptin levels and prostate cancer (45). In the Physicians' Health Study, a 25-year prospective study, no associations were found between leptin and prostate cancer risk and mortality (46). A genetic study demonstrated that the leptin polymorphism (-2548 G/A), leading to higher leptin levels, is associated with susceptibility to prostate cancer and risk of advanced disease (47). It is suggested that, regardless of these contradictory data, leptin may be associated with more advanced, hormone-refractory prostate cancer (48).

Leptin stimulates prostate growth and angiogenesis (43, 49, 50), and receptors for leptin are present in the prostate (51). *In vitro*, leptin stimulates growth of the androgen-insensitive, DU145 and PC3, prostate cancer cell lines but not androgen-sensitive cell lines, LNCaP (52, 53). Leptin has been reported to promote the proliferation and survival of DU145 and PC3 cell lines through the activation of the PI3K and the classical MAPK pathways; additionally, leptin mediates the growth effect through the JNK (c-Jun N-terminal kinase) MAP kinase pathway (52). Both the full-length and short forms of the leptin receptor have been implicated in these responses, and downstream targets of these pathways include c-jun, c-fos and other genes involved in cell proliferation.

Leptin can also promote neoangiogenesis in prostate cancer: the leptin gene can be induced under conditions of hypoxia, which often prevail inside solid tumours; it may have a role in vascular remodelling both on its own and through the induction and activation of a number of other pro-angiogenic factors, including vascular endothelial growth factor (VEGF), fibroblast growth factor 2 (FGF 2) and the matrix metalloproteinases 2 and 9 (54).

5.2. Breast cancer

The majority of breast cancers require action of oestrogens for their growth and progression. In addition, leptin in excess may also contribute to the pathogenesis of breast cancer. Vona-Davies and Rose summarises the contradictory results found in case-control studies (one of which was nested within a prospective study), with 3/10 showing positive correlations (55). More recently, another prospective observational study demonstrated that BMI and leptin were significantly correlated with pathological tumour classification and TNM stage in postmenopausal breast cancer patients (56). It seems that leptin may increase breast cancer risk in postmenopausal women specifically (56, 57), in which the only source of oestrogens is adipose tissue.

Leptin's effects on breast cancer can be explained by several mechanisms: 1) leptin is able to induce the growth of breast cancer cells through activation of the JAK/STAT3, MAPK-ERK1/2 (extracellular signal-regulated kinases 1/2), and/or PI3K (phosphoinositide 3-kinase) pathways; 2) leptin can mediate angiogenesis by inducing the expression of vascular endothelial growth factor (VEGF) (43); 3) leptin induces transactivation of human epidermal growth factor receptor 2 (ErbB-2), and interacts with insulin-like growth factor 1 (IGF-1) in triple negative breast cancer cells, transactivating the epidermal growth factor receptor (EGFR) and promoting invasion and migration; 4) leptin stimulates aromatase expression, increasing oestrogen levels and affecting the growth of oestrogen receptor (ER)-positive breast cancer cells; 5) leptin induces MAPK-dependent activation of ER (53, 58, 59); 6) leptin stimulates proteolytic cleavage of intercellular matrix, promoting cancer cell invasion (60).

In addition, leptin and its receptor are significantly overexpressed in human primary and metastatic breast cancer, being most abundant in less differentiated tumours (61). In breast cancer cells, the overexpression of leptin is associated with the leptin promoter polymorphism Lep-2548G/A (62). The subtype of leptin receptor seems to make a difference in the prognosis, since patients with elevated Ob-Ra expression have longer relapse-free survival, as compared to patients with high Ob-Rb/Ob-Ra ratio (63). The implication of leptin as a growth factor to breast cancer is further strengthened by the fact that leptin-deficient *ob/ob* (64) and leptin-resistant *db/db* mice (65) do not develop transgene-induced mammary tumours.

5.3. Colorectal cancer

Leptin levels were positively correlated with colorectal cancer in some prospective studies (66-68), but not in others (69, 70). It is questionable whether leptin is a cause or a mere bystander, since leptin is not as strongly associated with colorectal cancer in women, who have much higher leptin than men. Paradoxically, some studies observed significantly lower serum leptin concentration in patients with colorectal cancer, independent of BMI and weight loss (71, 72). In less differentiated colorectal cancers, the leptin expression is decreased (73), and the expression level may be positively correlated with a better prognosis (74).

Leptin receptors are expressed in human colon cancer cell lines and in human colon tumours, polyps and adjacent mucosa (75). In addition, leptin is overexpressed in primary colorectal cancers, which is significantly correlated with tumour grade and the presence of adenocarcinoma (76). Leptin stimulates growth of colon cancer cells via MAPK pathway (ERK 1/2 and JNK), JAK/STAT3 and PI3K/Akt, and reduces cell apoptosis (77-79). The pro-angiogenic effect of leptin plays an additional role on the pathogenesis of colorectal cancers (43). Moreover, leptin induces IL-6 production by Apc^{Min/+} colon epithelial cells which leads to the growth and proliferation of preneoplastic cells (80). However, *in vivo* studies are contradictory. In Apc^{Min/+}, *ob/ob* and *db/db*

mice, leptin supplementation did not affect tumorigenesis (81, 82). In a high-fat 1,2-dimethylhydrazine (a potent carcinogen)-treated rat model, growth of colonic cancer cells was enhanced by leptin (83), showing that leptin's effects may be synergistic to other environmental factors.

5.4. Thyroid cancer

There is evidence suggesting that obesity and increased BMI as positively correlated with thyroid cancer (84-86), but there are very few studies evaluating the association between leptin and thyroid cancer. In a small Turkish study, patients with papillary thyroid carcinoma had higher leptin levels as compared to controls (87).

Leptin receptors are expressed in the normal thyroid, and leptin treatment increases growth and secretion in the gland (88). In addition, leptin and leptin receptor are expressed in papillary thyroid cancer, and this expression is associated with aggressiveness (89, 90). All thyroid cancer cell lines – anaplastic (ARO), follicular (WRO) and papillary (CGTH-W3) – express long-form leptin receptors, but leptin stimulation does not alter the expression of the sodium-iodide symporter, cell growth or cell cycle. Leptin, however, promotes cell migration of papillary thyroid cancer cells and inhibits migration of anaplastic and follicular cancer cells (91). In a contradictory *in vitro* study, leptin stimulated cell proliferation and inhibited apoptosis of papillary carcinoma cells, via activation of PI3K/Akt (90).

5.5. Renal cancer

Studies associating leptin and renal cancer are scarce. In a case-control study that included 70 patients with renal cell carcinoma, leptin was inversely associated with cancer risk (OR: 0.53, CI: 0.28-0.99, *p*=0.05), which the authors attribute to leptin's pro-immunogenic effects (92). Similarly, higher serum leptin was an independent predictor of progression-free survival, but was also associated with tumour specimen venous invasion (93).

Leptin receptor is present in Caki-1, ACHN, 769P, A498, SKRC44 and SKRC49, and in the murine renal cancer cell line Renca. In the murine cell, leptin induces invasiveness (94). In another *in vitro* study, leptin increased the proliferation and mobility capabilities of Caki renal carcinoma cells by up-regulating the expression of the JAK/STAT3 and ERK1/2 signalling pathways (95). Leptin also induces collagen gel invasion of non-tumorigenic kidney MDCK epithelial cells through PI3K-, Rho-, and Rac-dependent signalling pathways (75). Leptin's effects on lymphangiogenesis, mediated by Akt and ERK1/2, and on lipid and protein biosynthesis, mediated by acyl-coenzyme A:cholesterol acyl transferase (ACAT), may explain the roles of leptin in the pathogenesis and in the phenotype of renal cancer (96). However, there is contradiction between the epidemiological and the molecular findings regarding leptin's role on kidney cancer.

5.6. Endometrial cancer

A few studies showed that leptin levels were correlated with the presence of endometrial cancer, but correlation disappeared when adjusted by BMI, suggesting

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that leptin may be a bystander and not the cause of endometrial cancer (97-99).

Leptin receptors are present in the normal and malignant human endometrium (more intensely at early secretory phases), and are inhibited by treatment with progesterone (100). Specifically in malignant tissues, the expression of leptin receptors is increased through the stimulatory effect of hypoxia-inducible factor 1 alpha (which is in turn induced by solid tumour-associated hypoxia) (101). In malignant tissues, lower expression levels of leptin receptor short form (Ob-Ra, which activates JAK2, IRS-1, and MAPK) is observed, suggesting that aberrant leptin receptor isoforms may be involved in the pathogenesis of endometrial cancer (98). In addition, higher-grade and less differentiated tumours have lower expression levels of leptin receptor (102). In ECC1 and Ishikawa endometrial adenocarcinoma cells, leptin promotes endometrial cancer growth and invasiveness through MAPK and Akt pathways, and also induces the expression of cyclooxygenase-2 (COX-2), which increases tumorigenesis, increases metastasis potential and promotes angiogenesis (103). Leptin also enhances cyclin D1 expression (a cell cycle regulator required for completion of the G1/S transition) through JAK/STAT, MAPK and PKA (protein kinase A) activation, leading to human endometrial cancer proliferation (104). Additionally, leptin's effects on the pathogenesis of endometrial adenocarcinoma are attributed to the increase of several angiogenic factors (105).

5.7. Pancreatic cancer

One study showed that leptin levels were lower in patients with pancreatic cancer, compared to the normal population. In the same study, patients with chronic pancreatitis also had lower leptin, and patients with autoimmune pancreatitis had high leptin levels (106). Similarly, another case-control study showed that pancreatic adenocarcinoma was associated with low leptin levels, but the causality remained unclear (107). Hypoleptinaemia may be just a consequence of weight loss, observed in many pancreatic cancer patients (108). If in fact hypoleptinaemia increases the risk of pancreatic cancer, it could be explained by the increase in insulin resistance, which is another risk factor for the disease (109).

Leptin inhibits growth of two human pancreatic cancer cell lines (PANC-1, Mia-PaCa), possibly through its proimmunogenic effects (53). Both in *ob/ob* and *db/db* mice, larger tumours, increased mortality and high of mice developing metastases were observed. This suggests that obesity, but not leptin, may not play a role in the pathogenesis of pancreatic cancer (110).

5.8. Oesophageal adenocarcinoma

There are no data showing an association between serum leptin levels and oesophageal cancer risk. In a case-control study, leptin was not associated with squamous cell carcinoma of the oesophagus (111). However, hyperleptinaemia has been shown to increase the risk of Barrett's oesophagus (BE), a precancerous lesion

(112). As in pancreatic cancer, hypoleptinaemia is observed in patients with weight loss and cachexia, independently of the presence of cancer (113).

Leptin receptors are overexpressed in oesophageal carcinoma cells (114, 115), and this overexpression indicates poorer prognosis. Leptin administration causes proliferation of human oesophageal squamous cancer cell line (KYSE410) with a mutant p53 gene, but not in a cell line without the mutant p53 (53). This effect seems to be mediated by the transactivation of the epidermal growth factor receptor, mediated by HB-EGF and transforming growth factor alpha (TGF-alpha) (115). Other studies confirmed leptin's anti-apoptotic and growth-promoting effects on oesophageal carcinoma cells (116), which are mediated by the activation of MAPK (through p38 and ERK), PI3K/Akt, JAK2 and EGFR (117, 118). In SEG-1 and BIC-1 cells lines, leptin has stimulatory actions on cell proliferation, but does not have anti-apoptotic effects (119).

6. ADIPONECTIN

Adiponectin is a 30-kDa peptide, present physiologically as trimers, hexamers or high-molecular-weight multimeric complexes (28). Adiponectin is expressed exclusively and in large quantities by adipocytes, making up approximately 0.01% of total human plasma protein content (120); it is negatively regulated by several other products of adipose tissue that become elevated during obesity, such as TNF-alpha, IL-6 and IL-18 (121). Adiponectin levels are negatively correlated with body fat and BMI. The two isoforms of the adiponectin receptor, AdipoR1 and R2, are most abundantly found in striated muscle and the liver, respectively, but are widely expressed in many types of normal and cancerous cells (122).

The main functions of adiponectin include anti-inflammation, anti-atherogenesis and insulin sensitisation. Circulating levels of adiponectin correlate inversely with inflammation states and insulin resistance (28), and a study by Ouchi *et al.* demonstrated that adiponectin modulates the inflammatory response in endothelial cells that is associated with coronary artery disease (123). Additionally, adiponectin has been identified as a likely angiogenic inhibitor that maintains the balance of quiescent vasculature present in adult adipose tissue (124).

However, as opposed to what is observed in the case of leptin, the role of adiponectin under physiological conditions appears to be redundant – neither its deletion nor its overexpression leads to any significant body weight differences in mice. Nonetheless, adiponectin has been shown to play an active role in regulating neovascularisation in tumours, suggesting that its effects may be more pronounced under pathological conditions, where there is more aggressive angiogenic activity than in slowly expanding adipose tissue (124).

6.1. Prostate cancer

Although the correlation between adiponectin levels and prostate cancer has not been consistently proven,

there is evidence to suggest that adiponectin is inversely related not only to the risk and incidence of prostate cancer, but also to the histological grade and disease stage (125, 126).

In vitro, adiponectin is a potent inhibitor of cell growth and proliferation in the metastatic prostate cancer cell lines, DU145, PC3 and LNCaP, through the downregulation of STAT3 signalling, which is constitutively active in androgen-independent DU145 cells. STAT3 increases the activity of the androgen receptors, leading to an amplification of the mitogenic signal; this forms the basis of hormone-refractory prostate cancer, and is crucial for the survival of DU145 cells (127).

Adiponectin also exerts an inhibitory effect on cell proliferation, regardless of androgen-dependence, through an AMPK-mediated pathway. Binding of adiponectin to its receptors activates AMPK, which regulates a diverse array of metabolic functions within the cell; notably, it phosphorylates and activates TSC2, which is an inhibitor of mTOR (mammalian target of rapamycin). mTOR and its downstream effectors, for example the PI3K/Akt pathway, have been implicated in several cancers, including prostate cancer, in which the phosphatase and tensin homolog (PTEN) gene is frequently inactivated (125).

6.2. Breast cancer

The association between adiponectin levels and breast cancer risk appears to be dependent on menopausal state. In postmenopausal women, there is evidence for a significant correlation between reduced adiponectin, as seen in obesity, and breast cancer incidence; however no such correlation was found in premenopausal women, in the same study (128).

Adiponectin inhibits breast cancer cell proliferation *in vitro*, in part by inhibiting the TNF-alpha-mediated NF-kappa B signalling pathway. TNF-alpha is a pro-inflammatory cytokine which promotes oestrogen biosynthesis; especially in obese, post-menopausal women, where adipose tissue is the main source of oestrogens (129), elevated oestrogen production stimulated by TNF-alpha supports the survival and growth of breast cancer cells. In the same way, adiponectin suppresses the induction of VEGF expression by TNF-alpha, disabling an important pro-angiogenic response (125). The lack of angiogenic support in a growing tumour can lead to considerable tumour cell apoptosis and reduction in tumour weights and volumes, as demonstrated in adiponectin-treated mice by Brakenhielm *et al.* (124).

Adiponectin inhibits the invasiveness of breast cancer cells by activating the AMPK pathway (130). In addition, adiponectin also stimulates the peroxisome proliferator-activated receptor (PPAR)-alpha pathway in breast cancer cells, inhibiting proliferation and promoting differentiation. Adiponectin-dependent activation of PPAR-gamma has also been shown to increase nuclear levels of BRCA1, a tumour suppressor protein involved in DNA repair, and therefore in reducing cancer risk (131).

The increased insulin resistance and hyperinsulinaemia due to a fall in adiponectin in obesity also has an impact on breast cancer. A number of studies support the role of insulin in stimulating proliferation in breast cancer cells, both through the insulin receptor signalling pathway and through interaction with oestrogens. Insulin may also serve to bolster the expression of VEGF. Therefore, the modulation of insulin levels is another indirect route through which adiponectin may protect against breast cancer.

6.3. Colorectal cancer

The association between colorectal cancer and adiponectin levels is still controversial (69, 132), but many studies have shown that low circulating adiponectin is correlated with an increased incidence of colorectal cancer (133-135), even after correction for BMI and insulin resistance.

Expression of the adiponectin receptors, AdipoR1 and R2, is elevated in colorectal cancer cells as compared to non-tumour colorectal tissue (136), giving support to the hypotheses that adiponectin causes colorectal cancer. On the other hand, adiponectin may indeed inhibit colorectal cancer cells, through the activation of the AMPK pathway, which inhibits mTOR, leading to anti-proliferative and pro-apoptotic effects of adiponectin (137). Furthermore, adiponectin can reduce lipogenic activity – which is essential to maintain membrane integrity in the rapidly dividing tumour cells – in colorectal cancer cells, by suppressing SREBP-1c activity. This in turn leads to a decrease in the expression of key lipogenic enzymes such as FAS (138).

Other roles of adiponectin, especially its interference with the proliferative and pro-inflammatory effects of leptin, TNF-alpha and IL-6, seem to operate independently of AMPK. As previously mentioned, adiponectin inhibits the activation of STAT3 by IL-6 and of the NF-kappa B pathway by TNF-alpha (134, 139). The study by Fenton *et al.* highlights the suppression by adiponectin of two separate leptin-mediated pathways. Adiponectin signals through the MAPK pathway to phosphorylate and activate I-kappa-K, an inhibitor of NF-kappa B. Additionally, adiponectin is able to suppress the pro-inflammatory IL-6 trans-signalling induced by leptin, by decreasing the availability of the soluble IL-6 receptor (sIL-6R) and increasing sgp130, the natural inhibitor of the IL6-sIL-6R complex (139).

6.4. Thyroid cancer

There are no studies evaluating the relationship between adiponectin and thyroid cancer. A follow-up study of patients with end-stage renal disease, who have a higher risk for developing cancer, showed that the most common site of cancer was the kidney (26.7%), followed by thyroid (13.3%) and stomach (13.3%). Lower adiponectin was an independent predictor of malignancy, but the associations between hypoadiponectinaemia and each type of cancer were not analysed (140). Further studies are necessary to evaluate whether hypoadiponectinaemia is associated with thyroid cancer.

6.5. Renal cancer

In general, renal cancer has a strong inverse association with serum adiponectin levels, as reported by a number of studies (141-143). Where the details of the disease are concerned, the correlation with adiponectin is less conclusive: while it is agreed that decreased adiponectin is related to the presence of metastasis in renal cell cancer, its association with tumour grade appears to be inconsistent. Evidence supports a role for adiponectin in renal cancer that is independent of BMI – in fact, Horiguchi *et al.* report a slight tendency for higher BMI to be associated with better clinicopathological features of the cancer, an observation may be due to cancer-induced cachexia, characterised by a preferential loss of skeletal muscle rather than adipose tissue (143). Spyridopoulos *et al.* suggest that an indicator of visceral adiposity, such as waist-to-hip ratio, may be a better parameter than BMI in gauging adiponectin's effects in renal cancer (142).

Both adiponectin receptors, especially AdipoR1, are known to be expressed in normal renal tissue as well as renal cancer tumour cells, and there are indications that the receptors may be downregulated in cancer tissue as compared to healthy tissue, reducing the protective potential of adiponectin in tumour cells (141). Few studies have investigated the specific molecular actions of adiponectin in renal cancer. Clear-cell renal cell carcinoma, the most common form of renal cancer, is well-known to be highly angiogenic, which suggests that adiponectin may have an important anti-angiogenic role in the development of the cancer (141). Low adiponectin levels are also implicated in the activation of several cancer-promoting factors, including STAT3, ERK 1/2 and Akt, whereas higher levels of adiponectin act to suppress them.

6.6. Endometrial cancer

The correlation between low adiponectin levels and endometrial cancer has been established independently of BMI (144, 145), although a combined decrease of adiponectin and obesity, as it often occurs, constitutes a greater risk (146). Significantly, this relationship is especially strong in premenopausal women, contrary to that of breast cancer risk (128, 145, 146). Possibly related to this is the finding that AdipoR1 and R2 are expressed in the human endometrium, and that their expression is increased during the midluteal period of the menstrual cycle.

The role that adiponectin plays in endometrial cancer is fairly similar to that which it plays in breast cancer. Both tissues respond to oestrogens, which promote cell proliferation; both tissues also express the pro-angiogenic factor VEGF. Adiponectin suppresses the biosynthesis of oestrogens and the expression of VEGF through its effects on the NF-kappa B signalling pathway (145). Excess insulin, such as in the case of insulin resistance, may augment the mitogenic effects of oestrogen, whereas adiponectin is an insulin sensitiser. The activation of AMPK is also an inhibiting effect of adiponectin on endometrial cancer (147).

A study by Cong *et al.* has expanded considerably the known mechanism of adiponectin through

the AMPK pathway, by observing that while adiponectin similarly induced cell cycle arrest and apoptosis in both investigated endometrial carcinoma cell lines, HEC-1-A and RL95-2, these effects occurred via different pathways: the protein kinase Akt is inactivated and cyclin D1 is downregulated in HEC-1-A cells, whereas in RL95-2 cells there was a reduction in cyclin E2 expression, as well as an inhibition of the p42/44 MAPK. These two cell lines differ genetically in that the RL95-2 line is deficient in the tumour suppressor PTEN, which regulates the PI3K signalling pathway; how this difference might explain the observations remains to be elucidated (148).

6.7 Pancreatic cancer

Evidence for the correlation between pancreatic cancer and serum adiponectin levels is conflicting. Adiponectin in pancreatic cancer patients has separately been observed to be elevated (149, 150), decreased (110, 151), and unchanged (106). An important caveat to consider in interpretation is that the results quoted above are not necessarily comparable in terms of study design and epidemiological relevance. Notably, case-control studies (149, 150) were liable to obtain evidence in support of increased adiponectin associated with pancreatic cancer, whereas the study on male Finnish smokers, which reported an inverse correlation, was of a prospective design (151).

This raises a host of possible explanations for the discrepancy, which tend to attribute the elevated adiponectin levels to reverse causation. Adiponectin has been tied to cancer cachexia, such that its increase may be due to weight-loss brought about by cancer progression (152); this is in agreement with the observation that adiponectin levels are increased in cases of anorexia nervosa or prolonged voluntary weight-loss (153). Elevated adiponectin in compensation for insulin resistance is also a suggested factor, as is resistance to adiponectin itself, as a consequence of a downregulation of either the adiponectin receptors or their downstream signalling pathways.

Regardless of the speculations, the direct action of adiponectin on pancreatic cancer cells has not yet been investigated. It has been observed that NF-kappa B activation occurs in cell cultures, animal models and human tissue of pancreatic cancer (154). Similarly, the activation of carcinogenic pathways involving STAT3, mTOR, PI3K and ERK, mediated by IL-6, have also been found to contribute to the survival and proliferation of pancreatic cancer cells (155). Although adiponectin is able to specifically counteract these pathways in other cancers, further studies are needed to conclusively establish adiponectin's role in pancreatic cancer.

6.8. Oesophageal adenocarcinoma

The association between oesophageal adenocarcinoma and adiponectin remains inconclusive, despite there being some evidence for an inverse correlation between adiponectin levels and oesophageal cancer, and between adiponectin and BE (156, 157). There has, however, been some doubt thrown on those associations (158).

Table 2. Possible physiopathological bases for the development of cancer in obese individuals

Hormonal changes	Increased oestrogens
	Increased insulin
	Increased insulin-like growth factors
Immune alterations	Disrupted immune response
	Increased activation of NF-kappa B
	Increased oxidative stress
	Increased peroxidation
Adipocytokines	Increased leptin
	Decreased adiponectin
	Other adipocytokines e.g. TNF-alpha, IL-6, PAI-1
Miscellaneous	Hypertension
	Chronic systemic inflammation
	Increased acid reflux
	Increased iodine uptake
	Decreased vitamin D bioavailability

Table 3. Effects of leptin on different types of cancer

Effects	Signalling pathway or molecular mechanism	Cancers involved
Growth	MAPK/ERK/JNK	Prostate Breast Colorectal Renal Endometrial Oesophageal
	JAK/STAT	Breast Colorectal Renal Endometrial Oesophageal
	PI3K/Akt	Prostate Breast Colorectal Thyroid Endometrial Oesophageal
	IL-6 induction	Colorectal
	PKA	Endometrial
	EGFR transactivation	Oesophageal
	Aromatase induction	Breast
	MAPK/ERK/JNK, JAK/STAT or PI3K/Akt	Colorectal Thyroid Oesophageal
Invasion/metastasis	MAPK/ERK/JNK, JAK/STAT or PI3K/Akt	Endometrial Renal
	EGFR transactivation	Breast
	Intercellular matrix proteolysis	Breast
Angiogenesis	MAPK/ERK	Renal Endometrial ¹
	PI3K/Akt	Renal Endometrial ¹
	VEGF induction	Prostate Breast Colorectal
	FGF 2 induction	Prostate
	Matrix metalloproteinases	Prostate
Lipid/protein biosynthesis	ACAT	Renal

¹ These pathways lead to the induction of COX-2 in endometrial cancer cells.

In contrast, while the question of whether serum adiponectin levels are related to the incidence of oesophageal adenocarcinoma remains unresolved, its receptors AdipoR1 and R2 have been identified in many oesophageal adenocarcinoma cell lines and BE tissue (114, 116, 159). Konturek *et al.* showed that the neoplastic tissue

had decreased expression of the adiponectin receptors, raising the possibility that adiponectin had reduced biological function, and thus less protective capacity, in tumour cells (159).

The anti-proliferative and pro-apoptotic effects of adiponectin on oesophageal adenocarcinoma have been demonstrated *in vitro*. Adiponectin was shown to increase apoptosis in a dose-dependent manner, with the concomitant increased expression of Bax and decreased expression of Bcl2, in the OE-19 cell line (159). Bax is pro-apoptotic whereas Bcl-2 is anti-apoptotic, such that the reciprocal regulation of the two proteins propels the adenocarcinoma cells towards apoptosis. In a study using four oesophageal adenocarcinoma cell lines and an additional non-tumorigenic cell line, globular adiponectin suppressed the proliferative effect of leptin. That effect was specifically mediated by AdipoR1, through AMPK and serine/threonine phosphatases that have not as yet been clearly identified (116). It has also been proposed that the abnormal activation of ERK 1/2, which promotes survival and proliferation in BE tissue, may be due to a deficiency in serum adiponectin (157).

7. PERSPECTIVES

There is strong epidemiological and molecular evidence showing that obesity is a risk factor for the development of several types of cancer. The pathophysiological bases for that association between obesity and cancer are summarised in Table 2.

The adipokines leptin and adiponectin, which levels are respectively elevated and decreased in obesity, may play important roles in the pathogenesis of obesity-related cancer. Both have crucial effects of cell proliferation, apoptosis, cell invasiveness and angiogenesis, regulating tumour formation. Tables 3 and 4 summarise the effects of leptin and adiponectin on the reviewed cancer types.

Obesity is also associated with increased levels of several other adipocytokines, such as interleukin-6 (IL-6) and tumour necrosis factor-alpha (TNF-alpha). These adipocytokines determine a proinflammatory state leading to DNA damage (160) and angiogenesis (161), and subsequently to the promotion of carcinogenesis and metastasis (162-164). They also increase insulin resistance and cause hyperinsulinaemia, which increases the risk for cancer. Moreover, each adipocytokine has additional specific effects. For example, IL-6 can cause growth of androgen receptor-positive prostate tumours through activation of the androgen receptor (165). TNF-alpha up-regulates CXCR4 expression, which, along with its ligand CXCL12, is implicated in metastases and tumour cell survival in a wide range of cancers (166). In the future, targeted therapies, such as IL-6 antibodies (siltuximab), may be clinically available for the treatment of cancer (167).

Although there has been considerable development in elucidating the actions of adiponectin and

Table 4. Effects of adiponectin on different types of cancer

Effects	Signalling pathway or mechanism	Cancers involved
Growth inhibition	MAPK/ERK inhibition	Renal Endometrial Oesophageal
	JAK/STAT inhibition	Prostate Colorectal Renal
	PI3K/Akt inhibition	Renal Endometrial
	NF-kappa B inhibition	Breast Colorectal
	PPAR activation	Breast
	AMPK activation	Endometrial Oesophageal Prostate Colorectal
	Oestrogen biosynthesis inhibition	Breast Endometrial
Apoptosis	PPAR activation	Breast
	AMPK activation	Oesophageal Colorectal Endometrial
Invasion/metastasis inhibition	AMPK	Breast
Anti-angiogenesis	VEGF downregulation	Breast Endometrial
	Caspase-mediated endothelial cell apoptosis	Renal
Anti-inflammation	IL-6 trans-signalling inhibition	Colorectal
Lipogenesis downregulation	AMPK-mediated SREBP-1c inhibition	Colorectal

leptin in several cancers, the precise pathways and downstream effectors that are called into play are still far from understood. There is also a need to reconcile the gap between *in vitro* and *in vivo* studies, to increase the relevance of experimental results to physiological systems; further studies using animal models, such as leptin *ob/ob* or adiponectin knockout mice, would be valuable in this respect. As many adipokines are thought to affect the process of cancer progression, a clearer understanding of the effects of leptin, adiponectin and other adipokines on cancer will allow the development of prophylactic or therapeutic therapies against cancer in obese individuals.

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9. REFERENCES

1. D. W. Haslam and W. P. James: Obesity. *Lancet*, 366(9492), 1197-209 (2005)
2. A. G. Renehan, D. L. Roberts and C. Dive: Obesity and cancer: pathophysiological and biological mechanisms. *Arch Physiol Biochem*, 114(1), 71-83 (2008)

3. A. G. Renehan, M. Tyson, M. Egger, R. F. Heller and M. Zwahlen: Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *Lancet*, 371(9612), 569-78 (2008)
4. E. A. Lew and L. Garfinkel: Variations in mortality by weight among 750,000 men and women. *J Chronic Dis*, 32(8), 563-76 (1979)
5. M. Wiseman: The second World Cancer Research Fund/American Institute for Cancer Research expert report. Food, nutrition, physical activity, and the prevention of cancer: a global perspective. *Proc Nutr Soc*, 67(3), 253-6 (2008)
6. G. K. Reeves, K. Pirie, V. Beral, J. Green, E. Spencer and D. Bull: Cancer incidence and mortality in relation to body mass index in the Million Women Study: cohort study. *BMJ*, 335(7630), 1134 (2007)
7. A. Bergstrom, P. Pisani, V. Tenet, A. Wolk and H. O. Adami: Overweight as an avoidable cause of cancer in Europe. *Int J Cancer*, 91(3), 421-30 (2001)
8. S. C. Larsson and A. Wolk: Obesity and colon and rectal cancer risk: a meta-analysis of prospective studies. *Am J Clin Nutr*, 86(3), 556-65 (2007)
9. D. J. Harriss, G. Atkinson, K. George, N. T. Cable, T. Reilly, N. Haboubi, M. Zwahlen, M. Egger and A. G. Renehan: Lifestyle factors and colorectal cancer risk (1): systematic review and meta-analysis of associations with body mass index. *Colorectal Dis*, 11(6), 547-63 (2009)
10. A. A. Moghaddam, M. Woodward and R. Huxley: Obesity and risk of colorectal cancer: a meta-analysis of 31 studies with 70,000 events. *Cancer Epidemiol Biomarkers Prev*, 16(12), 2533-47 (2007)
11. Z. Dai, Y. C. Xu and L. Niu: Obesity and colorectal cancer risk: a meta-analysis of cohort studies. *World J Gastroenterol*, 13(31), 4199-206 (2007)
12. S. C. Larsson and A. Wolk: Obesity and the risk of gallbladder cancer: a meta-analysis. *Br J Cancer*, 96(9), 1457-61 (2007)
13. S. C. Larsson and A. Wolk: Overweight, obesity and risk of liver cancer: a meta-analysis of cohort studies. *Br J Cancer*, 97(7), 1005-8 (2007)
14. D. Saunders, D. Seidel, M. Allison and G. Lyraopoulos: Systematic review: the association between obesity and hepatocellular carcinoma - epidemiological evidence. *Aliment Pharmacol Ther*, 31(10), 1051-63
15. A. Mathew, P. S. George and G. Ildaphonse: Obesity and kidney cancer risk in women: a meta-analysis (1992-2008). *Asian Pac J Cancer Prev*, 10(3), 471-8 (2009)
16. G. Ildaphonse, P. S. George and A. Mathew: Obesity and kidney cancer risk in men: a meta-analysis (1992-2008). *Asian Pac J Cancer Prev*, 10(2), 279-86 (2009)

17. S. C. Larsson, N. Orsini and A. Wolk: Body mass index and pancreatic cancer risk: A meta-analysis of prospective studies. *Int J Cancer*, 120(9), 1993-8 (2007)
18. A. A. Arslan, K. J. Helzlsouer, C. Kooperberg, X. O. Shu, E. Steplowski, H. B. Bueno-de-Mesquita, C. S. Fuchs, M. D. Gross, E. J. Jacobs, A. Z. Lacroix, G. M. Petersen, R. Z. Stolzenberg-Solomon, W. Zheng, D. Albanes, L. Amundadottir, W. R. Bamlet, A. Barricarte, S. A. Bingham, H. Boeing, M. C. Boutron-Ruault, J. E. Buring, S. J. Chanock, S. Clipp, J. M. Gaziano, E. L. Giovannucci, S. E. Hankinson, P. Hartge, R. N. Hoover, D. J. Hunter, A. Hutchinson, K. B. Jacobs, P. Kraft, S. M. Lynch, J. Manjer, J. E. Manson, A. McTiernan, R. R. McWilliams, J. B. Mendelsohn, D. S. Michaud, D. Palli, T. E. Rohan, N. Slimani, G. Thomas, A. Tjonneland, G. S. Tobias, D. Trichopoulos, J. Virtamo, B. M. Wolpin, K. Yu, A. Zeleniuch-Jacquotte and A. V. Patel: Anthropometric measures, body mass index, and pancreatic cancer: a pooled analysis from the Pancreatic Cancer Cohort Consortium (PanScan). *Arch Intern Med*, 170(9), 791-802 (2010)
19. C. M. Olsen, A. C. Green, D. C. Whiteman, S. Sadeghi, F. Kolahdooz and P. M. Webb: Obesity and the risk of epithelial ovarian cancer: a systematic review and meta-analysis. *Eur J Cancer*, 43(4), 690-709 (2007)
20. D. M. Purdie, C. J. Bain, P. M. Webb, D. C. Whiteman, S. Pirozzo and A. C. Green: Body size and ovarian cancer: case-control study and systematic review (Australia). *Cancer Causes Control*, 12(9), 855-63 (2001)
21. R. Suzuki, N. Orsini, S. Saji, T. J. Key and A. Wolk: Body weight and incidence of breast cancer defined by estrogen and progesterone receptor status--a meta-analysis. *Int J Cancer*, 124(3), 698-712 (2009)
22. R. J. MacInnis and D. R. English: Body size and composition and prostate cancer risk: systematic review and meta-regression analysis. *Cancer Causes Control*, 17(8), 989-1003 (2006)
23. S. C. Larsson and A. Wolk: Body mass index and risk of multiple myeloma: a meta-analysis. *Int J Cancer*, 121(11), 2512-6 (2007)
24. S. C. Larsson and A. Wolk: Obesity and risk of non-Hodgkin's lymphoma: a meta-analysis. *Int J Cancer*, 121(7), 1564-70 (2007)
25. A. Steffen, M. B. Schulze, T. Pischon, T. Dietrich, E. Molina, M. D. Chirlaque, A. Barricarte, P. Amiano, J. R. Quiros, R. Tumino, A. Mattiello, D. Palli, P. Vineis, C. Agnoli, G. Misirli, P. Boffetta, R. Kaaks, S. Rohrmann, H. B. Bueno-de-Mesquita, P. H. Peeters, A. M. May, E. A. Spencer, N. E. Allen, S. Bingham, A. Tjonneland, J. Halkjaer, K. Overvad, J. Stegger, J. Manjer, B. Lindkvist, G. Hallmanns, R. Stenling, E. Lund, E. Riboli, C. A. Gonzalez and H. Boeing: Anthropometry and esophageal cancer risk in the European prospective investigation into cancer and nutrition. *Cancer Epidemiol Biomarkers Prev*, 18(7), 2079-89 (2009)
26. D. P. Guh, W. Zhang, N. Bansback, Z. Amarsi, C. L. Birmingham and A. H. Anis: The incidence of co-morbidities related to obesity and overweight: a systematic review and meta-analysis. *BMC Public Health*, 9, 88 (2009)
27. J. S. Flier: Obesity wars: molecular progress confronts an expanding epidemic. *Cell*, 116(2), 337-50 (2004)
28. R. S. Ahima: Adipose tissue as an endocrine organ. *Obesity (Silver Spring)*, 14 Suppl 5, 242S-249S (2006)
29. O. A. MacDougald and C. F. Burant: The rapidly expanding family of adipokines. *Cell Metab*, 6(3), 159-61 (2007)
30. C. Garofalo and E. Surmacz: Leptin and cancer. *J Cell Physiol*, 207(1), 12-22 (2006)
31. T. Kelesidis, I. Kelesidis, S. Chou and C. S. Mantzoros: Narrative review: the role of leptin in human physiology: emerging clinical applications. *Ann Intern Med*, 152(2), 93-100 (2010)
32. C. L. Boguszewski, G. Paz-Filho and L. A. Velloso: Neuroendocrine body weight regulation: integration between fat tissue, gastrointestinal tract, and the brain. *Endokrynol Pol*, 61(2), 194-206 (2010)
33. G. Fruhbeck: Intracellular signalling pathways activated by leptin. *Biochem J*, 393(Pt 1), 7-20 (2006)
34. K. Baicy, E. D. London, J. Monterosso, M. L. Wong, T. Delibasi, A. Sharma and J. Licinio: Leptin replacement alters brain response to food cues in genetically leptin-deficient adults. *Proc Natl Acad Sci U S A*, 104(46), 18276-9 (2007)
35. I. S. Farooqi, G. Matarese, G. M. Lord, J. M. Keogh, E. Lawrence, C. Agwu, V. Sanna, S. A. Jebb, F. Perna, S. Fontana, R. I. Lechler, A. M. DePaoli and S. O'Rahilly: Beneficial effects of leptin on obesity, T cell hyporesponsiveness, and neuroendocrine/metabolic dysfunction of human congenital leptin deficiency. *J Clin Invest*, 110(8), 1093-103 (2002)
36. J. Licinio, S. Caglayan, M. Ozata, B. O. Yildiz, P. B. de Miranda, F. O'Kirwan, R. Whitby, L. Liang, P. Cohen, S. Bhasin, R. M. Krauss, J. D. Veldhuis, A. J. Wagner, A. M. DePaoli, S. M. McCann and M. L. Wong: Phenotypic effects of leptin replacement on morbid obesity, diabetes mellitus, hypogonadism, and behavior in leptin-deficient adults. *Proc Natl Acad Sci U S A*, 101(13), 4531-6 (2004)
37. J. A. Matochik, E. D. London, B. O. Yildiz, M. Ozata, S. Caglayan, A. M. DePaoli, M. L. Wong and J. Licinio: Effect of leptin replacement on brain structure in genetically leptin-deficient adults. *J Clin Endocrinol Metab*, 90(5), 2851-4 (2005)
38. G. Paz-Filho, K. Esposito, B. Hurwitz, A. Sharma, C. Dong, V. Andreev, T. Delibasi, H. Erol, A. Ayala, M. L.

- Wong and J. Licinio: Changes in insulin sensitivity during leptin replacement therapy in leptin-deficient patients. *Am J Physiol Endocrinol Metab*, 295(6), E1401-8 (2008)
39. G. J. Paz-Filho, T. Babikian, R. Asarnow, T. Delibasi, K. Esposito, H. K. Erol, M. L. Wong and J. Licinio: Leptin replacement improves cognitive development. *PLoS One*, 3(8), e3098 (2008)
40. G. J. Paz-Filho, A. Ayala, K. Esposito, H. K. Erol, T. Delibasi, B. E. Hurwitz, M. L. Wong and J. Licinio: Effects of leptin on lipid metabolism. *Horm Metab Res*, 40(8), 572-4 (2008)
41. A. Bouloumie, H. C. Drexler, M. Lafontan and R. Busse: Leptin, the product of Ob gene, promotes angiogenesis. *Circ Res*, 83(10), 1059-66 (1998)
42. P. Trayhurn and I. S. Wood: Adipokines: inflammation and the pleiotropic role of white adipose tissue. *Br J Nutr*, 92(3), 347-55 (2004)
43. M. R. Sierra-Honigsmann, A. K. Nath, C. Murakami, G. Garcia-Cardena, A. Papapetropoulos, W. C. Sessa, L. A. Madge, J. S. Schechner, M. B. Schwabb, P. J. Polverini and J. R. Flores-Riveros: Biological action of leptin as an angiogenic factor. *Science*, 281(5383), 1683-6 (1998)
44. G. Boden and G. I. Shulman: Free fatty acids in obesity and type 2 diabetes: defining their role in the development of insulin resistance and beta-cell dysfunction. *Eur J Clin Invest*, 32 Suppl 3, 14-23 (2002)
45. A. W. Hsing, L. C. Sakoda and S. Chua, Jr.: Obesity, metabolic syndrome, and prostate cancer. *Am J Clin Nutr*, 86(3), s843-57 (2007)
46. H. Li, M. J. Stampfer, L. Mucci, N. Rifai, W. Qiu, T. Kurth and J. Ma: A 25-year prospective study of plasma adiponectin and leptin concentrations and prostate cancer risk and survival. *Clin Chem*, 56(1), 34-43 (2010)
47. R. Ribeiro, A. Vasconcelos, S. Costa, D. Pinto, A. Morais, J. Oliveira, F. Lobo, C. Lopes and R. Medeiros: Overexpressing leptin genetic polymorphism (-2548 G/A) is associated with susceptibility to prostate cancer and risk of advanced disease. *Prostate*, 59(3), 268-74 (2004)
48. S. J. Freedland and E. A. Platz: Obesity and prostate cancer: making sense out of apparently conflicting data. *Epidemiol Rev*, 29, 88-97 (2007)
49. K. A. Frankenberry, P. Somasundar, D. W. McFadden and L. C. Vona-Davis: Leptin induces cell migration and the expression of growth factors in human prostate cancer cells. *Am J Surg*, 188(5), 560-5 (2004)
50. P. Somasundar, K. A. Frankenberry, H. Skinner, G. Vedula, D. W. McFadden, D. Riggs, B. Jackson, R. Vangilder, S. M. Hileman and L. C. Vona-Davis: Prostate cancer cell proliferation is influenced by leptin. *J Surg Res*, 118(1), 71-82 (2004)
51. D. Kielar, J. S. Clark, A. Ciechanowicz, G. Kurzawski, T. Sulikowski and M. Naruszewicz: Leptin receptor isoforms expressed in human adipose tissue. *Metabolism*, 47(7), 844-7 (1998)
52. M. Onuma, J. D. Bub, T. L. Rummel and Y. Iwamoto: Prostate cancer cell-adipocyte interaction: leptin mediates androgen-independent prostate cancer cell proliferation through c-Jun NH2-terminal kinase. *J Biol Chem*, 278(43), 42660-7 (2003)
53. P. Somasundar, A. K. Yu, L. Vona-Davis and D. W. McFadden: Differential effects of leptin on cancer *in vitro*. *J Surg Res*, 113(1), 50-5 (2003)
54. H. Y. Park, H. M. Kwon, H. J. Lim, B. K. Hong, J. Y. Lee, B. E. Park, Y. Jang, S. Y. Cho and H. S. Kim: Potential role of leptin in angiogenesis: leptin induces endothelial cell proliferation and expression of matrix metalloproteinases *in vivo* and *in vitro*. *Exp Mol Med*, 33(2), 95-102 (2001)
55. L. Vona-Davis and D. P. Rose: Adipokines as endocrine, paracrine, and autocrine factors in breast cancer risk and progression. *Endocr Relat Cancer*, 14(2), 189-206 (2007)
56. A. Maccio, C. Madeddu, G. Gramignano, C. Mulas, C. Floris, D. Massa, G. Astara, P. Chessa and G. Mantovani: Correlation of body mass index and leptin with tumor size and stage of disease in hormone-dependent postmenopausal breast cancer: preliminary results and therapeutic implications. *J Mol Med*, 88(7), 677-86 (2010)
57. D. P. Rose, D. Komninou and G. D. Stephenson: Obesity, adipocytokines, and insulin resistance in breast cancer. *Obes Rev*, 5(3), 153-65 (2004)
58. D. Cirillo, A. M. Rachiglio, R. la Montagna, A. Giordano and N. Normanno: Leptin signaling in breast cancer: an overview. *J Cell Biochem*, 105(4), 956-64 (2008)
59. K. A. Brown and E. R. Simpson: Obesity and breast cancer: progress to understanding the relationship. *Cancer Res*, 70(1), 4-7 (2010)
60. M. Castellucci, R. De Matteis, A. Meisser, R. Cancelli, V. Monsurro, D. Islami, R. Sarzani, D. Marziani, S. Cinti and P. Bischof: Leptin modulates extracellular matrix molecules and metalloproteinases: possible implications for trophoblast invasion. *Mol Hum Reprod*, 6(10), 951-8 (2000)
61. C. Garofalo, M. Koda, S. Cascio, M. Sulkowska, L. Kanczuga-Koda, J. Golaszewska, A. Russo, S. Sulkowski and E. Surmacz: Increased expression of leptin and the leptin receptor as a marker of breast cancer progression: possible role of obesity-related stimuli. *Clin Cancer Res*, 12(5), 1447-53 (2006)
62. M. Terrasi, E. Fiorio, A. Mercanti, M. Koda, C. A. Moncada, S. Sulkowski, S. Merali, A. Russo and E. Surmacz: Functional analysis of the -2548G/A leptin gene

polymorphism in breast cancer cells. *Int J Cancer*, 125(5), 1038-44 (2009)

63. F. Revillion, M. Charlier, V. Lhotellier, L. Hornez, S. Giard, M. C. Baranzelli, J. Djiane and J. P. Peyrat: Messenger RNA expression of leptin and leptin receptors and their prognostic value in 322 human primary breast cancers. *Clin Cancer Res*, 12(7 Pt 1), 2088-94 (2006)

64. M. P. Cleary, F. C. Phillips, S. C. Getzin, T. L. Jacobson, M. K. Jacobson, T. A. Christensen, S. C. Juneja, J. P. Grande and N. J. Maihle: Genetically obese MMTV-TGF- α /Lep(ob)Lep(ob) female mice do not develop mammary tumors. *Breast Cancer Res Treat*, 77(3), 205-15 (2003)

65. M. P. Cleary, S. C. Juneja, F. C. Phillips, X. Hu, J. P. Grande and N. J. Maihle: Leptin receptor-deficient MMTV-TGF- α /Lepr(db)Lepr(db) female mice do not develop oncogene-induced mammary tumors. *Exp Biol Med (Maywood)*, 229(2), 182-93 (2004)

66. P. Stattin, A. Lukanova, C. Biessy, S. Soderberg, R. Palmqvist, R. Kaaks, T. Olsson and E. Jellum: Obesity and colon cancer: does leptin provide a link? *Int J Cancer*, 109(1), 149-52 (2004)

67. P. Stattin, R. Palmqvist, S. Soderberg, C. Biessy, B. Ardnor, G. Hallmans, R. Kaaks and T. Olsson: Plasma leptin and colorectal cancer risk: a prospective study in Northern Sweden. *Oncol Rep*, 10(6), 2015-21 (2003)

68. K. Tamakoshi, H. Toyoshima, K. Wakai, M. Kojima, K. Suzuki, Y. Watanabe, N. Hayakawa, H. Yatsuya, T. Kondo, S. Tokudome, S. Hashimoto, S. Suzuki, M. Kawado, K. Ozasa, Y. Ito and A. Tamakoshi: Leptin is associated with an increased female colorectal cancer risk: a nested case-control study in Japan. *Oncology*, 68(4-6), 454-61 (2005)

69. T. E. Nakajima, Y. Yamada, T. Hamano, K. Furuta, T. Matsuda, S. Fujita, K. Kato, T. Hamaguchi and Y. Shimada: Adipocytokines as new promising markers of colorectal tumors: adiponectin for colorectal adenoma, and resistin and visfatin for colorectal cancer. *Cancer Sci*, 101(5), 1286-91 (2010)

70. L. Tessitore, B. Vizio, O. Jenkins, I. De Stefano, C. Ritossa, J. M. Argiles, C. Benedetto and A. Mussa: Leptin expression in colorectal and breast cancer patients. *Int J Mol Med*, 5(4), 421-6 (2000)

71. A. Kumor, P. Daniel, M. Pietruczuk and E. Malecka-Panas: Serum leptin, adiponectin, and resistin concentration in colorectal adenoma and carcinoma (CC) patients. *Int J Colorectal Dis*, 24(3), 275-81 (2009)

72. F. F. Bolukbas, H. Kilic, C. Bolukbas, M. Gumus, M. Horoz, N. S. Turhal and B. Kavakli: Serum leptin concentration and advanced gastrointestinal cancers: a case controlled study. *BMC Cancer*, 4, 29 (2004)

73. M. Koda, M. Sulkowska, L. Kanczuga-Koda, S. Cascio, G. Colucci, A. Russo, E. Surmacz and S. Sulkowski: Expression of the obesity hormone leptin and its receptor correlates with hypoxia-inducible factor-1 α in human colorectal cancer. *Ann Oncol*, 18 Suppl 6, vi116-9 (2007)

74. S. S. Paik, S. M. Jang, K. S. Jang, K. H. Lee, D. Choi and S. J. Jang: Leptin expression correlates with favorable clinicopathologic phenotype and better prognosis in colorectal adenocarcinoma. *Ann Surg Oncol*, 16(2), 297-303 (2009)

75. S. Attoub, V. Noe, L. Pirola, E. Bruyneel, E. Chastre, M. Mareel, M. P. Wymann and C. Gespach: Leptin promotes invasiveness of kidney and colonic epithelial cells via phosphoinositide 3-kinase-, rho-, and rac-dependent signaling pathways. *FASEB J*, 14(14), 2329-38 (2000)

76. M. Koda, M. Sulkowska, L. Kanczuga-Koda, E. Surmacz and S. Sulkowski: Overexpression of the obesity hormone leptin in human colorectal cancer. *J Clin Pathol*, 60(8), 902-6 (2007)

77. J. C. Hardwick, G. R. Van Den Brink, G. J. Offerhaus, S. J. Van Deventer and M. P. Peppelenbosch: Leptin is a growth factor for colonic epithelial cells. *Gastroenterology*, 121(1), 79-90 (2001)

78. S. Uddin, P. P. Bavi, A. R. Hussain, G. Alsbeih, N. Al-Sanea, A. Abduljabbar, L. H. Ashari, S. Alhomoud, F. Al-Dayel, M. Ahmed and K. S. Al-Kuraya: Leptin receptor expression in Middle Eastern colorectal cancer and its potential clinical implication. *Carcinogenesis*, 30(11), 1832-40 (2009)

79. O. O. Ogunwobi and I. L. Beales: The anti-apoptotic and growth stimulatory actions of leptin in human colon cancer cells involves activation of JNK mitogen activated protein kinase, JAK2 and PI3 kinase/Akt. *Int J Colorectal Dis*, 22(4), 401-9 (2007)

80. J. I. Fenton, N. G. Hord, J. A. Lavigne, S. N. Perkins and S. D. Hursting: Leptin, insulin-like growth factor-1, and insulin-like growth factor-2 are mitogens in ApcMin/+ but not Apc+/+ colonic epithelial cell lines. *Cancer Epidemiol Biomarkers Prev*, 14(7), 1646-52 (2005)

81. T. Aparicio, L. Kotelevets, A. Tsocas, J. P. Laigneau, I. Sobhani, E. Chastre and T. Lehy: Leptin stimulates the proliferation of human colon cancer cells *in vitro* but does not promote the growth of colon cancer xenografts in nude mice or intestinal tumorigenesis in Apc(Min/+) mice. *Gut*, 54(8), 1136-45 (2005)

82. K. N. Ealey, S. Lu and M. C. Archer: Development of aberrant crypt foci in the colons of ob/ob and db/db mice: evidence that leptin is not a promoter. *Mol Carcinog*, 47(9), 667-77 (2008)

83. Z. Liu, T. Uesaka, H. Watanabe and N. Kato: High fat diet enhances colonic cell proliferation and carcinogenesis

- in rats by elevating serum leptin. *Int J Oncol*, 19(5), 1009-14 (2001)
84. A. Engeland, S. Tretli, L. A. Akslen and T. Bjorge: Body size and thyroid cancer in two million Norwegian men and women. *Br J Cancer*, 95(3), 366-70 (2006)
 85. F. Clavel-Chapelon, G. Guillas, L. Tondeur, C. Kernaeguen and M. C. Boutron-Ruault: Risk of differentiated thyroid cancer in relation to adult weight, height and body shape over life: the French E3N cohort. *Int J Cancer*, 126(12), 2984-90 (2010)
 86. M. F. Leitzmann, A. Brenner, S. C. Moore, C. Koebnick, Y. Park, A. Hollenbeck, A. Schatzkin and E. Ron: Prospective study of body mass index, physical activity and thyroid cancer. *Int J Cancer*, 126(12), 2947-56 (2010)
 87. M. Akinci, F. Kosova, B. Cetin, S. Aslan, Z. Ari and A. Cetin: Leptin levels in thyroid cancer. *Asian J Surg*, 32(4), 216-23 (2009)
 88. K. W. Nowak, P. Kaczmarek, P. Mackowiak, A. Ziolkowska, G. Albertin, W. J. Ginda, M. Trejter, G. G. Nussdorfer and L. K. Malendowicz: Rat thyroid gland expresses the long form of leptin receptors, and leptin stimulates the function of the gland in euthyroid non-fasted animals. *Int J Mol Med*, 9(1), 31-4 (2002)
 89. S. P. Cheng, C. W. Chi, C. Y. Tzen, T. L. Yang, J. J. Lee, T. P. Liu and C. L. Liu: Clinicopathologic significance of leptin and leptin receptor expressions in papillary thyroid carcinoma. *Surgery*, 147(6), 847-53 (2010)
 90. S. Uddin, P. Bavi, A. K. Siraj, M. Ahmed, M. Al-Rasheed, A. R. Hussain, T. Amin, A. Alzahrani, F. Al-Dayel, J. Abubaker, R. Bu and K. S. Al-Kuraya: Leptin-R and its association with PI3K/Akt signaling pathway in papillary thyroid carcinoma. *Endocr Relat Cancer*, 17(1), 191-202 (2010)
 91. S. P. Cheng, P. H. Yin, Y. C. Chang, C. H. Lee, S. Y. Huang and C. W. Chi: Differential roles of leptin in regulating cell migration in thyroid cancer cells. *Oncol Rep*, 23(6), 1721-7 (2010)
 92. T. N. Spyridopoulos, E. T. Petridou, N. Dessypris, A. Terzidis, A. Skalkidou, C. Deliveliotis and G. P. Chrousos: Inverse association of leptin levels with renal cell carcinoma: results from a case-control study. *Hormones (Athens)*, 8(1), 39-46 (2009)
 93. A. Horiguchi, M. Sumitomo, J. Asakuma, T. Asano, R. Zheng, D. M. Nanus and M. Hayakawa: Increased serum leptin levels and over expression of leptin receptors are associated with the invasion and progression of renal cell carcinoma. *J Urol*, 176(4 Pt 1), 1631-5 (2006)
 94. A. Horiguchi, M. Sumitomo, J. Asakuma, T. Asano, R. Zheng, D. M. Nanus and M. Hayakawa: Leptin promotes invasiveness of murine renal cancer cells via extracellular signal-regulated kinases and rho dependent pathway. *J Urol*, 176(4 Pt 1), 1636-41 (2006)
 95. L. Li, Y. Gao, L. L. Zhang and D. L. He: Concomitant activation of the JAK/STAT3 and ERK1/2 signaling is involved in leptin-mediated proliferation of renal cell carcinoma Caki-2 cells. *Cancer Biol Ther*, 7(11), 1787-92 (2008)
 96. H. A. Drabkin and R. M. Gemmill: Obesity, cholesterol, and clear-cell renal cell carcinoma (RCC). *Adv Cancer Res*, 107, 39-56 (2010)
 97. E. Petridou, M. Belechri, N. Dessypris, P. Koukoulomatis, E. Diakomanolis, E. Spanos and D. Trichopoulos: Leptin and body mass index in relation to endometrial cancer risk. *Ann Nutr Metab*, 46(3-4), 147-51 (2002)
 98. S. S. Yuan, K. B. Tsai, Y. F. Chung, T. F. Chan, Y. T. Yeh, L. Y. Tsai and J. H. Su: Aberrant expression and possible involvement of the leptin receptor in endometrial cancer. *Gynecol Oncol*, 92(3), 769-75 (2004)
 99. A. Cymbaluk, A. Chudecka-Glaz and I. Rzepka-Gorska: Leptin levels in serum depending on Body Mass Index in patients with endometrial hyperplasia and cancer. *Eur J Obstet Gynecol Reprod Biol*, 136(1), 74-7 (2008)
 100. H. Koshiba, J. Kitawaki, H. Ishihara, N. Kado, I. Kusuki, K. Tsukamoto and H. Honjo: Progesterone inhibition of functional leptin receptor mRNA expression in human endometrium. *Mol Hum Reprod*, 7(6), 567-72 (2001)
 101. M. Koda, M. Sulkowska, A. Wincewicz, L. Kanczuga-Koda, B. Musiatowicz, M. Szymanska and S. Sulkowski: Expression of leptin, leptin receptor, and hypoxia-inducible factor 1 alpha in human endometrial cancer. *Ann N Y Acad Sci*, 1095, 90-8 (2007)
 102. M. Bogusiewicz, A. Semczuk, M. Gogacz, D. Skomra, J. A. Jakowicki and T. Rechberger: Lack of correlation between leptin receptor expression and PI3-K/Akt signaling pathway proteins immunostaining in endometrioid-type endometrial carcinomas. *Cancer Lett*, 238(1), 61-8 (2006)
 103. D. Sharma, N. K. Saxena, P. M. Vertino and F. A. Anania: Leptin promotes the proliferative response and invasiveness in human endometrial cancer cells by activating multiple signal-transduction pathways. *Endocr Relat Cancer*, 13(2), 629-40 (2006)
 104. S. Catalano, C. Giordano, P. Rizza, G. Gu, I. Barone, D. Bonofiglio, F. Giordano, R. Malivindi, D. Gaccione, M. Lanzino, F. De Amicis and S. Ando: Evidence that leptin through STAT and CREB signaling enhances cyclin D1 expression and promotes human endometrial cancer proliferation. *J Cell Physiol*, 218(3), 490-500 (2009)
 105. C. Carino, A. B. Olawaiye, S. Cherfils, T. Serikawa, M. P. Lynch, B. R. Rueda and R. R. Gonzalez: Leptin

regulation of proangiogenic molecules in benign and cancerous endometrial cells. *Int J Cancer*, 123(12), 2782-90 (2008)

106. R. Pezzilli, A. Barassi, M. M. Corsi, A. M. Morselli-Labate, D. Campana, R. Casadei, D. Santini, R. Corinaldesi and G. M. D'Eri: Serum leptin, but not adiponectin and receptor for advanced glycation end products, is able to distinguish autoimmune pancreatitis from both chronic pancreatitis and pancreatic neoplasms. *Scand J Gastroenterol*, 45(1), 93-9 (2010)

107. M. Dalamaga, K. Karmaniolas, A. Panagiotou, A. Hsi, J. Chamberland, C. Dimas, A. Lekka and C. S. Mantzoros: Low circulating adiponectin and resistin, but not leptin, levels are associated with multiple myeloma risk: a case-control study. *Cancer Causes Control*, 20(2), 193-9 (2009)

108. D. R. Brown, D. E. Berkowitz and M. J. Breslow: Weight loss is not associated with hyperleptinemia in humans with pancreatic cancer. *J Clin Endocrinol Metab*, 86(1), 162-6 (2001)

109. R. Z. Stolzenberg-Solomon, B. I. Graubard, S. Chari, P. Limburg, P. R. Taylor, J. Virtamo and D. Albanes: Insulin, glucose, insulin resistance, and pancreatic cancer in male smokers. *JAMA*, 294(22), 2872-8 (2005)

110. N. J. Zyromski, A. Mathur, H. A. Pitt, T. E. Wade, S. Wang, P. Nakshatri, D. A. Swartz-Basile and H. Nakshatri: Obesity potentiates the growth and dissemination of pancreatic cancer. *Surgery*, 146(2), 258-63 (2009)

111. T. E. Nakajima, Y. Yamada, T. Hamano, K. Furuta, I. Oda, H. Kato, K. Kato, T. Hamaguchi and Y. Shimada: Adipocytokines and squamous cell carcinoma of the esophagus. *J Cancer Res Clin Oncol*, 136(2), 261-6 (2010)

112. B. J. Kendall, G. A. Macdonald, N. K. Hayward, J. B. Prins, I. Brown, N. Walker, N. Pandeya, A. C. Green, P. M. Webb and D. C. Whiteman: Leptin and the risk of Barrett's oesophagus. *Gut*, 57(4), 448-54 (2008)

113. D. Diakowska, M. Krzystek-Korpacka, K. Markocka-Maczka, W. Diakowski, M. Matusiewicz and K. Grabowski: Circulating leptin and inflammatory response in esophageal cancer, esophageal cancer-related cachexia-anorexia syndrome (CAS) and non-malignant CAS of the alimentary tract. *Cytokine*, 51(2), 132-7 (2010)

114. J. M. Howard, P. Beddy, D. Ennis, M. Keogan, G. P. Pidgeon and J. V. Reynolds: Associations between leptin and adiponectin receptor upregulation, visceral obesity and tumour stage in oesophageal and junctional adenocarcinoma. *Br J Surg*, 97(7), 1020-7 (2010)

115. O. O. Ogunwobi and I. L. Beales: Leptin stimulates the proliferation of human oesophageal adenocarcinoma cells via HB-EGF and TGF- α mediated transactivation of the epidermal growth factor receptor. *Br J Biomed Sci*, 65(3), 121-7 (2008)

116. O. O. Ogunwobi and I. L. Beales: Globular adiponectin, acting via adiponectin receptor-1, inhibits leptin-stimulated oesophageal adenocarcinoma cell proliferation. *Mol Cell Endocrinol*, 285(1-2), 43-50 (2008)

117. I. L. Beales and O. O. Ogunwobi: Leptin synergistically enhances the anti-apoptotic and growth-promoting effects of acid in OE33 oesophageal adenocarcinoma cells in culture. *Mol Cell Endocrinol*, 274(1-2), 60-8 (2007)

118. O. Ogunwobi, G. Mutungi and I. L. Beales: Leptin stimulates proliferation and inhibits apoptosis in Barrett's esophageal adenocarcinoma cells by cyclooxygenase-2-dependent, prostaglandin-E2-mediated transactivation of the epidermal growth factor receptor and c-Jun NH2-terminal kinase activation. *Endocrinology*, 147(9), 4505-16 (2006)

119. P. Somasundar, D. Riggs, B. Jackson, L. Vona-Davis and D. W. McFadden: Leptin stimulates esophageal adenocarcinoma growth by nonapoptotic mechanisms. *Am J Surg*, 186(5), 575-8 (2003)

120. S. E. Wozniak, L. L. Gee, M. S. Wachtel and E. E. Frezza: Adipose tissue: the new endocrine organ? A review article. *Dig Dis Sci*, 54(9), 1847-56 (2009)

121. M. Liu and F. Liu: Transcriptional and post-translational regulation of adiponectin. *Biochem J*, 425(1), 41-52

122. T. Kadowaki and T. Yamauchi: Adiponectin and adiponectin receptors. *Endocr Rev*, 26(3), 439-51 (2005)

123. N. Ouchi, S. Kihara, Y. Arita, Y. Okamoto, K. Maeda, H. Kuriyama, K. Hotta, M. Nishida, M. Takahashi, M. Muraguchi, Y. Ohmoto, T. Nakamura, S. Yamashita, T. Funahashi and Y. Matsuzawa: Adiponectin, an adipocyte-derived plasma protein, inhibits endothelial NF- κ B signaling through a cAMP-dependent pathway. *Circulation*, 102(11), 1296-301 (2000)

124. E. Brakenhielm, N. Veitonmaki, R. Cao, S. Kihara, Y. Matsuzawa, B. Zhivotovsky, T. Funahashi and Y. Cao: Adiponectin-induced antiangiogenesis and antitumor activity involve caspase-mediated endothelial cell apoptosis. *Proc Natl Acad Sci U S A*, 101(8), 2476-81 (2004)

125. I. Kelesidis, T. Kelesidis and C. S. Mantzoros: Adiponectin and cancer: a systematic review. *Br J Cancer*, 94(9), 1221-5 (2006)

126. S. Goktas, M. I. Yilmaz, K. Caglar, A. Sonmez, S. Kilic and S. Bedir: Prostate cancer and adiponectin. *Urology*, 65(6), 1168-72 (2005)

127. J. D. Bub, T. Miyazaki and Y. Iwamoto: Adiponectin as a growth inhibitor in prostate cancer cells. *Biochem Biophys Res Commun*, 340(4), 1158-66 (2006)

128. C. Mantzoros, E. Petridou, N. Dessypris, C. Chavelas, M. Dalamaga, D. M. Alexe, Y. Papadiamantis, C. Markopoulos, E. Spanos, G. Chrousos and D. Trichopoulos: Adiponectin and breast cancer risk. *J Clin Endocrinol Metab*, 89(3), 1102-7 (2004)
129. M. P. Cleary and M. E. Grossmann: Minireview: Obesity and breast cancer: the estrogen connection. *Endocrinology*, 150(6), 2537-42 (2009)
130. K. Y. Kim, A. Baek, J. E. Hwang, Y. A. Choi, J. Jeong, M. S. Lee, D. H. Cho, J. S. Lim, K. I. Kim and Y. Yang: Adiponectin-activated AMPK stimulates dephosphorylation of Akt through protein phosphatase 2A activation. *Cancer Res*, 69(9), 4018-26 (2009)
131. A. M. Lorincz and S. Sukumar: Molecular links between obesity and breast cancer. *Endocr Relat Cancer*, 13(2), 279-92 (2006)
132. A. Lukanova, S. Soderberg, R. Kaaks, E. Jellum and P. Stattin: Serum adiponectin is not associated with risk of colorectal cancer. *Cancer Epidemiol Biomarkers Prev*, 15(2), 401-2 (2006)
133. E. K. Wei, E. Giovannucci, C. S. Fuchs, W. C. Willett and C. S. Mantzoros: Low plasma adiponectin levels and risk of colorectal cancer in men: a prospective study. *J Natl Cancer Inst*, 97(22), 1688-94 (2005)
134. S. Otake, H. Takeda, Y. Suzuki, T. Fukui, S. Watanabe, K. Ishihama, T. Saito, H. Togashi, T. Nakamura, Y. Matsuzawa and S. Kawata: Association of visceral fat accumulation and plasma adiponectin with colorectal adenoma: evidence for participation of insulin resistance. *Clin Cancer Res*, 11(10), 3642-6 (2005)
135. T. Yamaji, M. Iwasaki, S. Sasazuki and S. Tsugane: Interaction between adiponectin and leptin influences the risk of colorectal adenoma. *Cancer Res*, 70(13), 5430-7 (2010)
136. D. Barb, C. J. Williams, A. K. Neuwirth and C. S. Mantzoros: Adiponectin in relation to malignancies: a review of existing basic research and clinical evidence. *Am J Clin Nutr*, 86(3), s588-66 (2007)
137. M. Sugiyama, H. Takahashi, K. Hosono, H. Endo, S. Kato, K. Yoneda, Y. Nozaki, K. Fujita, M. Yoneda, K. Wada, H. Nakagawa and A. Nakajima: Adiponectin inhibits colorectal cancer cell growth through the AMPK/mTOR pathway. *Int J Oncol*, 34(2), 339-44 (2009)
138. A. Y. Kim, Y. S. Lee, K. H. Kim, J. H. Lee, H. K. Lee, S. H. Jang, S. E. Kim, G. Y. Lee, J. W. Lee, S. A. Jung, H. Y. Chung, S. Jeong and J. B. Kim: Adiponectin represses colon cancer cell proliferation via AdipoR1- and -R2-mediated AMPK activation. *Mol Endocrinol*, 24(7), 1441-52 (2010)
139. J. I. Fenton, J. M. Birmingham, S. D. Hursting and N. G. Hord: Adiponectin blocks multiple signaling cascades associated with leptin-induced cell proliferation in Apc Min/+ colon epithelial cells. *Int J Cancer*, 122(11), 2437-45 (2008)
140. J. T. Park, T. H. Yoo, T. I. Chang, D. H. Lee, J. H. Lee, J. E. Lee, H. Y. Choi, S. W. Kang, D. S. Han and D. R. Ryu: Insulin resistance and lower plasma adiponectin increase malignancy risk in nondiabetic continuous ambulatory peritoneal dialysis patients. *Metabolism* (2010)
141. J. H. Pinthus, N. Kleinmann, B. Tisdale, S. Chatterjee, J. P. Lu, A. Gillis, T. Hamlet, G. Singh, F. Farrokhyar and A. Kapoor: Lower plasma adiponectin levels are associated with larger tumor size and metastasis in clear-cell carcinoma of the kidney. *Eur Urol*, 54(4), 866-73 (2008)
142. T. N. Spyridopoulos, E. T. Petridou, A. Skalkidou, N. Dessypris, G. P. Chrousos and C. S. Mantzoros: Low adiponectin levels are associated with renal cell carcinoma: a case-control study. *Int J Cancer*, 120(7), 1573-8 (2007)
143. A. Horiguchi, K. Ito, M. Sumitomo, F. Kimura, T. Asano and M. Hayakawa: Decreased serum adiponectin levels in patients with metastatic renal cell carcinoma. *Jpn J Clin Oncol*, 38(2), 106-11 (2008)
144. P. T. Soliman, D. Wu, G. Tortolero-Luna, K. M. Schmeler, B. M. Slomovitz, M. S. Bray, D. M. Gershenson and K. H. Lu: Association between adiponectin, insulin resistance, and endometrial cancer. *Cancer*, 106(11), 2376-81 (2006)
145. A. E. Cust, R. Kaaks, C. Friedenreich, F. Bonnet, M. Lavielle, A. Lukanova, S. Rinaldi, L. Dossus, N. Slimani, E. Lundin, A. Tjonneland, A. Olsen, K. Overvad, F. Clavel-Chapelon, S. Mesrine, V. Joulin, J. Linseisen, S. Rohrmann, T. Pischon, H. Boeing, D. Trichopoulos, A. Trichopoulou, V. Benetou, D. Palli, F. Berrino, R. Tumino, C. Sacerdote, A. Mattiello, J. R. Quiros, M. A. Mendez, M. J. Sanchez, N. Larranaga, M. J. Tormo, E. Ardanaz, H. B. Bueno-de-Mesquita, P. H. Peeters, C. H. van Gils, K. T. Khaw, S. Bingham, N. Allen, T. Key, M. Jenab and E. Riboli: Plasma adiponectin levels and endometrial cancer risk in pre- and postmenopausal women. *J Clin Endocrinol Metab*, 92(1), 255-63 (2007)
146. L. Dal Maso, L. S. Augustin, A. Karalis, R. Talamini, S. Franceschi, D. Trichopoulos, C. S. Mantzoros and C. La Vecchia: Circulating adiponectin and endometrial cancer risk. *J Clin Endocrinol Metab*, 89(3), 1160-3 (2004)
147. Y. Takemura, Y. Osuga, T. Yamauchi, M. Kobayashi, M. Harada, T. Hirata, C. Morimoto, Y. Hirota, O. Yoshino, K. Koga, T. Yano, T. Kadowaki and Y. Taketani: Expression of adiponectin receptors and its possible implication in the human endometrium. *Endocrinology*, 147(7), 3203-10 (2006)
148. L. Cong, J. Gasser, J. Zhao, B. Yang, F. Li and A. Z. Zhao: Human adiponectin inhibits cell growth and induces apoptosis in human endometrial carcinoma cells, HEC-1-A and RL95 2. *Endocr Relat Cancer*, 14(3), 713-20 (2007)

149. M. C. Chang, Y. T. Chang, T. C. Su, W. S. Yang, C. L. Chen, Y. W. Tien, P. C. Liang, S. C. Wei and J. M. Wong: Adiponectin as a potential differential marker to distinguish pancreatic cancer and chronic pancreatitis. *Pancreas*, 35(1), 16-21 (2007)
150. M. Dalamaga, I. Migdalis, J. L. Fargnoli, E. Papadavid, E. Bloom, N. Mitsiades, K. Karmaniolas, N. Pelecanos, S. Tseleni-Balafouta, A. Dionyssiou-Asteriou and C. S. Mantzoros: Pancreatic cancer expresses adiponectin receptors and is associated with hypoleptinemia and hyperadiponectinemia: a case-control study. *Cancer Causes Control*, 20(5), 625-33 (2009)
151. R. Z. Stolzenberg-Solomon, S. Weinstein, M. Pollak, Y. Tao, P. R. Taylor, J. Virtamo and D. Albanes: Prediagnostic adiponectin concentrations and pancreatic cancer risk in male smokers. *Am J Epidemiol*, 168(9), 1047-55 (2008)
152. I. Wolf, S. Sadetzki, H. Kanety, Y. Kundel, C. Pariente, N. Epstein, B. Oberman, R. Catane, B. Kaufman and I. Shimon: Adiponectin, ghrelin, and leptin in cancer cachexia in breast and colon cancer patients. *Cancer*, 106(4), 966-73 (2006)
153. S. M. Brichard, M. L. Delporte and M. Lambert: Adipocytokines in anorexia nervosa: a review focusing on leptin and adiponectin. *Horm Metab Res*, 35(6), 337-42 (2003)
154. G. Garcea, A. R. Dennison, W. P. Steward and D. P. Berry: Role of inflammation in pancreatic carcinogenesis and the implications for future therapy. *Pancreatol*, 5(6), 514-29 (2005)
155. J. Chen and X. F. Huang: Interleukin-6 promotes carcinogenesis through multiple signal pathways. Comment on: Clinical significance of interleukin-6 gene polymorphism and IL-6 serum level in pancreatic adenocarcinoma and chronic pancreatitis. *Dig Dis Sci*, 54(6), 1373-4 (2009)
156. A. Yildirim, M. Bilici, K. Cayir, V. Yanmaz, S. Yildirim and S. B. Tekin: Serum adiponectin levels in patients with esophageal cancer. *Jpn J Clin Oncol*, 39(2), 92-6 (2009)
157. J. H. Rubenstein, A. Dahlkemper, J. Y. Kao, M. Zhang, H. Morgenstern, L. McMahon and J. M. Inadomi: A pilot study of the association of low plasma adiponectin and Barrett's esophagus. *Am J Gastroenterol*, 103(6), 1358-64 (2008)
158. T. Fujita: Adiponectin and Barrett's esophagus. *Am J Gastroenterol*, 104(1), 243; author reply 244-5 (2009)
159. P. C. Konturek, G. Burnat, T. Rau, E. G. Hahn and S. Konturek: Effect of adiponectin and ghrelin on apoptosis of Barrett adenocarcinoma cell line. *Dig Dis Sci*, 53(3), 597-605 (2008)
160. M. Jaiswal, N. F. LaRusso, L. J. Burgart and G. J. Gores: Inflammatory cytokines induce DNA damage and inhibit DNA repair in cholangiocarcinoma cells by a nitric oxide-dependent mechanism. *Cancer Res*, 60(1), 184-90 (2000)
161. J. R. Jackson, M. P. Seed, C. H. Kircher, D. A. Willoughby and J. D. Winkler: The codependence of angiogenesis and chronic inflammation. *FASEB J*, 11(6), 457-65 (1997)
162. F. Balkwill and A. Mantovani: Inflammation and cancer: back to Virchow? *Lancet*, 357(9255), 539-45 (2001)
163. G. Lazenec and A. Richmond: Chemokines and chemokine receptors: new insights into cancer-related inflammation. *Trends Mol Med*, 16(3), 133-44 (2010)
164. A. Mantovani, P. Allavena, A. Sica and F. Balkwill: Cancer-related inflammation. *Nature*, 454(7203), 436-44 (2008)
165. K. Malinowska, H. Neuwirt, I. T. Cavarretta, J. Bektic, H. Steiner, H. Dietrich, P. L. Moser, D. Fuchs, A. Hobisch and Z. Culig: Interleukin-6 stimulation of growth of prostate cancer *in vitro* and *in vivo* through activation of the androgen receptor. *Endocr Relat Cancer*, 16(1), 155-69 (2009)
166. F. Balkwill: The significance of cancer cell expression of the chemokine receptor CXCR4. *Semin Cancer Biol*, 14(3), 171-9 (2004)
167. T. B. Dorff, B. Goldman, J. K. Pinski, P. C. Mack, P. N. Lara, Jr., P. J. Van Veldhuizen, Jr., D. I. Quinn, N. J. Vogelzang, I. M. Thompson, Jr. and M. H. Hussain: Clinical and correlative results of SWOG S0354: a phase II trial of CNT0328 (siltuximab), a monoclonal antibody against interleukin-6, in chemotherapy-pretreated patients with castration-resistant prostate cancer. *Clin Cancer Res*, 16(11), 3028-34 (2010)

Abbreviations: ACAT: acyl-coenzyme A:cholesterol acyl transferase; Akt: protein kinase B; AMPK: 5' AMP-activated protein kinase; BE: Barrett's Oesophagus; BMI: body mass index; COX: cyclooxygenase; EGF: epidermal growth factor; EGFR: epidermal growth factor receptor; ER: oestrogen receptor; ERK: extracellular signal-regulated kinase; FGF: fibroblast growth factor; HIF: hypoxia-induced factor; IGF: insulin-like growth factor; IRS: insulin receptor substrate; IL: interleukin; JAK: Janus kinase; JNK: c-Jun N-terminal kinase; MAPK: mitogen-activated protein kinase; mTOR: mammalian target of rapamycin; NF-kappa B: nuclear factor kappa B; PAI: plasminogen activator inhibitor; PI3K: phosphoinositide 3-kinase; PKA: protein kinase A; PPAR: peroxisome proliferator-activated receptor; PTEN: phosphatase and tensin homolog; SREBP: sterol regulatory element binding protein; STAT: signal transducer and activator of transcription; TGF: transforming growth factor; TNF: tumour necrosis factor; VEGF: vascular endothelial growth factor.

Adipokines and cancer

Key Words: Adipocytokine, Adipokine, Adiponectin, Cancer, Leptin, Obesity, Review

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