

Role of nitric oxide in shock: the large animal perspective

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1. ABSTRACT

Excessive nitric oxide (NO) formation plays important roles in the pathogenesis of shock and multiple organ failure in sepsis and acute lung injury (ALI). Evidence from studies in large animal models of shock provide further insight into the role of NO and the varying nitric oxide synthase (NOS) isoforms. Nonselective NOS inhibition in sepsis models reversed sepsis-induced derangements in hemodynamic status, but was associated with side effects such as pulmonary vasoconstriction and decreases in global oxygen delivery. Results from studies on specific inhibition of inducible NOS (iNOS, NOS-2) and neuronal NOS (nNOS, NOS-1) in sepsis models remain inconclusive, but suggest that both isoenzymes are involved in the pathophysiological processes. While the long-term effects of NOS inhibition in models of burn and inhalation injury remain unknown, specific iNOS inhibition attenuated ALI without worsening injury-related pulmonary hypertension. Further investigation in large animal models is warranted to clarify the time course of increased expression and/or activity of different NOS isoenzymes and the effects of specific inhibition of the NOS isoforms at different time points.

2. INTRODUCTION

Nitric oxide (NO) is an endogenous vasodilator generated from L-arginine through catalysis by a family of enzymes called NO synthases (NOS). Three different genetic isoforms of NOS have been identified in mammals (1); in contrast to the constitutively synthesized isoenzymes endothelial NOS (eNOS, NOS-3) and neuronal NOS (nNOS, NOS-1), the inducible NOS (iNOS, NOS-2) is up-regulated by diverse stress stimuli such as oxidative burst and systemic inflammation. Constitutively produced NO is involved in various physiologic processes, including neurotransmission and the regulation of vascular tone and blood flow (2). Under pathophysiological conditions, however, endotoxin or inflammatory cytokines such as interleukin-1 (IL-1), interleukin-6 (IL-6), interferon gamma (IFN-gamma) and tumor necrosis factor alpha (TNF-alpha) may lead to an increased expression of iNOS. The resulting overproduction of NO is thought to be an important factor in the pathogenesis of shock and multiple organ failure resulting from sepsis and acute lung injury (2-4).

Importantly, a significant amount of research on the role of NO in shock states of various etiologies has

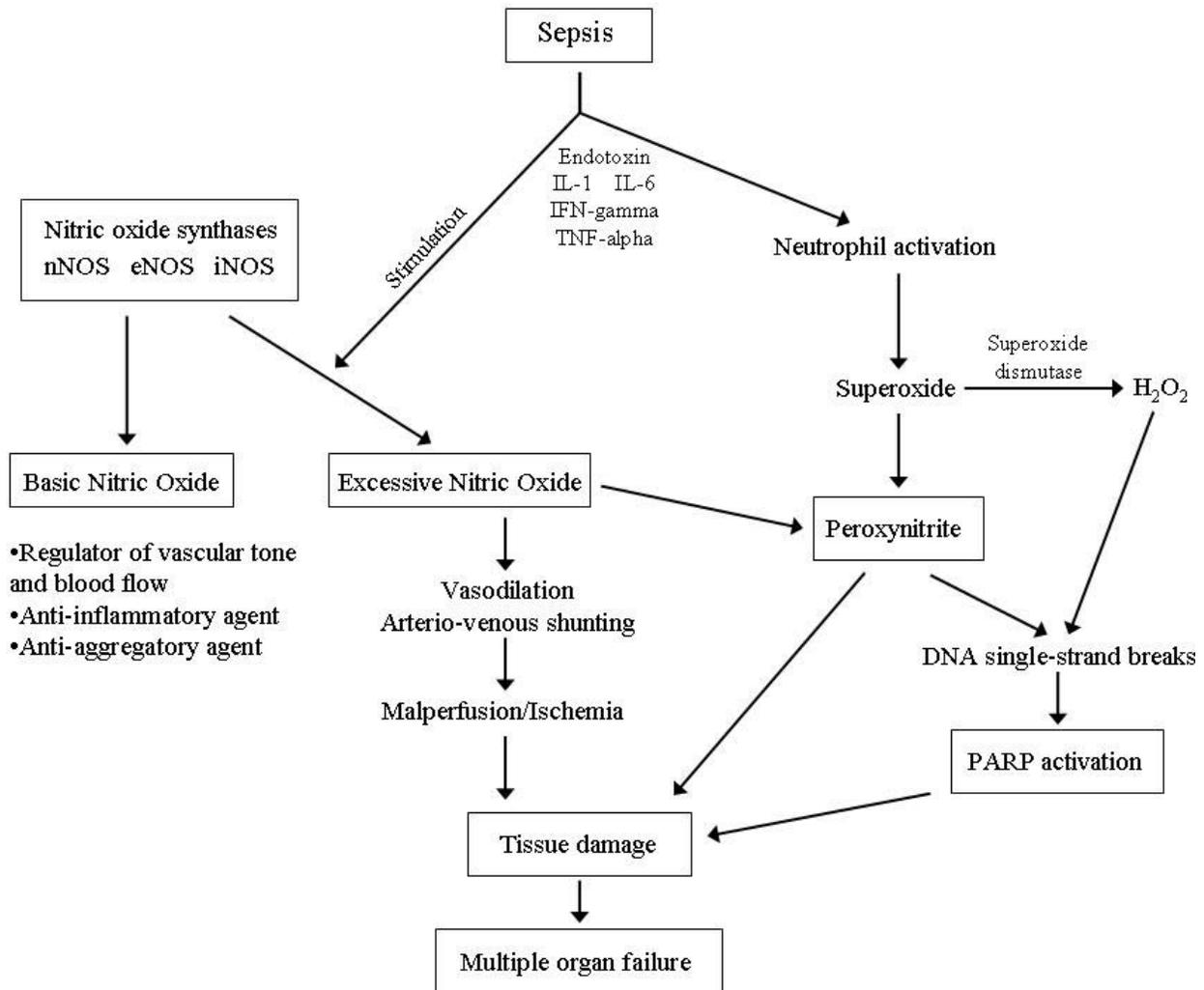


Figure 1. Possible role of nitric oxide in the pathophysiology of sepsis. Excessive nitric oxide production leads to vasodilation and arterio-venous shunting as well as peroxynitrite formation and poly(ADP-ribose) polymerase (PARP) activation, both contributing to tissue damage and multiple organ failure. The roles of the different nitric oxide synthase (NOS) isoenzymes at different time points of sepsis are not sufficiently identified.

been conducted in large animal models. This review examines the role of NO production and its pharmacological inhibition in large animal models of septic shock and/or acute lung injury.

3. ROLE OF NITRIC OXIDE IN ENDOTOXEMIA AND SEPSIS

Sepsis is a state of sustained infection, resulting in a severe systemic inflammatory response and, ultimately, shock. Despite significant improvements in critical care medicine during the last few decades, the mortality in septic shock remains high (5). The pathophysiological changes in patients with sepsis are typically characterized by systemic vasodilation to metabolically inactive tissues (6). The resultant systemic arterial hypotension reduces blood flow to organs that are metabolically active. The ensuing misdistribution of

systemic and microvascular blood flow leads to an impairment of tissue oxygenation, finally resulting in multiple organ failure (7, 8). Excessive formation of NO may be critically involved in these vascular changes (Figure 1). Via its secondary messenger, cyclic guanosine monophosphate, NO activates the myosin phosphatase and, by dephosphorylating myosin, causes vasodilation (8). Moreover, excessive NO formation may activate potassium channels in vascular smooth muscles, thereby causing vaso-relaxation (9, 10). The amount of NO production within the vascular system may vary at different anatomical sites, resulting in different degrees of vasodilation. Consequently, an underperfusion of metabolically active tissue and an overperfusion of metabolically inactive tissues may occur, possibly contributing to the deficient oxygen extraction, tissue hypoxia and lactic acidosis often observed in patients with septic shock (2).

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Increased plasma and urine levels of the stable NO byproducts nitrate and nitrite in septic patients, combined with the identification of the endothelium-derived relaxing factor as NO, led to the assumption that NO may be involved in the pathogenesis of cardiovascular changes in septic shock (4, 11). Since then, a vast number of studies have been performed, investigating the role of NO in the pathogenesis of septic shock. Most research in that field has been conducted in animal models. Large animal models are most suitable to study the pathophysiology of septic shock and the effects of various treatment strategies because they exert circulatory alterations that closely mimic the hemodynamic changes in patients with sepsis (12-14), whereas rodents produce NO at a much greater rate.

In response to continuous infusion of endotoxin or live bacteria, sheep typically develop significant decreases in systemic vascular resistance and blood pressure, while cardiac output and pulmonary pressure markedly increase. Furthermore, endotoxin infusion in sheep is associated with impairments of global oxygen transport and significantly increased regional blood flows (15-22). Notably, these endotoxin-related changes could be largely reversed by administration of the nonselective NOS inhibitor L-nitro-arginine-methylester (L-NAME), indicating that increased NO synthesis has a major role in the cardiovascular alterations in ovine endotoxemia. Infusion of L-NAME in endotoxemic ewes restored systemic vascular resistance and mean arterial pressure, while heart rate and cardiac index decreased. In addition, nonselective NOS inhibition in sheep reversed the endotoxin-induced elevation in cardiac output, oxygen delivery and regional blood flows (19-21).

Like sheep, pigs exert a hyperdynamic circulation and a substantial decrease in systemic vascular resistance in response to continuous infusion of endotoxin (23-25). In an animal model of endotoxic shock in swine, Santak *et al.* (25) confirmed the significant role of increased NO formation in endotoxin-induced cardiovascular changes. Infusion of the nonselective NOS inhibitor N-monomethyl-L-arginine (L-NMMA) reversed the hyperdynamic circulation close to pre-endotoxin levels. The authors also reported an attenuation of the endotoxin-related increase in NO₃ production by L-NMMA. However, despite hemodynamic stabilization, L-NMMA administration in pigs failed to beneficially influence the endotoxin-induced disturbances of both intestinal and liver energy balance (26, 27).

The role of NO in the pathogenesis of cardiovascular changes in response to endotoxin or tumor necrosis factor (TNF) has been investigated in a dog model (28-31). In anesthetized dogs, TNF, a cytotoxin produced by macrophages in reaction to bacterial endotoxin, induced a significant fall in mean arterial pressure, which was completely reversed by bolus infusion of L-NAME (28). Similarly, the decreases in systemic vascular resistance and mean arterial pressure following endotoxin infusion in dogs were inverted by bolus administration of L-NMMA (29). In the same animal model, Zhang *et al.* (30) tested the effects

of methylene blue, an inhibitor of soluble guanylate cyclase, on cardiopulmonary hemodynamics in endotoxic shock. Methylene blue increased systemic vascular resistance and arterial pressure in a dose-dependent manner, while organ blood flows decreased.

Taken together, these findings in dogs are in agreement with those made in sheep and pigs, suggesting that NO is critically involved in the pathophysiology of cardiovascular alterations due to endotoxemia in large animals. However, nonselective NOS inhibition in large animals was also associated with several unfavorable side effects. Inhibition of NOS by L-NAME and L-NMMA apparently reversed the endotoxin-related hyperdynamic circulation, as indicated by decreases in cardiac output, oxygen delivery and regional blood flows, including hepatic, portal, mesenteric and renal blood flow (15, 19-21, 30, 31). Nonetheless, it should be noted that NO is not only involved in pathophysiological processes (e.g., iNOS), but constitutively produced NO is an important physiological regulator of vascular tone and blood flow (e.g., nNOS, eNOS). Especially in sepsis, a condition of increased oxygen demand, it appears deleterious to inhibit all NOS isoforms to the same extent, because this process may lead to a further dysregulation of local vascular tone and regional perfusion, thereby possibly fostering tissue hypoxia and organ failure.

Furthermore, administration of L-NAME in sheep, dogs and pigs aggravated endotoxin-related pulmonary hypertension (15, 17, 20, 21, 25, 28-33) and pulmonary edema (17). This phenomenon may be explained by the blunt of vasodilatory effects of constitutively produced NO due to nonselective NOS inhibition. In this regard, it could be demonstrated that concomitant inhalation of NO decreased pulmonary hypertension (17, 32, 33) and ameliorated pulmonary edema (17) in experimental endotoxemia.

In interpreting these findings, it appears desirable to selectively inhibit iNOS without affecting the beneficial effects of constitutively expressed NOS. Several studies on selective iNOS inhibition using different compounds have been conducted. However, the results of these investigations remain largely inconclusive.

Booke *et al.* (34) investigated the effects of S-ethylisothiourea (S-EITU), a selective iNOS inhibitor *in vitro*, in healthy sheep and sheep exposed to continuous infusion of live bacteria. Since the overproduction of iNOS is believed to be responsible for septic vasodilation, S-EITU was expected to cause a more intense vasoconstriction under septic conditions. However, the effects of S-EITU on hemodynamics and regional blood flows were comparable in both septic and healthy sheep, suggesting either that S-EITU does not selectively inhibit iNOS or that other mediators besides NO play a significant role in septic vasodilation in sheep. Employing different selective iNOS inhibitors in an animal model of porcine endotoxemia also yielded controversial results. Administration of mercaptoethylguanidine (MEG) decreased the amount of expired NO and prevented the fall

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in systemic blood pressure without affecting cardiac output, but failed to improve the disturbances in hepatosplanchnic metabolism (24).

In contrast, the application of N-[3-(aminomethyl) benzyl] acetamide hydrochloride (1400W) reversed the endotoxin-associated disturbances in the systemic circulation and attenuated the impairment of intestinal and hepatocellular oxygenation and energy state (23, 35). Matejovic *et al.* (36) investigated the selective iNOS inhibitor L-N6-(1-iminoethyl)-lysine (L-NIL) in a model of continuous *Pseudomonas aeruginosa* infusion in pigs. Besides an inhibition of plasma nitrate/nitrite (NOx) levels and a stabilization of systemic hemodynamics, the authors observed several beneficial effects of L-NIL on hepatosplanchnic metabolism, including a mitigation of the sepsis-associated impairment of hepatosplanchnic redox state and liver lactate clearance, as well as an attenuation of mesenteric and hepatic venous acidosis.

Recently, an ovine model has been developed that induces sepsis by instillation of live *Pseudomonas aeruginosa* bacteria into the airways following acute lung injury by smoke inhalation (37, 38). This large animal model resembles the pathophysiological conditions of hyperdynamic sepsis in humans more closely than models of continuous intravenous endotoxin or bacteria infusion and may, therefore, provide further insight into the role of increased NO formation and the value of selective pharmacological NOS inhibition. However, while the administration of aminoguanidine, a specific inhibitor of iNOS, significantly inhibited the increase in plasma NOx concentrations in this model, it failed to prevent the drop in mean arterial pressure and pulmonary gas exchange and pulmonary shunt fraction. Likewise, the increases in lung wet-to-dry weight ratio and bronchial blood flow were not inhibited by aminoguanidine (38). It remains unclear why aminoguanidine was effective in reversing the endotoxin-induced changes in sheep, as reported by Evgenov *et al.* (39), but this may be related to the differences in the two animal models. Endotoxin infusion in sheep produces an acute and severe response, although the animals usually recover spontaneously after discontinuation of infusion. Instillation of live bacteria into the lungs produces a subacute response, likely with a different pathophysiology. In the same ovine model of sepsis following acute lung injury, Enkhbaatar *et al.* (40) investigated the effects of BBS-2, a newer and more potent selective iNOS inhibitor. Although BBS-2 significantly improved the pulmonary gas exchange and partially attenuated airway obstruction and increased ventilatory pressures, lung water content (lung wet-to-dry weight ratio) was not affected and septic vasodilation could not be reversed.

These results indicate that increased iNOS expression is only partially responsible for the pathophysiological alterations in sepsis induced by smoke inhalation and bacterial instillation in the airway in sheep. On the other hand, nonselective NOS inhibition improved those changes (38), suggesting that constitutively-produced NOS may be more critically involved in the septic process. Neuronal NOS is a constitutively expressed isoform of

NOS and its activation is regulated by the intracellular concentration of calcium. It is present in both the central and peripheral nervous system (41). The presence of nNOS in the airway epithelium, airway smooth muscle, submucosal glands, blood vessels, non-adrenergic non-cholinergic nerve endings, and in the airway intrinsic parasympathetic plexus has been described (42). We have also identified nNOS in the airway and goblet cells of the bronchi (43). This is another finding that differentiates large animals including humans from mice and rats since these small animals lack the large mucous secreting cells. The results of these studies led to the hypothesis that nNOS-derived NO in the lung could possibly participate in the pathogenesis of lung injury associated with sepsis. To test this hypothesis, Enkhbaatar *et al.* (44, 45) investigated the effects of 7-nitroindazole (7-NI), a specific inhibitor of nNOS in septic sheep. The administration of 7-NI significantly inhibited the increased plasma NOx levels, suggesting that the up-regulation of NO was, at least in part, due to the nNOS isoenzyme. In contrast to the specific iNOS inhibitor BBS-2, 7-NI significantly attenuated the drop in mean arterial pressure in sheep. Furthermore, nNOS blockade with 7-NI significantly improved the pulmonary gas exchange as well as reductions in lung water content, histological airway obstruction, and airway pressures. The fact that the specific nNOS inhibitor 7-NI reduced all these pathophysiological indices indicates that nNOS-derived NO could be an essential pathogenetic factor. Importantly, 7-NI inhibited the plasma NOx levels especially during the initial 12 hours after induction of sepsis, while the reduction was weaker during the second 12 hour interval after injury. These results suggest that early formation of NO was mainly derived from nNOS; and nNOS may be important in the up-regulation of iNOS during the later course of sepsis. In support of this relationship we found that iNOS mRNA is increased in a model of ARDS and that this increase was attenuated in animals that had been treated with a nNOS inhibitor (unpublished data). This hypothesis agrees with the finding that inhibition of plasma NOx levels by the selective iNOS inhibitor BBS-2 was greater at later time points (40).

Large amounts of NO exert potential cytotoxic and pro-inflammatory effects by reacting with superoxide radicals, yielding reactive nitrogen species such as peroxynitrite. Peroxynitrite exerts a deleterious influence by oxidizing/nitrating/nitrosating various other molecules or decaying and producing even more damaging species such as hydroxyl radicals (3, 46). Nitric oxide-mediated tissue injury may be related to DNA damage and subsequent activation of the nuclear enzyme poly (ADP-ribose) polymerase (PARP) (47, 48). After activation by DNA single-strand breaks, PARP catalyzes ADP-ribose subunits to nuclear proteins. This process depletes intracellular NAD⁺ and reduces the rate of glycolysis, electron transfer and ATP formation. Excessive PARP activation in response to immense oxidant-induced DNA strand breakage causes cell necrosis (47-49). It has been demonstrated that PARP activation can be induced by NO or its toxic products such as peroxynitrite (50) (Figure 1).

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There is good evidence of increased PARP activation and its detrimental effects in various large animal models of sepsis. Administration of the potent PARP inhibitor PJ34 in septic pigs following fecal peritonitis abolished injury-related poly (ADP-ribose) accumulation and formation of nitrotyrosine, a marker of oxidative/nitrative stress. In addition, inhibition of PARP synthesis by PJ34 significantly improved survival and attenuated both sepsis-induced hemodynamic changes as well as cytokine response (51). Murakami *et al.* (52) reported that administration of INO-1001, another PARP inhibitor, improved the acute lung injury induced by smoke inhalation and pneumonia in sheep. INO-1001 treatment attenuated the sepsis-induced worsening of pulmonary gas exchange and pulmonary shunt fraction. Moreover, INO-1001 reduced pulmonary histological injury and attenuated poly (ADP-ribose) formation in the lung.

4. ROLE OF NITRIC OXIDE IN BURN AND INHALATION INJURY

Despite the fact that care of burn victims has significantly improved with the use of broad-spectrum antibiotics, effective fluid resuscitation and early surgical removal of burned tissue, the mortality of burn victims with inhalation injury remains high (53, 54). In these patients, progressive pulmonary dysfunction and cardiovascular failure frequently occur, culminating in multiple organ failure and death.

Pulmonary edema formation after inhalation injury may be caused by critical changes in pulmonary blood flow and alterations in capillary permeability. Especially in patients with concomitant extensive cutaneous burns, vascular hyperpermeability occurs not only at the injured site, but also in regions distant from the injury (55, 56), leading to a fluid shift from the intravascular to the interstitial space. The loss of fluid from the circulation results in hypovolemic shock unless adequate fluid resuscitation is performed (57). The combination of capillary hyperpermeability and fluid resuscitation may lead to an excessive accumulation of fluid in the interstitial space of the lung. The ensuing pulmonary edema formation represents a major source of morbidity and mortality in burn patients (58).

In sheep, bronchial blood flow increases approximately 8-fold after smoke inhalation injury alone (59, 60), and tracheal blood flow increases approximately 20-fold after combined burn and inhalation injury, resulting in an impairment of pulmonary gas exchange and an increase in lung fluid content (61, 62). It has been demonstrated that these changes were all markedly improved by bronchial artery occlusion either by ligation or ethanol injection (63-65), suggesting that bronchial circulation also plays a crucial role in the pathophysiology of lung edema formation that occurs after smoke inhalation injury.

The pathophysiological response to combined smoke and inhalation injury in sheep has been described previously (43, 61, 62, 64-71). Acute lung injury in this

large animal model is characterized by significant increases in transpulmonary fluid flux and lung water content (wet-to-dry weight ratio), as well as significant decreases in PaO₂/FiO₂ (partial arterial O₂ pressure/inspired O₂ fraction) ratio. These changes are associated with the occurrence of marked airway obstruction and increases in ventilatory pressures.

As an essential regulator of vasotonus and microcirculatory blood flow, including vascular permeability (72), NO is thought to be critically involved in the regulation of bronchial blood flow (73). In acute lung injury, however, disturbances of NO synthesis in the lung tissue may, at least in part, account for the observed pathophysiological alterations (Figure 2). It has been demonstrated that human lung epithelium cells express iNOS (74), and that iNOS is up-regulated after burn and smoke inhalation injury in sheep (71).

Plasma NOx levels are known to be significantly increased in sheep exposed to burn and inhalation injury, as compared to uninjured control animals (61, 69). This increase in NOx levels can be eliminated by NOS inhibition (61, 71). Because inhalation injury mainly affects the lung, arginine metabolism in lung tissue has been measured by using the stable isotope (¹⁵N) arginine as a tracer in this ovine model. 24 hours after injury, lung arginine metabolism was markedly increased and could be significantly attenuated by administration of the nonselective NOS inhibitor L-NAME, suggesting that excessive NO may be responsible for the increased arginine metabolism (75). It has also been reported that the NOS enzymes can become uncoupled when arginine levels are reduced resulting in the formation of superoxide and peroxynitrite (76). We reported that the levels of arginine are markedly reduced following burn and inhalation injury but that the restoration of arginine levels reduced the pathophysiology seen with inhalation injury, suggesting that reactive oxygen and nitrogen species may be generated by NOS in this situation (77). To address these issues experiments were carried out in sheep with combined burn and smoke inhalation injury that were treated with vitamin E which scavenges reactive oxygen species. This treatment with tocopherols prevented increases in 3-nitrotyrosine, a peroxynitrite marker, as well as much of the pathophysiology that was noted in untreated subjects with burn and smoke inhalation injury (67, 68).

The effects of different specific iNOS inhibitors on pulmonary function and vascular permeability have been investigated in combined burn and smoke inhalation injury in sheep (61, 71, 78). Inhibition of iNOS in these studies congruently resulted in a significant amelioration of pathologically altered variables, providing evidence that iNOS is a key mediator of pulmonary pathology in this model. Administration of iNOS reversed the impairment of pulmonary gas exchange in addition to reducing pulmonary shunt fraction, tracheal blood flow, lung water content, and lung lymph flow. Furthermore, signs of histologically determined airway obstruction as well as increased ventilatory pressures were significantly attenuated. However, the exact mechanism by which NO formation contributes to the development of acute lung injury has not yet been clarified.

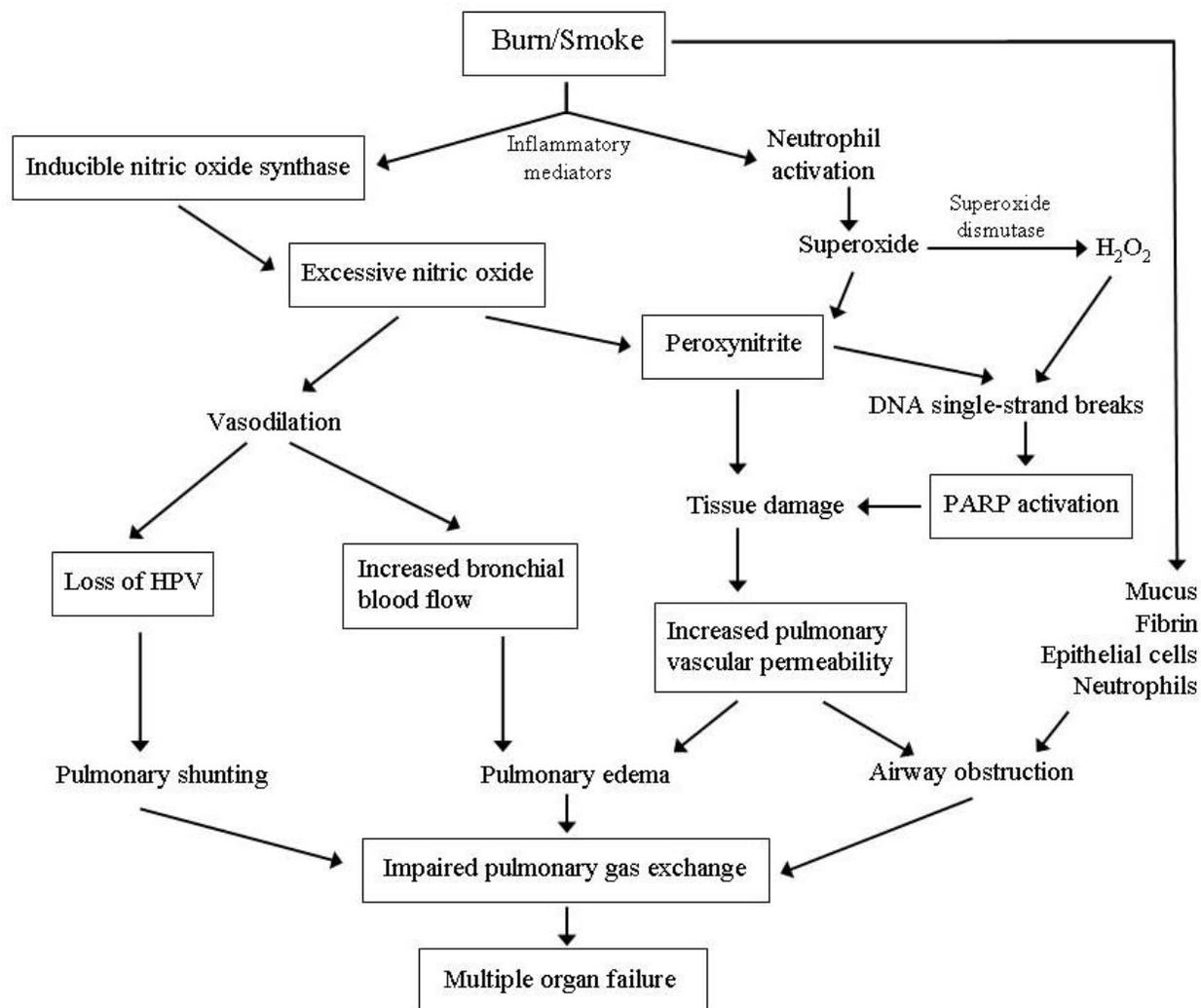


Figure 2. Possible role of nitric oxide in the pathophysiology of burn and inhalation injury. Excessive nitric oxide production by the inducible nitric oxide synthase causes pulmonary vasodilation, leading to the loss of hypoxic pulmonary vasoconstriction (HPV) and increased bronchial blood flow which, in turn, result in pulmonary shunting and pulmonary edema. Both nitric oxide-induced peroxynitrite formation and poly(ADP-ribose) polymerase (PARP) activation cause increased pulmonary vascular permeability. In combination with mucus secretion, fibrin clotting, as well as congregation of neutrophils and epithelial cell debris, airway obstruction occurs. Pulmonary shunting, pulmonary edema and airway obstruction result in impaired pulmonary gas exchange and ultimately multiple organ failure.

Hypoxic pulmonary vasoconstriction (HPV) is a physiologic reflex that matches lung perfusion to ventilation in order to optimize pulmonary gas exchange. Vasoconstriction occurs in under-ventilated, hypoxic areas of the lung, resulting in a diversion of blood flow from the unventilated to ventilated alveoli (79). Combined burn and smoke inhalation injury has recently been demonstrated to impair HPV in sheep (80). Excessive formation of NO may lead to a critical disturbance in pulmonary vasoregulation with a subsequent loss of HPV. This pathomechanism may play an important role in the pulmonary changes following inhalation injury (Figure 2).

As described above, large amounts of NO exert potential pro-inflammatory and cytotoxic effects by

reacting with superoxide radicals to form reactive nitrogen species such as peroxynitrite (81-83). Peroxynitrite may damage the alveolar capillary membrane, resulting in increased pulmonary vascular permeability and edema formation (83). Nitrotyrosine, a marker of peroxynitrite production, is markedly increased in lung tissue of sheep exposed to burn and smoke inhalation injury (80). This increase could be significantly attenuated by selective iNOS inhibition.

Excessive NO production and peroxynitrite formation cause DNA single-strand breakage with subsequent activation of PARP (50). To elucidate the role of PARP activation in burn and inhalation injury, Shimoda *et al.* (69) administered the selective PARP inhibitor INO-

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1001 in an ovine model. INO-1001 attenuated the observed deterioration in pulmonary gas exchange, lung edema formation, increases in airway blood flow and airway pressure, as well as histological lung injury. These findings suggest that PARP is involved in the lung damage caused by combined burn and inhalation injury in sheep (Figure 2).

Additional studies have been performed to assess the effects of selective iNOS inhibition on extrapulmonary co-morbidities in burn and smoke inhalation injury in sheep (66, 70). The injury induced systemic vascular leakage as evidenced by hemoconcentration and increased prefemoral lymph flow, an effect that could be reversed by iNOS inhibition. Moreover, iNOS inhibition attenuated both injury-associated myocardial depression and impaired renal function. These findings establish an important role of iNOS in the pathogenesis of systemic morbidity and multiple organ failure in combined burn and inhalation injury.

5. CONCLUSIONS

Results from large animal models prove the significant role of NO, subsequent peroxynitrite formation and PARP activation in the pathophysiology of shock and organ failure, resulting from both sepsis and combined burn and inhalation injury. Nonselective NOS inhibition has been reported to improve sepsis-related derangements in hemodynamic status, while simultaneously inducing significant adverse effects such as decreases in global oxygen delivery and organ blood flow, as well as increases in pulmonary vascular resistance. These results from large animal models are consistent with data available from clinical studies. Although administration of the nonselective NOS inhibitor 546C88 has been demonstrated to promote the resolution of shock in septic patients (84), the drug actually increased mortality in a recent phase III trial (85). Notably, the protocol of the latter study allowed a more rapid dose escalation of 546C88, resulting in the application of higher doses. These findings suggest that lower doses of a nonselective NOS inhibitor may be more beneficial in human septic shock, possibly even improving survival (85). However, since the only available phase III study in this field demonstrated increased mortality in patients treated with nonselective NOS inhibitors, further clinical trials will be problematic. Future large animal studies may provide valuable information in determining whether lower doses of nonspecific NOS inhibitors improve sepsis-related cardiopulmonary dysfunction with reduced side effects.

Furthermore, it may be crucial to know which NOS isoforms are involved in the pathophysiology of sepsis, as well as their respective time points, when considering possible treatment strategies. It has been generally believed that NO derived from constitutive NOS exerts physiological, regulatory effects, while NO from iNOS is detrimental. This assumption needs to be revised, as recent experimental studies indicate that constitutive NOS isoforms are also involved in the pathophysiology of sepsis (86, 87). However, different isoforms may be increasingly expressed at different time points.

Investigations on specific iNOS and nNOS inhibitors suggest that the early pathophysiological changes in ovine sepsis were induced by NO derived from nNOS, while NO from iNOS expression may account for the derangements in the later course of sepsis. Results from a rat model of sepsis following peritonitis indicate that the administration of a selective iNOS inhibitor improves survival only when given 12 hours after injury (88). The initiation of iNOS inhibition at an earlier time point in the same animal model even increased mortality. Future studies in large animal models are needed to shed light on the time course of different NOS isoenzyme expression in sepsis and the effects of their selective inhibition at different time points. Knowledge from these studies may allow for the development of more differentiated treatment strategies.

Existing evidence about the role of NO in burn and inhalation injury is more conclusive. Specific inhibition of iNOS attenuated acute lung injury without worsening injury-related pulmonary hypertension. However, the long-term effects of iNOS inhibition in burn and smoke inhalation injury have not yet been evaluated. In this regard, it is necessary to keep in mind that inhibition of iNOS may blunt the physiological, bactericidal properties of NO, possibly resulting in suppression of the host defense system with subsequent superinfection of burn tissue. Furthermore, NO physiologically exerts anti-aggregatory effects on different cell types, including platelets. Blunting these properties may increase the risk of blood clotting and embolism. Therefore, further studies should investigate the long-term effects of iNOS inhibition after burn and smoke inhalation injury in large animals.

7. REFERENCES

1. C. Nathan and Q. W. Xie: Nitric oxide synthases: roles, tolls, and controls. *Cell*, 78, 915-918 (1994)
2. M. A. Titheradge: Nitric oxide in septic shock. *Biochim Biophys Acta*, 1411, 437-455 (1999)
3. L. Liaudet, F. G. Soriano and C. Szabo: Biology of nitric oxide signaling. *Crit Care Med*, 28, N37-52 (2000)
4. C. Thiemeermann: Nitric oxide and septic shock. *Gen Pharmacol*, 29, 159-166 (1997)
5. D. C. Angus, W. T. Linde-Zwirble, J. Lidicker, G. Clermont, J. Carcillo and M. R. Pinsky: Epidemiology of severe sepsis in the United States: analysis of incidence, outcome, and associated costs of care. *Crit Care Med*, 29, 1303-1310 (2001)
6. R. C. Bone, W. J. Sibbald and C. L. Sprung: The ACCP-SCCM consensus conference on sepsis and organ failure: editorial; comment. *Chest*, 101, 1481-1483 (1992)
7. C. Ince and M. Sinaasappel: Microcirculatory oxygenation and shunting in sepsis and shock. *Crit Care Med*, 27, 1369-1377 (1999)

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8. D. W. Landry and J. A. Oliver: The pathogenesis of vasodilatory shock. *N Engl J Med*, 345, 588-595 (2001)
9. V. M. Bolotina, S. Najibi, J. J. Palacino, P. J. Pagano and R. A. Cohen: Nitric oxide directly activates calcium-dependent potassium channels in vascular smooth muscle. *Nature*, 368, 850-853 (1994)
10. M. E. Murphy and J. E. Brayden: Nitric oxide hyperpolarizes rabbit mesenteric arteries via ATP-sensitive potassium channels. *J Physiol*, 486 (Pt 1), 47-58 (1995)
11. R. M. Clancy, J. Leszczynska-Piziak and S. B. Abramson: Nitric oxide, an endothelial cell relaxation factor, inhibits neutrophil superoxide anion production via a direct action on the NADPH oxidase. *J Clin Invest*, 90, 1116-1121 (1992)
12. A. G. Gnidec, W. J. Sibbald, H. Cheung and C. A. Metz: Ibuprofen reduces the progression of permeability edema in an animal model of hyperdynamic sepsis. *J Appl Physiol*, 65, 1024-1032 (1988)
13. Q. Sun, G. Dimopoulos, D. N. Nguyen, Z. Tu, N. Nagy, A. D. Hoang, P. Rogiers, D. De Backer and J. L. Vincent: Low-dose vasopressin in the treatment of septic shock in sheep. *Am J Respir Crit Care Med*, 168, 481-486 (2003)
14. D. L. Traber: Animal models: the sheep. *Crit Care Med*, 28, 591-592 (2000)
15. H. G. Bone, R. Waurick, H. Van Aken, M. Booke, T. Prien and J. Meyer: Comparison of the haemodynamic effects of nitric oxide synthase inhibition and nitric oxide scavenging in endotoxaemic sheep. *Intensive Care Med*, 24, 48-54 (1998)
16. F. Hinder, J. Meyer, M. Booke, J. S. Ehardt, J. R. Salsbury, L. D. Traber and D. L. Traber: Endogenous nitric oxide and the pulmonary microvasculature in healthy sheep and during systemic inflammation. *Am J Respir Crit Care Med*, 157, 1542-1549 (1998)
17. F. Hinder, H. D. Stubbe, H. Van Aken, R. Waurick, M. Booke and J. Meyer: Role of nitric oxide in sepsis-associated pulmonary edema. *Am J Respir Crit Care Med*, 159, 252-257 (1999)
18. J. Meyer, M. Booke, R. Waurick, T. Prien and H. Van Aken: Nitric oxide synthase inhibition restores vasopressor effects of norepinephrine in ovine hyperdynamic sepsis. *Anesth Analg*, 83, 1009-1013 (1996)
19. J. Meyer, F. Hinder, J. Stothert, Jr., L. D. Traber, D. N. Herndon, J. T. Flynn and D. L. Traber: Increased organ blood flow in chronic endotoxemia is reversed by nitric oxide synthase inhibition. *J Appl Physiol*, 76, 2785-2793 (1994)
20. J. Meyer, C. W. Lentz, J. C. Stothert, Jr., L. D. Traber, D. N. Herndon and D. L. Traber: Effects of nitric oxide synthesis inhibition in hyperdynamic endotoxemia. *Crit Care Med*, 22, 306-312 (1994)
21. J. Meyer, L. D. Traber, S. Nelson, C. W. Lentz, H. Nakazawa, D. N. Herndon, H. Noda and D. L. Traber: Reversal of hyperdynamic response to continuous endotoxin administration by inhibition of NO synthesis. *J Appl Physiol*, 73, 324-328 (1992)
22. R. Waurick, H. G. Bone, J. Meyer, M. Booke, A. Meissner, T. Prien and H. Van Aken: Haemodynamic effects of dopexamine and nitric oxide synthase inhibition in healthy and endotoxaemic sheep. *Eur J Pharmacol*, 333, 181-186 (1997)
23. M. Matejovic, P. Radermacher, I. Tugtekin, A. Stehr, M. Theisen, J. Vogt, U. Wachter, F. Ploner, M. Georgieff and K. Trager: Effects of selective iNOS inhibition on gut and liver O₂-exchange and energy metabolism during hyperdynamic porcine endotoxemia. *Shock*, 16, 203-210 (2001)
24. F. Ploner, P. Radermacher, M. Theisen, I. F. Tugtekin, M. Matejovic, A. Stehr, C. Szabo, G. J. Southan, M. Georgieff, U. B. Bruckner and K. Trager: Effects of combined selective iNOS inhibition and peroxynitrite blockade during endotoxemia in pigs. *Shock*, 16, 130-136 (2001)
25. B. Santak, P. Radermacher, T. Iber, J. Adler, U. Wachter, D. Vassilev, M. Georgieff and J. Vogt: *In vivo* quantification of endotoxin-induced nitric oxide production in pigs from Na15NO₃-infusion. *Br J Pharmacol*, 122, 1605-1610 (1997)
26. K. Trager, P. Radermacher, K. M. Rieger, R. Grover, A. Vlatten, T. Iber, J. Adler, M. Georgieff and B. Santak: Norepinephrine and N(G)-monomethyl-L-arginine in hyperdynamic septic shock in pigs: effects on intestinal oxygen exchange and energy balance. *Crit Care Med*, 28, 2007-2014 (2000)
27. K. Trager, P. Radermacher, K. M. Rieger, A. Vlatten, J. Vogt, T. Iber, J. Adler, U. Wachter, R. Grover, M. Georgieff and B. Santak: Norepinephrine and nomega-monomethyl-L-arginine in porcine septic shock: effects on hepatic O₂ exchange and energy balance. *Am J Respir Crit Care Med*, 159, 1758-1765 (1999)
28. R. G. Kilbourn, S. S. Gross, A. Jubran, J. Adams, O. W. Griffith, R. Levi and R. F. Lodato: NG-methyl-L-arginine inhibits tumor necrosis factor-induced hypotension: implications for the involvement of nitric oxide. *Proc Natl Acad Sci U S A*, 87, 3629-3632 (1990)
29. J. C. Preiser, H. Zhang, D. Wachel, J. M. Boeynaems, W. Buurman and J. L. Vincent: Is endotoxin-induced hypotension related to nitric oxide formation? *Eur Surg Res*, 26, 10-18 (1994)
30. H. Zhang, P. Rogiers, J. C. Preiser, H. Spapen, P. Manikis, G. Metz and J. L. Vincent: Effects of methylene blue on oxygen availability and regional blood flow during endotoxic shock. *Crit Care Med*, 23, 1711-1721 (1995)

Role of nitric oxide in shock: the large animal perspective

31. H. Zhang, P. Rogiers, N. Smail, A. Cabral, J. C. Preiser, M. O. Peny and J. L. Vincent: Effects of nitric oxide on blood flow distribution and O₂ extraction capabilities during endotoxic shock. *J Appl Physiol*, 83, 1164-1173 (1997)
32. P. J. Offner, H. Ogura, B. S. Jordan, B. A. Pruitt, Jr. and W. G. Cioffi: Cardiopulmonary effects of combined nitric oxide inhibition and inhaled nitric oxide in porcine endotoxic shock. *J Trauma*, 41, 641-646 (1996)
33. H. Ogura, P. J. Offner, D. Saitoh, B. S. Jordan, A. A. Johnson, B. A. Pruitt, Jr. and W. G. Cioffi, Jr.: The pulmonary effect of nitric oxide synthase inhibition following endotoxemia in a swine model. *Arch Surg*, 129, 1233-1239 (1994)
34. M. Booke, F. Hinder, R. McGuire, L. D. Traber and D. L. Traber: Selective inhibition of inducible nitric oxide synthase: effects on hemodynamics and regional blood flow in healthy and septic sheep. *Crit Care Med*, 27, 162-167 (1999)
35. M. Siegemund, J. van Bommel, L. A. Schwarte, W. Studer, T. Girard, S. Marsch, P. Radermacher and C. Ince: Inducible nitric oxide synthase inhibition improves intestinal microcirculatory oxygenation and CO₂ balance during endotoxemia in pigs. *Intensive Care Med*, 31, 985-992 (2005)
36. M. Matejovic, A. Krouzicky, V. Martinkova, R. Rokyta, Jr., H. Kralova, V. Treska, P. Radermacher and I. Novak: Selective inducible nitric oxide synthase inhibition during long-term hyperdynamic porcine bacteremia. *Shock*, 21, 458-465 (2004)
37. K. Murakami, L. J. Bjertnaes, F. C. Schmalstieg, R. McGuire, R. A. Cox, H. K. Hawkins, D. N. Herndon, L. D. Traber and D. L. Traber: A novel animal model of sepsis after acute lung injury in sheep. *Crit Care Med*, 30, 2083-2090 (2002)
38. K. Murakami, R. McGuire, J. Jodoin, J. Katahira, F. C. Schmalstieg, L. D. Traber and D. L. Traber: Aminoguanidine did not attenuate septic changes and acute lung injury in sheep. *The 7th World Congress for Microcirculation*, Suppl, 465-469 (2001)
39. O. V. Evgenov, O. Hevroy, K. E. Bremnes and L. J. Bjertnaes: Effect of aminoguanidine on lung fluid filtration after endotoxin in awake sheep. *Am J Respir Crit Care Med*, 162, 465-470 (2000)
40. P. Enkhbaatar, K. Murakami, L. D. Traber, R. Cox, J. F. Parkinson, M. Westphal, A. Esehie, N. Morita, M. O. Maybauer, D. M. Maybauer, A. S. Burke, F. C. Schmalstieg, H. K. Hawkins, D. N. Herndon and D. L. Traber: The inhibition of inducible nitric oxide synthase in ovine sepsis model. *Shock*, 25, 522-527 (2006)
41. D. S. Bredt: Endogenous nitric oxide synthesis: biological functions and pathophysiology. *Free Radic Res*, 31, 577-596 (1999)
42. A. Fischer and B. Hoffmann: Nitric oxide synthase in neurons and nerve fibers of lower airways and in vagal sensory ganglia of man. Correlation with neuropeptides. *Am J Respir Crit Care Med*, 154, 209-216 (1996)
43. D. L. Traber, H. K. Hawkins, P. Enkhbaatar, R. A. Cox, F. C. Schmalstieg, J. B. Zwischenberger and L. D. Traber: The role of the bronchial circulation in the acute lung injury resulting from burn and smoke inhalation. *Pulm Pharmacol Ther*, 20, 163-166 (2007)
44. P. Enkhbaatar, K. Murakami, K. Shimoda, A. Mizutani, R. McGuire, F. Schmalstieg, R. Cox, H. Hawkins, J. Jodoin, S. Lee, L. Traber, D. Herndon and D. Traber: Inhibition of neuronal nitric oxide synthase by 7-nitroindazole attenuates acute lung injury in an ovine model. *Am J Physiol Regul Integr Comp Physiol*, 285, R366-372 (2003)
45. P. Enkhbaatar and D. Traber: Role of neuronal nitric oxide synthase in cardiopulmonary lesions of sepsis. In: Vincent, J.L. Yearbook of intensive care and emergency medicine 2004. Springer, Berlin (2004)
46. M. N. Hughes: Relationships between nitric oxide, nitroxyl ion, nitrosonium cation and peroxynitrite. *Biochim Biophys Acta*, 1411, 263-272 (1999)
47. L. Virag and C. Szabo: The therapeutic potential of poly(ADP-ribose) polymerase inhibitors. *Pharmacol Rev*, 54, 375-429 (2002)
48. J. Zhang, V. L. Dawson, T. M. Dawson and S. H. Snyder: Nitric oxide activation of poly(ADP-ribose) synthetase in neurotoxicity. *Science*, 263, 687-689 (1994)
49. C. Szabo, S. Cuzzocrea, B. Zingarelli, M. O'Connor and A. L. Salzman: Endothelial dysfunction in a rat model of endotoxic shock. Importance of the activation of poly(ADP-ribose) synthetase by peroxynitrite. *J Clin Invest*, 100, 723-735 (1997)
50. C. Szabo: DNA strand breakage and activation of poly-ADP ribosyltransferase: a cytotoxic pathway triggered by peroxynitrite. *Free Radic Biol Med*, 21, 855-869 (1996)
51. R. D. Goldfarb and C. Szabo: Free radical scavenging as a therapeutic strategy for bacteremia. *Crit Care Med*, 33, 1163-1166 (2005)
52. K. Murakami, P. Enkhbaatar, K. Shimoda, R. A. Cox, A. S. Burke, H. K. Hawkins, L. D. Traber, F. C. Schmalstieg, A. L. Salzman, J. G. Mabley, K. Komjati, P. Pacher, Z. Zsengeller, C. Szabo and D. L. Traber: Inhibition of poly(ADP-ribose) polymerase attenuates acute lung injury in an ovine model of sepsis. *Shock*, 21, 126-133 (2004)
53. R. E. Barrow, M. Spies, L. N. Barrow and D. N. Herndon: Influence of demographics and inhalation injury on burn mortality in children. *Burns*, 30, 72-77 (2004)

Role of nitric oxide in shock: the large animal perspective

54. D. N. Herndon and M. Spies: Modern burn care. *Semin Pediatr Surg*, 10, 28-31 (2001)
55. B. D. Bowen, J. L. Bert, X. Gu, T. Lund and R. K. Reed: Microvascular exchange during burn injury: III. Implications of the model. *Circ Shock*, 28, 221-233 (1989)
56. T. Lund, J. L. Bert, H. Onarheim, B. D. Bowen and R. K. Reed: Microvascular exchange during burn injury. I: A review. *Circ Shock*, 28, 179-197 (1989)
57. D. N. Herndon, J. G. Hilton, D. L. Traber and R. E. Barrow: Burn shock and its resuscitation. *Prog Clin Biol Res*, 236A, 539-557 (1987)
58. D. N. Herndon, R. E. Barrow, D. L. Traber, T. C. Rutan, R. L. Rutan and S. Abston: Extravascular lung water changes following smoke inhalation and massive burn injury. *Surgery*, 102, 341-349 (1987)
59. S. Abdi, D. N. Herndon, L. D. Traber, K. D. Ashley, J. C. Stothert, Jr., J. Maguire, R. Butler and D. L. Traber: Lung edema formation following inhalation injury: role of the bronchial blood flow. *J Appl Physiol*, 71, 727-734 (1991)
60. J. C. Stothert, Jr., K. D. Ashley, G. C. Kramer, D. N. Herndon, L. D. Traber, K. Deubel-Ashley and D. L. Traber: Intrapulmonary distribution of bronchial blood flow after moderate smoke inhalation. *J Appl Physiol*, 69, 1734-1739 (1990)
61. P. Enkhbaatar, K. Murakami, K. Shimoda, A. Mizutani, L. Traber, G. B. Phillips, J. F. Parkinson, R. Cox, H. Hawkins, D. Herndon and D. Traber: The inducible nitric oxide synthase inhibitor BBS-2 prevents acute lung injury in sheep after burn and smoke inhalation injury. *Am J Respir Crit Care Med*, 167, 1021-1026 (2003)
62. K. Soejima, F. C. Schmalstieg, H. Sakurai, L. D. Traber and D. L. Traber: Pathophysiological analysis of combined burn and smoke inhalation injuries in sheep. *Am J Physiol Lung Cell Mol Physiol*, 280, L1233-1241 (2001)
63. O. Efimova, A. B. Volokhov, S. Iliaifar and C. A. Hales: Ligation of the bronchial artery in sheep attenuates early pulmonary changes following exposure to smoke. *J Appl Physiol*, 88, 888-893 (2000)
64. H. Sakurai, R. Johnigan, Y. Kikuchi, M. Harada, L. D. Traber and D. L. Traber: Effect of reduced bronchial circulation on lung fluid flux after smoke inhalation in sheep. *J Appl Physiol*, 84, 980-986 (1998)
65. H. Sakurai, K. Soejima, M. Nozaki, L. D. Traber and D. L. Traber: Effect of ablated airway blood flow on systemic and pulmonary microvascular permeability after smoke inhalation in sheep. *Burns* (2007)
66. P. Enkhbaatar, K. Murakami, K. Shimoda, A. Mizutani, L. Traber, G. Phillips, J. Parkinson, J. R. Salisbury, N. Biondo, F. Schmalstieg, A. Burke, R. Cox, H. Hawkins, D. Herndon and D. Traber: Inducible nitric oxide synthase dimerization inhibitor prevents cardiovascular and renal morbidity in sheep with combined burn and smoke inhalation injury. *Am J Physiol Heart Circ Physiol*, 285, H2430-2436 (2003)
67. N. Morita, K. Shimoda, M. G. Traber, M. Westphal, P. Enkhbaatar, K. Murakami, S. W. Leonard, L. D. Traber and D. L. Traber: Vitamin E attenuates acute lung injury in sheep with burn and smoke inhalation injury. *Redox Rep*, 11, 61-70 (2006)
68. N. Morita, M. G. Traber, P. Enkhbaatar, M. Westphal, K. Murakami, S. W. Leonard, R. A. Cox, H. K. Hawkins, D. Herndon, L. D. Traber and D. L. Traber: Aerosolized alpha-tocopherol ameliorates acute lung injury following combined burn and smoke inhalation injury in sheep. *Shock*, 25, 277-282 (2006)
69. K. Shimoda, K. Murakami, P. Enkhbaatar, L. D. Traber, R. A. Cox, H. K. Hawkins, F. C. Schmalstieg, K. Komjati, J. G. Mabley, C. Szabo, A. L. Salzman and D. L. Traber: Effect of poly(ADP ribose) synthetase inhibition on burn and smoke inhalation injury in sheep. *Am J Physiol Lung Cell Mol Physiol*, 285, L240-249 (2003)
70. K. Soejima, F. C. Schmalstieg, L. D. Traber, C. Szabo, A. Salzman and D. L. Traber: Role of nitric oxide in myocardial dysfunction after combined burn and smoke inhalation injury. *Burns*, 27, 809-815 (2001)
71. K. Soejima, L. D. Traber, F. C. Schmalstieg, H. Hawkins, J. M. Jodoin, C. Szabo, E. Szabo, L. Virag, A. Salzman and D. L. Traber: Role of nitric oxide in vascular permeability after combined burns and smoke inhalation injury. *Am J Respir Crit Care Med*, 163, 745-752 (2001)
72. S. Moncada and A. Higgs: The L-arginine-nitric oxide pathway. *N Engl J Med*, 329, 2002-2012 (1993)
73. P. Carvalho, W. H. Thompson and N. B. Charan: Comparative effects of alpha-receptor stimulation and nitric oxide inhibition on bronchovascular tone. *J Appl Physiol*, 88, 1685-1689 (2000)
74. D. N. Watkins, D. J. Peroni, K. A. Basclain, M. J. Garlepp and P. J. Thompson: Expression and activity of nitric oxide synthases in human airway epithelium. *Am J Respir Cell Mol Biol*, 16, 629-639 (1997)
75. K. Murakami and D. L. Traber: Pathophysiological basis of smoke inhalation injury. *News Physiol Sci*, 18, 125-129 (2003)
76. Y. Xia and J. L. Zweier: Superoxide and peroxynitrite generation from inducible nitric oxide synthase in macrophages. *Proc Natl Acad Sci U S A*, 94, 6954-6958 (1997)
77. K. Murakami, P. Enkhbaatar, Y. M. Yu, L. D. Traber, R. A. Cox, H. K. Hawkins, R. G. Tompkins, D. Herndon and D. L. Traber: L-arginine attenuates acute lung injury after smoke inhalation and burn injury in sheep. *Shock* (2007)

Role of nitric oxide in shock: the large animal perspective

78. K. Soejima, R. McGuire, N. t. Snyder, T. Uchida, C. Szabo, A. Salzman, L. D. Traber and D. L. Traber: The effect of inducible nitric oxide synthase (iNOS) inhibition on smoke inhalation injury in sheep. *Shock*, 13, 261-266 (2000)

79. N. F. Voelkel: Mechanisms of hypoxic pulmonary vasoconstriction. [Review]. *Am Rev Respir Dis*, 133, 1186-1195 (1986)

80. M. Westphal, R. A. Cox, L. D. Traber, N. Morita, P. Enkhbaatar, F. C. Schmalstieg, H. K. Hawkins, D. M. Maybauer, M. O. Maybauer, K. Murakami, A. S. Burke, B. Westphal-Varghese, H. E. Rudloff, J. R. Salsbury, J. M. Jodoin, S. Lee and D. L. Traber: Combined burn and smoke inhalation injury impairs ovine hypoxic pulmonary vasoconstriction. *Crit Care Med*, 34, 1428-1436 (2006)

81. N. W. Kooy, J. A. Royall, Y. Z. Ye, D. R. Kelly and J. S. Beckman: Evidence for *in vivo* peroxynitrite production in human acute lung injury. *Am J Respir Crit Care Med*, 151, 1250-1254 (1995)

82. P. Pacher, J. S. Beckman and L. Liaudet: Nitric oxide and peroxynitrite in health and disease. *Physiol Rev*, 87, 315-424 (2007)

83. C. Szabo: The pathophysiological role of peroxynitrite in shock, inflammation, and ischemia-reperfusion injury. *Shock*, 6, 79-88 (1996)

84. J. Bakker, R. Grover, A. McLuckie, L. Holzapfel, J. Andersson, R. Lodato, D. Watson, S. Grossman, J. Donaldson and J. Takala: Administration of the nitric oxide synthase inhibitor NG-methyl-L-arginine hydrochloride (546C88) by intravenous infusion for up to 72 hours can promote the resolution of shock in patients with severe sepsis: results of a randomized, double-blind, placebo-controlled multicenter study (study no. 144-002). *Crit Care Med*, 32, 1-12 (2004)

85. A. Lopez, J. A. Lorente, J. Steingrub, J. Bakker, A. McLuckie, S. Willatts, M. Brockway, A. Anzueto, L. Holzapfel, D. Breen, M. S. Silverman, J. Takala, J. Donaldson, C. Arneson, G. Grove, S. Grossman and R. Grover: Multiple-center, randomized, placebo-controlled, double-blind study of the nitric oxide synthase inhibitor 546C88: effect on survival in patients with septic shock. *Crit Care Med*, 32, 21-30 (2004)

86. L. Connelly, M. Madhani and A. J. Hobbs: Resistance to endotoxic shock in endothelial nitric-oxide synthase (eNOS) knock-out mice: a pro-inflammatory role for eNOS-derived no *in vivo*. *J Biol Chem*, 280, 10040-10046 (2005)

87. N. C. Gocan, J. A. Scott and K. Tym: Nitric oxide produced via neuronal NOS may impair vasodilatation in septic rat skeletal muscle. *Am J Physiol Heart Circ Physiol*, 278, H1480-1489 (2000)

88. I. Okamoto, M. Abe, K. Shibata, N. Shimizu, N. Sakata, T. Katsuragi and K. Tanaka: Evaluating the role of inducible nitric oxide synthase using a novel and selective inducible nitric oxide synthase inhibitor in septic lung injury produced by cecal ligation and puncture. *Am J Respir Crit Care Med*, 162, 716-722 (2000)

Abbreviations: NO: nitric oxide; NOS: nitric oxide synthase; eNOS: endothelial NOS; nNOS: neuronal NOS; iNOS: inducible NOS; IL-1: interleukin 1; IL-2: interleukin 2; IFN: interferon gamma; TNF: tumor necrosis factor alpha; L-NAME: L-nitro-arginine-methylester; L-NMMA: N-monomethyl-L-arginine; S-EITU: S-ethylisothiourea; MEG: mercaptoethylguanidine; L-NIL: L-N6-(1-iminoethyl)-lysine; NOx: nitrate/nitrite; 7-NI: 7-nitroindazole; DNA: desoxyribo-nucleic acid; PARP: poly(ADP-ribose) polymerase; NAD: nicotinamide adenine dinucleotide; ATP: adenosine triphosphate; HPV: hypoxic pulmonary vasoconstriction

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