The role of death receptors in neural injury

Corina Lorz^{1,2}, Huseyin Mehmet^{1,3}

¹Weston Laboratory, Institute of Reproductive and Developmental Biology, Imperial College London, Hammersmith Hospital Campus, London, UK, ²Current address: Basic Research Department, Epithelial Biomedicine Division, Molecular Oncology Unit, Ciemat, Madrid, Spain, ³ Current address: Merck Research Laboratories, Rahway, USA

TABLE OF CONTENTS

1. Abstract

- 2. Introduction
- 3. Apoptosis and CNS disease
- 4. Role of death receptors in brain injury and disease
 - 4.1. Fas (CD95)
 - 4.2. Tumour necrosis factor receptor (TNFR)
 - 4.3. The p75 neurotrophin receptor (p75NTR)
 - 4.4. TNF-related apoptosis-inducing ligand receptors (TRAIL-R)
- 5. Perspectives
- 6. References

1. ABSTRACT

Programmed cell death is an essential process in the development of the central nervous system (CNS) and is fundamental for the control of the final number of neurons and glial cells. Excessive cell death has been implicated in a growing number of neurodegenerative diseases, such as Alzheimer's, Parkinson's, and multiple sclerosis as well as ischemic injury. We review the contribution of death receptors of the tumor necrosis factor (TNF)/nerve growth factor (NGF) family to cell death and survival in the context of CNS pathology, indicating the possible value of manipulating cell death induced by these receptors for the treatment of CNS diseases and injury.

2. INTRODUCTION

Programmed cell death is a key process during embryonic development and in the maintenance of tissue homeostasis. It is a mechanism that removes unwanted or damaged cells without triggering an inflammatory response. This highly conserved mechanism by which eukaryotic cells die follows a tightly regulated series of molecular events collectively called apoptosis. Any imbalance in the rate of apoptosis can result in disorders characterised by either excessive (e.g. neoplasia) or insufficient (e.g. neurodegenerative diseases) cell numbers.

In mammalian cells, apoptosis can be induced by the ligation of plasma membrane death receptors (the 'extrinsic' pathway) or by the perturbation of intracellular homeostasis (the 'intrinsic' pathway). In the latter case, cell organelles can act as sensors capable of detecting cell injury and activating the apoptotic machinery. Mitochondria are the best-characterised organelles known to trigger apoptosis (1) and not only participate in the initiation of cell death by the 'intrinsic' pathway, but also are required in some cell types to amplify the apoptotic signal triggered by death receptors (2, 3). Mitochondrial regulation of cell death involves the interaction among pro and anti apoptotic Bcl-2 family members, release of cytochrome c to the cytosol and apoptosome formation. Eventually, both pathways culminate in the activation by cleavage of a cascade of proteases, called caspases that are central activators and effectors of apoptosis.

Apoptosis is an essential process in the development of the central nervous system (CNS) and is fundamental for the control of the final number of neurons and glial cells. About half of all the neurons produced during development die before the completion of CNS maturation by this process of naturally occurring cell death. Two waves of apoptotic cell death affect CNS neurons at different stages of embryonic life. The first wave consists of the death of proliferating precursors and young postmitotic neuroblasts that appears to be independent from synaptogenesis and is closely linked to cell cycle regulation (4, 5). The second wave affects postmitotic neurons and glia, and is largely due to competition for limiting amounts of survival signals. While the late apoptotic death of postmitotic neurons is triggered through the mitochondrial pathway, death receptor activation may also be involved (6).

3. APOPTOSIS AND CNS DISEASE

Excessive cell death has been implicated in a growing number of neurodegenerative diseases. For example, an excessive rate of apoptosis occurs in Alzheimer's disease, Parkinson's disease, and multiple sclerosis (7).

Alzheimer's disease (AD) is a neurodegenerative disorder resulting from the progressive loss of neurons in areas critical for learning and memory. It is the most common neurodegenerative disease in the developed world and affects 4 million people in the United States alone. It seems to be caused by abnormal protein deposits accumulating in the brain, although inflammatory cytokines have also been implicated as important factors in the progression of neuronal damage. Pathological signs of apoptosis such as DNA fragmentation and cleavage of caspases have been detected in post-mortem human AD brains (8-10). A small protein called amyloid beta (AB), which originates from the γ -secretase-mediated processing of amyloid precursor protein (APP), is the major component of pathological plaques found in the brains of AD patients and it is capable of inducing apoptosis in cultured neurons (11). A β accumulation triggers caspase activation, leading to caspase-cleavage of the cytoskeletal protein tau that accumulates during AD (12). Also, APP can be cleaved by caspases at sites different to the classic secretase-processing sites, releasing a carboxy-terminal peptide that is a potent inducer of apoptosis (13). These studies suggest that therapeutics aimed at inhibiting tau and APP caspase-cleavage may prove beneficial in slowing AD cognitive decline. APP itself functions as a cell surface receptor (14), and mutations of this protein found in familial AD or its over expression can cause APP to signal aberrant neuronal DNA synthesis and apoptosis (15).

Parkinson's disease (PD) is the second most common neurodegenerative disease, after AD. Although

the aetiology of PD is unknown, the clinical symptoms are attributed to a deficiency in the neurotransmitter dopamine resulting from the selective loss of dopamine-producing neurons from the substantia nigra area of the brain. Approximately 5% of PD cases are familial, while the rest are sporadic and age related. Morphological hallmarks for apoptosis have been reported in dopaminergic neurons in post-mortem PD tissue (16, 17). Similarly, the proportion of cleaved caspase 3-positive dopaminergic neurons is significantly higher in PD patients and cleavage of caspase 8 and caspase 9 has also been reported (18-21).

Multiple sclerosis (MS) is a progressive inflammatory demyelinating disease of the CNS that affects over a million patients worldwide. The aetiology of MS is unclear, although current evidence suggests that a combination of viral and autoimmune factors may be involved. MS is characterised by multifocal areas of demyelination within the CNS, and oligodendrocytes (the myelin forming cells of the CNS) appear to be the primary target. There is a component of axonal loss, which is likely to be secondary to loss of trophic support from the myelin and may be the pathological correlate of the progressive neurological impairment associated to this disease (22). Oligodendrocyte (OL) progenitors are present in the normal adult human CNS and in MS lesions but fail to repair demyelinated regions (23). OL loss in MS tissue has been ascribed to both apoptotic and necrotic cell death (24, 25).

Ischemic stroke is the third most common cause of death and disability in the Western world. During cerebral ischemia, there is a gradient in the severity of hypoperfusion and in cerebral energy failure. In the core region of the infarction, blood flow may be close to zero and cell death is mainly necrotic. In the areas surrounding the core, the so-called ischemic penumbra, where the injury is less severe, the neurons suffer transiently reversible damage, and then ultimately undergo death by apoptosis (26). There is evidence of caspase 3 activity in ischaemic brain tissue (27, 28). During the first few hours, reversible neuronal injury of the ischemic penumbra offers an opportunity for therapeutic intervention and, in this context, caspase inhibitors significantly attenuate neuronal loss following stroke (29-31). Cerebral ischemia in newborn infants is a major cause of cerebral palsy, epilepsy, and mental retardation. After the initial transient ischemic insult, cell death by apoptosis accumulates resulting in a permanent loss of neural cells (32). In vivo studies have shown that the point at which cells are committed to apoptosis has not been reached by 6 hours after ischemia, and up to this point, cell death can be reduced with mild hypothermia (33, 34). Indeed, the results of a recent clinical trial have shown that head cooling can improve both survival and neurodevelopmental outcome in infants with neonatal encephalopathy (34, 35).

4. ROLE OF DEATH RECEPTORS IN BRAIN INJURY AND DISEASE

A total of eight human death receptors have been identified (36) (Figure 1), but the ones that have been shown to be most relevant to the CNS are Fas (CD95 or

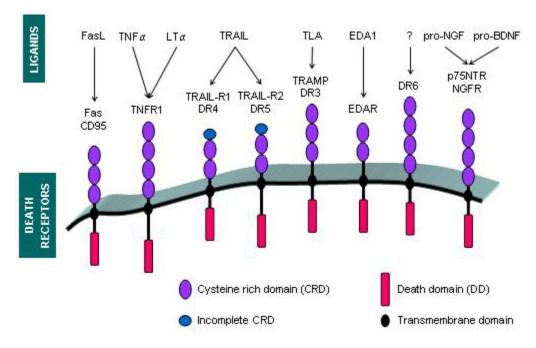


Figure 1. Apoptosis is induced by a subgroup of the tumor necrosis factor (TNF) receptor superfamily, the so called death receptors. Eight human death receptors are known to date. They are typically type I or type II transmembrane proteins that share a conserved 80 amino acid sequence, the death domain, in their cytoplasmic tail, and contain two to four cysteine rich extracellular domains that are involved in ligand binding. The known ligands for these receptors are denoted.

Apo-1), tumour necrosis factor receptor type 1 (TNFR1), the neurotrophin co-receptor p75 (p75NTR) and the TNF-related apoptosis-inducing ligand (TRAIL)receptors 1 (TRAIL-R1 or DR4) and 2 (TRAIL-R2 or DR5). Death receptors are all type I or type II transmembrane proteins with a cytoplasmic 'Death Domain' (DD) motif that couples the cell surface protein to intracellular signalling cascades. They are generally activated by ligand-mediated oligomerisation and some members of the family, such as TNFR1 and p75NTR, elicit a broad spectrum of activities in addition to triggering apoptosis. These include cell survival, proliferation, differentiation, neurite outgrowth or immune activation depending on the cell type (37, 38). In this review, we will focus on the role of death receptors in the brain during development and injury/disease.

4.1. Fas (CD95)

Perhaps the best characterised death receptor signalling pathway to date is that of Fas/CD95. Binding of FasL to its receptor triggers a conformational change in Fas that recruits the adaptor molecule Fas-associated deathdomain (FADD) to its intracellular death domain (DD). FADD recruits the death effector domain (DED)-containing caspases 8 and 10 to the receptor, forming the deathinducing signalling complex (DISC). Caspases recruited to the DISC proteolytically autoactivate themselves, initiating apoptosis by subsequent cleavage of downstream effector caspases (39) (Figure 2). Fas/CD95 can also induce cell death following recruitment of the receptor interacting protein (RIP) kinase to the DD of the receptor through a pathway that does not require apical caspase activation (40).

A crucial role for Fas on the developing CNS is unlikely, since Fas-deficient (lpr) mice and FasL-deficient (gld) mice show no neurological defects at birth (41, 42). Similarly, no developmental defects can be detected in genetically engineered Fas^{-/-} mice (43). In the two mouse strains, lpr and gld, the most frequently reported neurological effects are attributed to an autoimmune response rather than endogenous defects in Fas-mediated apoptosis because they appear after the onset of autoimmune disease between the second and third months of life (44, 45). Moreover, despite the lack of Fas and FasL expression in neurons, the effect of the gld and lpr mutations on neuronal populations is not significant compared to wild type mice (46). However, defects in downstream apoptosis-related genes result in readily apparent neural defects, mice deficient in caspase 9 die perinatally with a markedly enlarged and malformed cerebrum caused by reduced apoptosis during brain development, and caspase 3 knockout mice display a similar phenotype (47, 48). These data strongly suggest that Fas-mediated apoptosis is either not involved, or can be replaced by alternate mechanisms in the control of neuronal populations during CNS development.

Fas/CD95 has been implicated in protecting neural cells from autoimmune attack by immune privilege. The term "immune privilege" describes the lack of local immune responses within certain tissues. Local expression of FasL is essential for maintaining immune privilege in the eye and the placenta through the deletion of early infiltrating inflammatory cells (49, 50), similarly, FasL has been reported to mediate immune privilege in the testis (51). It follows that any failure in immune privilege might

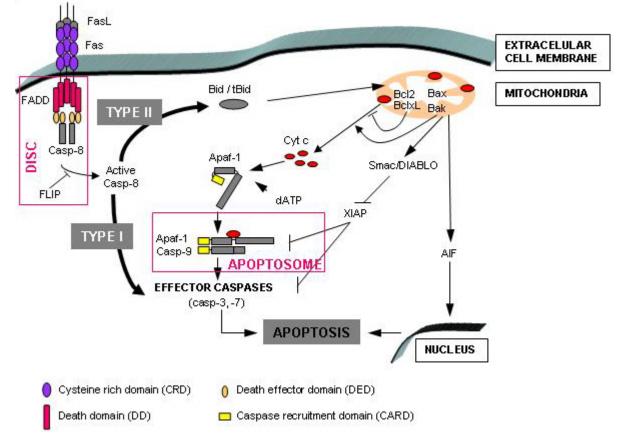


Figure 2. Fas/FasL-triggered apoptosis is the best characterised cell death receptor induced pathway. Upon Fas oligomerisation the adaptor molecule FADD and the pro-form of caspase 8 are sequentially recruited to the intracellular potion of the receptor via DD and DED interactions. The regulator c-FLIP can also be recruited to this complex named death-inducing signaling complex (DISC). The resulting oligomerisation of procaspase 8 results in its autoproteolytic processing and activation. The ensuing apoptotic program kills cells via two different pathways: either active caspase 8 directly cleaves and activates caspase 3 (type I) or caspase 3 cleavage is induced indirectly through cleavage of Bid and activation of the mitochondrial pathway (type II).

lead to inflammatory brain disease. For example, breakdown of the blood-brain barrier is a primary event in MS and other demyelinating diseases. This specialised endothelium is thought to be one of the factors responsible for the relative immune privilege of the CNS (52). In this context, the Fas-FasL system is a double-edge sword, acting as a protective and beneficial machinery against the endogenous immune responses, or as a destructive force in situations of aberrant immunity. FasL is expressed on macrophages, oligodendrocytes, microglia and lymphocytes in MS lesions (53, 54). Soluble FasL can also be recovered from cerebrospinal fluid (CSF) obtained from MS patients (55). Perhaps not surprisingly, it has been found that disruption of the Fas-FasL system improves clinical signs of experimental autoimmune encephalomyelitis (EAE), a widely studied animal model of MS. In both lpr and gld mutations the clinical signs of EAE are ameliorated while neutralisation of FasL during the progression phase of EAE significantly reduced the severity of the disease in wild type animals (56, 57). Paradoxically, when recombinant FasL was infused before EAE onset it suppressed acute EAE; however, injection of specific antibodies against FasL prevented spontaneous remission of EAE (57, 58). Thus, the Fas-FasL system may play a dual role in MS: a protective role against augmented immune responses to self antigens by triggering the (activation-induced) death of infiltrating inflammatory cells (59) and a cytotoxic role during the acute phase of the disease, causing the death of oligodendrocytes. In fact, mice lacking Fas expression in oligodendrocytes are partially protected from EAE (60).

In contrast to the exacerbated immune response in MS, Alzheimer's disease might result from defective immune surveillance mechanisms. Activated T cells, neutrophils, and immunoglobulins are notably absent in the affected areas of the AD brain. Instead, activated microglia are associated with $A\beta$ deposits in AD brains, but seem unable to clear them properly (61). FasL expression is significantly elevated in senile plaques and neurofilamentpositive dystrophic neurites in AD (62), which might explain the scarcity of T-cell infiltration. Increased levels of Fas have been detected in post-mortem AD brains and in the CSF of AD patients (63, 64). Disruption of Fas-FasL signalling by means of a fusion protein consisting of the ligand binding domain of Fas and the Fc domain of IgG (FasFc) has been found to protect primary cortical and cerebellar neurons against AB neurotoxicity (65). Moreover, neurons isolated from gld or lpr mice are also resistant to the apoptotic effects of A β (62). An intriguing finding came from Ethell *et al.* who observed that a broad-spectrum metalloproteinase inhibitor acted synergistically with A β to exacerbate neuronal cell death and this effect was mediated by FasL (66). Metalloproteinases facilitate the processing of APP to produce AB, and regulate the availability of FasL at the cell surface by shedding the Fasbinding ectodomain. While membrane-bound FasL is highly toxic, the released soluble form competes for Fas availability and has anti-apoptotic activity (67). The picture drawn by all these data is difficult to interpret; future research should elucidate the molecular mechanisms of AD and whether FasL plays a central role.

Soluble Fas and FasL have also emerged as candidates for prognostic markers of HIV-associated dementia (68, 69). Despite effective anti-retroviral therapy, about 11% of late-stage HIV-1 patients develop a syndrome of neurological deterioration known as HIV-associated dementia. Neurons are not productively infected by HIV-1, so neuronal injury most likely is the outcome of the microglial activation and the battery of pro-inflammatory and neurotoxic mediators released by activated mononuclear phagocytes. In this sense, activated astrocytes express FasL (70, 71). While neurons express Fas, they also depend on close contact with astrocytes for survival signals. Thus FasL expressed by activated astrocytes is likely to be deleterious to neurons.

Fas signalling has also been implicated in ischemic damage. Both Fas and FasL are up regulated following cerebral hypoxic-ischaemic injury to the developing and adult brain (26, 72). Studies using lpr and gld mice have shown that disruption of the Fas-FasL signalling pathway protects against cerebral ischemia (73). Inflammation plays an important role in brain damage progression after acute stroke, thus, one caveat of studies using gld or lpr mice is that the immune system is grossly dysregulated, and experimental data derived from these mice requires careful interpretation (74). Nevertheless, neutralisation of FasL protects primary cultures of cortical neurons from hypoxia-induced cell death, and FasL neutralising antibodies reduce infarct volume in mice subjected to focal ischaemic injury (75). These data suggest that Fas-FasL is implicated in ischemia-induced neuronal apoptosis, and that Fas-mediated neuronal apoptosis in this situation is deleterious. Similarly, detrimental effects of FasL have been recently described in a model of spinal cord injury, where neutralisation of FasL, but not TNF, promoted both axonal regeneration and functional improvement (76).

4.2. Tumour necrosis factor receptor (TNFR)

TNF exerts its effects through two distinct receptors, TNFR1 and TNFR2, but only TNFR1 has an

intracellular death domain (36). Similar to Fas-FasL interaction, binding of trimeric TNF α to TNFR1 induces receptor cross linking. The first protein recruited to TNFR1 is TNFR1-associated death domain protein (TRADD), which serves as a platform to recruit additional mediators that trigger distinct biological responses. Recruitment of adaptor proteins such as FADD allows the binding and auto-activation of caspase 8, which leads to apoptosis. On the other hand, recruitment of TNFR associating factor 2 (TRAF2) and RIP leads to NF-kB and JNK activation. Once activated, JNKs can phosphorylate transcription factors (e.g. c-Jun) that then transactivate pro-inflammatory and anti-apoptotic genes. In the other arm of this protective pathway, TNFR1 suppression of apoptosis is largely dependent on NF-KB activation, which augments the inflammatory response to TNF (77).

TNF is readily detected in active MS lesions, largely produced by macrophages and microglia (78). TNFR1 expression has been found in OLs in the MS lesions (79) this might explain, at least in part, the demise of the OL lineage in MS pathology (60, 79). In accordance, transgenic over expression of TNF within the CNS results in chronic inflammation and demvelination and converts the acute phase of EAE into a more chronic phenotype (80-82); however, in the absence of TNF, myelin-specific T cells accumulate in the spleen of immunised animals that develop a chronic form of EAE (83). These findings point to a dual role of TNF: despite its harmful pro-inflammatory properties, it may also provide beneficial functions in autoimmune diseases by exerting immune-suppressive effects. This may provide an explanation for augmented immune activation and disease progression evidenced in clinical trials with MS patients treated with anti-TNF agents (84-86). Research using knockout mice for the different TNF receptors has revealed that demvelination and disease severity depend largely on TNFR1, while inmunosuppressive functions of TNF are exerted through both TNF receptors (82, 87-89). It is therefore likely that specific anti-TNFR1 strategies will be more advantageous in the treatment of human immunopathologies than therapies conducted to block TNF activity.

A different scenario may apply to stroke. It is well documented that following cerebral ischemia, TNF expression in the ischemic penumbra increases (90-92). Studies using TNF knockout mice or strategies that neutralise TNF after ischemia have been shown to reduce infarct volume (75, 93), while mice lacking TNFR1, TNFR2 or both, show enhanced ischemic damage (94), thus, an apparent paradox arises from these studies. Interestingly, increased levels of TNF prior to the ischemic insult significantly reduced infarct size in mice (95, 96). This notion is strengthened by data obtained in humans where it has been found that ischemic tolerance in acute stroke is associated with increased plasma TNF levels (97). In summary, there appears to be a protective effect of TNF, possibly mediated by TNFR1 (98), if present before the ischemic injury and a deleterious one after the insult. This has clear implications in the development of TNF-based strategies for stroke treatment.

4.3. The p75 Neurotrophin Receptor (p75NTR)

The low affinity neurotrophin receptor p75 (p75NTR) is involved in several different biological pathways including cell death, survival, proliferation, differentiation. axonal elongation and synaptic transmission. Although p75NTR was among the earliest identified neurotrophin receptors, its biological role and mode of action have proved elusive. Much of the controversy about the role of this receptor stems from the fact that it binds all of the known neurotrophins with similar affinity (therefore its early name low-affinity nerve growth factor receptor). The association of p75NTR with receptors of the trk family (tropomyosin related receptor tyrosine kinases), TrkA, TrkB or TrkC, modulates its binding affinity for neurotrophins (99-101). The p75NTR also associates with the Nogo receptor complex, signalling responses to Nogo, myelin associated glycoprotein (MAG) and myelin oligodendrocyte glycoprotein (MOG) (102,103). In the context of apoptosis, p75NTR seems to mediate cell death in response to neurotrophin binding rather than ligand withdrawal, as suggested by initial p75NTR over expression experiments (104-106). Pro-NGF and pro-BDNF are the most effective inducers of apoptosis via the activation of a receptor complex of p75NTR and sortilin (107-109).

There are two isoforms of the p75NTR, fulllength p75NTR (FL-p75NTR) and a short isoform (sp75NTR) that differ in the number of extracellular cysteine-rich repeats but both contain an intracellular death domain. Recently, a relative of p75NTR designated PLAIDD (p75-like apoptosis-inducing death domain) was also identified (110). Unlike Fas or TNFR, the p75NTR death domain exhibits a type II structure. Multiple adaptor proteins can be recruited to the intracellular domain of p75NTR being responsible for its different biological activities (111). Among these are TNF receptor associated factor 6 (TRAF6), neurotrophin receptor interacting factors 1 and 2 (NRIF 1 and NRIF2), melanoma associated antigen (MAGE), neurotrophin-interacting MAGE homologue (NRAGE or MAGE-D1), Schwann cell factor 1 (SC-1), RhoGDI, protein kinases such as the interleukin-1 receptor associated kinase (IRAK) and the mitogen-activated protein kinases ERK1 and ERK2 and others.

Developmental studies have established a role for p75NTR in programmed cell death associated with neurogenesis. Studies in mice carrying mutations in the NGF and p75NTR genes indicate that a significant amount of the early cell death observed in the developing retina and in the spinal cord is mediated by NGF acting through p75NTR (112). Under certain pathological conditions such as traumatic brain damage, cerebral ischemia, axotomy, AD, and epileptic seizures, p75NTR is re-expressed by CNS cells at levels comparable to those present in early development (113, 114). p75NTR re-expression under pathological conditions, when Trk receptors may be down regulated, suggests that an imbalance of neurotrophin receptor signalling may be involved in some diseases of the nervous system. The fact that p75NTR induces cell death upon binding toxic peptides, such as the neurotoxic prion protein fragment PrP (aa 26-106) and the $A\beta$ peptide (115, 116), reinforce this idea and suggest that research in p75NTR signalling may identify promising drug targets for preventing cell death in some CNS pathologies.

4.4. TNF-related apoptosis-inducing ligand receptors (TRAIL-R)

The TNF-related apoptosis-inducing ligand, TRAIL, is capable of binding four homologous receptor molecules with comparable affinities (117). Two of them, TRAIL-R1 (DR4) and TRAIL-R2 (DR5), induce apoptosis upon ligand binding (118-120). In contrast, the other two receptors are unable to transduce a death signal; TRAIL-R3 (DcR1) lacks the intracellular portion, and TRAIL-R4 (DcR2) has a truncated cytoplasmic DD. These receptors are preferentially expressed on healthy cells and act as "decoys", protecting normal tissues against TRAIL-induced apoptosis (121, 122). The absence of these decoy receptors on tumour cells, together with the expression of functional TRAIL-R1 or -R2 (123), may represent the underlying basis for the sensitivity of many cancer cells to TRAILmediated apoptosis. This has triggered a flurry of research into the use of TRAIL as a chemotherapeutic adjunct molecule. TRAIL itself is not constitutively expressed in adult human brain (124), while the healthy brain and the majority of brain tumours and glioma cell lines co-express TRAIL-R1 and -R2, and the "decoy" TRAIL-R3, suggesting that there might not be a significant role for this system in the apoptotic regulation of brain tumours (125). However, recent studies suggest that brain tumours may be more susceptible to TRAIL-induced apoptosis than was first thought (126-128). Resistance of some neural tumours to TRAIL therapy might be due to the loss of activity of some downstream TRAIL effectors (129, 130). TRAIL-based strategies aiming at enhanced apoptosis of chemotherapy-resistant brain cancer cells have been proposed as new therapeutic alternatives for the treatment of primary brain tumours (131).

The fact that TRAIL selectively induces death of human oligodendrocytes isolated from adult brain and in MS lesions (132, 133), suggests a putative role for TRAIL as an effector molecule in inflammatory or demyelinating diseases such as MS (134). In fact, very recently it has been shown that this is exactly the case. Aktas and co-workers found that TRAIL-deficient myelin-specific lymphocytes induced less severe EAE when transferred to wild-type mice, while intracerebral delivery of TRAIL to animals with EAE increased the clinical deficits (135). These findings suggest that an anti-TRAIL approaches may be useful in the treatment of MS (136).

5. PERPECTIVES

Death receptors of the TNF superfamily are capable of eliciting multiple cell responses that range from proliferation, differentiation, and inflammation to cell death. Since they are differentially expressed during brain development their role in neurogenesis might be expected to differ, and this is indeed the case. So, while mutations in Fas/CD95, or the TNF and TRAIL receptors do not have an overt neural phenotype, p75NTR has been implicated in the NGF-dependent elimination of spinal cord motor neurons.

In contrast to a lack of function in normal brain development, Fas/CD95 and the TNF and TRAIL receptors are strongly implicated in a number of CNS pathologies. The most promising strategies aimed to block the Fas-FasL system are in the context of cerebral ischemia and spinal cord injury. The benefit of blocking TNF in CNS injury is not so straightforward. TNF, through its two receptors, has been implicated in protective immune responses as well as cell death. Because of this dual role, blocking TNF can be either beneficial or harmful depending on the type or stage of injury. In this context, the most promising therapeutic strategies might be those aimed at interfering specifically with one or other of the TNF receptors, especially TNFR1. The study of TRAIL in brain development and pathology is in its infancy and results obtained using TRAIL as an antitumour agent have yielded mixed results in gliomas and neuroblastomas. On the other hand, there is a growing body of evidence for the involvement of TRAIL and other death receptors in CNS inflammation and in demyelinating diseases in particular. Clearly, this is an area or research where further investigation is warranted.

6. REFERENCES

1. K. F. Ferri and G. Kroemer: Organelle-specific initiation of cell death pathways. *Nat Cell Biol*, 3 (11), E255-63 (2001)

2. X. Luo, I. Budihardjo, H. Zou, C. Slaughter and X. Wang: Bid, a Bcl2 interacting protein, mediates cytochrome c release from mitochondria in response to activation of cell surface death receptors. *Cell*, 94 (4), 481-90 (1998)

3. L. Lossi, S. Mioletti and A. Merighi: Synapseindependent and synapse-dependent apoptosis of cerebellar granule cells in postnatal rabbits occur at two subsequent but partly overlapping developmental stages. *Neuroscience*, 112 (3), 509-23 (2002)

4. D. Thomaidou, M. C. Mione, J. F. Cavanagh and J. G. Parnavelas: Apoptosis and its relation to the cell cycle in the developing cerebral cortex. *J Neurosci*, 17 (3), 1075-85 (1997)

5. L. Lossi and A. Merighi: *In vivo* cellular and molecular mechanisms of neuronal apoptosis in the mammalian CNS. *Prog Neurobiol*, 69, 287-312 (2003)

6. M. Vila and S. Przedborski: Targeting programmed cell death in neurodegenerative diseases. *Nat Rev Neurosci*, 4, 365-375 (2003)

7. J. H. Su, A. J. Anderson, B. J. Cummings and C. W. Cotman: Immunohistochemical evidence for apoptosis in Alzheimer's disease. *Neuroreport*, 5, 2529-2533 (1994)

8. L. A. Selznick, D. M. Holtzman, B. H. Han, M. Gokden, A. N. Srinivasan, E. M. J. Johnson and K. A. Roth: *In situ* immunodetection of neuronal caspase-3 activation in Alzheimer disease. *J Neuropathol Exp Neurol*, 58, 1020-1026 (1999) 9. T. T. Rohn, E. Head, W. H. Nesse, C. W. Cotman and D. H. Cribbs: Activation of caspase-8 in the Alzheimer's disease brain. *Neurobiol Dis. 8:*, 8, 1006-1016 (2001)

10. D. T. Loo, A. Copani, C. J. Pike, E. R. Whittemore, A. J. Walencewicz and C. W. Cotman: Apoptosis is induced by beta-amyloid in cultured central nervous system neurons. *Proc Natl Acad Sci USA*, 90, 7951-7955 (1993)

11. R. A. Rissman, W. W. Poon, M. Blurton-Jones, S. Oddo, R. Torp, M. P. Vitek, F. M. LaFerla, T. T. Rohn and C. W. Cotman: Caspase-cleavage of tau is an early event in Alzheimer disease tangle pathology. *J Clin Invest*, 114 (1), 121-30 (2004)

12. D. C. Lu, S. Rabizadeh, S. Chandra, R. F. Shayya, L. M. Ellerby, X. Ye, G. S. Salvesen, E. H. Koo and D. E. Bredesen: A second cytotoxic proteolytic peptide derived from amyloid beta-protein precursor. *Nat Med* 6, 397-404 (2000)

13. E. Brouillet, A. Trembleau, D. Galanaud, M. Volovitch, C. Bouillot, C. Valenza, A. Prochiantz and B. Allinquant: The amyloid precursor protein interacts with Go heterotrimeric protein within a cell compartment specialized in signal transduction. *J Neurosci*, 19 (5), 1717-27 (1999)

14. R. L. Neve and D. L. McPhie: Dysfunction of amyloid precursor protein signaling in neurons leads to DNA synthesis and apoptosis. *Biochim Biophys Acta*, 1772 (4), 430-7 (2007)

15. H. Mochizuki, K. Goto, H. Mori and Y. Mizuno: Histochemical detection of apoptosis in Parkinson's disease. *J Neurol Sci*, 137, 120-123 (1996)

16. P. Anglade, S. Vyas, F. Javoy-Agid, M. T. Herrero, P. P. Michel, J. Marquez, A. Mouatt-Prigent, M. Ruberg, E. C. Hirsch and Y. Agid: Apoptosis and autophagy in nigral neurons of patients with Parkinson's disease. *Histol Histopathol*, 12 (1), 25-31 (1997)

17. N. A. Tatton, A. Maclean-Fraser, W. G. Tatton, D. P. Perl and C. W. Olanow: A fluorescent double-labeling method to detect and confirm apoptotic nuclei in Parkinson's disease. *Ann Neurol*, 44, S142-148 (1998)

18. A. Hartmann, S. Hunot, P. P. Michel, M. P. Muriel, S. Vyas, B. A. Faucheux, A. Mouatt-Prigent, H. Turmel, A. Srinivasan, M. Ruberg, G. I. Evan, Y. Agid and E. C. Hirsch: Caspase-3: A vulnerability factor and final effector in apoptotic death of dopaminergic neurons in Parkinson's disease. *Proc Natl Acad Sci U S A*, 97 (6), 2875-80 (2000)

19. A. Hartmann, J. D. Troadec, S. Hunot, K. Kikly, B. A. Faucheux, A. Mouatt-Prigent, M. Ruberg, Y. Agid and E. C. Hirsch: Caspase-8 is an effector in apoptotic death of dopaminergic neurons in Parkinson's disease, but pathway inhibition results in neuronal necrosis. *J Neurosci*, 21 (7), 2247-55 (2001)

20. V. Viswanath, Y. Wu, R. Boonplueang, S. Chen, F. F. Stevenson, F. Yantiri, L. Yang, M. F. Beal and J. K. Andersen: Caspase-9 activation results in downstream caspase-8 activation and bid cleavage in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinson's disease. *J Neurosci*, 21, 9519-9528 (2001)

21. E. M. Frohman, M. K. Racke and C. S. Raine: Multiple sclerosis--the plaque and its pathogenesis. *N Engl J Med*, 354 (9), 942-55 (2006)

22. G. Wolswijk: Oligodendrocyte survival, loss and birth in lesions of chronic-stage multiple sclerosis. *Brain 123*, 123, 105-115 (2000)

23. B. Bonetti and C. S. Raine: Multiple sclerosis: oligodendrocytes display cell death-related molecules *in situ* but do not undergo apoptosis. *Ann Neurol*, 42, 74-84 (1997)

24. P. Dowling, W. Husar, J. Menonna, H. Donnenfeld, S. Cook and M. Sidhu: Cell death and birth in multiple sclerosis brain. *J Neurol Sci*, 149, 1-11 (1997)

25. T. Sairanen, M. L. Karjalainen-Lindsberg, A. Paetau, P. Ijas and P. J. Lindsberg: Apoptosis dominant in the periinfarct area of human ischaemic stroke--a possible target of antiapoptotic treatments. Brain, 129 (Pt 1), 189-99 (2006)

26. H. Nawashiro, D. Martin and J. M. Hallenbeck: Neuroprotective effects of TNF binding protein in focal cerebral ischemia. Brain Res, 778, 265-271 (1997)

27. S. Love, R. Barber, A. Srinivasan and G. K. Wilcock: Activation of caspase-3 in permanent and transient brain ischaemia in man. *Neuroreport*, 11, 2495-2499 (2000)

28. C. Wiessner, D. Sauer, D. Alaimo and P. R. Allegrini: Protective effect of a caspase inhibitor in models for cerebral ischemia *in vitro* and *in vivo*. *Cell Mol Biol*, 46, 53-62 (2000)

29. M. Rabuffetti, C. Sciorati, G. Tarozzo, E. Clementi, A. A. Manfredi and M. Beltramo: Inhibition of caspase-1-like activity by Ac-Tyr-Val-Ala-Asp-chloromethyl ketone induces long-lasting neuroprotection in cerebral ischemia through apoptosis reduction and decrease of proinflammatory cytokines. *J Neurosci*, 20, 4398-4404 (2000)

30. S. Renolleau, S. Fau, C. Goyenvalle, L. M. Joly, D. Chauvier, E. Jacotot, J. Mariani and C. Charriaut-Marlangue: Specific caspase inhibitor Q-VD-OPh prevents neonatal stroke in P7 rat: a role for gender. *J Neurochem*, 100 (4), 1062-71 (2007)

31. D. L. Taylor, A. Edwards and H. Mehmet: Oxidative metabolism, apoptosis and perinatal brain injury. *Brain Pathol*, 9, 93-117 (1999)

32. C. Zhu, X. Wang, X. Cheng, L. Qiu, F. Xu, G. Simbruner and K. Blomgren: Post-ischemic hypothermiainduced tissue protection and diminished apoptosis after neonatal cerebral hypoxia-ischemia. *Brain Res*, 996, 67-75 (2004)

33. P. D. Gluckman, J. S. Wyatt, D. Azzopardi, R. Ballard, A. D. Edwards, D. M. Ferriero, R. A. Polin, C. M. Robertson, M. Thoresen, A. Whitelaw and A. J. Gunn: Selective head cooling with mild systemic hypothermia after neonatal encephalopathy: multicentre randomised trial. *Lancet*, 365 (9460), 663-70 (2005)

34. J. S. Wyatt, P. D. Gluckman, P. Y. Liu, D. Azzopardi, R. Ballard, A. D. Edwards, D. M. Ferriero, R. A. Polin, C. M. Robertson, M. Thoresen, A. Whitelaw and A. J. Gunn: Determinants of outcomes after head cooling for neonatal encephalopathy. *Pediatrics*, 119 (5), 912-21 (2007)

35. A. Ashkenazi and V. Dixit: Death receptors: signaling and modulation. *Science*, 281, 1305-1308 (1998)

36. V. Baud and M. Karin: Signal transduction by tumor necrosis factor and its relatives. *Trends Cell Biol*, 11, 372-377 (2001)

37. R. M. Locksley, N. Killeen and M. J. Lenardo: The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell*, 104, 487-501 (2001)

38. A. Thorburn: Death receptor-induced cell killing. *Cell Signal*, 16, 139-144 (2004)

39. N. Holler, R. Zaru, O. Micheau, M. Thome, A. Attinger, S. Valitutti, J. L. Bodmer, P. Schneider, B. Seed and J. Tschopp: Fas triggers an alternative, caspase-8-independent cell death pathway using the kinase RIP as effector molecule. *Nat Immunol*, 1 (6), 489-95 (2000)

40. R. Watanabe-Fukunaga, C. I. Brannan, N. G. Copeland, N. Jenkins and S. Nagata: Lymphoproliferation disorder in mice explained by defects in Fas antigen that mediates apoptosis. *Nature*, 356, 314-317 (1992)

41. T. Takahashi, M. Tanaka, C. I. Brannan, N. Jenkins, N. Copeland, T. Suda and S. Nagata: Generalized lymphoproliferative disease in mice, caused by a point mutation in the Fas ligand. *Cell*, 76, 969-976 (1994)

42. M. Adachi, S. Suematsu, T. Kondo, J. Ogasawara, T. Tanaka, N. Yoshida and S. Nagata: Targeted mutation in the Fas gene causes hyperplasia in peripheral lymphoid organs and liver. *Nat Genet*, 11, 294-300 (1995)

43. C. M. Vogelweid, G. C. Johnson, C. L. Besch-Williford, J. Basler and S. E. Walker: Inflammatory central nervous system disease in lupus-prone MRL/lpr mice: comparative histologic and immunohistochemical findings. *J Neuroimmunol*, 35, 89-99 (1991)

44. B. Sakic, B. Kolb, I. Q. Whishaw, G. Gorny, H. Szechtman and J. A. Denburg: Immunosuppression prevents neuronal atrophy in lupus-prone mice: evidence for brain damage induced by autoimmune disease? *J Neuroimmunol*, 111, 93-101 (2000)

45. A. D. Kovac, J. Grammig, J. Mahlo, B. Steiner, K. Roth, R. Nitsch and I. Bechmann: Comparison of neuronal density and subfield sizes in the hippocampus of CD95L-deficient (gld), CD95-deficient (lpr) and nondeficient mice. *Eur J Neurosci*, 16, 159-163 (2002)

46. K. Kuida, T. S. Zheng, S. Na, C. Y. Kuan, D. Yang, H. Karasuyama, P. Rakic and R. A. Flavell: Decreased apoptosis in the brain and premature lethality in CPP32-deficient mice. *Nature*, 384, 368-372 (1996)

47. K. Kuida, T. F. Haydar, C. Y. Kuan, Y. Gu, C. Taya, H. Karasuyama, M. S. Su, P. Rakic and R. A. Flavell: Reduced apoptosis and cytochrome c-mediated caspase activation in mice lacking caspase 9. *Cell*, 94, 325-337 (1998)

48. T. S. Griffith, T. Brunner, S. M. Fletcher, D. R. Green and T. A. Ferguson: Fas ligand-induced apoptosis as a mechanism of immune privilege. Science, 270, 1189-1192 (1995)

49. J. S. Hunt, D. Vassmer, T. A. Ferguson and L. Miller: Fas ligand is positioned in mouse uterus and placenta to prevent trafficking of activated leukocytes between the mother and the conceptus. J Immunol, 158, 4122-4128 (1997)

50. D. Bellgrau, D. Gold, H. Selawry, J. Moore, A. Franzusoff and R. C. Duke: A role for CD95 ligand in preventing graft rejection. *Nature*, 377, 630-632 (1995)

51. K. Wosik, K. Biernacki, M. P. Khouzam and A. Prat: Death receptor expression and function at the human blood brain barrier. *J Neurol Sci*, 259 (1-2), 53-60 (2007)

52. S. D. D'Souza, B. Bonetti, V. Balasingam, N. R. Cashman, P. A. Barker, A. B. Troutt, C. S. Raine and J. P. Antel: Multiple sclerosis: Fas signaling in oligodendrocyte cell death. *J Exp Med* 184, 2361-2370 (1996)

53. P. Dowling, G. Shang, S. Raval, J. Menonna, S. Cook and W. Husar: Involvement of the CD95 (APO-1/Fas) receptor/ligand system in multiple sclerosis brain. *J Exp Med*, 184, 1513-1518 (1996)

54. E. Ciusani, S. Frigerio, M. Gelati, E. Corsini, A. Dufour, A. Nespolo, L. La Mantia, C. Milanese, G. Massa and A. Salmaggi: Soluble Fas (Apo-1) levels in cerebrospinal fluid of multiple sclerosis patients. *J Neuroimmunol* 82, 5-12 (1998)

55. H. Waldner, R. A. Sobel, E. Howard and V. K. Kuchroo: Fas- and FasL-deficient mice are resistant to

induction of autoimmune encephalomyelitis. *J Immunol*, 159 (7), 3100-3 (1997)

56. Y. Okuda, S. Sakoda, H. Fujimura, S. Nagata, T. Yanagihara and C. C. Bernard: Intrathecal administration of neutralizing antibody against Fas ligand suppresses the progression of experimental autoimmune encephalomyelitis. *Biochem Biophys Res Commun*, 275 (1), 164-8 (2000)

57. B. Zhu, L. Luo, Y. Chen, D. W. Paty and M. S. Cynader: Intrathecal Fas ligand infusion strengthens immunoprivilege of central nervous system and suppresses experimental autoimmune encephalomyelitis. *J Immunol*, 169 (3), 1561-9 (2002)

58. Y. Okuda, B. R. Apatoff and D. N. Posnett: Apoptosis of T cells in peripheral blood and cerebrospinal fluid is associated with disease activity of multiple sclerosis. *J Neuroimmunol*, 171 (1-2), 163-70 (2006)

59. N. Hovelmeyer, Z. Hao, K. Kranidioti, G. Kassiotis, T. Buch, F. Frommer, L. von Hoch, D. Kramer, L. Minichiello, G. Kollias, H. Lassmann and A. Waisman: Apoptosis of oligodendrocytes via Fas and TNF-R1 is a key event in the induction of experimental autoimmune encephalomyelitis. *J Immunol*, 175 (9), 5875-84 (2005)

60. W. J. Streit: Microglia and Alzheimer's disease pathogenesis. *J Neurosci Res* 77, 1-8 (2004)

61. J. H. Su, A. J. Anderson, D. H. Cribbs, C. Tu, L. Tong, P. Kesslack and C. W. Cotman: Fas and Fas ligand are associated with neuritic degeneration in the AD brain and participate in beta-amyloid-induced neuronal death. *Neurobiol Dis* 12, 182-193 (2003)

62. M. Martinez, E. Fernandez-Vivancos, A. Frank, M. De la Fuente and A. Hernanz: Increased cerebrospinal fluid fas (Apo-1) levels in Alzheimer's disease. Relationship with IL-6 concentrations. *Brain Res*, 869 (1-2), 216-9 (2000)

63. S. M. de la Monte, Y. K. Sohn and J. R. Wands: Correlates of p53- and Fas (CD95)-mediated apoptosis in Alzheimer's disease. *J Neurol Sci* 152, 73-83 (1997)

64. Y. Morishima, Y. Gotoh, J. Zieg, T. Barrett, H. Takano, R. A. Flavell, R. J. Davis, Y. Shirasaki and M. E. Greenberg: Beta-amyloid induces neuronal apoptosis via a mechanism that involves the c-Jun N-terminal kinase pathway and the induction of Fas ligand. *J Neurosci*, 21, 7551-7560 (2001)

65. D. W. Ethell, R. Kinloch and D. R. Green: Metalloproteinase shedding of Fas ligand regulates betaamyloid neurotoxicity. *Curr Biol* 12, 1595-1600 (2002)

66. M. Tanaka, T. Itai, M. Adachi and S. Nagata: Downregulation of Fas ligand by shedding. *Nat Med* 4, 31-36 (1998)

67. F. Sabri, A. De Milito, R. Pirskanen, I. Elovaara, L. Hagberg, P. Cinque, R. Price and F. Chiodi: Elevated levels of

soluble Fas and Fas ligand in cerebrospinal fluid of patients with AIDS dementia complex. *J Neuroimmunol*, 114 (1-2), 197-206 (2001)

68. A. Towfighi, R. L. Skolasky, C. St Hillaire, K. Conant and J. C. McArthur: CSF soluble Fas correlates with the severity of HIV-associated dementia. *Neurology*, 62 (4), 654-6 (2004)

69. A. Ghorpade, S. Holter, K. Borgmann, R. Persidsky and L. Wu: HIV-1 and IL-1 beta regulate Fas ligand expression in human astrocytes through the NF-kappa B pathway. *J Neuroimmunol*, 141, 141-149 (2003)

70. M. Deshpande, J. Zheng, K. Borgmann, R. Persidsky, L. Wu, C. Schellpeper and A. Ghorpade: Role of activated astrocytes in neuronal damage: potential links to HIV-1-associated dementia. *Neurotox Res*, 7 (3), 183-92 (2005)

71. U. Felderhoff-Mueser, D. L. Taylor, K. Greenwood, M. Kozma, D. Stibenz, U. C. Joashi, A. D. Edwards and H. Mehmet Fas/CD95/APO-1 can function as a death receptor for neuronal cells *in vitro* and *in vivo* and is up regulated following cerebral hypoxic-ischemic injury to the developing rat brain. *Brain Pathol* 10, 17-29 (2000)

72. A. Martin-Villalba, I. Herr, I. Jeremias, M. Hahne, R. Brandt, J. Vogel, J. Schenkel, T. Herdegen and K. M. Debatin: CD95 ligand (Fas-L/APO-1L) and tumor necrosis factor-related apoptosis-inducing ligand mediate ischemia-induced apoptosis in neurons. *J Neurosci* 19, 3809-3817 (1999)

73. H. Mehmet: Stroke treatment enters the Fas lane. *Cell Death Differ* 8, 659-661 (2001)

74. A. Martin-Villalba, M. Hahne, S. Kleber, J. Vogel, W. Falk, J. Schenkel and P. H. Krammer: Therapeutic neutralization of CD95-ligand and TNF attenuates brain damage in stroke. *Cell Death Differ* 8, 679-686 (2001)

75. D. Demjen, S. Klussmann, S. Kleber, C. Zuliani, B. Stieltjes, C. Metzger, U. A. Hirt, H. Walczak, W. Falk, M. Essig, L. Edler, P. H. Krammer and A. Martin-Villalba: Neutralization of CD95 ligand promotes regeneration and functional recovery after spinal cord injury. *Nat Med*, 10 (4), 389-95 (2004)

76. O. Micheau and J. Tschopp: Induction of TNF receptor I-mediated apoptosis via two sequential signaling complexes. *Cell*, 114 (2), 181-90 (2003)

77. K. Selmaj, C. S. Raine, B. Cannella and C. F. Brosnan: Identification of lymphotoxin and tumor necrosis factor in multiple sclerosis lesions. *J Clin Invest*, 87 (3), 949-54 (1991)

78. C. S. Raine, B. Bonetti and B. Cannella: Multiple sclerosis: expression of molecules of the tumor necrosis factor ligand and receptor families in relationship to the demyelinated plaque. *Rev Neurol (Paris)* 154, 577-585 (1998)

79. L. Probert, K. Akassoglou, M. Pasparakis, G. Kontogeorgos and G. Kollias: Spontaneous inflammatory demyelinating disease in transgenic mice showing central nervous system-specific expression of tumor necrosis factor alpha. *Proc Natl Acad Sci USA*, 92, 11294-11298 (1995)

80. L. Probert, K. Akassoglou, G. Kassiotis, M. Pasparakis, L. Alexopoulou and G. Kollias: TNF-alpha transgenic and knockout models of CNS inflammation and degeneration. *J Neuroimmunol*, 72, 137-141 (1997)

81. K. Akassoglou, J. Bauer, G. Kassiotis, M. Pasparakis, H. Lassmann, G. Kollias and L. Probert: Oligodendrocyte apoptosis and primary demyelination induced by local TNF/p55TNF receptor signaling in the central nervous system of transgenic mice: models for multiple sclerosis with primary oligodendrogliopathy. *Am J Pathol* (153), 801-813 (1998)

82. G. Kassiotis and G. Kollias: Uncoupling the proinflammatory from the immunosuppressive properties of tumor necrosis factor (TNF) at the p55 TNF receptor level: implications for pathogenesis and therapy of autoimmune demyelination. *J Exp Med*, 193, 427-434 (2001)

83. B. W. van Oosten, F. Barkhof, L. Truyen, J. B. Boringa, F. W. Bertelsmann, B. M. von Blomberg, J. N. Woody, H. P. Hartung and C. H. Polman: Increased MRI activity and immune activation in two multiple sclerosis patients treated with the monoclonal anti-tumor necrosis factor antibody cA2. *Neurology*, 47, 1531-1534 (1996)

84. TNF neutralization in MS: results of a randomized, placebo-controlled multicenter study. The Lenercept Multiple Sclerosis Study Group and The University of British Columbia MS/MRI Analysis Group. *Neurology*, 53 (3), 457-65 (1999)

85. R. Gold, R. A. Linker and A. Chan: Termination of inflammation in the nervous system. *Rev Neurol (Paris)*, 163 (6-7), 672-6 (2007)

86. K. Akassoglou, E. Douni, J. Bauer, H. Lassmann, G. Kollias and L. Probert: Exclusive tumor necrosis factor (TNF) signaling by the p75TNF receptor triggers inflammatory ischemia in the CNS of transgenic mice. *Proc Natl Acad Sci USA*, 100, 709-714 (2003)

87. H. P. Eugster, K. Frei, R. Bachmann, H. Bluethmann, H. Lassmann and A. Fontana: Severity of symptoms and demyelination in MOG-induced EAE depends on TNFR1. *Eur J Immunol*, 29, 626-632 (1999)

88. G. Kollias and D. Kontoyiannis: Role of TNF/TNFR in autoimmunity: specific TNF receptor blockade may be advantageous to anti-TNF treatments. *Cytokine Growth Factor Rev*, 13, 315-321 (2002)

89. T. Liu, R. K. Clark, P. C. McDonnell, P. R. Young, R. F. White, F. C. Barone and G. Z. Feuerstein: Tumor

necrosis factor-alpha expression in ischemic neurons. Stroke, 25, 1481-1488 (1994)

90. M. Buttini, K. Appel, A. Sauter, P. J. Gebicke-Haerter and H. W. Boddeke: Expression of tumor necrosis factor alpha after focal cerebral ischaemia in the rat. *Neuroscience*, 71, 1-16 (1996)

91. K. Saito, K. Suyama, K. Nishida, Y. Sei and A. S. Basile: Early increases in TNF-alpha, IL-6 and IL-1 beta levels following transient cerebral ischemia in gerbil brain. *Neurosci Lett*, 206, 149-152 (1996)

92. X. Wang, G. Z. Feuerstein, L. Xu, H. Wang, W. A. Schumacher, M. L. Ogletree, R. Taub, J. J. Duan, C. P. Decicco and R. Q. Liu: Inhibition of tumor necrosis factoralpha-converting enzyme by a selective antagonist protects brain from focal ischemic injury in rats. *Mol Pharmacol*, 65, 890-896 (2004)

93. A. J. Bruce, W. Boling, M. S. Kindy, J. Peschon, P. J. Kraemer, M. K. Carpenter, F. W. Holtsberg and M. P. Mattson: Altered neuronal and microglial responses to excitotoxic and ischemic brain injury in mice lacking TNF receptors. *Nat Med* 2, 788-794 (1996)

94. H. Nawashiro, K. Tasaki, C. A. Ruetzler and J. M. Hallenbeck: TNF-alpha pretreatment induces protective effects against focal cerebral ischemia in mice. *J Cereb Blood Flow Metab* 17, 4834-90 (1997)

95. H. L. Rosenzweig, M. Minami, N. S. Lessov, S. C. Coste, S. L. Stevens, D. C. Henshall, R. Meller, R. P. Simon and M. P. Stenzel-Poore: Endotoxin preconditioning protects against the cytotoxic effects of TNFalpha after stroke: a novel role for TNFalpha in LPS-ischemic tolerance. *J Cereb Blood Flow Metab* (2007)

96. J. Castillo, M. A. Moro, M. Blanco, R. Leira, J. Serena, I. Lizasoain and A. Davalos: The release of tumor necrosis factor-alpha is associated with ischemic tolerance in human stroke. *Ann Neurol* 54, 811-819 (2003)

97. E. Taoufik, S. Valable, G. J. Muller, M. L. Roberts, D. Divoux, A. Tinel, A. Voulgari-Kokota, V. Tseveleki, F. Altruda, H. Lassmann, E. Petit and L. Probert: FLIP (L) protects neurons against *in vivo* ischemia and *in vitro* glucose deprivation-induced cell death. *J Neurosci*, 27 (25), 6633-46 (2007)

98. R. Klein, V. Nanduri, S. A. Jing, F. Lamballe, P. Tapley, S. Bryant, C. Cordon-Cardo, K. R. Jones, L. F. Reichardt and M. Barbacid: The trkB tyrosine protein kinase is a receptor for brain-derived neurotrophic factor and neurotrophin-3. *Cell* 66, 395-403 (1991)

99. R. Klein, S. Q. Jing, V. Nanduri, E. O'Rourke and M. Barbacid: The trk proto-oncogene encodes a receptor for nerve growth factor. *Cell*, 65 (1), 189-97 (1991)

100. F. Lamballe, R. Klein and M. Barbacid: TrkC, a new member of the trk family of tyrosine protein kinases, is a receptor for neurotrophin-3. *Cell* 66, 967-979 (1991)

101. K. C. Wang, J. A. Kim, R. Sivasankaran, R. Segal and Z. He: P75 interacts with the Nogo receptor as a coreceptor for Nogo, MAG and OMgp. *Nature* 420, 74-78 (2002)

102. L. Dupuis, M. Pehar, P. Cassina, F. Rene, R. Castellanos, C. Rouaux, M. Gandelman, L. Dimou, M. E. Schwab, J. P. Loeffler, L. Barbeito and J. L. Gonzalez de Aguilar: Nogo receptor antagonizes p75NTR-dependent motor neuron death. *Proc Natl Acad Sci U S A*, 105 (2), 740-5 (2008)

103. S. Rabizadeh, J. Oh, L. T. Zhong, J. Yang, C. M. Bitler, L. L. Butcher and D. E. Bredesen: Induction of apoptosis by the low-affinity NGF receptor. *Science* 16, 345-348 (1993)

104. J. M. Frade, A. Rodriguez-Tebar and Y. A. Barde: Induction of cell death by endogenous nerve growth factor through its p75 receptor. *Nature* 383, 166-168 (1996)

105. P. Casaccia-Bonnefil, B. D. Carter, R. T. Dobrowsky and M. V. Chao: Death of oligodendrocytes mediated by the interaction of nerve growth factor with its receptor p75. *Nature* 383, 716-719 (1996)

106. H. K. Teng, K. K. Teng, R. Lee, S. Wright, S. Tevar, R. D. Almeida, P. Kermani, R. Torkin, Z. Y. Chen, F. S. Lee, R. T. Kraemer, A. Nykjaer and B. L. Hempstead: ProBDNF induces neuronal apoptosis via activation of a receptor complex of p75NTR and sortilin. *J Neurosci* 25, 5455-5463 (2005)

107. A. Nykjaer, R. Lee, K. K. Teng, P. Jansen, P. Madsen, M. S. Nielsen, C. Jacobsen, M. Kliemannel, E. Schwarz, T. E. Willnow, B. L. Hempstead and C. M. Petersen: Sortilin is essential for proNGF-induced neuronal cell death. *Nature*, 427 (6977), 843-8 (2004)

108. P. Jansen, K. Giehl, J. R. Nyengaard, K. Teng, O. Lioubinski, S. S. Sjoegaard, T. Breiderhoff, M. Gotthardt, F. Lin, A. Eilers, C. M. Petersen, G. R. Lewin, B. L. Hempstead, T. E. Willnow and A. Nykjaer: Roles for the pro-neurotrophin receptor sortilin in neuronal development, aging and brain injury. *Nat Neurosci*, 10 (11), 1449-57 (2007)

109. K. C. Kanning, M. Hudson, P. S. Amieux, J. C. Wiley, M. Bothwell and L. C. Schecterson: Proteolytic processing of the p75 neurotrophin receptor and two homologs generates C-terminal fragments with signaling capability. *J Neurosci*, 23 (13), 5425-36 (2003)

110. J. C. Arevalo and S. H. Wu: Neurotrophin signaling: many exciting surprises! *Cell Mol Life Sci*, 63 (13), 1523-37 (2006)

111. J. M. Frade and Y. A. Barde: Genetic evidence for cell death mediated by nerve growth factor and the neurotrophin receptor p75 in the developing mouse retina and spinal cord. *Development* 126, 683-690 (1999)

112. P. Podlesniy, A. Kichev, C. Pedraza, J. Saurat, M. Encinas, B. Perez, I. Ferrer and C. Espinet: Pro-NGF from Alzheimer's disease and normal human brain displays distinctive abilities to induce processing and nuclear translocation of intracellular domain of p75NTR and apoptosis. *Am J Pathol*, 169 (1), 119-31 (2006)

113. N. F. Schor: The p75 neurotrophin receptor in human development and disease. *Prog Neurobiol*, 77 (3), 201-14 (2005)

114. Y. Hashimoto, Y. Kaneko, E. Tsukamoto, H. Frankowski, K. Kouyama, Y. Kita, T. Niikura, S. Aiso, D. E. Bredesen, M. Matsuoka and I. Nishimoto: Molecular characterization of neurohybrid cell death induced by Alzheimer's amyloid-beta peptides via p75NTR/PLAIDD. *J Neurochem*, 90 (3), 549-58 (2004)

115. V. Della-Bianca, F. Rossi, U. Armato, I. Dal-Pra, C. Costantini, G. Perini, V. Politi and G. Della Valle: Neurotrophin p75 receptor is involved in neuronal damage by prion peptide- (106-126). *J Biol Chem* 276, 38929-38933 (2001)

116. P. Schneider, J. L. Bodmer, M. Thome, K. Hofmann, N. Holler and J. Tschopp: Characterization of two receptors for TRAIL. *FEBS Lett* 416, 329-334 (1997)

117. P. Schneider, M. Thome, K. Burns, J. L. Bodmer, K. Hofmann, T. Kataoka, N. Holler and J. Tschopp: TRAIL receptors 1 (DR4) and 2 (DR5) signal FADD-dependent apoptosis and activate NF-kappaB. *Immunity* 7, 831-836 (1997)

118. G. Pan, K. O'Rourke, A. M. Chinnaiyan, R. Gentz, R. Ebner, J. Ni and V. M. Dixit: The receptor for the cytotoxic ligand TRAIL. *Science* 276, 111-113 (1997)

119. P. M. Chaudhary, M. Eby, A. Jasmin, A. Bookwalter, J. Murray and L. Hood: Death receptor 5, a new member of the TNFR family, and DR4 induce FADD-dependent apoptosis and activate the NF-kappaB pathway. *Immunity* 7, 821-830 (1997)

120. G. Pan, J. Ni, Y. F. Wei, G. Yu, R. Gentz and V. M. Dixit: An antagonist decoy receptor and a death domaincontaining receptor for TRAIL. *Science* 277, 815-818 (1997)

121. J. P. Sheridan, S. A. Marsters, R. M. Pitti, A. Gurney, M. Skubatch, D. Baldwin, L. Ramakrishnan, C. L. Gray, K. Baker, W. I. Wood, A. D. Goddard, P. Godowski and A. Ashkenazi: Control of TRAIL-induced apoptosis by a family of signaling and decoy receptors. *Science*, 277 (5327), 818-21 (1997) 122. H. Walczak, R. E. Miller, K. Ariail, B. Gliniak, T. S. Griffith, M. Kubin, W. Chin, J. Jones, A. Woodward, T. Le, C. Smith, 123. P. Smolak, R. G. Goodwin, C. T. Rauch, J. C. Schuh and D. H. Lynch: Tumoricidal activity of tumor necrosis factor-related apoptosis-inducing ligand *in vivo*. *Nat Med*, 5 (2), 157-63 (1999)

124. J. Dorr, I. Bechmann, S. Waiczies, O. Aktas, H. Walczak, P. H. Krammer, R. Nitsch and F. Zipp: Lack of tumor necrosis factor-related apoptosis-inducing ligand but presence of its receptors in the human brain. *J Neurosci*, 22 (4), RC209 (2002)

125. S. Frank, U. Kohler, G. Schackert and H. K. Schackert: Expression of TRAIL and its receptors in human brain tumors. *Biochem Biophys Res Commun* 257, 454-459 (1999)

126. K. Shah, G. Hsich and X. O. Breakefield: Neural precursor cells and their role in neuro-oncology. *Dev Neurosci*, 26, 118-130 (2004)

127. C. J. Hawkins: TRAIL and malignant glioma. *Vitam* Horm 67, 427-452 (2004)

128. J. M. Kuijlen, J. J. Mooij, I. Platteel, E. W. Hoving, W. T. van der Graaf, M. M. Span, H. Hollema and W. F. den Dunnen: TRAIL-receptor expression is an independent prognostic factor for survival in patients with a primary glioblastoma multiforme. *J Neurooncol*, 78 (2), 161-71 (2006)

129. S. Fulda, W. Wick, M. Weller and K. M. Debatin: Smac agonists sensitize for Apo2L/TRAIL- or anticancer drug-induced apoptosis and induce regression of malignant glioma *in vivo*. *Nat Med*, 8 (8), 808-15 (2002)

130. S. Hopkins-Donaldson, J. L. Bodmer, K. B. Bourloud, C. B. Brognara, J. Tschopp and N. Gross: Loss of caspase-8 expression in highly malignant human neuroblastoma cells correlates with resistance to tumor necrosis factorrelated apoptosis-inducing ligand-induced apoptosis. *Cancer Res* 60, 4315-4319 (2000)

131. J. P. Steinbach and M. Weller: Apoptosis in Gliomas: Molecular Mechanisms and Therapeutic Implications. *J Neurooncol*, 70 (2), 247-256 (2004)

132. M. Matysiak, A. Jurewicz, D. Jaskolski and K. Selmaj: TRAIL induces death of human oligodendrocytes isolated from adult brain. *Brain* 125, 2469-2480 (2002)

133. B. Cannella, S. Gaupp, K. M. Omari and C. S. Raine: Multiple sclerosis: Death receptor expression and oligodendrocyte apoptosis in established lesions. *J Neuroimmunol*, 188 (1-2), 128-37 (2007)

134. F. Zipp and O. Aktas: The brain as a target of inflammation: common pathways link inflammatory and neurodegenerative diseases. *Trends Neurosci*, 29 (9), 518-27 (2006)

135. O. Aktas, A. Smorodchenko, S. Brocke, C. Infante-Duarte, U. Schulze Topphoff, J. Vogt, T. Prozorovski, S. Meier, V. Osmanova, E. Pohl, I. Bechmann, R. Nitsch and F. Zipp: Neuronal damage in autoimmune neuroinflammation mediated by the death ligand TRAIL. *Neuron*, 46 (3), 421-32 (2005)

136. O. Aktas, S. Waiczies and F. Zipp: Neurodegeneration in autoimmune demyelination: recent mechanistic insights reveal novel therapeutic targets. *J Neuroimmunol*, 184 (1-2), 17-26 (2007)

Abbreviations: Ab: Amyloid beta, AD: Alzheimer's disease, APP: Amyloid precursor protein, BDNF: Brain derived neurotrophic factor. CNS: Central nervous system. CSF: Cerebrospinal fluid, DD: Death domain, DED: Death effector domain, DISC: Death-inducing signalling EAE: Experimental complex, autoimmune encephalomyelitis, FADD: Fas-associated death domain, MS: Multiple sclerosis, NF-kB: Nuclear factor kappa B, NGF: Nerve growth factor, OL: Oligodendrocyte, PD: Parkinson's disease, p75NTR: Neurotrophin receptor p75, TNF: Tumour necrosis factor, TNFR1: TNF receptor type 1, TNFR2: TNF receptor type 2, TRAIL: TNF-related apoptosis-inducing ligand, TRAIL-R (1 to 4): TRAIL receptor (1 to 4)

Key Words : CNS, Injury, Apoptosis, Death Receptors, Review

Send correspondence to: Corina Lorz, Molecular Oncology Unit, Edificio 70, Ave. Complutense 22, 28040 Madrid, Spain, Tel: 34 91 3460865, Fax: 34 91 3466484, E-mail: clorz@ciemat.es

http://www.bioscience.org/current/vol14.htm