

Effect of soy proteins and isoflavones on lipid metabolism and involved gene expression

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1. ABSTRACT

Clinical trials and animal studies showed that ingestion of soy proteins improves blood lipid profiles including lowering triglyceride, total and LDL cholesterol levels and increasing HDL cholesterol content. However, the effective components in the soy and the mechanisms involved in the hypolipidemic actions are not fully understood. Increasing evidence from animal studies have suggested that soy components may regulate lipid metabolism by modulating the activities of key transcription factors and thereby changing the downstream gene expression involved in lipogenesis or lipolysis. It has been shown that intake of soy proteins alters the expression of genes for sterol regulatory element binding protein, peroxisomal proliferator activated receptor, and liver X receptor. Dietary soy proteins suppress the DNA binding activities of hepatic nuclear receptors for thyroid hormones and retinoic acid, and alter the activities of key enzymes including cholesterol 7 α hydroxylase and ATPase/ATP synthase through post-translational protein modifications. This paper reviews the current understanding of the cellular and molecular events by which soy components affect lipid levels, especially focusing on modulation of transcription factors and regulation of gene expression involved in lipid metabolism by soy proteins and associated isoflavones.

2. INTRODUCTION

Soy foods have been consumed for over 5000 years in Asian countries. The epidemiological evidence suggest that soy consumption is linked to a lower incidence of chronic diseases including coronary heart disease, atherosclerosis, type 2 diabetes, osteoporosis, and certain types of cancers such as prostate cancer and breast cancer. Clinical trials (1) and animal studies (2-4) also showed that dietary soy proteins or isoflavones (ISF) reduce the risk factors for cardiovascular diseases including lowering triglyceride, total and LDL cholesterol levels and increasing the ratio of HDL/LDL cholesterol. Consequently, these studies have led to the approval of a food labeling health claim for soy proteins in the prevention of coronary heart disease by the U.S. Food and Drug Administration in 1999. Since then, soy food consumption in the U.S. has doubled, and over 2500 new foods containing soy have been introduced by U.S. food manufacturers.

Similar health petitions for soy proteins have also been approved thereafter in the United Kingdom, South Africa, Philippines, Brazil, Indonesia, Korea and Malaysia. However, the purported beneficial health effects were quite variable in different clinical studies. The Nutrition

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Committee of the American Heart Association has recently assessed 22 randomized trials conducted since 1999, and found that isolated soy proteins with ISF only slightly decreased LDL cholesterol but had no effect on HDL cholesterol, triglyceride, lipoprotein(a) or blood pressure. Soy ISF failed to change HDL cholesterol and other lipid risk factors in 19 studies. The effects of soy proteins and ISF on vasomotor symptoms of menopause, postmenopausal bone loss, and prevention of breast, endometrial, and prostate cancers were not substantiated (5).

More recent results suggest that a variety of factors could cause variations among different studies. These include processing procedures of soy protein isolate (SPI) which have been shown to affect the intactness of the protein subunits (6), different forms of ISF (conjugated or unconjugated to sugar) (7), and the ability of the subjects to metabolize daidzein, one of the major soy ISF, to produce equol (8). In addition, lack of understanding of the bioactive components in the soy and the molecular mechanisms by which lipid metabolism is impacted may contribute a major part to the discrepancies. Existing literature is quite controversial regarding which component (proteins or associated ISF) in soy has the lipid-lowering actions. This paper reviews the current understanding of the molecular events by which soy components affect lipid levels, especially focusing on the modulation of transcription factors and regulation of gene expression involved in lipid metabolism by soy proteins and associated ISF.

3. EFFECT OF SOY PROTEINS AND ISF ON LIPID PROFILES

Soybeans contain about 40% protein, and depending on the processing procedure, the protein content can reach over 90% as in SPI that is usually used in soy-based infant formulas. Most of the attention in soy studies has been focused on the proteins and their associated ISF.

3.1. Human clinical trials

The first human study on the lipid-lowering effect of soy proteins was reported in 1967 (9). It was shown that a replacement of mixed proteins by vegetable proteins, mainly isolated soy proteins, remarkably reduced the average blood cholesterol levels by more than 100 mg/ml, from approximately 295 mg/ml to 172 mg/ml in hypercholesterolemic men. The hypolipidemic action of soy proteins has been further demonstrated in the following clinical studies.

A meta-analysis of 38 controlled clinical trials published between 1977 and 1994 showed that average intake of 47 g/day of soy proteins (ranged from 17 to 124 g/day) significantly decreased total cholesterol by 9.3%, LDL cholesterol by 12.9%, triglycerides by 10.5%, and increased HDL cholesterol by 2.4% but this increase was not statistically significant. All of these studies used either isolated or textured soy proteins. Among them, 30 studies were conducted with hypercholesterolemic subjects (1). More recent results suggest that intact soy proteins with

ISF (3.0~185.0 mg/d) were associated with significant decreases in serum total cholesterol by 3.8%, LDL cholesterol by 5.3%, and triglycerides by 7.3% and significant increases in serum HDL cholesterol by 3.0% in 23 randomized controlled studies published from 1995 to 2002. The effects on total and LDL cholesterol were greater in men than in women (10).

The effects of soy ISF on blood lipid profile appear to be inconsistent among different studies. For instance, a meta-analysis showed that ISF-enriched soy proteins (61.7~317.9 mg/d ISF) markedly decreased serum total cholesterol by 1.8%, and LDL cholesterol by 3.6%, but no changes in HDL cholesterol and triglycerides were significant compared to the ISF-depleted soy proteins (1.2~14.6 mg/d ISF). ISF-depleted soy proteins significantly decreased LDL cholesterol by 2.8%, while ISF-enriched soy proteins significantly decreased LDL cholesterol by 5.0% and increased HDL cholesterol by 3.0% compared to animal proteins (11). With equal amounts of soy protein intake, high ISF intake (averagely 96 mg/d) resulted in significantly greater decreases in serum LDL cholesterol than low ISF intake (6 mg/d), suggesting that the LDL cholesterol-lowering effects of ISF are independent of soy proteins (12).

However, the hypolipidemic benefits of soy ISF in other studies using a range of 40-150 mg/d ISF in either normocholesterolemic or hypercholesterolemic men or women were insignificant. For example, total and LDL cholesterol levels in healthy perimenopausal women receiving 61.8 mg/d of ISF or placebo for 4 weeks had no significant difference (13). The LDL cholesterol levels among the moderately hypercholesterolemic women supplemented with 42 g/d milk protein, soy plus ISF (80 mg/d), or soy without ISF did not differ (14). When hypercholesterolemic postmenopausal women were given a much higher dosage of ISF (150 mg/d) for 6 months, total and LDL cholesterol levels were lower, but not statistically different (15). Similar results were observed in the study with healthy men given 55 mg/d of ISF (16). A recent meta-analysis of 10 randomized controlled trials from 1995 to 2002 indicated that 36 g/d of soy proteins in combination with 52 mg of soy-associated ISF lowered LDL cholesterol by 4%; however, despite large increases in blood ISF concentrations, no significant correlation between soy ISF and change in LDL cholesterol was detected. It has been concluded that ISF did not have an independent effect on lowering LDL cholesterol levels in the presence of soy proteins (17). This conclusion was supported by another meta-analysis showing that supplementation with extracted soy ISF had no significant effect on total cholesterol reduction (10).

Nevertheless, soy proteins, regardless of ISF content, have been shown to lower the ratios of total to HDL cholesterol, and LDL to HDL cholesterol compared to the milk protein in healthy young men (18), and significantly reduced total and LDL cholesterol and triglyceride concentrations in hypercholesterolemic men and women (19). However, the added ISF had no effect on plasma lipid levels. In addition, the hypocholesterolemic

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effects of soy proteins were closely associated with the initial blood cholesterol levels. The reductions in total or LDL cholesterol levels are much greater in hypercholesterolemic subjects than in normocholesterolemic subjects (1,10,11).

3.2. Animal studies

Obese Zucker rats fed soy proteins with high ISF (5.8 mg/g protein) for 8 or 11 weeks had lower liver weights, lower liver cholesterol and triglyceride content than those fed either casein or soy proteins with low amounts of ISF (0.1 mg/g protein) (20). Plasma cholesterol concentrations in the rats fed either ethanol-washed SPI or SPI with an ethanol extract of soy were comparable and significantly lower than those fed casein. Addition of ethanol extract to SPI or casein did not influence plasma cholesterol level, suggesting that the cholesterol-lowering effect of SPI in rats was attributed to its protein component but not to the ethanol-extractable minor constituents including ISF (21). Our study in Sprague Dawley rats also showed that intake of 20% alcohol-washed SPI with or without added ISF markedly lowered the plasma triglyceride levels compared to a casein diet, however the added ISF had no additional effects (22).

3.3. Possible effective components

Soybeans contain two major storage globulins, beta-conglycinin (7S) and glycinin (11S). Beta-conglycinin has three subunits (α' , α , and β), and is believed to be essential for the hypolipidemic actions of soy. Consumption of beta-conglycinin decreased serum triglycerides in both normal and genetic obese mice (3) as well as in humans (23). Addition of beta-conglycinin into cultured HepG2 cells up-regulated LDL receptor activity measured by the uptake and degradation of 125 I-labeled LDL (24). However, absence of α' subunit in beta-conglycinin derived from an α' mutated soy cultivar had no effect (25). This suggests that the α' subunit in the beta-conglycinin may be a bioactive component that mediates the hypolipidemic activities of soy.

Different amino acid composition in soy proteins is believed to play a role in mediating the hypolipidemic actions of soy. It has been shown that lysine and methionine have moderate hypercholesterolemic effects (26,27), whereas arginine lowers cholesterol concentrations (28). Soy proteins contain a higher ratio of arginine to lysine and methionine, which may be at least partially responsible for the hypocholesterolemic effects of soy proteins (29,30).

The major soy-derived ISF such as genistein and daidzein are potent regulators of many genes involved in lipid metabolism (20,31-33), however the lipid-lowering actions of soy-derived ISF are quite variable among different studies. The contributing factors causing these inconsistencies are not fully understood, but it has been suggested that the initial lipid levels and the ability of the subjects to metabolize the daidzein or daidzin to equol as well as the forms of ISF taken (glucosides or aglycones) may play a role in determining the bioavailabilities of the ISF and the responsiveness of the subjects to ISF (7,10,12).

4. REGULATION OF TRANSCRIPTION FACTORS RELATED TO LIPID METABOLISM BY SOY PROTEINS AND ISF

Although the molecular mechanism(s) by which soy components affect lipid metabolism is not fully understood, several hypotheses have been proposed to date based on the information obtained from studies in animals, humans, and *in vitro* tissue cultures. These include suppressed cholesterol absorption (34), increased fecal excretion of bile acids (35,36), up-regulated hepatic LDL receptor activity (37), and elevated serum thyroxine (T4) levels (38).

Recent studies suggest that dietary soy components may influence physiological functions via regulation of the expression of genes involved in various biochemical pathways. DNA-microarray analyses show that 33% out of 8000 liver genes were differentially expressed in soy-fed rats as compared to casein-fed rats. Most of those genes were involved in lipid metabolism, transcriptional regulation and energy metabolism. Among those, the ones involved in the fatty acid synthesis were down-regulated, whereas the ones related to cholesterol synthesis or steroid catabolism were up-regulated in soy-fed animals (39,40). More importantly, dietary soy proteins were shown to differentially regulate or modify transcription factors such as sterol regulatory element binding protein (SREBP) (3,41) and several nuclear receptors (42-44).

4.1. Sterol regulatory element binding protein

SREBP is synthesized in the endoplasmic reticulum as a precursor protein, and becomes active after cleavage of the NH_2 -terminal domain by two specific proteases, SP1 and SP2 in the Golgi apparatus. The active SREBP translocates to the nucleus, where it activates transcription of multiple target genes by binding to sterol response elements in their promoter regions (45). Transcriptional controls of fatty acid and cholesterol synthesis as well as cholesterol uptake genes are partially mediated by SREBP (46).

There are three SREBP isoforms (SREBP-1a, -1c and SREBP-2) with specific and overlapping transcriptional targets. SREBP-1a and -1c preferentially regulate the enzymes involved in fatty acid and triglyceride biosynthesis such as acetyl-CoA carboxylase (ACC), fatty acid synthase (FAS) and microsomal triglyceride transfer protein and are activated in response to insulin. In addition, SREBP-1 is shown to reduce the mRNA expression of stearoyl-CoA desaturase 1 (SCD-1), an important enzyme to catalyze desaturation of long-chain fatty acids and enhance their incorporation into very low density lipoprotein (VLDL) assembly (47). SREBP-2 preferentially binds to promoters of the genes involved in cholesterol biosynthesis and uptake such as hydroxymethylglutaryl-CoA synthase (HMG-CoAs), HMG-CoA reductase (HMG-CoAr) and LDL receptor, and is activated by a low cholesterol content of the cell (45).

Consumption of soy proteins reduces hepatic SREBP-1 mRNA and protein expression and concurrently reduces FAS and malic enzyme (ME) mRNA abundance as well as endogenous fatty acid biosynthesis (4,48-50).

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Although the underlying molecular mechanism(s) are not fully understood, regulation of SREBP-1 gene expression by dietary soy proteins may be mediated through either insulin-dependent or -independent pathways. It has been shown that insulin stimulates SREBP-1 expression in liver (4) and rat primary hepatocytes (51), and that soy proteins decrease insulin secretion and increase hepatic insulin removal (52). However, recent evidence indicates that despite high insulin concentrations in hyperinsulinemic obese fa/fa rats, rats fed soy proteins have lower SREBP-1 expression than rats fed casein (47). Further investigation demonstrated that reduction in SREBP-1 by soy proteins might be through the negative regulation of liver X receptor (LXR) mRNA expression (47). Interaction of SREBP with nuclear factors is a common mechanism through which SREBP activates gene transcription. For example, SREBP-mediated activation of the genes for HMG-CoAs (53) and FAS (54) is dependent on the interaction between SREBP and nuclear factor-Y. Other studies have shown that SREBP activation of LDL receptor gene is dependent on the interaction between SREBP and sterol regulatory element-1 (55).

Daidzein, genistein, or soy ISF have no effects on SREBP-1 content in cultured hepatocytes (56) nor in rats (47). However, a study in HepG2 cells demonstrated that genistein reduced the expression of site-1 protease which is responsible for the processing and activation of SREBP-1, and the processing of SREBP-1 but failed to change the expression of SREBP-1 mRNA. Meanwhile, the expression of SREBP-1-regulated lipogenic genes including SCD1, ACCalpha and ACCbeta were suppressed (31).

SREBP-2 is a transcription factor regulating the transcriptional activation of HMG-CoAr and LDL receptor (45). Consumption of ISF-poor SPI decreased the mRNA levels of SREBP-2, HMG-CoAr, CYP7A1, and LDL receptor in the liver of rats and markedly reduced liver cholesterol and triglyceride content and plasma triglyceride concentrations (50). In contrast, ISF-containing soy extract and the individual ISF increased the mature form of SREBP-2 and HMG-CoAr protein as well as HMGCoAs mRNA levels in cultured HepG2 cells (56) and in rats (4,57), and elevated serum cholesterol clearance (4,57). The discrepancy between these studies may attribute to the variable content of ISF in SPI.

4.2. Peroxisomal proliferator activated receptor

Peroxisomal proliferator activated receptor (PPAR) is a ligand-activated transcription factor (58) and has three isoforms (alpha, gamma and delta). These isoforms are approximately 60-80% homologous in their ligand- and DNA-binding domains and have overlapping tissue distribution. PPARs are important in the regulation of lipid metabolism. For example, PPARalpha controls fatty acid oxidative metabolism through transcriptional induction of carnitine-palmitoyl transferase 1 (CPT-1). CPT-1 is a key enzyme involved in the transfer of fatty acids into the mitochondria to promote β -oxidation in the liver (59). PPARalpha knockout in mice remarkably elevates blood and hepatic cholesterol and triglyceride levels (60,61).

Intake of soy proteins significantly decreased hepatic triglyceride levels and epididymal adipose tissue weight, and increased skeletal muscle CPT1 activity as well as CPT1 and PPARalpha mRNA levels in male Sprague-Dawley rats (62). Up-regulation of CPT-1 mRNA expression by soy proteins was shown to be mediated through activation of PPAR in rat liver and thereby increased fatty acid oxidation (47).

The effect of ISF on lipid metabolism might be mediated through both PPARalpha-dependent and independent pathways. The wild-type and PPARalpha knockout mice were fed soy proteins with or without ISF for 6 weeks. Soy proteins with ISF activated hepatic PPARalpha in wild-type mice and decreased serum triglyceride levels in both wild-type and PPARalpha knockout mice (61). Additionally, an ISF-containing soy extract doubled PPAR-directed gene expression in RAW 264.7 cells containing either a PPARalpha or PPARgamma expression plasmid. A similar induction was observed when the soy ISF genistein or daidzein were used to treat cells (20).

PPARgamma is a key regulator of glucose homeostasis and adipogenesis and is required for normal adipocyte differentiation. Activation of PPARgamma or its obligate heterodimer PPARgamma:RXR inhibited cholesterol accumulation by enhancing cholesterol efflux (63). Dietary soy proteins increased PPARgamma mRNA steady-state levels (47) and protein content (64) in adipose tissue of Zucker diabetic fatty fa/fa rats compared to a casein diet, and reduced the total and liver adiposity and maintained a lower number of dysfunctional adipocytes.

4.3 Thyroid hormone receptor

It has been established that thyroid hormones play important roles in the regulation of lipid metabolism. A deficiency in thyroid hormones results in elevated cholesterol levels in blood, however, it can be normalized by thyroid hormone substitution (65,66). Thyroid hormones influence lipid metabolism at several critical steps in the liver, including (a) regulation of fatty acid synthesis via ACC (67), FAS (68) and ME (69); (b) control of cholesterol biosynthesis via HMG-CoAr (70); (c) regulation of bile acid synthesis via CYP7A1 (71,72); and (d) mediation of cholesterol uptake from the circulation via LDL receptor (70,73).

Increased T4 levels were proposed as a putative mechanism responsible for the hypocholesterolemic actions of soy proteins (38). T4 levels were negatively correlated with plasma cholesterol and triglyceride concentrations in gerbils (74), rats (75), and hamsters (76) fed soy protein-based diets. However, this notion appears to be inconsistent with the results obtained from human clinical studies. Daily intake of SPI with either 1.64 or 61.7 mg ISF for 57 d had no effect on total triiodothyronine (T3), free T3, total T4, free T4, thyroid stimulating hormone (TSH) and thyroid binding globulin in healthy young men (77). The blood lipid levels including total cholesterol, LDL cholesterol, HDL cholesterol, triglycerides, apolipoprotein (apo) B, and

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apo A-I were not different among three dietary groups. However, the ratios of total to HDL cholesterol, LDL to HDL cholesterol, and apo B to apo A-I were significantly lower with both SPI treatments than with milk protein isolate treatment (18).

Of the seven trials (77-83) assessing the effects of soy proteins on serum TSH, total T3, and T4, only one study (82) showed elevated serum TSH, T3 and T4 by soy proteins compared with casein group. However, when compared to the normal baseline level, these increases were minimal. Another study also found that soy intake significantly decreased the plasma cholesterol concentration, but had no effect on T4 level in postmenopausal women with type 2 diabetes (79). In addition, although SPI and soy protein concentrates are both hypocholesterolemic in hamsters, only SPI increases T4 levels (84). These results indicate that modulation of thyroid hormone status may not be the only mechanism responsible for the cholesterol-lowering action of soy proteins.

Thyroid hormone receptor (TR) mediates the thyroid hormone-regulated physiological functions. Mammalian TR is encoded by two genes, TRalpha and TRbeta. The primary transcript from each gene can be alternatively spliced to form different isoforms. Four TR isoforms have been reported to date and they are TRalpha1, TRalpha2, TRbeta1, and TRbeta2 (85). TRalpha and TRbeta differ slightly in structure and substantially in tissue distribution. TRalpha regulates heart rate and the speed and force of systolic contraction (86). TRbeta has diverse effects on lipid metabolism and hypothyroidism (87) and is highly expressed in liver (88). They are key regulators of many genes involved in lipid metabolism.

We have shown that consumption of alcohol-washed SPI markedly increased hepatic TRbeta1 protein content in rats compared with a casein-based diet, but had no effect on TRbeta1 mRNA abundances. Supplemental ISF had no additional effect compared with SPI alone. We further demonstrated that effects of SPI on TR were isoform- and tissue-specific (43). Interestingly, the binding ability of nuclear TR to the target genes measured by electrophoretic mobility shift assay was remarkably suppressed by dietary SPI (42).

TRbeta isoform is the major functional TR in liver, accounting for 80% of hepatic T3-binding activity (89), and plays a unique role in the regulation of cholesterol metabolism. For instance, the normal stimulation of T3 on CYP7A1, a rate limiting enzyme in bile acid biosynthesis, was lost in TRbeta^{-/-} but not in TRalpha^{-/-} mice (90), and overexpression of TRalpha1 could not substitute for absence of TRbeta (91).

Inhibition of TRbeta1 binding to target genes by SPI could block the stimulatory effects of thyroid hormones and alter downstream gene expression. In addition, suppression of TRbeta DNA binding may also affect lipid metabolism through altering PPARgamma-mediated gene expression involved in lipid homeostasis. It has been

recently demonstrated that TRbeta can compete with PPARgamma and bind to the peroxisome proliferator response element (PPRE) as homodimers and heterodimers with PPARgamma or the retinoid X receptor (RXR) (92), thereby inhibiting PPAR-mediated gene expression at the level of PPAR binding to PPRE (93). Decreased DNA binding ability of TRbeta by SPI may result in increased binding of PPARgamma to PPRE and up-regulation of PPARgamma-mediated gene expression.

4.4. Retinoic acid receptor

RAR family is activated by both all-trans and 9-cis retinoic acid (94). Three RAR subtypes (alpha, beta, and gamma) have been characterized (94,95) and are encoded by distinct genes. Several isoforms are produced from each gene. For example, RARbeta has four isoforms (beta1, beta2, beta3 and beta4) which are generated via alternative gene splicing of primary transcripts initiated from 2 promoters (96,97). Retinoic acid (RA), a metabolite of vitamin A, is important in the prevention and treatment of various cancers (98,99) and also plays important roles in controlling lipid metabolism (100,101).

However, induction of hypertriglyceridemia is one of the major adverse effects in the treatment of cancers with retinoids in both rats (102) and humans (98,103-105). Replacement of dietary casein with SPI markedly reduced the severity of RA-induced hypertriglyceridemia in rats (100,106). Although the involved molecular events are unclear, the RA-induced hypertriglyceridemia was shown to be mediated via retinoid receptors (100,107) and the serum retinoid level in RA-treated rats was not affected by dietary SPI (106).

We have recently reported that ingestion of 20% alcohol-washed SPI significantly elevated hepatic RARbeta2 protein content, but had no effect on its mRNA abundance in rats, compared with a casein diet. However, the DNA binding abilities of hepatic RARbeta2 to its target genes were significantly suppressed. Increasing amounts of added soy ISF had no effect on RARbeta. We further demonstrated that dietary SPI induces modification of the receptor protein, which is believed to be responsible for the functional inhibition of RARbeta (44). RARbeta2 is the most abundant RARbeta isoform in the body (108). Suppression of RARbeta DNA binding activity may be an important cellular event by which dietary soy proteins alleviate the retinoid-induced hypertriglyceridemia.

4.5. Liver X receptor

LXR is predominantly expressed in the liver and can be activated by oxysterols. LXR regulates a variety of genes involved in the catabolism, transport, and uptake of cholesterol and its metabolites. CYP7A1 is one of those genes and contains a LXR response element in its promoter region (109). LXR acts as a cholesterol sensor and up-regulates the expression of CYP7A1, which results in increased bile acid synthesis and subsequently excretion of cholesterol (110). Through regulation of ATP-binding cassette-A1 transporter system in extrahepatic cells, LXR controls the efflux of cholesterol from these cells which will then be taken up by HDL and transported back to the liver for further catabolism and elimination in bile (111). In

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addition, LDL receptor and SREBP-1c genes possess the LXR response element through which LXR mediates endocytic uptake of LDL cholesterol in the liver (112) and insulin-induced SREBP-1 gene expression (113). Ingestion of non-extracted soy proteins (containing normal level of ISF) reduced hepatic cholesterol concentrations and down-regulated the expression of liver LXR, CYP7A1, ATP-binding cassette-A1 transporter and SREBP-1 mRNA in obese rats (47).

5. REGULATION OF GENES RELATED TO LIPID METABOLISM BY SOY PROTEINS AND ISF

5.1. Fatty acid biosynthesis

Three key enzymes are involved in fatty acid synthesis. ACC catalyzes the carboxylation of acetyl-CoA to form malonyl-CoA, which is the first step of the long-chain fatty acid biosynthesis (114). FAS catalyzes malonyl-CoA to generate long-chain fatty acids, a complex NADPH-dependent seven-step reaction (115). ME catalyzes oxidative decarboxylation of malate to pyruvate and CO₂, with the concomitant reduction of the cofactor NAD(P)(+) to NAD(P)H (116), a process providing the source of NADPH for FAS. Alteration in the gene expression or enzymatic activity in any of these enzymes may affect fatty acid synthesis.

5.1.1. ACC

Two ACC isoforms have been identified and are encoded by distinct genes. ACCalpha is a cytosolic protein, highly expressed in liver and adipose tissue that exhibit high rates of fatty acid synthesis, whereas ACCbeta is located in mitochondria and is predominantly expressed in heart and muscle, and to a lesser extent in liver. ACCbeta plays a pivotal role in the regulation of mitochondrial fatty acid beta-oxidation (117) through feedback inhibition of CPT-1, an enzyme that controls the entry of long-chain fatty acid CoA esters into the mitochondria for degradation (118). Suppression of ACCbeta increases CPT-1 activity and enhances fatty acid beta-oxidation.

It has been found that feeding soy proteins reduces hepatic ACCalpha mRNA content in fatty rats (119). This is further supported by our study in normal rats showing that intake of alcohol-washed SPI significantly reduced ACCalpha and ACCbeta mRNA and protein expression in the liver (22). Moreover, the ratio of phospho-ACCalpha/ACCalpha and phospho-ACCbeta/ACCbeta were not different in liver, kidney and heart between rats fed SPI- or casein-based diet in our study (22), suggesting that dietary SPI may affect ACC mainly through transcriptional regulation rather than phosphorylation or dephosphorylation.

Regulation of ACC gene expression by dietary SPI might be mediated through at least two different pathways which could be TR-dependent or SREBP-dependent. ACC gene contains thyroid hormone response element in its promoter region and its expression is regulated by thyroid hormones (120). In addition, SREBP-1 is a transcription factor of the ACCalpha gene and is down-regulated by dietary SPI. However, a recent study showed that dietary SPI significantly decreased hepatic

ACCalpha mRNA, but did not affect hepatic SREBP-1 mRNA content (121). Further study revealed that ACCalpha expression was controlled by two promoters, PI and PII (121). Dietary SPI significantly decreased PI-generated ACCalpha mRNA expression through inhibiting the binding ability of SREBP-1 to ACCalpha promoter, but did not affect PII and PII-generated ACCalpha mRNA (122), suggesting that SPI regulates ACCalpha mRNA mainly by regulating SREBP-1 binding to PI via nuclear factors other than SREBP-1 itself.

5.1.2. FAS and ME

Mammalian FAS is the product of a single non-duplicated gene that generates a 250 kDa polypeptide chain. FAS is active as a homodimer (115) and its activity is regulated by diet, glucose, T4 and SREBP-1c through transcriptional control (123). Consumption of soy proteins reduced the mRNA expression of FAS and ME, and lowered hepatic triglyceride depots in rats (4,48,49). This regulation is suggested to be mediated through down-regulation of SREBP-1 (4,48,49). However, FAS (124) and ME (125) also contain a thyroid hormone response element in their promoters. Inhibition of the DNA binding activity of TR by SPI (43) may be an alternative pathway for controlling gene expression and lipid synthesis.

5.1.3. ATPase/ATP synthase

ATPase/ATP synthase is an enzymatic complex responsible for ATP synthesis and hydrolysis in mitochondria. It consists of a membrane-bound F₀ portion and a soluble F₁ portion. F₁ has five subunits (alpha, beta, gamma, delta, epsilon), and the beta subunit contains the catalytic sites of the ATP synthesis. F₁ catalyzes the synthesis of ATP from adenosine diphosphate and inorganic phosphate. ATPase/ATP synthase plays important roles in the regulation of carbohydrate, protein and lipid metabolism via modulating energy homeostasis. Its beta subunit on the hepatocyte plasma membrane was recently identified as a high-affinity HDL receptor (126). HDL mediates the efflux and transport of cholesterol from peripheral cells to the liver for further metabolism, suggesting that ATPase/ATP synthase may play a role in the regulation of cholesterol metabolism.

Dietary supplementation with soy isolated proteins containing ISF prevented reduction of Na⁺, K⁺-ATPase activity, another member of the ATPase family, in diabetic rats (127). Genistein, one of the main soy ISF, suppressed brain and hepatic mitochondrial ATP synthase activity *in vitro* (128). We have recently shown that hepatic mitochondrial ATPase activity was significantly higher in the rats fed alcohol-washed SPI than in those fed casein. Addition of ISF to SPI eliminated the action of SPI. ATPase/ATP synthase beta protein content in the liver were unchanged; however its patterns (isoelectric points) measured by 2 dimensional Western blot were different among dietary groups. We further demonstrated that dietary SPI increases the dephosphorylation of the mitochondrial ATPase/ATP synthase beta subunit, which is responsible for the increase in the enzymatic activity (129). However, the physiological importance of this cellular response to the dietary SPI remains to be investigated.

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5.2. Cholesterol metabolism

5.2.1. Synthesis

In most cells, cholesterol is derived either from endogenous synthesis via the mevalonate pathway or from the uptake of circulating cholesterol-rich LDL. HMG-CoAr is the rate-limiting enzyme in cholesterol biosynthesis and catalyzes the formation of mevalonate from HMG-CoA. Genistein has been shown to decrease cholesterol synthesis in cultured HepG2 (32) and inhibit HMG-CoAr activity in MCF-7 human breast cancer cells (33). Consumption of SPI significantly decreased HMG-CoAr mRNA abundance and cholesterol synthesis in the liver of nephrotic rats (49). We also found that hepatic HMG-CoAr protein content was significantly lower in the rats fed SPI with ISF supplement compared to casein control (unpublished data).

5.2.2. Catabolism

One of the mechanisms proposed to explain the hypocholesterolemic effect of soy proteins is through an increase in bile acid secretion. Rabbits fed casein diet excreted less neutral steroids and produced less bile acid than animals fed a soy protein diet. In addition, rabbits fed casein excreted mainly cholesterol whereas those fed soy proteins excreted unabsorbable coprostanol (35). Peptides prepared by the *in vitro* digestion of soy proteins were found to stimulate fecal steroid excretion and consequently lower serum cholesterol in rats (130) as well as in human volunteers (131).

Bile acids are amphipathic and polar derivatives of cholesterol. The liver excretes bile acids into the intestine where ~ 95% of bile acids are then reabsorbed in the terminal ileum and returned to the liver. If bile acids are no longer available for transport back to the liver, they are lost in the faeces. As a result, there is an up-regulation of hepatic enzymes involved in bile acid biosynthesis such as CYP7A1.

The activity of CYP7A1 is negatively regulated by bile acid, diet, drug and hormones. The effects of consumption of soy proteins on hepatic CYP7A1 gene expression are inconsistent in different studies. One study showed that dietary soy proteins increased CYP7A1 gene expression in the liver (30), whereas other studies reported opposite effects (47,50,132). We have recently demonstrated that ingestion of 20% alcohol-washed SPI (containing minimal amount of ISF) remarkably reduced the total cholesterol levels in the blood and liver of the rats compared with a casein diet. However, the hepatic CYP7A1 protein contents measured by Western blot were not different between dietary groups. Our further study showed that SPI appears to increase the phosphorylation of hepatic CYP7A1, which may lead to a higher enzymatic activity (unpublished data). Although this remains to be confirmed, other studies have already demonstrated that increased protein phosphorylation is correlated with enhanced activity of CYP7A1 (133).

5.2.3. Uptake

LDL cholesterol is mainly from three sources: peripheral cholesterol synthesis, hepatic cholesterol

synthesis, and intestinal cholesterol absorption. LDL receptor is a major regulator of circulating LDL cholesterol. It has been suggested that the hypocholesterolemic effects of soy proteins may be mediated through up-regulation of LDL receptors (37,134-137). Soy proteins enhance the expression of the LDL receptor in cultured human hepatoma cells (37,134), animals (136,137), and hypercholesterolemic type 2 diabetic patients (135). A similar effect has been observed with a soy protein polypeptide in cultured HepG2 cells (25). However, results obtained from a study with LDL receptor null mice did not support this hypothetical mechanism, in which SPI lowered the plasma LDL and VLDL cholesterol concentrations and inhibited atherosclerosis despite the absence of the LDL receptor in the mice (138). This suggests that LDL receptor-independent pathway(s) by which dietary SPI lowers cholesterol levels may exist.

6. SUMMARY

The hypolipidemic actions of soy components (mainly proteins and associated ISF) appear to be consistent in both animal and human studies. However, the size of the effect is variable, which might be attributed to the variation in the processing procedures of SPI, the content of ISF, the amount of unknown bioactive component(s) or the physiology of the subjects. It has been shown that different processing procedures affect the intactness of soy protein subunits that might be crucial for their physiological functions (6). Although the *in vitro* and animal studies showed that beta-conglycinin, particularly the alpha' subunit, has most of the functions of the soy, all the human trials and animal studies were based on the amount of total soy proteins rather than the amount of the bioactive components which might be very different among different SPI preparations. In addition, the ability of the subjects to metabolize daidzein or daidzin to equol is suggested to be associated with their responsiveness to dietary soy. However, only 30-50% of the adult population can excrete equol in urine when challenged daily with soy foods (8). Soy proteins and ISF regulate lipid metabolism via multiple cellular pathways (fatty acid biosynthesis, cholesterol synthesis, catabolism and uptake) and through modulation of the key transcription factors including SREBP, PPAR, TR, RAR, and LXR. In addition to regulation at transcriptional levels, soy proteins have also been shown to posttranslationally modify the receptor (such as RAR) and enzymatic (ATPase/ATP synthase and CYP7A1) proteins via a phosphorylation and dephosphorylation mechanism, and thereby altering the DNA binding ability of the receptor or activity of the enzymes. Since most of the mechanism studies on soy actions have been conducted in either cultured cells or animal models, whether the same mechanisms are shared in humans remains to be verified using appropriate biomarkers.

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8. REFERENCES

1. J. W. Anderson, B. M. Johnstone and Newell Cook: Meta-analysis of the effects of soy protein intake on serum lipids. *N Engl J Med* 333, 276-282 (1995)
2. Y. Lin, G. W. Meijer, M. A. Vermeer and E. A. Trautwein: Soy protein enhances the cholesterol-lowering effect of plant sterol esters in cholesterol-fed hamsters. *J Nutr* 134, 143-148 (2004)
3. T. Moriyama, K. Kishimoto, K. Nagai, R. Urade, T. Ogawa, S. Utsumi, N. Maruyama and M. Maebuchi: Soybean beta-conglycinin diet suppresses serum triglyceride levels in normal and genetically obese mice by induction of beta-oxidation, downregulation of fatty acid synthase, and inhibition of triglyceride absorption. *Biosci Biotechnol Biochem* 68, 352-359 (2004)
4. C. Ascencio, N. Torres, F. Isoard-Acosta, F. J. Gomez-Perez, R. Hernandez-Pando and A. R. Tovar: Soy protein affects serum insulin and hepatic SREBP-1 mRNA and reduces fatty liver in rats. *J Nutr* 134, 522-529 (2004)
5. F. M. Sacks, A. Lichtenstein, L. Van Horn, W. Harris, P. Kris-Etherton and M. Winston: Soy protein, isoflavones, and cardiovascular health. An American Heart Association Science Advisory for Professionals From the Nutrition Committee. *Circulation* 113, 1034-1044 (2006)
6. E. Gianazza, I. Eberini, A. Arnoldi, R. Wait and C. R. Sirtori: A proteomic investigation of isolated soy proteins with variable effects in experimental and clinical studies. *J Nutr* 133, 9-14 (2003)
7. K. D. Setchell, N. M. Brown, L. Zimmer-Nechemias, W. T. Brashear, B. E. Wolfe, A. S. Kirschner and J. E. Heubi: Evidence for lack of absorption of soy isoflavone glycosides in humans, supporting the crucial role of intestinal metabolism for bioavailability. *Am J Clin Nutr* 76, 447-453 (2002)
8. K. D. Setchell, N. M. Brown and E. Lydeking-Olsen: The clinical importance of the metabolite equol-a clue to the effectiveness of soy and its isoflavones. *J Nutr* 132, 3577-3584 (2002)
9. R. E. Hodges, W. A. Krehl, D. B. Stone and A. Lopez: Dietary carbohydrates and low cholesterol diets: effects on serum lipids on man. *Am J Clin Nutr* 20, 198-208 (1967)
10. S. Zhan and S. C. Ho: Meta-analysis of the effects of soy protein containing isoflavones on the lipid profile. *Am J Clin Nutr* 81, 397-408 (2005)
11. K. Taku, K. Umegaki, Y. Sato, Y. Taki, K. Endoh and S. Watanabe: Soy isoflavones lower serum total and LDL cholesterol in humans: a meta-analysis of 11 randomized controlled trials. *Am J Clin Nutr* 85, 1148-1156 (2007)
12. X. G. Zhuo, M. K. Melby and S. Watanabe: Soy isoflavone intake lowers serum LDL cholesterol: a meta-analysis of 8 randomized controlled trials in humans. *J Nutr* 134, 2395-2400 (2004)
13. T. Uesugi, Y. Fukui and Y. Yamori: Beneficial effects of soybean isoflavone supplementation on bone metabolism and serum lipids in postmenopausal Japanese women: a four-week study. *J Am Coll Nutr* 21, 97-102 (2002)
14. C. D. Gardner, K. A. Newell, R. Cherin and W. L. Haskell: The effect of soy protein with or without isoflavones relative to milk protein on plasma lipids in hypercholesterolemic postmenopausal women. *Am J Clin Nutr* 73, 728-735 (2001)
15. A. Dewell, C. B. Hollenbeck and B. Bruce: The effects of soy-derived phytoestrogens on serum lipids and lipoproteins in moderately hypercholesterolemic postmenopausal women. *J Clin Endocrinol Metab* 87, 118-121 (2002)
16. J. M. Hodgson, I. B. Puddey, L. J. Beilin, T. A. Mori and K. D. Croft: Supplementation with isoflavonoid phytoestrogens does not alter serum lipid concentrations: a randomized controlled trial in humans. *J Nutr* 128, 728-732 (1998)
17. R. M. Weggemans and E. A. Trautwein: Relation between soy-associated isoflavones and LDL and HDL cholesterol concentrations in humans: a meta-analysis. *Eur J Clin Nutr* 57, 940-946 (2003)
18. B. L. McVeigh, B. L. Dillingham, J. W. Lampe and A. M. Duncan: Effect of soy protein varying in isoflavone content on serum lipids in healthy young men. *Am J Clin Nutr* 83, 244-251 (2006)
19. A. H. Lichtenstein, S. M. Jalbert, H. Adlercreutz, B. R. Goldin, H. Rasmussen, E. J. Schaefer and L. M. Ausman: Lipoprotein response to diets high in soy or animal protein with and without isoflavones in moderately hypercholesterolemic subjects. *Arterioscler Thromb Vasc Biol* 22, 1852-1858 (2002)
20. O. Mezei, W. J. Banz, R. W. Steger, M. R. Peluso, T. A. Winters and N. Shay: Soy isoflavones exert antidiabetic and hypolipidemic effects through the PPAR pathways in obese Zucker rats and murine RAW 264.7 cells. *J Nutr* 133, 1238-1243 (2003)
21. K. Fukui, N. Tachibana, Y. Fukuda, K. Takamatsu and M. Sugano: Ethanol washing does not attenuate the hypocholesterolemic potential of soy protein. *Nutrition* 20, 984-990 (2004)
22. C. W. Xiao, C. Wood, W. Huang, M. R. L'Abbe, G. S. Gilani, G. M. Cooke and I. Curran: Tissue-specific regulation of acetyl-CoA carboxylase gene expression by dietary soya protein isolate in rats. *Br J Nutr* 95, 1048-1054 (2006)
23. M. Kohno, M. Hirotsuka, M. Kito and Y. Matsuzawa: Decreases in serum triacylglycerol and visceral fat

Soy affects lipid metabolism

- mediated by dietary soybean beta-conglycinin. *J Atheroscler Thromb* 13, 247-255 (2006)
24. C. Manzoni, M. Duranti, I. Eberini, H. Scharnag, W. Marz, S. Castiglioni and M. R. Lovati: Subcellular localization of soybean 7S globulin in HepG2 cells and LDL receptor up-regulation by its alpha' constituent subunit. *J Nutr* 133, 2149-2155 (2003)
25. C. Manzoni, M. R. Lovati, E. Gianazza and C. R. Sirtori: Soybean protein products as regulators of liver low-density lipoprotein receptors. II. a-a' rich commercial soy concentrate and a' deficient mutant differently affect low-density lipoprotein receptor activation. *J Agric Food Chem* 46, 2481-2484 (1998)
26. M. W. Huff, R. M. Hamilton and K. K. Carroll: Plasma cholesterol levels in rabbits fed low fat, cholesterol-free, semipurified diets: effects of dietary proteins, protein hydrolysates and amino acid mixtures. *Atherosclerosis* 28, 187-195 (1977)
27. E. M. Kurowska and K. K. Carroll: Hypercholesterolemic responses in rabbits to selected groups of dietary essential amino acids. *J Nutr* 124, 364-370 (1994)
28. J. D. Radcliffe and D. M. Czajka-Narins: Use of arginine to reduce the severity of retinoid-induced hypertriglyceridemia. *Nutr Cancer* 36, 200-206 (2000)
29. T. Morita, A. Oh-hashii, K. Takei, M. Ikai, S. Kasaoka and S. Kiriyaama: Cholesterol-lowering effects of soybean, potato and rice proteins depend on their low methionine contents in rats fed a cholesterol-free purified diet. *J Nutr* 127, 470-477 (1997)
30. O. A. Gudbrandsen, H. Wergedahl, B. Liaset, M. Espe and R. K. Berge: Dietary proteins with high isoflavone content or low methionine-glycine and lysine-arginine ratios are hypocholesterolaemic and lower the plasma homocysteine level in male Zucker fa/fa rats. *Br J Nutr* 94, 321-330 (2005)
31. E. S. Shin, H. H. Lee, S. Y. Cho, H. W. Park, S. J. Lee and T. R. Lee: Genistein downregulates SREBP-1 regulated gene expression by inhibiting site-1 protease expression in HepG2 cells. *J Nutr* 137, 1127-1131 (2007)
32. N. M. Borradaile, L. E. de Dreu, L. J. Wilcox, J. Y. Edwards and M. W. Huff: Soya phytoestrogens, genistein and daidzein, decrease apolipoprotein B secretion from HepG2 cells through multiple mechanisms. *Biochem J* 366, 531-539 (2002)
33. Jason R. B. Dyck, Naomi Kudo, Amy J. Barr, Stephen P. Davies, D. Grahame Hardie and Gary D. Lopaschuk: Phosphorylation control of cardiac acetyl-CoA carboxylase by cAMP-dependent protein kinase and 5'-AMP activated protein kinase. *Eur J Biochem* 262, 184-190 (1999)
34. K. A. Greaves, M. D. Wilson, L. L. Rudel, J. K. Williams and J. D. Wagner: Consumption of soy protein reduces cholesterol absorption compared to casein protein alone or supplemented with an isoflavone extract or conjugated equine estrogen in ovariectomized cynomolgus monkeys. *J Nutr* 130, 820-826 (2000)
35. M. W. Huff and K. K. Carroll: Effects of dietary protein on turnover, oxidation, and absorption of cholesterol, and on steroid excretion in rabbits. *J Lipid Res* 21, 546-548 (1980)
36. S. K. Choi, M. Adachi and S. Utsumi: Improved bile acid-binding ability of soybean glycinin A1a polypeptide by the introduction of a bile acid-binding peptide (VAWWMY). *Biosci Biotechnol Biochem* 68, 1980-1983 (2004)
37. M. R. Lovati, C. Manzoni, E. Gianazza and C. R. Sirtori: Soybean protein products as regulators of liver low-density lipoprotein receptors. I. identification of active b-conglycinin subunits. *J Agric Food Chem* 46, 2474-2480 (1998)
38. W. A. Forsythe: Soy protein, thyroid regulation and cholesterol metabolism. *J Nutr* 125, 619S-623S (1995)
39. W. J. Banz, J. Davis, R. Peterson and M. J. Iqbal: Gene expression and adiposity are modified by soy protein in male Zucker diabetic fatty rats. *Obes Res* 12, 1907-1913 (2004)
40. N. Tachibana, I. Matsumoto, K. Fukui, S. Arai, H. Kato, K. Abe and K. Takamatsu: Intake of soy protein isolate alters hepatic gene expression in rats. *J Agric Food Chem* 53, 4253-4257 (2005)
41. A. R. Tovar, C. Ascencio and N. Torres: Soy protein, casein, and zein regulate histidase gene expression by modulating serum glucagon. *Am J Physiol Endocrinol Metab* 283, E1016-E1022 (2002)
42. W. Huang, C. Wood, M. R. L'Abbe, G. S. Gilani, K. A. Cockell and C. W. Xiao: Soy protein isolate increases hepatic thyroid hormone receptor content and inhibits its binding to target genes in rats. *J Nutr* 135, 1631-1635 (2005)
43. C. W. Xiao, M. R. L'Abbe, G. S. Gilani, G. M. Cooke, I. H. Curran and S. A. Papademetriou: Dietary soy protein isolate and isoflavones modulate hepatic thyroid hormone receptors in rats. *J Nutr* 134, 743-749 (2004)
44. C. W. Xiao, J. Mei, W. Huang, C. Wood, M. R. L'Abbe, G. S. Gilani, G. M. Cooke and I. H. Curran: Dietary Soy Protein Isolate Modifies Hepatic Retinoic Acid Receptor- β Proteins and Inhibits Their DNA Binding Activity in Rats. *J Nutr* 137, 1-6 (2007)
45. J. D. Horton, J. L. Goldstein and M. S. Brown: SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. *J Clin Invest* 109, 1125-1131 (2002)

Soy affects lipid metabolism

46. M. S. Brown and J. L. Goldstein: The SREBP pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor. *Cell* 89, 331-340 (1997)
47. A. R. Tovar, I. Torre-Villalvazo, M. Ochoa, A. L. Elias, V. Ortiz, C. A. Aguilar-Salinas and N. Torres: Soy protein reduces hepatic lipotoxicity in hyperinsulinemic obese Zucker fa/fa rats. *J Lipid Res* 46, 1823-1832 (2005)
48. A. Nagasawa, K. Fukui, M. Kojima, K. Kishida, N. Maeda, H. Nagaretani, T. Hibuse, H. Nishizawa, S. Kihara, M. Waki, K. Takamatsu, T. Funahashi and Y. Matsuzawa: Divergent effects of soy protein diet on the expression of adipocytokines. *Biochem Biophys Res Commun* 311, 909-914 (2003)
49. A. R. Tovar, F. Murguia, C. Cruz, R. Hernandez-Pando, C. A. Aguilar-Salinas, J. Pedraza-Chaverri, R. Correa-Rotter and N. Torres: A soy protein diet alters hepatic lipid metabolism gene expression and reduces serum lipids and renal fibrogenic cytokines in rats with chronic nephrotic syndrome. *J Nutr* 132, 2562-2569 (2002)
50. A. Shukla, C. Brandsch, A. Bettzieche, F. Hirche, G. I. Stangl and K. Eder: Isoflavone-poor soy protein alters the lipid metabolism of rats by SREBP-mediated down-regulation of hepatic genes. *J Nutr Biochem* 18, 313-321 (2007)
51. I. Shimomura, Y. Bashmakov and J. D. Horton: Increased levels of nuclear SREBP-1c associated with fatty livers in two mouse models of diabetes mellitus. *J Biol Chem* 274, 30028-30032 (1999)
52. C. Lavigne, A. Marette and H. Jacques: Cod and soy proteins compared with casein improve glucose tolerance and insulin sensitivity in rats. *Am J Physiol Endocrinol Metab* 278, E491-E500 (2000)
53. K. A. Dooley, S. Millinder and T. F. Osborne: Sterol regulation of 3-hydroxy-3-methylglutaryl-coenzyme A synthase gene through a direct interaction between sterol regulatory element binding protein and the trimeric CCAAT-binding factor/nuclear factor Y. *J Biol Chem* 273, 1349-1356 (1998)
54. M. M. Magana, S. H. Koo, H. C. Towle and T. F. Osborne: Different sterol regulatory element-binding protein-1 isoforms utilize distinct co-regulatory factors to activate the promoter for fatty acid synthase. *J Biol Chem* 275, 4726-4733 (2000)
55. H. B. Sanchez, L. Yieh and T. F. Osborne: Cooperation by sterol regulatory element-binding protein and Sp1 in sterol regulation of low density lipoprotein receptor gene. *J Biol Chem* 270, 1161-1169 (1995)
56. E. Mullen, R. M. Brown, T. F. Osborne and N. F. Shay: Soy isoflavones affect sterol regulatory element binding proteins (SREBPs) and SREBP-regulated genes in HepG2 cells. *J Nutr* 134, 2942-2947 (2004)
57. N. Torres, I. Torre-Villalvazo and A. R. Tovar: Regulation of lipid metabolism by soy protein and its implication in diseases mediated by lipid disorders. *J Nutr Biochem* 17, 365-373 (2006)
58. B. P. Kota, T. H. Huang and B. D. Roufogalis: An overview on biological mechanisms of PPARs. *Pharmacol Res* 51, 85-94 (2005)
59. E. Ip, G. C. Farrell, G. Robertson, P. Hall, R. Kirsch and I. Leclercq: Central role of PPARalpha-dependent hepatic lipid turnover in dietary steatohepatitis in mice. *Hepatology* 38, 123-132 (2003)
60. F. J. Gonzalez: Recent update on the PPAR alpha-null mouse. *Biochimie* 79, 139-144 (1997)
61. O. Mezei, Y. Li, E. Mullen, J. S. Ross-Viola and N. F. Shay: Dietary isoflavone supplementation modulates lipid metabolism via PPARalpha-dependent and -independent mechanisms. *Physiol Genomics* 26, 8-14 (2006)
62. M. Morifuji, C. Sanbongi and K. Sugiura: Dietary soya protein intake and exercise training have an additive effect on skeletal muscle fatty acid oxidation enzyme activities and mRNA levels in rats. *Br J Nutr* 96, 469-475 (2006)
63. C. A. Argmann, C. G. Sawyetz, C. J. McNeil, R. A. Hegele and M. W. Huff: Activation of peroxisome proliferator-activated receptor gamma and retinoid X receptor results in net depletion of cellular cholesteryl esters in macrophages exposed to oxidized lipoproteins. *Arterioscler Thromb Vasc Biol* 23, 475-482 (2003)
64. J. Davis, A. Higginbotham, T. O'connor, N. Moustaid-Moussa, A. Tebbe, Y. C. Kim, K. W. Cho, N. Shay, S. Adler, R. Peterson and W. Banz: Soy Protein and Isoflavones Influence Adiposity and Development of Metabolic Syndrome in the Obese Male ZDF Rat. *Ann Nutr Metab* 51, 42-52 (2007)
65. K. M. Kutty, D. G. Bryant and N. R. Farid: Serum lipids in hypothyroidism—a re-evaluation. *J Clin Endocrinol Metab* 46, 55-56 (1978)
66. C. G. Roberts and P. W. Ladenson: Hypothyroidism. *Lancet* 363, 793-803 (2004)
67. A. Katsurada, N. Iritani, H. Fukuda, Y. Matsumura, N. Nishimoto, T. Noguchi and T. Tanaka: Effects of nutrients and hormones on transcriptional and post-transcriptional regulation of acetyl-CoA carboxylase in rat liver. *Eur J Biochem* 190, 435-441 (1990)
68. A. Katsurada, N. Iritani, H. Fukuda, Y. Matsumura, N. Nishimoto, T. Noguchi and T. Tanaka: Effects of nutrients and hormones on transcriptional and post-transcriptional regulation of fatty acid synthase in rat liver. *Eur J Biochem* 190, 427-433 (1990)
69. A. Katsurada, N. Iritani, H. Fukuda, T. Noguchi and T. Tanaka: Transcriptional and posttranscriptional regulation

Soy affects lipid metabolism

- of malic enzyme synthesis by insulin and triiodothyronine. *Biochim Biophys Acta* 950, 113-117 (1988)
70. G. C. Ness, L. C. Pendleton, Y. C. Li and J. Y. Chiang: Effect of thyroid hormone on hepatic cholesterol 7 alpha hydroxylase, LDL receptor, HMG-CoA reductase, farnesyl pyrophosphate synthetase and apolipoprotein A-I mRNA levels in hypophysectomized rats. *Biochem Biophys Res Commun* 172, 1150-1156 (1990)
71. L. Johansson, M. Rudling, T. S. Scanlan, T. Lundasen, P. Webb, J. Baxter, B. Angelin and P. Parini: Selective thyroid receptor modulation by GC-1 reduces serum lipids and stimulates steps of reverse cholesterol transport in euthyroid mice. *Proc Natl Acad Sci U S A* 102, 10297-10302 (2005)
72. W. M. Pandak, D. M. Heuman, K. Redford, R. T. Stravitz, J. Y. Chiang, P. B. Hylemon and Z. R. Vlahcevic: Hormonal regulation of cholesterol 7alpha-hydroxylase specific activity, mRNA levels, and transcriptional activity *in vivo* in the rat. *J Lipid Res* 38, 2483-2491 (1997)
73. G. C. Ness and Z. Zhao: Thyroid hormone rapidly induces hepatic LDL receptor mRNA levels in hypophysectomized rats. *Arch Biochem Biophys* 315, 199-202 (1994)
74. W. A. Forsythe: Comparison of dietary casein or soy protein effects on plasma lipids and hormone concentrations in the gerbil (*Meriones unguiculatus*). *J Nutr* 116, 1165-1171 (1986)
75. C. A. Barth, Ahrens Scholz, Vrese de and A. Hotze: Difference of plasma amino acids following casein or soy protein intake: significance for differences of serum lipid concentrations. *J Nutr Sci Vitaminol (Tokyo)* 36 Suppl 2, S111-S117 (1990)
76. S. M. Wright and A. M. Salter: The effect of dietary casein and soyprotein on cholesterol metabolism in hamsters. *Biochem Soc Trans* 21, 155S (1993)
77. B. L. Dillingham, B. L. McVeigh, J. W. Lampe and A. M. Duncan: Soy protein isolates of varied isoflavone content do not influence serum thyroid hormones in healthy young men. *Thyroid* 17, 131-137 (2007)
78. B. Bruce, M. Messina and G. A. Spiller: Isoflavone supplements do not affect thyroid function in iodine-replete postmenopausal women. *J Med Food* 6, 309-316 (2003)
79. V. Jayagopal, P. Albertazzi, E. S. Kilpatrick, E. M. Howarth, P. E. Jennings, D. A. Hepburn and S. L. Atkin: Beneficial effects of soy phytoestrogen intake in postmenopausal women with type 2 diabetes. *Diabetes Care* 25, 1709-1714 (2002)
80. R. Mackey, A. Ekanagaki and J. A. Eden: The effects of soy protein in women and men with elevated plasma lipids. *Biofactors* 12, 251-257 (2000)
81. Z. K. Roughead, J. R. Hunt, L. K. Johnson, T. M. Badger and G. I. Lykken: Controlled substitution of soy protein for meat protein: effects on calcium retention, bone, and cardiovascular health indices in postmenopausal women. *J Clin Endocrinol Metab* 90, 181-189 (2005)
82. V. W. Persky, M. E. Turyk, L. Wang, S. Freels, R. Chatterton, Jr., S. Barnes, J. Erdman, Jr., D. W. Sepkovic, H. L. Bradlow and S. Potter: Effect of soy protein on endogenous hormones in postmenopausal women. *Am J Clin Nutr* 75, 145-153 (2002)
83. E. Lydeking-Olsen, J. E. Beck-Jensen, K. D. Setchell and T. Holm-Jensen: Soymilk or progesterone for prevention of bone loss--a 2 year randomized, placebo-controlled trial. *Eur J Nutr* 43, 246-257 (2004)
84. S. M. Potter, J. Pertile and Jimenez Berber: Soy protein concentrate and isolated soy protein similarly lower blood serum cholesterol but differently affect thyroid hormones in hamsters. *J Nutr* 126, 2007-2011 (1996)
85. M. A. Lazar: Thyroid hormone receptors: multiple forms, multiple possibilities. *Endocr Rev* 14, 184-193 (1993)
86. S. Fazio, E. A. Palmieri, G. Lombardi and B. Biondi: Effects of thyroid hormone on the cardiovascular system. *Recent Prog Horm Res* 59, 31-50 (2004)
87. P. M. Yen: Physiological and molecular basis of thyroid hormone action. *Physiol Rev* 81, 1097-1142 (2001)
88. R. E. Weiss, Y. Murata, K. Cua, Y. Hayashi, H. Seo and S. Refetoff: Thyroid hormone action on liver, heart, and energy expenditure in thyroid hormone receptor beta-deficient mice. *Endocrinology* 139, 4945-4952 (1998)
89. H. L. Schwartz, K. A. Strait, N. C. Ling and J. H. Oppenheimer: Quantitation of rat tissue thyroid hormone binding receptor isoforms by immunoprecipitation of nuclear triiodothyronine binding capacity. *J Biol Chem* 267, 11794-11799 (1992)
90. H. Gullberg, M. Rudling, D. Forrest, B. Angelin and B. Vennstrom: Thyroid hormone receptor beta-deficient mice show complete loss of the normal cholesterol 7alpha-hydroxylase (CYP7A) response to thyroid hormone but display enhanced resistance to dietary cholesterol. *Mol Endocrinol* 14, 1739-1749 (2000)
91. H. Gullberg, M. Rudling, C. Salto, D. Forrest, B. Angelin and B. Vennstrom: Requirement for thyroid hormone receptor beta in T3 regulation of cholesterol metabolism in mice. *Mol Endocrinol* 16, 1767-1777 (2002)
92. O. Araki, H. Ying, F. Furuya, X. Zhu and S. Y. Cheng: Thyroid hormone receptor beta mutants: Dominant negative regulators of peroxisome proliferator-activated receptor gamma action. *Proc Natl Acad Sci U S A* 102, 16251-16256 (2005)

Soy affects lipid metabolism

93. T. Miyamoto, A. Kaneko, T. Kakizawa, H. Yajima, K. Kamijo, R. Sekine, K. Hiramatsu, Y. Nishii, T. Hashimoto and K. Hashizume: Inhibition of peroxisome proliferator signaling pathways by thyroid hormone receptor. Competitive binding to the response element. *J Biol Chem* 272, 7752-7758 (1997)
94. P. Chambon: A decade of molecular biology of retinoic acid receptors. *FASEB J* 10, 940-954 (1996)
95. A. Krust, P. Kastner, M. Petkovich, A. Zelent and P. Chambon: A third human retinoic acid receptor, hRAR-gamma. *Proc Natl Acad Sci U S A* 86, 5310-5314 (1989)
96. A. Zelent, C. Mendelsohn, P. Kastner, A. Krust, J. M. Garnier, F. Ruffenach, P. Leroy and P. Chambon: Differentially expressed isoforms of the mouse retinoic acid receptor beta generated by usage of two promoters and alternative splicing. *EMBO J* 10, 71-81 (1991)
97. S. Nagpal, A. Zelent and P. Chambon: RAR- β 4, a retinoic acid receptor isoform is generated from RAR- β 2 by alternative splicing and usage of a CUG initiator codon. *PNAS* 89, 2718-2722 (1992)
98. S. Toma, L. Bonelli, A. Sartoris, E. Mira, A. Antonelli, F. Beatrice, C. Giordano, M. Benazzo, A. Caroggio, A. L. Cavalot, S. Gandolfo, A. Garozzo, G. Margarino, G. Schenone, N. Spadini, Z. Sirotova, F. Zibordi, F. Balzarini, I. Serafini, P. Miani and G. Cortesina: 13-cis retinoic acid in head and neck cancer chemoprevention: results of a randomized trial from the Italian Head and Neck Chemoprevention Study Group. *Oncol Rep* 11, 1297-1305 (2004)
99. W. Jeffrey Petty, Na Li, Adrian Biddle, Rebecca Bounds, Christopher Nitkin, Yan Ma, Konstantin H. Dragnev, Sarah J. Freemantle and Ethan Dmitrovsky: A novel retinoic acid receptor β isoform and retinoid resistance in lung carcinogenesis. *J Natl Cancer Inst* 97, 1645-1651 (2005)
100. A. M. Standeven, R. L. Beard, A. T. Johnson, M. F. Boehm, M. Escobar, R. A. Heyman and R. A. Chandraratna: Retinoid-induced hypertriglyceridemia in rats is mediated by retinoic acid receptors. *Fundam Appl Toxicol* 33, 264-271 (1996)
101. F. Villarroya, R. Iglesias and M. Giral: Retinoids and retinoid receptors in the control of energy balance: novel pharmacological strategies in obesity and diabetes. *Curr Med Chem* 11, 795-805 (2004)
102. L. E. Gerber and J. W. Erdman, Jr.: Hyperlipidemia in rats fed retinoic acid. *Lipids* 16, 496-501 (1981)
103. J. R. Marsden: Effect of dietary fish oil on hyperlipidaemia due to isotretinoin and etretinate. *Hum Toxicol* 6, 219-222 (1987)
104. J. R. Marsden, T. R. Trinick, M. F. Laker and S. Shuster: Effects of isotretinoin on serum lipids and lipoproteins, liver and thyroid function. *Clin Chim Acta* 143, 243-251 (1984)
105. B. Cartmel, J. Dziura, M. R. Cullen, S. Vegso, G. S. Omenn, G. E. Goodman and C. A. Redlich: Changes in cholesterol and triglyceride concentrations in the Vanguard population of the Carotene and Retinol Efficacy Trial (CARET). *Eur J Clin Nutr* 59, 1173-1180 (2005)
106. J. D. Radcliffe, V. L. Imrhan and A. M. Hsueh: The use of soy protein isolate to reduce the severity of 13-cis retinoic acid-induced hypertriglyceridemia. *Cancer Detect Prev* 22, 526-532 (1998)
107. N. Vu-Dac, P. Gervois, I. P. Torra, J. C. Fruchart, V. Kosykh, T. Kooistra, H. M. Princen, J. Dallongeville and B. Staels: Retinoids increase human apo C-III expression at the transcriptional level via the retinoid X receptor. Contribution to the hypertriglyceridemic action of retinoids. *J Clin Invest* 102, 625-632 (1998)
108. Xiao chun Xu, J. Jack Lee, Tsung Teh Wu, Ashraful Hoque, Jeffer A. Ajani and Scott M. Lippman: Increased retinoic acid receptor- β 4 correlates *in vivo* with reduced retinoic acid receptor- β 2 in esophageal squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev* 14, 826-829 (2005)
109. J. M. Lehmann, S. A. Kliewer, L. B. Moore, T. A. Smith-Oliver, B. B. Oliver, J. L. Su, S. S. Sundseth, D. A. Winegar, D. E. Blanchard, T. A. Spencer and T. M. Willson: Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway. *J Biol Chem* 272, 3137-3140 (1997)
110. D. J. Peet, S. D. Turley, W. Ma, B. A. Janowski, J. M. Lobaccaro, R. E. Hammer and D. J. Mangelsdorf: Cholesterol and bile acid metabolism are impaired in mice lacking the nuclear oxysterol receptor LXR alpha. *Cell* 93, 693-704 (1998)
111. R. N. Redinger: The coming of age of our understanding of the enterohepatic circulation of bile salts. *Am J Surg* 185, 168-172 (2003)
112. K. Ishimoto, K. Tachibana, M. Sumitomo, S. Omote, I. Hanano, D. Yamasaki, Y. Watanabe, T. Tanaka, T. Hamakubo, J. Sakai, T. Kodama and T. Doi: Identification of human low-density lipoprotein receptor as a novel target gene regulated by liver X receptor alpha. *FEBS Lett* 580, 4929-4933 (2006)
113. T. Yoshikawa, H. Shimano, M. Amemiya-Kudo, N. Yahagi, A. H. Hasty, T. Matsuzaka, H. Okazaki, Y. Tamura, Y. Iizuka, K. Ohashi, J. Osuga, K. Harada, T. Gotoda, S. Kimura, S. Ishibashi and N. Yamada: Identification of liver X receptor-retinoid X receptor as an activator of the sterol regulatory element-binding protein 1c gene promoter. *Mol Cell Biol* 21, 2991-3000 (2001)
114. S. J. Wakil, J. K. Stoops and V. C. Joshi: Fatty acid synthesis and its regulation. *Annu Rev Biochem* 52, 537-579 (1983)

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115. G. V. Ronnett, E. K. Kim, L. E. Landree and Y. Tu: Fatty acid metabolism as a target for obesity treatment. *Physiol Behav* 85, 25-35 (2005)
116. Y. Xu, G. Bhargava, H. Wu, G. Loeber and L. Tong: Crystal structure of human mitochondrial NAD(P)(+)-dependent malic enzyme: a new class of oxidative decarboxylases. *Structure Fold Des* 7, 877-889 (1999)
117. F. B. Hillgartner, T. Charron and K. A. Chesnut: Alterations in nutritional status regulate acetyl-CoA carboxylase expression in avian liver by a transcriptional mechanism. *Biochem J* 319 (Pt 1), 263-268 (1996)
118. L. Abu-Elheiga, W. R. Brinkley, L. Zhong, S. S. Chirala, G. Woldegiorgis and S. J. Wakil: The subcellular localization of acetyl-CoA carboxylase 2. *Proc Natl Acad Sci U S A* 97, 1444-1449 (2000)
119. N. Iritani, H. Hosomi, H. Fukuda, K. Tada and H. Ikeda: Soybean protein suppresses hepatic lipogenic enzyme gene expression in Wistar fatty rats. *J Nutr* 126, 380-388 (1996)
120. C. Huang and H. C. Freake: Thyroid hormone regulates the acetyl-CoA carboxylase PI promoter. *Biochem Biophys Res Commun* 249, 704-708 (1998)
121. X. C. Luo, K. Park, F. Lopez-Casillas and K. H. Kim: Structural features of the acetyl-CoA carboxylase gene: mechanisms for the generation of mRNAs with 5' end heterogeneity. *Proc Natl Acad Sci U S A* 86, 4042-4046 (1989)
122. H. Aoki, K. Kimura, K. Igarashi and A. Takenaka: Soy protein suppresses gene expression of acetyl-coA carboxylase alpha from promoter PI in rat liver. *Biosci Biotechnol Biochem* 70, 843-849 (2006)
123. J. B. Kim, P. Sarraf, M. Wright, K. M. Yao, E. Mueller, G. Solanes, B. B. Lowell and B. M. Spiegelman: Nutritional and insulin regulation of fatty acid synthetase and leptin gene expression through ADD1/SREBP1. *J Clin Invest* 101, 1-9 (1998)
124. S. Xiong, S. S. Chirala, M. H. Hsu and S. J. Wakil: Identification of thyroid hormone response elements in the human fatty acid synthase promoter. *Proc Natl Acad Sci U S A* 95, 12260-12265 (1998)
125. K. J. Petty, B. Desvergne, T. Mitsuhashi and V. M. Nikodem: Identification of a thyroid hormone response element in the malic enzyme gene. *J Biol Chem* 265, 7395-7400 (1990)
126. L. O. Martinez, S. Jacquet, J. P. Esteve, C. Rolland, E. Cabezon, E. Champagne, T. Pineau, V. Georgeaud, J. E. Walker, F. Terce, X. Collet, B. Perret and R. Barbaras: Ectopic beta-chain of ATP synthase is an apolipoprotein A-I receptor in hepatic HDL endocytosis. *Nature* 421, 75-79 (2003)
127. R. Franzon, F. Chiarani, R. H. Mendes, A. Bello-Klein and A. T. Wyse: Dietary soy prevents brain Na⁺, K⁽⁺⁾-ATPase reduction in streptozotocin diabetic rats. *Diabetes Res Clin Pract* 69, 107-112 (2005)
128. J. Zheng and V. D. Ramirez: Inhibition of mitochondrial proton F₀F₁-ATPase/ATP synthase by polyphenolic phytochemicals. *Br J Pharmacol* 130, 1115-1123 (2000)
129. J. Mei, C. Wood, M. R. L'Abbe, G. S. Gilani, G. M. Cooke, I. H. Curran and C. W. Xiao: Consumption of soy protein isolate modulates the phosphorylation status of hepatic ATPase/ATP synthase beta protein and increases ATPase activity in rats. *J Nutr* 137, 2029-2035 (2007)
130. M. Sugano, S. Goto, Y. Yamada, K. Yoshida, Y. Hashimoto, T. Matsuo and M. Kimoto: Cholesterol-lowering activity of various undigested fractions of soybean protein in rats. *J Nutr* 120, 977-985 (1990)
131. M. F. Wang, S. Yamamoto, H. M. Chung, S. Y. Chung, S. Miyatani, M. Mori, T. Okita and M. Sugano: Antihypercholesterolemic effect of undigested fraction of soybean protein in young female volunteers. *J Nutr Sci Vitaminol (Tokyo)* 41, 187-195 (1995)
132. A. Tovar, N. Manzano and N. Torres: Metabolism of cholesterol and fatty acids in nephrotic syndrome and its regulation by sterol regulatory element binding proteins (SREBP's). Effect of soy protein consumption. *Gac Med Mex* 141, 407-415 (2005)
133. L. B. Nguyen, S. Shefer, G. Salen, J. Y. Chiang and M. Patel: Cholesterol 7alpha-hydroxylase activities from human and rat liver are modulated *in vitro* posttranslationally by phosphorylation/dephosphorylation. *Hepatology* 24, 1468-1474 (1996)
134. M. R. Lovati, C. Manzoni, E. Gianazza, A. Arnoldi, E. Kurowska, K. K. Carroll and C. R. Sirtori: Soy protein peptides regulate cholesterol homeostasis in Hep G2 cells. *J Nutr* 130, 2543-2549 (2000)
135. M. R. Lovati, C. Manzoni, A. Canavesi, M. Sirtori, V. Vaccarino, M. Marchi, G. Gaddi and C. R. Sirtori: Soybean protein diet increases low density lipoprotein receptor activity in mononuclear cells from hypercholesterolemic patients. *J Clin Invest* 80, 1498-1502 (1987)
136. M. R. Lovati, C. Manzoni, P. Agostinelli, S. Ciappellano, L. Mannucci and C. R. Sirtori: Studies on the mechanism of the cholesterol lowering activity of soy proteins. *Nutr Metab Cardiovasc Dis* 1, 18-24 (1991)
137. C. R. Sirtori, G. Galli, M. R. Lovati, P. Carrara, E. Bosisio and M. G. Kienle: Effects of dietary proteins on the regulation of liver lipoprotein receptors in rats. *J Nutr* 114, 1493-1500 (1984)

Soy affects lipid metabolism

138. M. R. Adams, D. L. Golden, M. S. Anthony, T. C. Register and J. K. Williams: The inhibitory effect of soy protein isolate on atherosclerosis in mice does not require the presence of LDL receptors or alteration of plasma lipoproteins. *J Nutr* 132, 43-49 (2002)

Abbreviations: ACC: acetyl-CoA carboxylase; CYP7A1: cholesterol-7alpha hydroxylase; FAS: fatty acid synthase; HMG-CoAr: hydroxymethylglutaryl-CoA reductase; HMG-CoAs: hydroxymethylglutaryl-CoA synthase; ISF: isoflavones; LDL: low density lipoprotein; LXR: liver X receptor; ME: malic enzyme; PPAR: peroxisomal proliferators activated receptor; RAR: retinoic acid receptor; SPI: soy protein isolate; SREBP: sterol regulatory element binding protein; T3: triiodothyronine; T4: thyroxine; TR: thyroid hormone receptor.

Key Words: Soy proteins, Isoflavones, Transcription factors, Lipid metabolism, Gene expression, Review

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