# Therapeutic implications of the nuclear factor-kappaB/nuclear receptor cross-talk

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# 1. ABSTRACT

More and more evidence reveals that the transcription factor NF-kappaB plays a critical role in tumor development and progression and that it may constitute the missing link between inflammation and cancer. It turned out that many of the well known cancer drugs exert their anti-tumoral effect at least in part through modulating the activity of NF-kappaB. The potential of nuclear receptors to modulate the activity of this widespread transcription factor has repeatedly been reported and illustrates their enormous therapeutic potential. However, the efficacy of these liganded receptors is overshadowed by the occurrence of unwanted effects owing to their broad range of actions. Accordingly, researchers pursue the ambition to improve the specificity of nuclear receptor modulators. In this review we have explored the molecular mechanisms by which nuclear receptors interfere with NF-kappaB signalling and quoted the therapeutic implications of their cross-coupling. Strategies that are explored at the moment and that may hold great potential for the future are extensively reviewed.

# 2. NUCLEAR RECEPTORS

Nuclear receptors (NRs) are transcription factors (TFs) with essential and non-redundant roles in fundamental biological processes, including growth, development, homeostasis and cell death. The fact that these receptors are evolutionary related is reflected by their structural similarities. Indeed, most nuclear receptor family members contain three functional domains, i.e. an N-terminal transactivation domain, a DNAbinding domain (DBD) and a C-terminal ligand-binding domain (LBD) (1). Sequencing of the human genome has led to the identification of 48 NRs that can be divided in two main groups. Firstly, the orphan receptors, which can be further divided into two subgroups, being 1) true orphans, for which ligands are unknown or may not exist, or 2) adopted orphans, for which candidate ligands have only recently been identified (e.g. peroxisome proliferator-activated receptors, PPAR). Secondly, the liganded receptors, which contain 24 members, including the glucocorticoid receptor (GR), the estrogen receptor (ER), the mineralocorticoid receptor (MR), the progesterone receptor (PR) and the androgen receptor (AR) (1).

GR is expressed in almost all tissues of the human body. However, the levels of GR protein, of which different splice- and translation variants occur, are tissueand cell cycle-specifically regulated (2-6). Cortisol or hydrocortisone is the major glucocorticoid (GC) in the human body. Due to its prominent role in a variety of biological actions, including carbohydrate, protein and fat metabolism, as well as its modulatory role in the central nervous system, hematopoietic, renal and immune systems, its expression by the adrenal cortex is tightly controlled via a negative feedback mechanism on the hypothalamopituitary-adrenal axis (7). Although cortisol can also bind the MR, this binding is inhibited in mineralocorticoid-target tissues due to the presence of 11beta-hydroxysteroid dehydrogenase 2, which converts cortisol into the inactive metabolite cortisone (8), thereby making GR its primary target in those tissues. Besides GRs, ERs, as well as PPARs, are known to regulate the inflammatory response. The effects of estrogens span further than the immune system. This is exemplified by their influence not only on reproduction, which is widely recognized, but also on skeletal, cardiovascular and central nervous systems (9-11). These hormones act through activation of two receptors, namely ERalpha and ERbeta (12), which can form homoor heterodimers. PPARs, on the other hand, form a family of three nuclear receptors (PPARalpha, PPARbeta/delta, PPARgamma). They respond to various fatty acids (13-15) and are differentially distributed to distinct tissues (16). To be able to exert their effects on specific metabolic processes. PPARs form a heterodimer with the retinoid X receptor (RXR). Furthermore, PPARs function as regulators of cell proliferation and apoptosis, two cellular mechanisms of which deregulation can have detrimental outcomes and can result in diseases, such as cancer (15).

In the absence of ligands, GR, MR, PR and ER are kept inactive in the cytoplasm of the cell by chaperone proteins, such as heat shock proteins (hsp) and immunophilins (17). However, recently it became clear that this is a simplified model and that, in fact, a continuous shuttling of the receptors between the two cellular compartments occurs (18, 19). Only after binding of ligands to their corresponding receptor the dynamic equilibrium is disturbed, thereby resulting in a net nuclear shift. Indeed, due to their small, lipophilic nature, ligands such as steroid hormones (e.g. GCs, estrogens, progesterone, mineralocorticoids, androgens), fatty acids and prostaglandins diffuse freely through the cell membrane. Once inside the cell, these hormonal and metabolic substances bind to their corresponding receptor and induce a conformational change. A nuclear localization signal is exposed, thereby allowing the receptor to translocate to the nucleus and to influence transcription. More specifically, this conformational change allows nuclear receptors to recruit coactivator complexes, via their activation function 2 (AF-2) domain, which consists of a short conserved helical sequence within the C-terminus of the LBD (20). These coactivator complexes are composed of chromatin-modifying proteins, such as factors with ATPdependent chromatin remodelling (e.g. BRG-1, BRM) or histone arginine methyltransferase activities (21), molecular scaffolds that assemble cofactor complexes (e.g.

PPARγ coactivator-1 (PGC-1)), as well as members of the p160 family (e.g. steroid receptor coactivator 1 (SRC-1), transcription intermediary factor 2 (TIF2/GRIP-1) and p300/cAMP-responsive element-binding protein (CBP)). Coactivator molecules such as CBP, p300 and SRC-1 modulate the activity of the transcription apparatus through their histone acetyltransferase (HAT) activity (22, 23). Core histones are posttranslationally modified, thereby changing their electrical charge and pushing the DNA in a more relaxed chromatin structure. It is believed that the inverse process, deacetylation of histones by HDACs (histone deacetylases) results in a more condensed chromatin structure, thereby reducing the access of TFs to their binding sites and repressing transcription of target genes (24). It has been suggested that at the end of this initial chromatin-modifying step, the p160 family members are acetylated, thereby loosing their ability to interact with the receptor, or alternatively, that these coactivators are degraded by the proteasome (25, 26). Once the first cofactor complex is disassembled, nuclear receptors interact with members of the Mediator complex (TRAP/DRIP/ARC), which directly contact the basal transcriptional machinery (27). A bridge is formed with the RNA polymerase II holoenzyme that can subsequently be recruited onto the promoter. However, it has been shown that different ligands (28-30) or different NRs (31, 32) can exhibit a preference when it comes to the recruitment of different coactivators. In addition, the spatial and temporal modes in which the process of cofactor recruitment occurs can vary for different NRs and for different promoters. For example. Métivier and colleagues (33) suggested that at the ER-responsive pS2 promoter, the coactivators p300 and SRC-1 were first recruited to induce histone acetylation, followed by the recruitment of ERalpha and the Mediator complex. On the other hand, recruitment of AR and cofactors onto the PSA regulatory regions (34) support a combinatorial model. A similar model was already available for the cathepsin D promoter onto which p160 proteins, ERalpha and DRIP/TRAP are recruited in a combinatorial mode (35). Furthermore, when ligand is added for longer time periods, a cyclic and dynamic recruitment of coactivator complexes to the ER-responsive promoter can be observed (33). In contrast, association of the AR-coactivator complex on PSA regulatory regions gradually increases, with a maximum occupancy at 16 hr, followed by a gradual decline (34). Taking into account that specific cell types (36) or diverse promoters can display alternative requirements of different coactivators that can be recruited sequentially, combinatorially or in parallel, it seems logic that in this way a code for obtaining tissue and gene specificity is created. Moreover, the possibility of a rapid NR/DNA interaction turnover (37-39), together with the fact that coactivators and NRs themselves are targets for modification by different signal transduction pathways (reviewed in (40-42)) creates a frame in which it is possible for a cell to very quickly respond to changing environmental factors.

It has been proposed that the segregation of nuclear receptors in different subcellular compartments acts as an important regulatory checkpoint. It is hypothesized, e.g. for ER, that this mechanism of cellular segregation

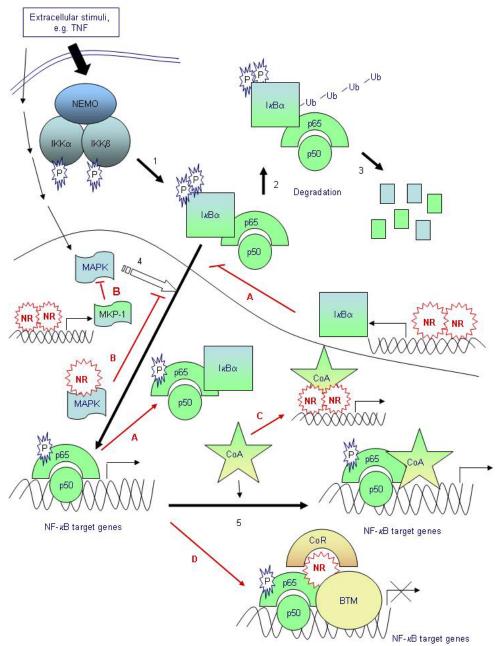


Figure 1. Interference of nuclear receptors (NRs) with the canonical pathway of NF-kappaB activation. Upon triggering with stimuli, such as TNFalpha, IkappaBalpha is phosphorylated by the activated IKK complex (1), and subsequently ubiquitinated (2) and degraded (3). NF-kappaB (p65/p50) enters the nucleus and after being post-translationally modified by kinase pathways, such as the TNF-activated MAPK pathway (4), it induces gene transcription (5). Different models have been hypothesized to describe the cross-coupling between NF-kappaB and NRs. Note that some of these mechanisms may be very cell type-dependent and this complication is discussed in more detail in the text: (A) NRs may inhibit NF-kappaB release through up-regulation of IkappaBalpha, thereby inhibiting NF-kappaB/DNA binding, (B) NRs may inhibit MAPK signalling via upregulation of the MAP kinase phosphatase MKP-1 or via direct interactions, thereby preventing post-translational modifications of NF-kappaB, (C) NRs may compete with NF-kappaB for co-activators, and (D) NRs inhibit the formation of a functionally active transcription complex and important non-exclusive mechanistic aspects hereof may include: a direct physical interaction between NRs and NF-kappaB, interference with the basal transcription machinery (BTM) and a cofactor exchange, namely the removal of coactivators (CoA) and the recruitment of corepressors (CoR). Non-genomic effects are at this moment not depicted in the figure due to the lack of a conclusive model.

may allow the receptor to exert both genomic (nuclear) and non-genomic (cytoplasmic) activities (43, 44). Recently, Carrigan and co-workers (45) have defined a nuclear retention signal in the hinge region of GR. Their data suggest that active nuclear retention of GR acts as a strong inducer of GR transcriptional activity (45). However, for other NRs, for instance PPARs, which can be found constitutively in the nucleus, even in the absence of ligand, a different mechanism must occur. In the unliganded state, DNA-bound PPAR is kept inactive through interaction with a nuclear corepressor complex, containing the nuclear corepressor (NCoR), silencing mediator of retinoic-acid and thyroid hormone receptors (SMRT) and HDACs, which keep the chromatin in a condensed state. In analogy with the previous model, ligand induction results in a corepressor/coactivator switch (46), a process called "derepression" (47). The repressor complex is dissociated, through ubiquitination and degradation of NCoR, and substituted by a coactivator complex with HAT activity, thereby allowing transcriptional activation (47).

In general, multiple modes of action have been revealed and several common mechanisms exist. Ligandactivated NRs can homo- or heterodimerize and can activate the transcription of responsive genes via direct DNA binding to responsive elements in the promoter (transactivation). In contrast, liganded receptors can also regulate the activities of other major signalling pathways (transrepression) (48, 49), such as those driven by nuclear factor-kappa B (NF-kappaB), activator protein 1 (AP-1) (50-52), cAMP-responsive element-binding protein (CREB), signal transducers and activators of transcription (STATs) (53) or interferon regulatory factor 3 (IRF3) (54). It is mainly the latter characteristic of the NRs, in particular their interference with the functionality of NF-kappaB, that explains their success as drug targets, and this topic will be discussed further in this review.

# 3. NUCLEAR FACTOR-KAPPA B

The mammalian NF-kappaB family consists of a heterogeneous and commonly expressed group of TFs. The family contains 5 members that can be divided in two subgroups. The first class is composed of members that comprise a C-terminal transcription activation domain (RelA (p65), RelB and c-Rel), whereas the second class (NF-kappaB1 (p50/p105), NF-kappaB2 (p52/p100)) does not. Each member contains a Rel homology domain (RHD) that mediates DNA binding, dimerization, as well as interaction with members of the "inhibitor of NF-kappaB" (IkappaB) family, which keep NF-kappaB dimers in the cytoplasm of the cell. NF-kappaB can be activated by a wide array of extracellular stimuli, including cytokines, viruses, oxidative stress, phorbol esters, lipopolysaccharide and B- and T-lymphocyte activation (55-57). After activation of cell surface receptors, distinct signal transduction pathways may be activated, but most of these stimuli seem to converge on the level of IkappaB (56, 57). Whether it is via activation of the IkappaB kinase complex (IKK) (canonical pathway) or via the activation of the p38activated serine/threonine kinase casein kinase II (CKII) (after exposure to UV-light; atypical pathway) (58, 59),

IκB is phosphorylated and subsequently degraded (60-62). Indeed, although both kinases phosphorylate IkappaBalpha different residues, i.e. IKK phosphorylates IkappaBalpha on N-terminal sites, while CKII targets a cluster of C-terminal sites, both pathways finally converge on the level of IkappaBalpha (59). The liberated NFkappaB, which is predominantly composed of a p65 and p50 heterodimer, enters the nucleus and regulates the transcription of a diverse subset of genes encoding, others, cytokines, chemokines, amongst metalloproteinases (MMPs), cell adhesion molecules, inducible NO-synthase (iNOS) and cyclooxygenase 2 (COX-2) (63-67). Alternatively, a select group of stimuli, e.g. B-cell activating factor (BAFF), lymphotoxin beta (LTbeta), CD40 ligand and viruses, such as human T-cell leukaemia (HTLV) and Epstein-Barr (EBV) activate NFkappaB2 via an alternative, non-canonical pathway, i.e. in an IkappaB-independent manner, but via activation of NFkappaB-inducing kinase (NIK) and IKKalpha, p100 is processed into p52 (58). Finally, recent studies highlighted the existence of another NF-kappaB-activating pathway that is signalling inside-out, i.e. from the nucleus to the cytoplasm. Indeed, although it was previously thought that DNA-damaging agents signalled via the same pathway, referred to as the atypical pathway, it became clear that these stimuli could not be grouped and in fact activate different patways. Recently gained insights are extensively reviewed in (58, 68). In brief, in contrast to UV-light that signals to NF-kappaB in an IKK-independent manner (as described above), other DNA-damaging agents, including genotoxic stress (i.e. the formation of double stranded breaks), oxidative stress, heat or electric shocks, do signal via a member of the IKK complex, namely via the NFkappaB essential modulator (NEMO or IKKgamma). It is believed that, upon DNA damage, p53-inducible death domain-containing protein (PIDD) and receptor-interacting protein 1 (RIP-1) translocate to the nucleus were they sumoylate NEMO (69). In parallel, the ataxia telangiectasia mutated (ATM) kinase is activated, which recognizes and phosphorylates the sumoylated NEMO. Subsequently, NEMO gets mono-ubiquitinated (70, 71) a modification that serves as a nuclear export signal instead of being a tag for degradation by the proteasome. Finally, NEMO returns to the cytoplasm, where it triggers the activation of the IKK complex (72).

Different studies agree that NF-kappaB is highly activated at sites of inflammation (73-75), thereby further supporting the role of NF-kappaB as an important regulator of inflammation. However, it is only recently that NFkappaB was pinpointed as being the missing link between inflammation and cancer (76). Indeed, several proinflammatory cytokines and chemokines (e.g. TNF, IL-6, IL-1, CXCL8) are under the transcriptional control of the IKKbeta-dependent NF-kappaB activation pathway and are associated with tumor development and progression (77-80). Furthermore, the transcription of proteins that are crucial for tumor cells to proliferate, to invade and to metastasize, such as anti-apoptotic proteins, growth factors (e.g. vascular endothelial growth factor (VEGF)) and MMPs, is NF-kappaB-regulated (77, 78, 80-83). As an example, the NF-kappaB-regulated pleiotropic cytokine IL-

6 is believed to be a crucial player, standing at the crossroads between inflammation and cancer. This is due to its ability to act as a paracrine/autocrine growth factor of many tumor cells and is further supported by its implication in tumor progression, angiogenesis, invasion and motility (84). Furthermore, IL-6 can epigenetically modulate gene expression via reduction of the miRNA, miR-370, that controls the expression of the oncogene MAP3K8 (85). The recently gained insights that NF-kappaB is essential for promoting inflammation-associated cancer, paves the way for the use of anti-inflammatory agents also into anti-cancer protocols. As such, agents that target the NF-kappaB activation pathway, such as proteasome inhibitors and upstream kinase inhibitors, display anti-cancer properties in clinical or preclinical studies (86). Alternatively, a variety of studies have shown a mutually antagonistic cross-talk between activated NRs and NF-kappaB (51). The mutual repressive activities between NF-kappaB and PR were recently confirmed and this antagonism is shown to be important for the downregulation of cytokine expression in human leukocyte cells (87, 88). More information is available concerning the cross-coupling of GR, ER, AR and PPAR with NF-kappaB. The importance of ER/NF-kappaB cross-talk has been elucidated using ER-positive and ERnegative breast cancer cells. Whereas active DNA-bound NF-kappaB was absent in the ER-positive cells, a constitutively activated transcription factor could be found in ER-negative cells (89). In the same line, in AR-negative prostate cancer cells NF-kappaB was also found to be constitutively active (90). In addition, research has revealed that GR overexpression in the epidermis of transgenic mice dramatically inhibited skin carcinogenesis. The mechanistic basis of the tumor-suppressor effect of GR lies in the interference of GR with NF-kappaB (91). A similar study was performed in prostate cancer cells and suggested that GR inhibits multiple signalling pathways and TFs involved in proliferation and transformation, including NF-kappaB, thereby explaining the tumor-suppressive role of GR in the prostate (92).

# 4. CROSS-TALK MECHANSIMS

### 4.1. Cytoplasmic Models

# 4.1.1. Upregulation of inhibitors

One convenient way of how NRs could counteract the activity of NF-kappaB would be through upregulation of the inhibitor protein IkappaBalpha. Concomitantly, more than a decade ago Scheinman (93, 94) and Auphan (95) proposed that, after administration of GC, p65 is sequestered in an inactive cytoplasmic form, thereby reducing the NF-kappaB/DNA binding. These observations found follow-ups in other NR research fields, such as for ER, AR, PPAR (extensively reviewed in (50)) and retinoidrelated orphan receptor alpha I (96). For instance, IkappaBalpha levels were found to be higher in ER-positive compared to ER-negative breast cancer cell lines (97). In addition, in rats suffering a transient cerebral ischemia, an increase in phosphorylated IkappaBalpha is coupled to activation of NF-kappaB (98). However, more and more evidence is arising showing that IkappaBalpha upregulation is not the main mechanism by which NRs exert their antiinflammatory effects. First, the question still remains how

GC could up-regulate IkappaBalpha when no classical glucocorticoid response element (GRE) can be found in the promoter region. However, an alternative mechanism, which does not require DNA-binding, may occur. Such a mechanism has been described before for PPARalpha, showing that this NR requires DRIP205, recruited onto the Sp1 sites flanking the kappaB site, to regulate IkappaBalpha expression. These data indicate that even in the absence of its functional response elements, PPARalpha may positively regulate gene expression (99). It seems not unreasonable that a similar mechanism may occur for other NRs. Nevertheless, it has been shown that even in the absence of new protein synthesis GR was able to efficiently repress NF-kappaB activity (100). Furthermore, using in vivo footprinting and Chromatin Immunoprecipitation (ChIP), it was revealed that NF-kappaB remains bound to the ICAM and IL-8 promoter respectively, even under conditions of gene repression by GCs (101, 102). Also the initial finding that estradiol (E2) inhibits the expression of NF-kappaB-driven genes by interfering with the binding of NF-kappaB to the DNA (103), could not be generalized. In fact, recent ChIP data showed that E2 treatment did not impair the binding of the p50 or p65 subunit of NF-kappaB to the TNFalpha promoter (104).

The seemingly contradictory results obtained by different research groups may reflect the tissue specificity of this mechanism, whereby different characteristics prone to the cell (e.g. a cell-specific subset of cofactors, differential GC effects on alternative pathways in different cells) eventually define the importance of IkappaBalpha upregulation (50).

# 4.1.2. Interference with other signal transduction pathways

Although the picture of NF-kappaB regulation seemed clarified for years and deemed relatively simple, it is only a few years ago that research has unravelled a more complex and sophisticated image of NF-kappaB activation. Post-translational modifications of the different members of the NF-kappaB family seem to be responsible for finetuning NF-kappaB activity (reviewed in (58)). This additional level of regulation provides a point for cross-talk with other signalling pathways, which, for some examples, may be under the tight control of NRs. Indeed, previous studies reported on the importance of ERK and p38, as well as the subsequent phosphorylation of p65 at Ser276 by the downstream nuclear kinase MSK-1, for obtaining fullblown NF-kappaB activation (105, 106). Since GC are able to upregulate MKP-1, a dual specificity phosphatase (DUSP) which in turn de-phosphorylates and thus inactivates ERK and p38, this might constitute an alternative mechanism by which GC exert their antiinflammatory role (107-109). In addition, it is thought that p38 MAPK can post-transcriptionally regulate the expression of a variety of pro-inflammatory genes. Indeed, although steroids may influence the levels and activity of tristetraprolin (TTP), a factor that regulates mRNA stability and thus the expression of certain inflammatory genes (109-111), steroids can alternatively alter the stability of pro-inflammatory mRNAs via inhibition of p38. Through activation of its substrate MAPKAPK-2, p38 can stabilize

various mRNAs, such as IL-6, IL-8, COX-2 and TNFalpha, and this process seems to depend upon the presence of an A/U-rich repeat (ARE) (112-114). The role of MKP-1 induction after GC treatment has been reported in synovial fibroblasts (115) and its significance was further elaborated by Abraham and colleagues, who show that in DUSP knock-out mice the in vivo anti-inflammatory effects of DEX on zymosan-induced inflammation is impaired (116). However, taking into account that the inhibition of novel protein synthesis does not hamper repression of IL-6 production by GC (100), reasons that GC-induced MKP-1 upregulation is not the only mechanism explaining the transrepressive effects of GC or, alternatively, that the importance of this mechanism is again cell type- and/or gene-dependent. This is in agreement with the observation that the dependency of DEX-mediated MKP-1 upregulation for the concomitant repression of inflammatory mediators can in fact vary for the different markers. Indeed, it is shown for IL-1alpha, COX-2 and IL-12 that the MKP-1 dependency decreases from highly dependent to not at all dependent (116), reviewed in (117). Alternatively to MKP-1 upregulation, GR might regulate the function of the MAPK family members through direct protein-protein interactions. Such a mechanism has already been described by Bruna and co-workers who have characterized a hormone-regulated JNK docking site in the GR ligandbinding domain. From their results these authors conclude that by binding to JNK, GR inhibits the interaction of JNK with its upstream kinase MKK-7, thereby inhibiting the activation of this MAPK. As a consequence inactive JNK accumulates on the AP-1-bound response elements of the cjun gene (118). A direct interaction mechanism has also been described for ER-mediated activation of the Src/p21/ERK pathway, this via interaction of estrogenactivated ER with c-Src (119). However, such a direct interaction has, to our knowledge, not yet been described between p38 and ERK on the one hand and GR on the other hand.

Vice versa, the activity of NRs can be modulated by other pathways, such as the cAMP-dependent protein kinase (PKA) pathway. Not only has it been shown that GR is an effective substrate for phosphorylation mediated by the catalytical subunit of PKA (PKAc) (120), in addition, it has been reported that PKAc/retinoid acid receptor crosstalk occurs through direct phosphorylation of the receptor (121, 122), and furthermore, that PKAc regulates dimerization of human ER-alpha (123). A more recent study however, reported on the importance of PKAc in the cross-talk between GR and NF-kappaB (124). From their data. Doucas and co-workers conclude that while PKAc potentiates GR-dependent transcription, it attenuates the cross-repression between NF-kappaB and GR. Moreover, they suggest that both TFs may already interact in the cytoplasm of the cell (124). A similar observation was made by Widén et al., who showed by means of immunoaffinity chromatography or immunoprecipitation and western blotting that the p65/p50/IkappaBalpha complex already interacts with GR in the cytoplasm, even in the absence of a hormonal ligand or a pro-inflammatory signal (125). However, just as for the IkappaBalpha and MKP-1 mechanisms, data exist claiming that a restricted cytoplasmic event is unlikely to be the main mechanism explaining the transrepression activity of GR. Indeed, by means of an exlusively nuclear set-up using a Gal4 (DBD)-p65 fusion protein, which could constitutively activate a Gal4-dependent luciferase construct, it was shown that the repressive effects of DEX were not influenced by the lack of interferences by upstream events (100). Although this experiment does not rule out the occurrence of an interaction between NF-kappaB and GR in the cytoplasm of the cell, unarguably it does indicate that this cytoplasmic encounter may not entirely explain the cross-coupling between both TFs. Further evidence supporting nuclear models will be discussed below.

### 4.2. Nuclear models

# 4.2.1. From physical interaction to the basal transcription machinery

In analogy with the transactivation function of GR, it was proposed almost 20 years ago that transcriptional repression by GCs occurs via interaction of GR with a negative GC response element (nGRE) in the promoter regions of repressed genes, analogous to the prototype described in the proximal region of the proopiomelanocotin (POMC) promoter (126, 127). However, in 1997, together with the characterization of a Nur response element (NurRE) which binds Nur77 (NGFI-B) dimers but not GR molecules, a novel POMC-promoter target for GC-mediated repression was identified (128). Later on it turned out that at the level of the NurRE, NGFI-B and GR interact via their DBDs; in the absence of GR DNA binding and GR homo-dimerization (129). Furthermore, it was shown that this is not an exclusive mechanism for NGFI-B, but instead, that all members of the orphan nuclear receptor Nur77 subfamily are able to interact with GR (129). At the prostate-specific antigen promoter an alternative mechanism has been described for AR. In this setting, the cross-modulatory activity of AR with p65 results from the binding of both factors to a common cis-DNA element (130).

Actually, the above-described physiological interaction between Nur77 and GR, closely resembles a model that is proposed for transrepression between GR and NF-kappaB (129). Indeed, first evidence concerning a direct physical interaction between the latter 2 TFs came from co-immunoprecipitation studies (131). By means of point mutants it became clear that GR interacts with the RHD and C-terminal transactivation domain of RelA via its DBD (100, 132). More specifically, the GR zinc-binding region (ZBR), which includes the DNAbinding and dimerization functions of the receptor, is sufficient to associate with the RelA subunit of NF-kappaB in vivo (102). However, in analogy with the model described for Nur77, also here DNA binding of the receptor per se was not necessary (131, 133). Some evidence is gathered that a similar mechanism also exists in the field of ER. One group has reported the ability of ERalpha to stably associate with DNA-bound NF-kappaB in gel shift experiments under conditions that NF-kappaB-driven gene expression is inhibited (134). Another group has elucidated that the NF-kappaB repressive activity of ER is fully dependent on the complete ER DBD. Again a mutation

approach pointed out that nucleotide residues within or overlapping the DBD of ER are essential to maintain ER's ability to repress the expression of the NF-κB-driven IL-6 gene (135). In a recent manuscript it was clarified that the RHD of p65 is necessary for the interaction of p65 with ER *in vitro* and *in vivo* (136). The occurrence of protein-protein interactions between PPARalpha and NF-kappaB was first brought to the attention by Delerive and co-workers in 1999 (137). In addition, it has been suggested that a weak interaction, as observed for AR and RelA, may explain their reciprocal negative cross-talk (138).

These pieces of evidence were used to further elucidate the role of this protein-protein interaction and it was questioned whether there would be a role for the basal transcription machinery (BTM) in the transrepression activity of NRs. Indeed, it was found by Meyer and colleagues that the repression of the osteocalcin gene is the result of a GRE overlapping with the TATA-box, thereby preventing the assembly of a functional basal transcription machinery (139). The significance of the TATA-box determining responsiveness towards GR-mediated transrepression was confirmed using NF-kappaB-driven recombinant constructs (140). Conclusive evidence for an interfering role of GR in the assembly of a functional BTM was given by Nissen & Yamamoto (102). Since TNF-alpha stimulated the formation of a pre-initiation complex (PIC) at the IL-8 and ICAM promoters, as well as the phosphorylation of the largest subunit of RNA polymerase II (pol II) at serines 2 and 5 situated in its carboxy-terminal domain (CTD), these authors reasoned that GR might interfere with these essential modifications. Indeed, although GR did not interfere with PIC assembly under repressive conditions, it did interfere with the phosphorylation of Ser 2 of the pol II CTD. In a follow-up paper the group of Yamamoto described that promoterspecific gene regulation by the glucocorticoid receptor resulted from the formation of diverse regulatory complexes at the different promoters (141). These results strongly suggest that GR might interfere with the recruitment of different cofactors to efficiently repress gene transcription (102).

### 4.2.2. Getting specificity: a role for cofactors?

Upon realizing that NRs counteract the repression of only a subset of NF-kappaB-driven genes we are left with the question of how signal-, promoter- and cellspecific modulation of the inflammatory response is obtained by these receptors (142). TFs themselves cannot drive inflammatory responses, but instead need the help of an overlapping set of coactivator proteins, thereby raising the possibility that these cofactors may carry the secret of context-dependent regulation. Since at least some of these coactivator proteins are present in cells at functionally limiting concentrations (143), it has been proposed that NRs compete with NF-kappaB for coactivators, such as CBP/p300. This cofactor competition model found support by data gathered in the AR field and the GR field (144, 145). However, a major draw-back of this model is its incapability to explain the gene specificity of transrepression, since a plethora of TFs that are homed in a cell converge at the level of CBP/p300 for their

transcriptional activity (51, 140). Experimental evidence minimizing the importance of cofactor competition came from the observation that GC repression of p65-mediated gene expression is not relieved by overexpression of the coactivator molecules CBP/p300 and SRC-1. In addition, a nuclear GAL4-p65 point mutant, defective in CBP recruitment, could still be functionally repressed by GR (140).

Instead of competing for coactivators, it has been proposed that NRs exert their repressive effects on gene expression through the recruitment of HDACs. A role for these corepressors was elaborated using the HDAC inhibitor trichostatin A (146, 147). Subsequently, by means of a GR antagonist, namely RU486, Ito and co-workers were able to pin-point the importance of HDAC-2. By showing that the recruitment of HDAC-2 to the NF-kappaB complex was impaired after RU486 induction, these authors concluded that HDAC-2 recruitment is an essential step in the transrepression activity of GR (148). More recently, the importance of HDAC-2 was further highlighted through the use of RNA interference. Loss of HDAC-2 inhibited the association between GR and NFkappaB. This can be understood by taking into account that GR becomes acetylated after ligand binding and by accepting that HDAC-2-mediated GR deacetylation is necessary to enable binding of GR to NF-kappaB (149). In the same study it was shown that overexpression of HDAC-2 in GC-insensitive alveolar macrophages from patients with COPD is able to restore GC sensitivity. A similar role for HDAC-2 was described in the cross-repression between NGFI-B and GR. The group of Drouin has reported on the importance of Brg1, the ATPase subunit of the Swi/Snf complex, for in vivo stabilization of the interactions between GR and NGFI-B on the one hand and between GR and HDAC-2 on the other hand. Whereas Brg1 resides constitutively on the POMC promoter, ligand induction is necessary for the recruitment of GR and HDAC-2, thereby resulting in histone H4 deacetylation and inhibition of pol II clearance from the promoter. In addition, there is convincing evidence showing that the lack of nuclear expression of both Brg1 and HDAC-2 gives rise to the occurrence of GC resistance (150). The significance of corepressors in NR-mediated transrepression mechanisms was further supported by studies of PPARgamma in macrophages. Genes that are subject to transrepression by PPARgamma have promoters that interact in their basal state with corepressor complexes containing NCoR-HDAC-3-TBL. In the absence of PPARgamma ligand, LPS signalling results in the clearance of this repressor complex from the promoter, thereby enabling NF-kappaB and its coactivator complex to bind. However, when PPARgamma is activated by ligand, a conformational change is elicited that enables SUMOylation of its ligand-binding domain, thereby stabilizing the interaction between the receptor and the NCoR/HDAC complex. As a consequence, LPS signalling fails to relieve the repressive effects on transcription (151).

Since different promoters may have a preference for different coactivators and corepressors, the above findings may in part explain the gene specificity of this mechanism. In transactivation mechanisms, the mediator subunits MED14 and MED1 have been implicated in transcriptional regulation and seem to be used by the GR in a gene-specific manner (152). Evidence accumulates that different NR target genes may require different activation functions of the receptor (153), thereby creating the possibility of interaction with different coactivator proteins (such as the Mediator complex). Secondly, also for mechanisms, the promoter-specific transrepression recruitment of cofactors has been described. In a first example it was shown that regulatory complexes formed at the IL-8 and IkappaB promoters were distinguished by differential recruitment of the Ser2 CTD kinase, P-TEFb. This might be an explanation for the differential expression of two genes of which the proximal kappaB-elements are distinguished by a single base pair difference only (141). In addition, Ogawa and colleagues (154) describe a mechanism in which GR is able to repress the activation of functionally related NF-kappaB-driven genes by disturbing p65/interferon regulatory factor (IRF) complexes. NFkappaB-driven genes, which are not negatively regulated by GR, seem to use other proteins as coactivators, e.g. Bcl-3. To make the story even more complex, it has been described that some cofactors can exert both transactivation and transrepression effects in a single cell type, albeit not at the same moment and in a response element-specific manner. The cofactor GRIP, for instance, displays different activity domains that make a coactivator/corepressor switch possible (155). A similar coactivator/corepressor switch is described for ER. A recent study shows that unliganded ERalpha behaves as a TNFalpha-induced coactivator that becomes a corepressor in the presence of E<sub>2</sub> by recruiting GRIP-1 (104).

The observation that GR-mediated repression of NF-kappaB-driven genes can only be observed after tolllike receptor (TLR)-4 and TLR-9 triggering, but on the contrary fails after TLR-3 triggering (154), brings us to a point of signal specificity. What will be the final outcome of transrepression in a cell where different pathways interact and influence each other and how are these messages combined into a combinatorial code? Indeed, it is shown for GR and NF-kappaB that both TFs only reside transiently on their DNA-binding sites. It would thus be interesting to explore what the turn-over is of both factors in repressive conditions, as a rapid turn-over would make continuous sampling of the cell environment possible (51). Moreover, NRs cannot be seen as functionally completely separated molecules, but in fact they can modulate the expression of distinct, as well as of an overlapping set of target genes. An example is the synergistic negative effect of GR and PPARgamma on iNOS expression, which probably resides in a simultaneous targeting of NF-kappaB by GR and of NCoR complexes by PPARgamma. The phenomenon that NRs block NF-kappaB can thus not be considered as a general mechanism, since only a subset of NF-kappaB-driven genes are influenced, but should be considered a more subtle encounter in which cell-, signaland promoter-specific mechanisms play an essential role for defining the final response. Taken together, all these arguments favour a cofactor exchange model instead of a cofactor competition model (142, 143).

#### 4.3. Alternative mechanisms

Since the genomic effects of NRs depend on transrepression of inflammatory gene expression and/or upregulation of anti-inflammatory genes, this mechanism takes a few hours to days. However, more and more evidence accumulates, stating that NRs can have an effect on cellular responses, which happen in a time frame from seconds to an hour (156, 157). Which receptor/pathway is responsible for mediating these non-genomic effects is yet to be elucidated. For GCs and estrogens for instance, their interaction with a specific membrane receptor, namely a Gprotein-coupled receptor, has been described (158-160). Secondly, an alternative mechanism has been reported to explain the GC-mediated immunosuppressive effects on Tcell activation. As reviewed by Löwenberg and colleagues, it has been proposed that GR physically interacts with the T-cell receptor (TCR) complex and that this interaction gets disturbed after ligand activation of GR, leading to impaired T-cell signalling (157). Thirdly, it has been postulated that for some steroid ligands a plasma membrane-bound NR exists (159, 161-163). Alternatively, Buttgereit and colleagues showed that GC at high concentrations intercalate into the plasma membrane of immune cells, thereby interfering with calcium and sodium cycling across the membranes (164, 165). Although the mechanistic details of these rapid actions are still lacking, it seems that steroids can induce an increase in several second messengers such as inositol triphosphate, cAMP, Ca<sup>2+</sup>. Furthermore, it has been proposed that phytoestrogenic isoflavones can selectively, in an ER-independent manner, block nuclear transactivation of NF-kappaB via successive attenuation of MEK/ERK and MSK-1 activity, hereby preventing the phosphorylation of p65 and histone H3 (166). By showing that steroids can rapidly interfere with the action of kinase pathways it becomes clear that genomic and non-genomic effects cannot completely be seen as two diverse mechanisms. Indeed, evidence is arising that membrane-initiated actions of NRs may provide an additional mechanism for the regulation of gene transcription. Firstly, research has unravelled that the PKA pathway induces the degradation of GRIP-1 through the ubiquitin-proteasome pathway, thereby providing a novel regulatory mechanism by which hormones down-regulate a cofactor (167). Secondly, a recent report showed that E<sub>2</sub> induces cell proliferation via a non-genomic pathway, through activation of ERK1/2 and subsequent upregulation of cyclin D1 (via mER and G protein) (168). In contrast, ER might cooperate with NF-kappaB for the expression of COX-2 via a non-genomic effect (169). This in turn results in the production of PGI2 and may thus explain the protective effects of estrogens on the cardiovascular system (52, 170). It would thus be very interesting to further define which mechanism (genomic versus non-genomic) and/or receptor type contributes most to the immunosuppressive actions and which mechanism lies at the basis of currently observed unwanted systemic effects. Furthermore, it is of the utmost importance to define which factors are crucial to determine whether TFs, such as ER and NF-kappaB, will cooperate or will antagonize each other's functions (52). Indeed, a synergistic cooperation between ER and NFkappaB, whereby E2 mediates an increase in activated

ERK, subsequently leading to NF-kappaB activation and expression of anti-oxidant enzymes, such as Mn-superoxide dismutase and gluthathion peroxidase seems highly promoter-specific (171).

### 5. THERAPEUTIC IMPLICATIONS

### 5.1. GR ligands

The broad-range anti-inflammatory effects of GCs are reflected by the many mechanisms by which their corresponding receptor can interfere with pro-inflammatory pathways. As such, GR ligands are able to inhibit the expression of a battery of pro-inflammatory mediators, thereby explaining their therapeutical relevance. Indeed, in contrast to e.g. antibody therapy, which targets only one cytokine or chemokine and is used in treatment regimens of inflammatory diseases such as rheumatoid arthritis and Crohn's disease, GCs display a much broader target range. A very attractive model has been described by Smolen and colleagues to explain the relevance of targeting more than one pro-inflammatory mediator in rheumatoid arthritis (172). These authors use the picture of an inflammatory house of cards to explain that once an inflammation process is initiated and a cascade of pro-inflammatory gene expression is started, targeting the initial cytokine, that triggered the process, will not be sufficient anymore for an optimal therapeutic response. On the contrary, to bring the inflammatory house of cards to full collapse and to avoid remission, multiple mediators should be targeted (172). However, this multiplicity of GC treatment is not only its strength, but at the same time it is responsible for the main weakness of cortisone treatment, as it originates from the essential role GCs play in the regulation of metabolic and stress responses. The frequent administration of GCs leads to a systemic increase of hormone levels and this in turn can lead to a dysregulation of sugar and lipid metabolism, as well as fluid and salt retention, resulting in the occurrence of unwanted effects, including diabetes, glaucoma, fat redistribution, hypertension, but also osteoporosis, muscle wasting, insomnia and psychiatric disorders (173). Keeping the unwanted effects of GC in mind, it has to be stated that, if cortisone would be discovered today, it would probably never get approved by the regulatory authorities. However, despite the scientific progress made in the last years concerning the molecular mechanisms of GR function and despite the continuous efforts of the pharmaceutical industry, no other drug has been able to kick GC-based therapy from its pedestal when it comes to therapeutic benefits. Consequently, it is still the most commonly used drug that does improve the quality of live of patients with chronic inflammatory and autoimmune diseases, as well as cancer. Different directions are however explored to optimize treatment protocols based on GR ligands, in the hope to find drugs that display the same anti-inflammatory potential as classical steroid hormones, but lacking the unwanted secondary effects (173). Firstly, although MR-containing tissues are protected from cortisol effects by an enzymatic activity converting cortisol to the weaker cortisone, synthetic GR ligands, such as DEX, are not sensitive to 11β-hydroxysteroid dehydrogenase 2 and thus retain a full capacity to bind to MR in certain tissues, e.g. the kidneys. As such, DEX imposes its effects on the kidney via both MR and GR and this mechanism is believed to form the basis of hypertension (173). Improvements thus came with the knowledge that local administration (e.g. in aerosols or creams for topical usage) instead of systemic usage would already limit the occurrence of some of the side-effects. Secondly, pro-drugs might be developed, which only release their active substances at sites of inflammation (174). Alternatively, GCs can be marketed as soft-drugs, which are rapidly metabolized and inactivated after exerting their pharmacologic effects (175). Nevertheless, localized therapy can only be used in certain conditions and therefore there is still an urgent need for drugs with a better benefitrisk profile (176, 177).

The observation that different (ant)agonists of GR can induce only a subset of the functions elicited by the natural ligands raised the intriguing possibility that it might be possible to find or create ligands, able to separate the beneficial effects from the side-effects (48, 178). Indeed, recent developments in the GR field have allowed the identification of so-called dissociated GR ligands. The first dissociated GC described was able to inhibit AP-1dependent transcription, but failed to promote GRE-driven transcription and was called RU24858 (179). However, subsequent work showed that the transactivation, as well as the transrepression effects of this compound are strongly cell type-dependent (180, 181) and these combined data may explain the poor separation of wanted and unwanted effects in vivo (117, 182). A second promising selective GR agonist (SEGRA), was described by Schäcke et al. in 2004 and constitutes the non-steroidal compound ZK216348 (183). In a murine croton oil-induced ear inflammation model, this compound was able to suppress inflammation to the same extent as prednisolone does. In contrast, this compound showed a markedly superior side-effect profile regarding blood glucose levels, spleen involution, skin atrophy and osteoporosis (183, 184). However, the main draw-back of this compound arises from the fact that it is not solely modulating the activity of GR, but in addition exerts effects at the PR and MR level (183).

Recently, a plant-derived compound, referred to as CompoundA (CpdA), was discovered to selectively interfere with NF-kappaB-driven pro-inflammatory gene expression in a solely GR-dependent manner, yet fails to transactivate GRE-driven genes. Furthermore, CpdA is able to inhibit inflammation in a model of acute paw swelling without increasing blood glucose concentrations (185). To elaborate if CpdA would also be able to inhibit inflammation in a therapeutic protocol under chronic inflammatory conditions, the collagen-induced arthritis model was used to show that the therapeutic potential of CpdA is indeed present under conditions lacking the unwanted diabetogenic effects (186). It was additionally shown that these observations arise from the fact that CpdA actively induces the formation of a GR monomer (186). These data are in line with the observations made by Reichardt and co-workers, who found that knock-in mice which express a GR that is unable to dimerize and to bind DNA on classical GRE-elements were still able to suppress inflammation. In these GRdim mice, induction of

gluconeogenic enzymes in the liver could not be observed (187, 188). Conclusively, it is postulated that the antiinflammatory action of GR can be separated from its
unwanted effects if a ligand is found or synthesised that is
able to specifically induce the GR monomer. However,
recently it has been shown that GR can transactivate some
genes even in the absence of dimerization (153). Since
CpdA can induce a differential conformation of the
receptor as compared to DEX (185), a large scale gene
expression array will allow to compare both GRmodulating compounds and extend this knowledge to
different cell types and *in vivo* models.

Indeed, after a more detailed research it appeared that many of the compounds originally categorized as being dissociated, turned out to rather be gene-selective. An example is the characterization of AL-438, a compound that represses and activates only a subset of genes normally regulated by classical steroids. These characteristics do not affect the anti-inflammatory potential of the compound in vivo, but correlate with a decrease of the negative effects on bone metabolism, chondrocyte proliferation and glucose levels (30, 184, 189). The causal molecular mechanism resides in a differential cofactor recruitment in response to ligand. While prednisolone-induced GR interacts with both cofactors, i.e. PGC-1, a cofactor critical for upregulating glucose levels after steroid induction, and GRIP-1, induction of GR by AL-438 reduces the interaction of GR with PGC-1, while maintaining the interaction with GRIP-1 (30).

The complexity of the whole story is brought further to the attention by the realization that only subtle differences in ligand structure can have profound effects on the outcome of gene-expression (190). The knowledge that endogenous promoters behave as very sensitive detectors of only subtle differences pops the question if it would be feasable to design cell- or tissue-specific ligands (51, 117, 190). Furthermore, it is not inconceivable that in order to reach efficient treatment regimens in all patients, it might be necessary to adapt therapies to patient-specific needs. Indeed, our preliminary data using fibroblast-like synoviocytes (FLS) isolated from the inflamed tissues of patients with RA, show that different patients show a promoter- and ligand-dependent differential response to the GR modulators DEX and CpdA (191).

#### 5.2. ER ligands

The role of estrogens in cancer is dual and complex. While the transcriptional activation of ER target genes, which promote cell proliferation, drives cancer progression (192-194), the ability of ER to interfere with the NF-kappaB activity is believed to be responsible for the protective anti-inflammatory and anti-tumoral effects (84). The search for safer drugs has resulted in the identification of so-called selective estrogen receptor modulators (SERM), that exert tissue-selective activities (i.e. they exert ER agonist effects in one tissue, while ER antagonist effects in another) and may display a preference for one of the two ER isoforms (195, 196). Due to its ER antagonistic effect in breast tissue, tamoxifen is a generally applied SERM, used for the treatment of ER-expressing breast

cancer cells. Although it was previously assumed that tamoxifen displays a safer side-effect profile, this compound seems to display agonistic effects in the uterus, thereby elevating the risk of developing endometrial cancer (197). However, as small changes in ligand structure can lead to differential cofactor recruitment, thus resulting in a ligand-dependent cell type- and promoter-specific response, there are various trials ongoing, in order to develop promising non-steroidal ER modulators, which display an improved activity, as well as a higher tissue specificity (extensively reviewed by Barker (196) and Harnish (198)). For instance, the tissue-selective ER down-regulator GW5638 still exerts some of the agonist actions of tamoxifen, yet behaves as an antagonist in breast tissue and displays no effects in the uterus (194, 199, 200).

With the exception of raloxifene, which is currently used in osteoporosis therapy but under reviewing for its utility in treatment regimens for breast cancer and which is able to inhibit NF-kappaB activity in myeloma cells via the removal of p65 from its binding sites in an ERalpha-dependent manner, most other currently used SERMs do not interfere with NF-kappaB activity (201, 202). However, the relation of excessive NF-kappaB signalling with aberrant cell death pathways and continuous cell proliferation has repeatedly been reported, and higher levels of c-Rel, p65 and p50 have been reported in over 90% of breast cancers (89, 97, 203, 204). Furthermore, it has recently been reported that ERalpha acts as an important regulator to control epithelial to mesenchymal transition (EMT) by controlling de de novo synthesis of RelB, which in turn controls the expression of Bcl-2. This mechanism may explain the more invasive character of ERalpha-negative breast cancer cells and explains the need for ligands that can block NF-kappaB activity (204). Therefore, the recently characterized WAY-169916, which selectively antagonizes NF-kappaB activity, without stimulating uterine proliferation or ER-mediated gene expression, may hold great promise (205).

Another trail that is explored is the possibility of targeting one specific ER isoform. Indeed, while both ERalpha and ERbeta can contribute to the transrepressive effects on inflammatory genes, it is ERalpha that promotes proliferation of breast cancer cells, while ERbeta behaves as a tumor suppressor (104). Furthermore, ERbeta is believed to be more potent in inhibiting NF-kappaB than ERalpha (206). As a consequence, ERbeta-selective estrogens, including phytoestrogens, such as the herbal extract MF101, may constitute a safer alternative to estrogens (84, 207). In addition, as already mentioned above, soy isoflavones, e.g. genestein, daidzein and biochanin, inhibit the NF-kappaB pathway in an ERindependent manner, making them attractive candidates for the treatment of ER-negative breast cancers (166). However, the *in vivo* efficacy of these natural compounds in cancer treatment has still to be determined.

Conclusively, the high degree of tissue-specific responses following the ligand-dependent conformational change of ER has made the search for new targets extra difficult. As with GR, cofactors play a crucial role in

mediating these differential effects. Currently, peptides that selectively target ER/cofactor interactions are under intensive investigation and might hold promise for the development of a more direct mechanism to modulate ER activity (reviewed in (208)). The potential of these peptidomimetics are not restricted to the ER field, but in addition found their way into PPAR research (209).

### 5.3. PPAR ligands

As for GR and ER, PPARs exert their antiinflammatory effects mainly by interfering with activities of other TFs, including nuclear factor of activated T-cells (NFAT), signal transducer and activator of transcription (STAT)-3, CCAAT/enhancer-binding protein (C/EBPbeta), AP-1 and NF-kappaB. As such, fibrates can inhibit in a PPARalpha-dependent manner the production of IL-6, a key player in inflammation and tumorigenesis, via the blockage of AP-1 and NF-kappaB (137). However, the anti-inflammatory effects of PPARgamma-ligands have more extensively been reported. For instance, Arnold and co-workers showed that the respiratory syncytial virus (RSV)-induced DNA binding of NF-kappaB could be inhibited by PPARgamma ligands, thus correlating with a decrease in IL-6, IL-8 and ICAM-1 mRNA expression (210, 211). A decrease in LPS-induced IL-8, as well as COX-2 mRNA expression was also observed in colon cells after concomitant treatment with PPARgamma ligands and may involve a delayed IkappaBalpha degradation (212). Alternatively, Rosiglitazone (ROSI) may attenuate acute colitis through modulation of the NF-kappaB and p38 MAPK pathways, subsequently leading to decreased COX-2 levels (213). Furthermore, although different causal molecular mechanisms are described, there is unanimity about the expression of iNOS being under the transcriptional control of PPARgamma (151, 214).

Despite the clear potential of PPAR ligands to interfere with the activity of pro-inflammatory and protumorigenic TFs, e.g. NF-kappaB, in vitro (215) and despite the promising results obtained in in vivo models of inflammation (216-220) and in patients suffering ulcerative colitis (221), the role of their corresponding receptors in cancer remains controversial and totally conflicting results have been reported by different research groups (extensively reviewed in (15)). The complexity of PPAR signalling is underscored by the observation that PPAR ligands may exert their effects via NR-independent mechanisms (222, 223). It would thus be of high interest to further explore to what level exactly the different mechanisms of PPAR ligand action mediate either beneficial or detrimental effects of PPAR function. Indeed, as for the previously described receptors, signalling to transcription results from integrating cellular pathways via differential cofactor recruitment and post-translational modifications of cofactor, nuclear receptor and other TFs that may bind adjacent promoter sites (15). For instance, interactions between hormone receptors, e.g. GR, on the one hand and PPARs on the other hand allows the cell to integrate local and systemic responses (142). This interaction may hold great therapeutic potential as, according to Nie and colleagues, a synergistic inhibition of chemokine expression occurs upon combined treatment with PPARgamma agonists and GC (224). In analogy, the simultaneous activation of PPARalpha and GR dose-dependently enhances the repression of NF-kappaB-driven genes (225). These results suggest that a combination therapy with PPARalpha/gamma ligands may allow lowering the doses of synthetic GC in treatment regimens without affecting their anti-inflammatory potential.

# 6. GENERAL CONCLUSIONS AND PERSPECTIVES

More and more evidence reveals that many of the well known cancer drugs exert their anti-tumoral effect at least in part through modulating the activity of NF-kappaB, thereby stressing the importance of NF-kappaB in tumor development and progression (reviewed in (226)). However, we should not forget the central role NF-kappaB plays (when regulated normally) in maintaining immune homeostasis. Complete blockage of NF-kappaB might thus have detrimental effects by undermining the important function of the immune system, not only when it comes to pathogen infection, but also in the light of immune surveillance to prevent tumor development (227). Therefore, NR ligands that down-modulate the activity of this widespread TF, instead of completely blocking it, are of enormous value due to their therapeutic potential. However, the efficacy of NR ligands is overshadowed by the occurrence of unwanted effects owing to their broad range of actions. Accordingly, the main goal of NR research remains the improvement of specificity, and molecular research is advancing to pursue this ambition. In this review we have quoted several strategies explored at the moment. Firstly, we reviewed interesting data obtained with different NR modulators, showing that different NR ligands could manipulate gene expression in different signal-, cell- and gene-specific ways on account of differential cofactor recruitment. The importance of these cofactors is further highlighted by their role in alternative splicing and elucidates that these crucial proteins not only influence the abundance, but also the nature of their products (228, 229). Since splicing deregulation is associated with different pathologies, including cancer, it might thus be interesting to identify which splice variants are associated with cancer and are regulated by hormones (229). The story is even more complex as it becomes clear that cross-talk between different cellular pathways results in diverse patterns of post-translational modifications (PTMs) of cofactors as well as TFs, referred to as the "protein code", hereby modifying their activity (41). As discussed above, nuclear receptors can modulate cell signalling pathways via genomic and non-genomic pathways and it is therefore crucial to further elaborate to what extent both mechanisms influence NR signalling. Furthermore, drugs that target the pathways responsible for these PTMs can drastically interfere with NR function and might thus be of importance for usage in combination protocols. Next to the combination of agents that act on different NR pathways, as discussed for GR and PPAR ligands, therapeutic benefit may also come from combination with ligands that act on other key signalling pathways, e.g. the MAPK, Akt, PKA, PKC pathways. As an example, it has recently been shown that PKA-induced phosphorylation of ERalpha induces resistance to

tamoxifen in breast cancer cells (230). In analogy, it is described that phosphorylation of GR after excessive p38 MAPK activation interferes with the affinity of GR for corticosteroids (231). Drug cocktails including kinase inhibitors might thus reduce steroid resistance, allowing to lower the concentrations of NR ligands used.

New fields that are still at the beginning of exploration when it comes to NR signalling include epigenetics and micro-RNAs (miRNAs). Indeed, specificity of gene regulation can also be obtained at the chromatin level and is reflected by the cross-talk of different individual modifications of the histone tails (histon code) and the DNA itself (in general epigenetics). Increasing evidence reveals the importance of epigenetics in NFkappaB and NR signalling and it might thus hold a great challenge to reveal novel epigenetic targets that determine specificity (232). Lastly, miRNAs, which constitute a subset of non-coding RNAs important in controlling the stability of mRNAs, are reported to be aberrantly expressed in cancer tissues (233-235). Since these miRNAs may constitute an extra gene regulatory mechanism it will be exciting to learn how these molecules might interfere with NF-kappaB/NR-signalling.

To conclude, blocking NF-kappaB seems a twosided sword. While some reports claim an anti-apoptotic role of NF-kappaB, other reports declare that NF-kappaB exhibits a pro-apoptotic role after DNA-damage. It is thus of extreme importance to unravel under which conditions NF-kappaB behaves as a stimulus for apoptosis (68). Additionally, it may be interesting to explore if NR ligands could cooperate with the pro-apoptotic capacity of NFkappaB, which may add great value to cancer therapies.

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### 8. REFERENCES

- 1. Germain, P., B. Staels, C. Dacquet, M. Spedding & V. Laudet: Overview of nomenclature of nuclear receptors. *Pharmacol Rev*, 58, 685-704 (2006)
- 2. Oakley, R. H., M. Sar & J. A. Cidlowski: The human glucocorticoid receptor beta isoform. Expression, biochemical properties, and putative function. *J Biol Chem*, 271, 9550-9 (1996)
- 3. Yudt, M. R. & J. A. Cidlowski: Molecular identification and characterization of a and b forms of the glucocorticoid receptor. *Mol Endocrinol*, 15, 1093-103 (2001)
- 4. Lu, N. Z. & J. A. Cidlowski: Translational regulatory mechanisms generate N-terminal glucocorticoid receptor isoforms with unique transcriptional target genes. *Mol Cell*, 18, 331-42 (2005)

- 5. Turner, J. D., A. B. Schote, J. A. Macedo, L. P. Pelascini & C. P. Muller: Tissue specific glucocorticoid receptor expression, a role for alternative first exon usage? *Biochem Pharmacol*, 72, 1529-37 (2006)
- 6. Lu, N. Z. & J. A. Cidlowski: Glucocorticoid receptor isoforms generate transcription specificity. *Trends Cell Biol*, 16, 301-7 (2006)
- 7. De Kloet, E. R., M. Joels & F. Holsboer: Stress and the brain: from adaptation to disease. *Nat Rev Neurosci*, 6, 463-75 (2005)
- 8. Hammer, F. & P. M. Stewart: Cortisol metabolism in hypertension. *Best Pract Res Clin Endocrinol Metab*, 20, 337-53 (2006)
- 9. Klouche, M.: Estrogens in human vascular diseases. *Ann N Y Acad Sci*, 1089, 431-43 (2006)
- 10. Simoncini, T., P. Mannella & A. R. Genazzani: Rapid estrogen actions in the cardiovascular system. *Ann N Y Acad Sci*, 1089, 424-30 (2006)
- 11. McDonnell, D. P. & J. D. Norris: Connections and regulation of the human estrogen receptor. *Science*, 296, 1642-4 (2002)
- 12. Nilsson, S., S. Makela, E. Treuter, M. Tujague, J. Thomsen, G. Andersson, E. Enmark, K. Pettersson, M. Warner & J. A. Gustafsson: Mechanisms of estrogen action. *Physiol Rev*, 81, 1535-65 (2001)
- 13. Forman, B. M., J. Chen & R. M. Evans: The peroxisome proliferator-activated receptors: ligands and activators. *Ann N Y Acad Sci*, 804, 266-75 (1996)
- 14. Kliewer, S. A. & T. M. Willson: The nuclear receptor PPARgamma bigger than fat. *Curr Opin Genet Dev*, 8, 576-81 (1998)
- 15. Feige, J. N., L. Gelman, L. Michalik, B. Desvergne & W. Wahli: From molecular action to physiological outputs: peroxisome proliferator-activated receptors are nuclear receptors at the crossroads of key cellular functions. *Prog Lipid Res*, 45, 120-59 (2006)
- 16. Kliewer, S. A., B. M. Forman, B. Blumberg, E. S. Ong, U. Borgmeyer, D. J. Mangelsdorf, K. Umesono & R. M. Evans: Differential expression and activation of a family of murine peroxisome proliferator-activated receptors. *Proc Natl Acad Sci U S A*, 91, 7355-9 (1994)
- 17. Pratt, W. B. & D. O. Toft: Steroid receptor interactions with heat shock protein and immunophilin chaperones. *Endocr Rev*, 18, 306-60 (1997)
- 18. Savory, J. G., B. Hsu, I. R. Laquian, W. Giffin, T. Reich, R. J. Hache & Y. A. Lefebvre: Discrimination between NL1- and NL2-mediated nuclear localization of the glucocorticoid receptor. *Mol Cell Biol*, 19, 1025-37 (1999)

- 19. Kakar, M., C. Kanwal, J. R. Davis, H. Li & C. S. Lim: Geldanamycin, an inhibitor of Hsp90, blocks cytoplasmic retention of progesterone receptors and glucocorticoid receptors via their respective ligand binding domains. *Aaps J*, 8, E718-28 (2006)
- 20. Bourguet, W., P. Germain & H. Gronemeyer: Nuclear receptor ligand-binding domains: three-dimensional structures, molecular interactions and pharmacological implications. *Trends Pharmacol Sci*, 21, 381-8 (2000)
- 21. Rosenfeld, M. G. & C. K. Glass: Coregulator codes of transcriptional regulation by nuclear receptors. *J Biol Chem*, 276, 36865-8 (2001)
- 22. Chan, H. M. & N. B. La Thangue: p300/CBP proteins: HATs for transcriptional bridges and scaffolds. *J Cell Sci*, 114, 2363-73 (2001)
- 23. Sterner, D. E. & S. L. Berger: Acetylation of histones and transcription-related factors. *Microbiol Mol Biol Rev*, 64, 435-59 (2000)
- 24. Davie, J. R. & V. A. Spencer: Control of histone modifications. *J Cell Biochem*, Suppl 32-33, 141-8 (1999)
- 25. Chen, H., R. J. Lin, W. Xie, D. Wilpitz & R. M. Evans: Regulation of hormone-induced histone hyperacetylation and gene activation via acetylation of an acetylase. *Cell*, 98, 675-86 (1999)
- 26. Yan, F., X. Gao, D. M. Lonard & Z. Nawaz: Specific ubiquitin-conjugating enzymes promote degradation of specific nuclear receptor coactivators. *Mol Endocrinol*, 17, 1315-31 (2003)
- 27. Malik, S. & R. G. Roeder: Transcriptional regulation through Mediator-like coactivators in yeast and metazoan cells. *Trends Biochem Sci*, 25, 277-83 (2000)
- 28. Kraichely, D. M., J. Sun, J. A. Katzenellenbogen & B. S. Katzenellenbogen: Conformational changes and coactivator recruitment by novel ligands for estrogen receptor-alpha and estrogen receptor-beta: correlations with biological character and distinct differences among SRC coactivator family members. *Endocrinology*, 141, 3534-45 (2000)
- 29. Bramlett, K. S., Y. Wu & T. P. Burris: Ligands specify coactivator nuclear receptor (NR) box affinity for estrogen receptor subtypes. *Mol Endocrinol*, 15, 909-22 (2001)
- 30. Coghlan, M. J., P. B. Jacobson, B. Lane, M. Nakane, C. W. Lin, S. W. Elmore, P. R. Kym, J. R. Luly, G. W. Carter, R. Turner, C. M. Tyree, J. Hu, M. Elgort, J. Rosen & J. N. Miner: A novel antiinflammatory maintains glucocorticoid efficacy with reduced side effects. *Mol Endocrinol*, 17, 860-9 (2003)

- 31. He, Y. & S. S. Simons, Jr.: STAMP, a novel predicted factor assisting TIF2 actions in glucocorticoid receptor-mediated induction and repression. *Mol Cell Biol*, 27, 1467-85 (2007)
- 32. Li, X., J. Wong, S. Y. Tsai, M. J. Tsai & B. W. O'Malley: Progesterone and glucocorticoid receptors recruit distinct coactivator complexes and promote distinct patterns of local chromatin modification. *Mol Cell Biol*, 23, 3763-73 (2003)
- 33. Metivier, R., G. Penot, M. R. Hubner, G. Reid, H. Brand, M. Kos & F. Gannon: Estrogen receptor-alpha directs ordered, cyclical, and combinatorial recruitment of cofactors on a natural target promoter. *Cell*, 115, 751-63 (2003)
- 34. Wang, Q., J. S. Carroll & M. Brown: Spatial and temporal recruitment of androgen receptor and its coactivators involves chromosomal looping and polymerase tracking. *Mol Cell*, 19, 631-42 (2005)
- 35. Shang, Y., X. Hu, J. DiRenzo, M. A. Lazar & M. Brown: Cofactor dynamics and sufficiency in estrogen receptor-regulated transcription. *Cell*, 103, 843-52 (2000)
- 36. Grenier, J., A. Trousson, A. Chauchereau, J. Cartaud, M. Schumacher & C. Massaad: Differential recruitment of p160 coactivators by glucocorticoid receptor between Schwann cells and astrocytes. *Mol Endocrinol*, 20, 254-67 (2006)
- 37. McNally, J. G., W. G. Muller, D. Walker, R. Wolford & G. L. Hager: The glucocorticoid receptor: rapid exchange with regulatory sites in living cells. *Science*, 287, 1262-5 (2000)
- 38. Stavreva, D. A., W. G. Muller, G. L. Hager, C. L. Smith & J. G. McNally: Rapid glucocorticoid receptor exchange at a promoter is coupled to transcription and regulated by chaperones and proteasomes. *Mol Cell Biol*, 24, 2682-97 (2004)
- 39. Rayasam, G. V., C. Elbi, D. A. Walker, R. Wolford, T. M. Fletcher, D. P. Edwards & G. L. Hager: Ligand-specific dynamics of the progesterone receptor in living cells and during chromatin remodeling *in vitro*. *Mol Cell Biol*, 25, 2406-18 (2005)
- 40. Duma, D., C. M. Jewell & J. A. Cidlowski: Multiple glucocorticoid receptor isoforms and mechanisms of post-translational modification. *J Steroid Biochem Mol Biol*, 102, 11-21 (2006)
- 41. Ito, K.: Impact of post-translational modifications of proteins on the inflammatory process. *Biochem Soc Trans*, 35, 281-3 (2007)
- 42. Wu, R. C., C. L. Smith & B. W. O'Malley: Transcriptional regulation by steroid receptor coactivator phosphorylation. *Endocr Rev*, 26, 393-9 (2005)

- 43. Losel, R. & M. Wehling: Nongenomic actions of steroid hormones. *Nat Rev Mol Cell Biol*, 4, 46-56 (2003) 44. Leclercq, G., M. Lacroix, I. Laios & G. Laurent: Estrogen receptor alpha: impact of ligands on intracellular shuttling and turnover rate in breast cancer cells. *Curr Cancer Drug Targets*, 6, 39-64 (2006)
- 45. Carrigan, A., R. F. Walther, H. A. Salem, D. Wu, E. Atlas, Y. A. Lefebvre & R. J. Haché: An Active Nuclear Retention Signal in the Glucocorticoid Receptor Functions as a Strong Inducer of Transcriptional Activation. *J Biol Chem*, 282, 10963-10971 (2007)
- 46. Glass, C. K. & M. G. Rosenfeld: The coregulator exchange in transcriptional functions of nuclear receptors. *Genes Dev.*, 14, 121-41 (2000)
- 47. Perissi, V., A. Aggarwal, C. K. Glass, D. W. Rose & M. G. Rosenfeld: A corepressor/coactivator exchange complex required for transcriptional activation by nuclear receptors and other regulated transcription factors. *Cell*, 116, 511-26 (2004)
- 48. Resche-Rigon, M. & H. Gronemeyer: Therapeutic potential of selective modulators of nuclear receptor action. *Curr Opin Chem Biol*, 2, 501-7 (1998)
- 49. Wintermantel, T. M., S. Berger, E. F. Greiner & G. Schütz: Evaluation of steroid receptor function by gene targeting in mice. *J Steroid Biochem Mol Biol*, 93, 107-12 (2005)
- 50. De Bosscher, K., W. Vanden Berghe & G. Haegeman: The interplay between the glucocorticoid receptor and nuclear factor-kappaB or activator protein-1: molecular mechanisms for gene repression. *Endocr Rev*, 24, 488-522 (2003)
- 51. De Bosscher, K., W. Vanden Berghe & G. Haegeman: Cross-talk between nuclear receptors and nuclear factor kappaB. *Oncogene*, 25, 6868-86 (2006)
- 52. Kalaitzidis, D. & T. D. Gilmore: Transcription factor cross-talk: the estrogen receptor and NF-kappaB. *Trends Endocrinol Metab*, 16, 46-52 (2005)
- 53. Buser, A. C., E. K. Gass-Handel, S. L. Wyszomierski, W. Doppler, S. A. Leonhardt, J. Schaack, J. M. Rosen, H. Watkin, S. M. Anderson & D. P. Edwards: Progesterone receptor repression of prolactin/signal transducer and activator of transcription 5-mediated transcription of the beta-casein gene in mammary epithelial cells. *Mol Endocrinol*, 21, 106-25 (2007)
- 54. Reily, M. M., C. Pantoja, X. Hu, Y. Chinenov & I. Rogatsky: The GRIP1:IRF3 interaction as a target for glucocorticoid receptor-mediated immunosuppression. *EMBO J.* 25, 108-17 (2006)
- 55. Gloire, G., E. Dejardin & J. Piette: Extending the nuclear roles of IkappaB kinase subunits. *Biochem Pharmacol*, 72, 1081-9 (2006)

- 56. Baldwin, A. S., Jr.: The NF-kappa B and I kappa B proteins: new discoveries and insights. *Annu Rev Immunol*, 14, 649-83 (1996)
- 57. Baldwin, A. S., Jr.: Series introduction: the transcription factor NF-kappaB and human disease. *J Clin Invest*, 107, 3-6 (2001)
- 58. Neumann, M. & M. Naumann: Beyond IkappaBs: alternative regulation of NF-kappaB activity. *FASEB J* (2007)
- 59. Viatour, P., M. P. Merville, V. Bours & A. Chariot: Phosphorylation of NF-kappaB and IkappaB proteins: implications in cancer and inflammation. *Trends Biochem Sci*, 30, 43-52 (2005)
- 60. Kato, T., Jr., M. Delhase, A. Hoffmann & M. Karin: CK2 Is a C-Terminal IkappaB Kinase Responsible for NF-kappaB Activation during the UV Response. *Mol Cell*, 12, 829-39 (2003)
- 61. Hayden, M. S. & S. Ghosh: Signaling to NF-kappaB. *Genes Dev*, 18, 2195-224 (2004)
- 62. Gilmore, T. D.: Introduction to NF-kappaB: players, pathways, perspectives. *Oncogene*, 25, 6680-4 (2006)
- 63. Crofford, L. J., B. Tan, C. J. McCarthy & T. Hla: Involvement of nuclear factor kappa B in the regulation of cyclooxygenase-2 expression by interleukin-1 in rheumatoid synoviocytes. *Arthritis Rheum*, 40, 226-36 (1997)
- 64. Vincenti, M. P., C. I. Coon & C. E. Brinckerhoff: Nuclear factor kappaB/p50 activates an element in the distal matrix metalloproteinase 1 promoter in interleukin-1beta-stimulated synovial fibroblasts. *Arthritis Rheum*, 41, 1987-94 (1998)
- 65. Bond, M., A. H. Baker & A. C. Newby: Nuclear factor kappaB activity is essential for matrix metalloproteinase-1 and -3 upregulation in rabbit dermal fibroblasts. *Biochem Biophys Res Commun*, 264, 561-7 (1999)
- 66. Li, P., I. Sanz, R. J. O'Keefe & E. M. Schwarz: NF-kappa B regulates VCAM-1 expression on fibroblast-like synoviocytes. *J Immunol*, 164, 5990-7 (2000)
- 67. Tak, P. P. & G. S. Firestein: NF-kappaB: a key role in inflammatory diseases. *J Clin Invest*, 107, 7-11 (2001)
- 68. Janssens, S. & J. Tschopp: Signals from within: the DNA-damage-induced NF-kappaB response. *Cell Death Differ*, 13, 773-84 (2006)
- 69. Janssens, S., A. Tinel, S. Lippens & J. Tschopp: PIDD mediates NF-kappaB activation in response to DNA damage. *Cell*, 123, 1079-92 (2005)
- 70. Huang, T. T., S. M. Wuerzberger-Davis, Z. H. Wu & S. Miyamoto: Sequential modification of NEMO/IKKgamma

- by SUMO-1 and ubiquitin mediates NF-kappaB activation by genotoxic stress. *Cell*, 115, 565-76 (2003)
- 71. Hay, R. T.: Modifying NEMO. *Nat Cell Biol*, 6, 89-91 (2004)
- 72. Wu, C. J., D. B. Conze, T. Li, S. M. Srinivasula & J. D. Ashwell: Sensing of Lys 63-linked polyubiquitination by NEMO is a key event in NF-kappaB activation [corrected]. *Nat Cell Biol*, 8, 398-406 (2006)
- 73. Han, Z., D. L. Boyle, A. M. Manning & G. S. Firestein: AP-1 and NF-kappaB regulation in rheumatoid arthritis and murine collagen-induced arthritis. *Autoimmunity*, 28, 197-208 (1998)
- 74. Hart, L. A., V. L. Krishnan, I. M. Adcock, P. J. Barnes & K. F. Chung: Activation and localization of transcription factor, nuclear factor-kappaB, in asthma. *Am J Respir Crit Care Med*, 158, 1585-92 (1998)
- 75. Van Den Brink, G. R., F. J. ten Kate, C. Y. Ponsioen, M. M. Rive, G. N. Tytgat, S. J. van Deventer & M. P. Peppelenbosch: Expression and activation of NF-kappa B in the antrum of the human stomach. *J Immunol*, 164, 3353-9 (2000)
- 76. Pikarsky, E., R. M. Porat, I. Stein, R. Abramovitch, S. Amit, S. Kasem, E. Gutkovich-Pyest, S. Urieli-Shoval, E. Galun & Y. Ben-Neriah: NF-kappaB functions as a tumour promoter in inflammation-associated cancer. *Nature*, 431, 461-6 (2004)
- 77. Balkwill, F. & A. Mantovani: Inflammation and cancer: back to Virchow? *Lancet*, 357, 539-45 (2001)
- 78. Karin, M. & F. R. Greten: NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nat Rev Immunol*, 5, 749-59 (2005)
- 79. Richmond, A.: NF-kappa B, chemokine gene transcription and tumour growth. *Nat Rev Immunol*, 2, 664-74 (2002)
- 80. Aggarwal, B. B., S. Shishodia, S. K. Sandur, M. K. Pandey & G. Sethi: Inflammation and cancer: how hot is the link? *Biochem Pharmacol*, 72, 1605-21 (2006)
- 81. Kucharczak, J., M. J. Simmons, Y. Fan & C. Gelinas: To be, or not to be: NF-kappaB is the answerrole of Rel/NF-kappaB in the regulation of apoptosis. *Oncogene*, 22, 8961-82 (2003)
- 82. Escarcega, R. O., S. Fuentes-Alexandro, M. Garcia-Carrasco, A. Gatica & A. Zamora: The transcription factor nuclear factor-kappa B and cancer. *Clin Oncol (R Coll Radiol)*, 19, 154-61 (2007)
- 83. Coussens, L. M. & Z. Werb: Inflammation and cancer. *Nature*, 420, 860-7 (2002)

- 84. Dijsselbloem, N., W. Vanden Berghe, A. De Naeyer & G. Haegeman: Soy isoflavone phyto-pharmaceuticals in interleukin-6 affections. Multi-purpose nutraceuticals at the crossroad of hormone replacement, anti-cancer and anti-inflammatory therapy. *Biochem Pharmacol*, 68, 1171-85 (2004)
- 85. Meng, F., H. Wehbe-Janek, R. Henson, H. Smith & T. Patel: Epigenetic regulation of microRNA-370 by interleukin-6 in malignant human cholangiocytes. *Oncogene* (2007)
- 86. Van Waes, C.: Nuclear factor-kappaB in development, prevention, and therapy of cancer. *Clin Cancer Res*, 13, 1076-82 (2007)
- 87. Kalkhoven, E., S. Wissink, P. T. van der Saag & B. van der Burg: Negative interaction between the RelA (p65) subunit of NF-kappaB and the progesterone receptor. *J Biol Chem*, 271, 6217-24 (1996)
- 88. Srivastava, M. D. & D. J. Anderson: Progesterone receptor expression by human leukocyte cell lines: molecular mechanisms of cytokine suppression. *Clin Exp Obstet Gynecol*, 34, 14-24 (2007)
- 89. Biswas, D. K., Q. Shi, S. Baily, I. Strickland, S. Ghosh, A. B. Pardee & J. D. Iglehart: NF-kappa B activation in human breast cancer specimens and its role in cell proliferation and apoptosis. *Proc Natl Acad Sci U S A*, 101, 10137-42 (2004)
- 90. Suh, J., F. Payvandi, L. C. Edelstein, P. S. Amenta, W. X. Zong, C. Gelinas & A. B. Rabson: Mechanisms of constitutive NF-kappaB activation in human prostate cancer cells. *Prostate*, 52, 183-200 (2002)
- 91. Budunova, I. V., D. Kowalczyk, P. Perez, Y. J. Yao, J. L. Jorcano & T. J. Slaga: Glucocorticoid receptor functions as a potent suppressor of mouse skin carcinogenesis. *Oncogene*, 22, 3279-87 (2003)
- 92. Yemelyanov, A., J. Czwornog, D. Chebotaev, A. Karseladze, E. Kulevitch, X. Yang & I. Budunova: Tumor suppressor activity of glucocorticoid receptor in the prostate. *Oncogene*, 26, 1885-96 (2007)
- 93. Scheinman, R. I., P. C. Cogswell, A. K. Lofquist & A. S. Baldwin, Jr.: Role of transcriptional activation of I kappa B alpha in mediation of immunosuppression by glucocorticoids. *Science*, 270, 283-6 (1995)
- 94. Scheinman, R. I., A. Gualberto, C. M. Jewell, J. A. Cidlowski & A. S. Baldwin, Jr.: Characterization of mechanisms involved in transrepression of NF-kappa B by activated glucocorticoid receptors. *Mol Cell Biol*, 15, 943-53 (1995)
- 95. Auphan, N., J. A. DiDonato, C. Rosette, A. Helmberg & M. Karin: Immunosuppression by glucocorticoids: inhibition of NF-kappa B activity through induction of I kappa B synthesis. *Science*, 270, 286-90 (1995)

- 96. Delerive, P., D. Monte, G. Dubois, F. Trottein, J. Fruchart-Najib, J. Mariani, J. C. Fruchart & B. Staels: The orphan nuclear receptor ROR alpha is a negative regulator of the inflammatory response. *EMBO Rep.*, 2, 42-8 (2001)
- 97. Nakshatri, H., P. Bhat-Nakshatri, D. A. Martin, R. J. Goulet, Jr. & G. W. Sledge, Jr.: Constitutive activation of NF-kappaB during progression of breast cancer to hormone-independent growth. *Mol Cell Biol*, 17, 3629-39 (1997)
- 98. Wen, Y., S. Yang, R. Liu, E. Perez, K. D. Yi, P. Koulen & J. W. Simpkins: Estrogen attenuates nuclear factor-kappa B activation induced by transient cerebral ischemia. *Brain Res.* 1008, 147-54 (2004)
- 99. Delerive, P., K. De Bosscher, W. Vanden Berghe, J. C. Fruchart, G. Haegeman & B. Staels: DNA binding-independent induction of IkappaBalpha gene transcription by PPARalpha. *Mol Endocrinol*, 16, 1029-39 (2002)
- 100. De Bosscher, K., M. L. Schmitz, W. Vanden Berghe, S. Plaisance, W. Fiers & G. Haegeman: Glucocorticoid-mediated repression of nuclear factor-kappaB-dependent transcription involves direct interference with transactivation. *Proc Natl Acad Sci U S A*, 94, 13504-9 (1997)
- 101. Lidén, J., I. Rafter, M. Truss, J. A. Gustafsson & S. Okret: Glucocorticoid effects on NF-kappaB binding in the transcription of the ICAM-1 gene. *Biochem Biophys Res Commun*, 273, 1008-14 (2000)
- 102. Nissen, R. M. & K. R. Yamamoto: The glucocorticoid receptor inhibits NFkappaB by interfering with serine-2 phosphorylation of the RNA polymerase II carboxyterminal domain. *Genes Dev*, 14, 2314-29 (2000)
- 103. Galien, R. & T. Garcia: Estrogen receptor impairs interleukin-6 expression by preventing protein binding on the NF-kappaB site. *Nucleic Acids Res*, 25, 2424-9 (1997)
- 104. Cvoro, A., C. Tzagarakis-Foster, D. Tatomer, S. Paruthiyil, M. S. Fox & D. C. Leitman: Distinct roles of unliganded and liganded estrogen receptors in transcriptional repression. *Mol Cell*, 21, 555-64 (2006)
- 105. Vanden Berghe, W., S. Plaisance, E. Boone, K. De Bosscher, M. L. Schmitz, W. Fiers & G. Haegeman: p38 and extracellular signal-regulated kinase mitogen-activated protein kinase pathways are required for nuclear factor-kappaB p65 transactivation mediated by tumor necrosis factor. *J Biol Chem*, 273, 3285-90 (1998)
- 106. Vermeulen, L., G. De Wilde, P. Van Damme, W. Vanden Berghe & G. Haegeman: Transcriptional activation of the NF-kappaB p65 subunit by mitogen- and stress-activated protein kinase-1 (MSK1). *EMBO J*, 22, 1313-24 (2003)
- 107. Kassel, O., A. Sancono, J. Kratzschmar, B. Kreft, M. Stassen & A. C. Cato: Glucocorticoids inhibit MAP kinase

- via increased expression and decreased degradation of MKP-1. *EMBO J*, 20, 7108-16 (2001)
- 108. Lasa, M., S. M. Abraham, C. Boucheron, J. Saklatvala & A. R. Clark: Dexamethasone causes sustained expression of mitogen-activated protein kinase (MAPK) phosphatase 1 and phosphatase-mediated inhibition of MAPK p38. *Mol Cell Biol*, 22, 7802-11 (2002)
- 109. Clark, A. R.: Anti-inflammatory functions of glucocorticoid-induced genes. *Mol Cell Endocrinol*, 275, 79-97 (2007)
- 110. Jalonen, U., A. Lahti, R. Korhonen, H. Kankaanranta & E. Moilanen: Inhibition of tristetraprolin expression by dexamethasone in activated macrophages. *Biochem Pharmacol*, 69, 733-40 (2005)
- 111. Smoak, K. & J. A. Cidlowski: Glucocorticoids regulate tristetraprolin synthesis and posttranscriptionally regulate tumor necrosis factor alpha inflammatory signaling. *Mol Cell Biol*, 26, 9126-35 (2006)
- 112. Lasa, M., K. R. Mahtani, A. Finch, G. Brewer, J. Saklatvala & A. R. Clark: Regulation of cyclooxygenase 2 mRNA stability by the mitogen-activated protein kinase p38 signaling cascade. *Mol Cell Biol*, 20, 4265-74 (2000)
- 113. Nishimori, T., H. Inoue & Y. Hirata: Involvement of the 3'-untranslated region of cyclooxygenase-2 gene in its post-transcriptional regulation through the glucocorticoid receptor. *Life Sci.*, 74, 2505-13 (2004)
- 114. Clark, A. R., J. L. Dean & J. Saklatvala: Post-transcriptional regulation of gene expression by mitogenactivated protein kinase p38. *FEBS Lett*, 546, 37-44 (2003)
- 115. Toh, M. L., Y. Yang, M. Leech, L. Santos & E. F. Morand: Expression of mitogen-activated protein kinase phosphatase 1, a negative regulator of the mitogenactivated protein kinases, in rheumatoid arthritis: upregulation by interleukin-1beta and glucocorticoids. *Arthritis Rheum*, 50, 3118-28 (2004)
- 116. Abraham, S. M., T. Lawrence, A. Kleiman, P. Warden, M. Medghalchi, J. Tuckermann, J. Saklatvala & A. R. Clark: Antiinflammatory effects of dexamethasone are partly dependent on induction of dual specificity phosphatase 1. *J Exp Med*, 203, 1883-9 (2006)
- 117. Catley, M.: Dissociated steroids. *ScientificWorldJournal*, 7, 421-30 (2007)
- 118. Bruna, A., M. Nicolas, A. Muñoz, J. M. Kyriakis & C. Caelles: Glucocorticoid receptor-JNK interaction mediates inhibition of the JNK pathway by glucocorticoids. *EMBO J*, 22, 6035-44 (2003)
- 119. Castoria, G., A. Migliaccio, A. Bilancio, M. Di Domenico, A. de Falco, M. Lombardi, R. Fiorentino, L. Varricchio, M. V. Barone & F. Auricchio: P13-kinase in

- concert with Src promotes the S-phase entry of oestradiolstimulated MCF-7 cells. *EMBO J*, 20, 6050-9 (2001)
- 120. Haske, T., M. Nakao & V. K. Moudgil: Phosphorylation of immunopurified rat liver glucocorticoid receptor by the catalytic subunit of cAMP-dependent protein kinase. *Mol Cell Biochem*, 132, 163-71 (1994)
- 121. Huggenvik, J. I., M. W. Collard, Y. W. Kim & R. P. Sharma: Modification of the retinoic acid signaling pathway by the catalytic subunit of protein kinase-A. *Mol Endocrinol*, 7, 543-50 (1993)
- 122. Rochette-Egly, C., M. Oulad-Abdelghani, A. Staub, V. Pfister, I. Scheuer, P. Chambon & M. P. Gaub: Phosphorylation of the retinoic acid receptor-alpha by protein kinase A. *Mol Endocrinol*, 9, 860-71 (1995)
- 123. Chen, D., P. E. Pace, R. C. Coombes & S. Ali: Phosphorylation of human estrogen receptor alpha by protein kinase A regulates dimerization. *Mol Cell Biol*, 19, 1002-15 (1999)
- 124. Doucas, V., Y. Shi, S. Miyamoto, A. West, I. Verma & R. M. Evans: Cytoplasmic catalytic subunit of protein kinase A mediates cross-repression by NF-kappa B and the glucocorticoid receptor. *Proc Natl Acad Sci U S A*, 97, 11893-8 (2000)
- 125. Widén, C., J. A. Gustafsson & A. C. Wikstrom: Cytosolic glucocorticoid receptor interaction with nuclear factor-kappa B proteins in rat liver cells. *Biochem J*, 373, 211-20 (2003)
- 126. Drouin, J., M. A. Trifiro, R. K. Plante, M. Nemer, P. Eriksson & O. Wrange: Glucocorticoid receptor binding to a specific DNA sequence is required for hormone-dependent repression of pro-opiomelanocortin gene transcription. *Mol Cell Biol*, 9, 5305-14 (1989)
- 127. Drouin, J., Y. L. Sun, M. Chamberland, Y. Gauthier, A. De Lean, M. Nemer & T. J. Schmidt: Novel glucocorticoid receptor complex with DNA element of the hormone-repressed POMC gene. *EMBO J*, 12, 145-56 (1993)
- 128. Philips, A., M. Maira, A. Mullick, M. Chamberland, S. Lesage, P. Hugo & J. Drouin: Antagonism between Nur77 and glucocorticoid receptor for control of transcription. *Mol Cell Biol*, 17, 5952-9 (1997)
- 129. Martens, C., S. Bilodeau, M. Maira, Y. Gauthier & J. Drouin: Protein-protein interactions and transcriptional antagonism between the subfamily of NGFI-B/Nur77 orphan nuclear receptors and glucocorticoid receptor. *Mol Endocrinol*, 19, 885-97 (2005)
- 130. Cinar, B., F. Yeung, H. Konaka, M. W. Mayo, M. R. Freeman, H. E. Zhau & L. W. Chung: Identification of a negative regulatory cis-element in the enhancer core region of the prostate-specific antigen promoter: implications for

- intersection of androgen receptor and nuclear factor-kappaB signalling in prostate cancer cells. *Biochem J*, 379, 421-31 (2004)
- 131. Caldenhoven, E., J. Lidén, S. Wissink, A. Van de Stolpe, J. Raaijmakers, L. Koenderman, S. Okret, J. A. Gustafsson & P. T. Van der Saag: Negative cross-talk between RelA and the glucocorticoid receptor: a possible mechanism for the antiinflammatory action of glucocorticoids. *Mol Endocrinol*, 9, 401-12 (1995)
- 132. Wissink, S., E. C. van Heerde, M. L. Schmitz, E. Kalkhoven, B. van der Burg, P. A. Baeuerle & P. T. van der Saag: Distinct domains of the RelA NF-kappaB subunit are required for negative cross-talk and direct interaction with the glucocorticoid receptor. *J Biol Chem*, 272, 22278-84 (1997)
- 133. Adcock, I. M., R. Newton & P. J. Barnes: NF-kappa B involvement in IL-1 beta-induction of GM-CSF and COX-2: inhibition by glucocorticoids does not require I-kappa B. *Biochem Soc Trans*, 25, 154S (1997)
- 134. Tzagarakis-Foster, C., R. Geleziunas, A. Lomri, J. An & D. C. Leitman: Estradiol represses human T-cell leukemia virus type 1 Tax activation of tumor necrosis factor-alpha gene transcription. *J Biol Chem*, 277, 44772-7 (2002)
- 135. Ray, A., K. E. Prefontaine & P. Ray: Down-modulation of interleukin-6 gene expression by 17 beta-estradiol in the absence of high affinity DNA binding by the estrogen receptor. *J Biol Chem*, 269, 12940-6 (1994)
- 136. Quaedackers, M. E., C. E. van den Brink, P. T. van der Saag & L. G. Tertoolen: Direct interaction between estrogen receptor alpha and NF-kappaB in the nucleus of living cells. *Mol Cell Endocrinol*, 273, 42-50 (2007)
- 137. Delerive, P., K. De Bosscher, S. Besnard, W. Vanden Berghe, J. M. Peters, F. J. Gonzalez, J. C. Fruchart, A. Tedgui, G. Haegeman & B. Staels: Peroxisome proliferator-activated receptor alpha negatively regulates the vascular inflammatory gene response by negative crosstalk with transcription factors NF-kappaB and AP-1. *J Biol Chem*, 274, 32048-54 (1999)
- 138. Palvimo, J. J., P. Reinikainen, T. Ikonen, P. J. Kallio, A. Moilanen & O. A. Jänne: Mutual transcriptional interference between RelA and androgen receptor. *J Biol Chem*, 271, 24151-6 (1996)
- 139. Meyer, T., J. A. Gustafsson & J. Carlstedt-Duke: Glucocorticoid-dependent transcriptional repression of the osteocalcin gene by competitive binding at the TATA box. *DNA Cell Biol*, 16, 919-27 (1997)
- 140. De Bosscher, K., W. Vanden Berghe, L. Vermeulen, S. Plaisance, E. Boone & G. Haegeman: Glucocorticoids repress NF-kappaB-driven genes by disturbing the interaction of p65 with the basal transcription machinery,

- irrespective of coactivator levels in the cell. *Proc Natl Acad Sci U S A*, 97, 3919-24 (2000)
- 141. Luecke, H. F. & K. R. Yamamoto: The glucocorticoid receptor blocks P-TEFb recruitment by NFkappaB to effect promoter-specific transcriptional repression. *Genes Dev*, 19, 1116-27 (2005)
- 142. Glass, C. K. & S. Ogawa: Combinatorial roles of nuclear receptors in inflammation and immunity. *Nat Rev Immunol*, 6, 44-55 (2006)
- 143. Rosenfeld, M. G., V. V. Lunyak & C. K. Glass: Sensors and signals: a coactivator/corepressor/epigenetic code for integrating signal-dependent programs of transcriptional response. *Genes Dev*, 20, 1405-28 (2006)
- 144. Aarnisalo, P., J. J. Palvimo & O. A. Jänne: CREB-binding protein in androgen receptor-mediated signaling. *Proc Natl Acad Sci U S A*, 95, 2122-7 (1998)
- 145. Sheppard, K. A., K. M. Phelps, A. J. Williams, D. Thanos, C. K. Glass, M. G. Rosenfeld, M. E. Gerritsen & T. Collins: Nuclear integration of glucocorticoid receptor and nuclear factor-kappaB signaling by CREB-binding protein and steroid receptor coactivator-1. *J Biol Chem*, 273, 29291-4 (1998)
- 146. Furumai, R., Y. Komatsu, N. Nishino, S. Khochbin, M. Yoshida & S. Horinouchi: Potent histone deacetylase inhibitors built from trichostatin A and cyclic tetrapeptide antibiotics including trapoxin. *Proc Natl Acad Sci U S A*, 98, 87-92 (2001)
- 147. Kagoshima, M., T. Wilcke, K. Ito, L. Tsaprouni, P. J. Barnes, N. Punchard & I. M. Adcock: Glucocorticoid-mediated transrepression is regulated by histone acetylation and DNA methylation. *Eur J Pharmacol*, 429, 327-34 (2001)
- 148. Ito, K., E. Jazrawi, B. Cosio, P. J. Barnes & I. M. Adcock: p65-activated histone acetyltransferase activity is repressed by glucocorticoids: mifepristone fails to recruit HDAC2 to the p65-HAT complex. *J Biol Chem*, 276, 30208-15 (2001)
- 149. Ito, K., S. Yamamura, S. Essilfie-Quaye, B. Cosio, M. Ito, P. J. Barnes & I. M. Adcock: Histone deacetylase 2-mediated deacetylation of the glucocorticoid receptor enables NF-kappaB suppression. *J Exp Med*, 203, 7-13 (2006)
- 150. Bilodeau, S., S. Vallette-Kasic, Y. Gauthier, D. Figarella-Branger, T. Brue, F. Berthelet, A. Lacroix, D. Batista, C. Stratakis, J. Hanson, B. Meij & J. Drouin: Role of Brg1 and HDAC2 in GR trans-repression of the pituitary POMC gene and misexpression in Cushing disease. *Genes Dev*, 20, 2871-86 (2006)
- 151. Pascual, G., A. L. Fong, S. Ogawa, A. Gamliel, A. C. Li, V. Perissi, D. W. Rose, T. M. Willson, M. G. Rosenfeld & C. K. Glass: A SUMOylation-dependent pathway

- mediates transrepression of inflammatory response genes by PPAR-gamma. *Nature*, 437, 759-63 (2005)
- 152. Chen, W., I. Rogatsky & M. J. Garabedian: MED14 and MED1 differentially regulate target-specific gene activation by the glucocorticoid receptor. *Mol Endocrinol*, 20, 560-72 (2006)
- 153. Rogatsky, I., J. C. Wang, M. K. Derynck, D. F. Nonaka, D. B. Khodabakhsh, C. M. Haqq, B. D. Darimont, M. J. Garabedian & K. R. Yamamoto: Target-specific utilization of transcriptional regulatory surfaces by the glucocorticoid receptor. *Proc Natl Acad Sci U S A*, 100, 13845-50 (2003)
- 154. Ogawa, S., J. Lozach, C. Benner, G. Pascual, R. K. Tangirala, S. Westin, A. Hoffmann, S. Subramaniam, M. David, M. G. Rosenfeld & C. K. Glass: Molecular determinants of crosstalk between nuclear receptors and toll-like receptors. *Cell*, 122, 707-21 (2005)
- 155. Rogatsky, I., H. F. Luecke, D. C. Leitman & K. R. Yamamoto: Alternate surfaces of transcriptional coregulator GRIP1 function in different glucocorticoid receptor activation and repression contexts. *Proc Natl Acad Sci U S A*, 99, 16701-6 (2002)
- 156. Norman, A. W., M. T. Mizwicki & D. P. Norman: Steroid-hormone rapid actions, membrane receptors and a conformational ensemble model. *Nat Rev Drug Discov*, 3, 27-41 (2004)
- 157. Löwenberg, M., A. P. Verhaar, G. R. van den Brink & D. W. Hommes: Glucocorticoid signaling: a nongenomic mechanism for T-cell immunosuppression. *Trends Mol Med*, 13, 158-63 (2007)
- 158. Maier, C., D. Runzler, J. Schindelar, G. Grabner, W. Waldhausl, G. Kohler & A. Luger: G-protein-coupled glucocorticoid receptors on the pituitary cell membrane. *J Cell Sci*, 118, 3353-61 (2005)
- 159. Tasker, J. G., S. Di & R. Malcher-Lopes: Minireview: rapid glucocorticoid signaling via membrane-associated receptors. *Endocrinology*, 147, 5549-56 (2006)
- 160. Manavathi, B. & R. Kumar: Steering estrogen signals from the plasma membrane to the nucleus: two sides of the coin. *J Cell Physiol*, 207, 594-604 (2006)
- 161. Kelly, M. J. & E. R. Levin: Rapid actions of plasma membrane estrogen receptors. *Trends Endocrinol Metab*, 12, 152-6 (2001)
- 162. Bartholome, B., C. M. Spies, T. Gaber, S. Schuchmann, T. Berki, D. Kunkel, M. Bienert, A. Radbruch, G. R. Burmester, R. Lauster, A. Scheffold & F. Buttgereit: Membrane glucocorticoid receptors (mGCR) are expressed in normal human peripheral blood mononuclear cells and up-regulated after *in vitro* stimulation and in patients with rheumatoid arthritis. *FASEB J*, 18, 70-80 (2004)

- 163. Revankar, C. M., D. F. Cimino, L. A. Sklar, J. B. Arterburn & E. R. Prossnitz: A transmembrane intracellular estrogen receptor mediates rapid cell signaling. *Science*, 307, 1625-30 (2005)
- 164. Buttgereit, F. & A. Scheffold: Rapid glucocorticoid effects on immune cells. *Steroids*, 67, 529-34 (2002)
- 165. Stahn, C., M. Lowenberg, D. W. Hommes & F. Buttgereit: Molecular mechanisms of glucocorticoid action and selective glucocorticoid receptor agonists. *Mol Cell Endocrinol*, 275, 71-8 (2007)
- 166. Vanden Berghe, W., N. Dijsselbloem, L. Vermeulen, N. Ndlovu, E. Boone & G. Haegeman: Attenuation of mitogen- and stress-activated protein kinase-1-driven nuclear factor-kappaB gene expression by soy isoflavones does not require estrogenic activity. *Cancer Res*, 66, 4852-62 (2006)
- 167. Hoang, T., I. S. Fenne, C. Cook, B. Borud, M. Bakke, E. A. Lien & G. Mellgren: cAMP-dependent protein kinase regulates ubiquitin-proteasome-mediated degradation and subcellular localization of the nuclear receptor coactivator GRIP1. *J Biol Chem*, 279, 49120-30 (2004)
- 168. Fu, X. D., Y. H. Cui, G. P. Lin & T. H. Wang: Nongenomic effects of 17beta-estradiol in activation of the ERK1/ERK2 pathway induces cell proliferation through upregulation of cyclin D1 expression in bovine artery endothelial cells. *Gynecol Endocrinol*, 23, 131-7 (2007)
- 169. Pedram, A., M. Razandi, M. Aitkenhead, C. C. Hughes & E. R. Levin: Integration of the non-genomic and genomic actions of estrogen. Membrane-initiated signaling by steroid to transcription and cell biology. *J Biol Chem*, 277, 50768-75 (2002)
- 170. Egan, K. M., J. A. Lawson, S. Fries, B. Koller, D. J. Rader, E. M. Smyth & G. A. Fitzgerald: COX-2-derived prostacyclin confers atheroprotection on female mice. *Science*, 306, 1954-7 (2004)
- 171. Borras, C., J. Gambini, M. C. Gomez-Cabrera, J. Sastre, F. V. Pallardo, G. E. Mann & J. Vina: 17beta-oestradiol up-regulates longevity-related, antioxidant enzyme expression via the ERK1 and ERK2[MAPK]/NFkappaB cascade. *Aging Cell*, 4, 113-8 (2005)
- 172. Smolen, J. S., K. Redlich, J. Zwerina, D. Aletaha, G. Steiner & G. Schett: Pro-inflammatory cytokines in rheumatoid arthritis: pathogenetic and therapeutic aspects. *Clin Rev Allergy Immunol*, 28, 239-48 (2005)
- 173. Rosen, J. & J. N. Miner: The search for safer glucocorticoid receptor ligands. *Endocr Rev*, 26, 452-64 (2005)
- 174. Winkler, J., G. Hochhaus & H. Derendorf: How the lung handles drugs: pharmacokinetics and

- pharmacodynamics of inhaled corticosteroids. *Proc Am Thorac Soc*, 1, 356-63 (2004)
- 175. Bodor, N. & P. Buchwald: Corticosteroid design for the treatment of asthma: structural insights and the therapeutic potential of soft corticosteroids. *Curr Pharm Des*, 12, 3241-60 (2006)
- 176. Schäcke, H., H. Rehwinkel, K. Asadullah & A. C. Cato: Insight into the molecular mechanisms of glucocorticoid receptor action promotes identification of novel ligands with an improved therapeutic index. *Exp Dermatol*, 15, 565-73 (2006)
- 177. Schäcke, H., M. Berger, R. H. & K. Asadullah: Selective Glucocorticoid Receptor Agonists (SEGRAs): Novel ligands with an improved therapeutic index. *Mol Cell Endocrinol*, 275, 109-17 (2007)
- 178. Vanden Berghe, W., E. Francesconi, K. De Bosscher, M. Resche-Rigon & G. Haegeman: Dissociated glucocorticoids with anti-inflammatory potential repress interleukin-6 gene expression by a nuclear factor-kappaB-dependent mechanism. *Mol Pharmacol*, 56, 797-806 (1999)
- 179. Vayssière, B. M., S. Dupont, A. Choquart, F. Petit, T. Garcia, C. Marchandeau, H. Gronemeyer & M. Resche-Rigon: Synthetic glucocorticoids that dissociate transactivation and AP-1 transrepression exhibit antiinflammatory activity *in vivo*. *Mol Endocrinol*, 11, 1245-55 (1997)
- 180. Tanigawa, K., K. Tanaka, H. Nagase, H. Miyake, M. Kiniwa & K. Ikizawa: Cell type-dependent divergence of transactivation by glucocorticoid receptor ligand. *Biol Pharm Bull*, 25, 1619-22 (2002)
- 181. Janka-Junttila, M., E. Moilanen, H. Hasala, X. Zhang, I. Adcock & H. Kankaanranta: The glucocorticoid RU24858 does not distinguish between transrepression and transactivation in primary human eosinophils. *J Inflamm (Lond)*, 3, 10 (2006)
- 182. Belvisi, M. G., S. L. Wicks, C. H. Battram, S. E. Bottoms, J. E. Redford, P. Woodman, T. J. Brown, S. E. Webber & M. L. Foster: Therapeutic benefit of a dissociated glucocorticoid and the relevance of *in vitro* separation of transrepression from transactivation activity. *J Immunol*, 166, 1975-82 (2001)
- 183. Schäcke, H., A. Schottelius, W. D. Docke, P. Strehlke, S. Jaroch, N. Schmees, H. Rehwinkel, H. Hennekes & K. Asadullah: Dissociation of transactivation from transrepression by a selective glucocorticoid receptor agonist leads to separation of therapeutic effects from side effects. *Proc Natl Acad Sci U S A*, 101, 227-32 (2004)
- 184. Humphrey, E. L., J. H. Williams, M. W. Davie & M. J. Marshall: Effects of dissociated glucocorticoids on OPG and RANKL in osteoblastic cells. *Bone*, 38, 652-61 (2006)
- 185. De Bosscher, K., W. Vanden Berghe, I. M. Beck, W. Van Molle, N. Hennuyer, J. Hapgood, C. Libert, B. Staels, A. Louw & G. Haegeman: A fully dissociated compound of

- plant origin for inflammatory gene repression. *Proc Natl Acad Sci U S A*, 102, 15827-32 (2005)
- 186. Dewint, P., V. Gossye, K. De Bosscher, W. Vanden Berghe, P. Deforce, S. Van Calenbergh, U. Müller-Ladner, B. Vander Cruyssen, G. Verbruggen, G. Haegeman & D. Elewaut: A plant-derived ligand favouring monomeric glucocorticoid receptor confirmation with impaired transactivation potential attenuates collagen-induced arthritis. *J Immunol*, under revision, (2007)
- 187. Reichardt, H. M., K. H. Kaestner, O. Wessely, P. Gass, W. Schmid & G. Schütz: Analysis of glucocorticoid signalling by gene targeting. *J Steroid Biochem Mol Biol*, 65, 111-5 (1998)
- 188. Reichardt, H. M., J. P. Tuckermann, M. Göttlicher, M. Vujic, F. Weih, P. Angel, P. Herrlich & G. Schütz: Repression of inflammatory responses in the absence of DNA binding by the glucocorticoid receptor. *EMBO J*, 20, 7168-73 (2001)
- 189. Owen, H. C., J. N. Miner, S. F. Ahmed & C. Farquharson: The growth plate sparing effects of the selective glucocorticoid receptor modulator, AL-438. *Mol Cell Endocrinol*, 264, 164-70 (2007)
- 190. Wang, J. C., N. Shah, C. Pantoja, S. H. Meijsing, J. D. Ho, T. S. Scanlan & K. R. Yamamoto: Novel arylpyrazole compounds selectively modulate glucocorticoid receptor regulatory activity. *Genes Dev*, 20, 689-99 (2006)
- 191. Gossye, V., D. Elewaut, W. Vanden Berghe, G. Haegeman & K. De Bosscher: Anti-inflammatory effects of two GR-modulators on fibroblast-like synoviocytes isolated from different rheumatoid arthritis patients (in preparation)
- 192. Foster, J. S., D. C. Henley, S. Ahamed & J. Wimalasena: Estrogens and cell-cycle regulation in breast cancer. *Trends Endocrinol Metab*, 12, 320-7 (2001)
- 193. Foster, J. S., D. C. Henley, A. Bukovsky, P. Seth & J. Wimalasena: Multifaceted regulation of cell cycle progression by estrogen: regulation of Cdk inhibitors and Cdc25A independent of cyclin D1-Cdk4 function. *Mol Cell Biol*, 21, 794-810 (2001)
- 194. Wu, Y. L., X. Yang, Z. Ren, D. P. McDonnell, J. D. Norris, T. M. Willson & G. L. Greene: Structural basis for an unexpected mode of SERM-mediated ER antagonism. *Mol Cell*, 18, 413-24 (2005)
- 195. Barker, S.: Non-steroidal anti-estrogens in the treatment of breast cancer. *Curr Opin Investig Drugs*, 7, 1085-91 (2006)
- 196. Barker, S., S. D. Malouitre, H. R. Glover, J. R. Puddefoot & G. P. Vinson: Comparison of effects of 4-hydroxy tamoxifen and trilostane on oestrogen-regulated gene expression in MCF-7 cells: up-regulation of oestrogen receptor beta. *J Steroid Biochem Mol Biol*, 100, 141-51 (2006)

- 197. Katzenellenbogen, B. S. & J. A. Katzenellenbogen: Biomedicine. Defining the "S" in SERMs. *Science*, 295, 2380-1 (2002)
- 198. Harnish, D. C.: Estrogen receptor ligands in the control of pathogenic inflammation. *Curr Opin Investig Drugs*, 7, 997-1001 (2006)
- 199. Bentrem, D., R. Dardes, H. Liu, J. MacGregor-Schafer, J. Zapf & V. Jordan: Molecular mechanism of action at estrogen receptor alpha of a new clinically relevant antiestrogen (GW7604) related to tamoxifen. *Endocrinology*, 142, 838-46 (2001)
- 200. Connor, C. E., J. D. Norris, G. Broadwater, T. M. Willson, M. M. Gottardis, M. W. Dewhirst & D. P. McDonnell: Circumventing tamoxifen resistance in breast cancers using antiestrogens that induce unique conformational changes in the estrogen receptor. *Cancer Res*, 61, 2917-22 (2001)
- 201. Olivier, S., P. Close, E. Castermans, L. de Leval, S. Tabruyn, A. Chariot, M. Malaise, M. P. Merville, V. Bours & N. Franchimont: Raloxifene-induced myeloma cell apoptosis: a study of nuclear factor-kappaB inhibition and gene expression signature. *Mol Pharmacol*, 69, 1615-23 (2006)
- 202. Biswas, D. K., S. Singh, Q. Shi, A. B. Pardee & J. D. Iglehart: Crossroads of estrogen receptor and NF-kappaB signaling. *Sci STKE*, 2005, pe27 (2005)
- 203. Sovak, M. A., R. E. Bellas, D. W. Kim, G. J. Zanieski, A. E. Rogers, A. M. Traish & G. E. Sonenshein: Aberrant nuclear factor-kappaB/Rel expression and the pathogenesis of breast cancer. *J Clin Invest*, 100, 2952-60 (1997)
- 204. Wang, X., K. Belguise, N. Kersual, K. H. Kirsch, N. D. Mineva, F. Galtier, D. Chalbos & G. E. Sonenshein: Oestrogen signalling inhibits invasive phenotype by repressing RelB and its target BCL2. *Nat Cell Biol*, 9, 470-8 (2007)
- 205. Chadwick, C. C., S. Chippari, E. Matelan, L. Borges-Marcucci, A. M. Eckert, J. C. Keith, Jr., L. M. Albert, Y. Leathurby, H. A. Harris, R. A. Bhat, M. Ashwell, E. Trybulski, R. C. Winneker, S. J. Adelman, R. J. Steffan & D. C. Harnish: Identification of pathway-selective estrogen receptor ligands that inhibit NF-kappaB transcriptional activity. *Proc Natl Acad Sci U S A*, 102, 2543-8 (2005)
- 206. Harrington, W. R., S. Sheng, D. H. Barnett, L. N. Petz, J. A. Katzenellenbogen & B. S. Katzenellenbogen: Activities of estrogen receptor alpha- and beta-selective ligands at diverse estrogen responsive gene sites mediating transactivation or transrepression. *Mol Cell Endocrinol*, 206, 13-22 (2003)
- 207. Cvoro, A., S. Paruthiyil, J. O. Jones, C. Tzagarakis-Foster, N. J. Clegg, D. Tatomer, R. T. Medina, M. Tagliaferri, F. Schaufele, T. S. Scanlan, M. I. Diamond, I. Cohen & D. C. Leitman: Selective activation of estrogen

- receptor-beta transcriptional pathways by an herbal extract. *Endocrinology*, 148, 538-47 (2007)
- 208. Hall, J. M. & D. P. McDonnell: Coregulators in nuclear estrogen receptor action: from concept to therapeutic targeting. *Mol Interv*, 5, 343-57 (2005)
- 209. Mettu, N. B., T. B. Stanley, M. A. Dwyer, M. S. Jansen, J. E. Allen, J. M. Hall & D. P. McDonnell: The Nuclear Receptor-Coactivator Interaction Surface as a Target for Peptide Antagonists of the Peroxisome Proliferator Activated Receptors. *Mol Endocrinol* (2007)
- 210. Arnold, R. & W. König: Peroxisome-proliferatoractivated receptor-gamma agonists inhibit the release of proinflammatory cytokines from RSV-infected epithelial cells. *Virology*, 346, 427-39 (2006)
- 211. Arnold, R., M. Neumann & W. König: Peroxisome proliferator-activated receptor-gamma agonists inhibit respiratory syncytial virus-induced expression of intercellular adhesion molecule-1 in human lung epithelial cells. *Immunology*, 121, 71-81 (2007)
- 212. Eun, C. S., D. S. Han, S. H. Lee, C. H. Paik, Y. W. Chung, J. Lee & J. S. Hahm: Attenuation of colonic inflammation by PPARgamma in intestinal epithelial cells: effect on Toll-like receptor pathway. *Dig Dis Sci*, 51, 693-7 (2006)
- 213. Sanchez-Hidalgo, M., A. R. Martin, I. Villegas & C. A. de la Lastra: Rosiglitazone, a PPARgamma ligand, modulates signal transduction pathways during the development of acute TNBS-induced colitis in rats. *Eur J Pharmacol*, 562, 247-58 (2007)
- 214. Neve, B. P., J. C. Fruchart & B. Staels: Role of the peroxisome proliferator-activated receptors (PPAR) in atherosclerosis. *Biochem Pharmacol*, 60, 1245-50 (2000)
- 215. Su, C. G., X. Wen, S. T. Bailey, W. Jiang, S. M. Rangwala, S. A. Keilbaugh, A. Flanigan, S. Murthy, M. A. Lazar & G. D. Wu: A novel therapy for colitis utilizing PPAR-gamma ligands to inhibit the epithelial inflammatory response. *J Clin Invest*, 104, 383-9 (1999)
- 216. Desreumaux, P., L. Dubuquoy, S. Nutten, M. Peuchmaur, W. Englaro, K. Schoonjans, B. Derijard, B. Desvergne, W. Wahli, P. Chambon, M. D. Leibowitz, J. F. Colombel & J. Auwerx: Attenuation of colon inflammation through activators of the retinoid X receptor (RXR)/peroxisome proliferator-activated receptor gamma (PPARgamma) heterodimer. A basis for new therapeutic strategies. *J Exp Med*, 193, 827-38 (2001)
- 217. Feinstein, D. L., E. Galea, V. Gavrilyuk, C. F. Brosnan, C. C. Whitacre, L. Dumitrescu-Ozimek, G. E. Landreth, H. A. Pershadsingh, G. Weinberg & M. T. Heneka: Peroxisome proliferator-activated receptor-gamma agonists prevent experimental autoimmune encephalomyelitis. *Ann Neurol*, 51, 694-702 (2002)

- 218. Diab, A., C. Deng, J. D. Smith, R. Z. Hussain, B. Phanavanh, A. E. Lovett-Racke, P. D. Drew & M. K. Racke: Peroxisome proliferator-activated receptor-gamma agonist 15-deoxy-Delta (12,14)-prostaglandin J (2) ameliorates experimental autoimmune encephalomyelitis. *J Immunol*, 168, 2508-15 (2002)
- 219. Ellis, C. N., J. Varani, G. J. Fisher, M. E. Zeigler, H. A. Pershadsingh, S. C. Benson, Y. Chi & T. W. Kurtz: Troglitazone improves psoriasis and normalizes models of proliferative skin disease: ligands for peroxisome proliferator-activated receptor-gamma inhibit keratinocyte proliferation. *Arch Dermatol*, 136, 609-16 (2000)
- 220. Setoguchi, K., Y. Misaki, Y. Terauchi, T. Yamauchi, K. Kawahata, T. Kadowaki & K. Yamamoto: Peroxisome proliferator-activated receptor-gamma haploinsufficiency enhances B cell proliferative responses and exacerbates experimentally induced arthritis. *J Clin Invest*, 108, 1667-75 (2001)
- 221. Lewis, J. D., G. R. Lichtenstein, R. B. Stein, J. J. Deren, T. A. Judge, F. Fogt, E. E. Furth, E. J. Demissie, L. B. Hurd, C. G. Su, S. A. Keilbaugh, M. A. Lazar & G. D. Wu: An open-label trial of the PPAR-gamma ligand rosiglitazone for active ulcerative colitis. *Am J Gastroenterol*, 96, 3323-8 (2001)
- 222. Feinstein, D. L., A. Spagnolo, C. Akar, G. Weinberg, P. Murphy, V. Gavrilyuk & C. Dello Russo: Receptor-independent actions of PPAR thiazolidinedione agonists: is mitochondrial function the key? *Biochem Pharmacol*, 70, 177-88 (2005)
- 223. Weng, J. R., C. Y. Chen, J. J. Pinzone, M. D. Ringel & C. S. Chen: Beyond peroxisome proliferator-activated receptor gamma signaling: the multi-facets of the antitumor effect of thiazolidinediones. *Endocr Relat Cancer*, 13, 401-13 (2006)
- 224. Nie, M., L. Corbett, A. J. Knox & L. Pang: Differential regulation of chemokine expression by peroxisome proliferator-activated receptor gamma agonists: interactions with glucocorticoids and beta2-agonists. *J Biol Chem*, 280, 2550-61 (2005)
- 225. Bougarne, N., R. Paumelle, S. Caron, B. Staels, G. Haegeman & K. De Bosscher: Cross-talk between PPARalpha and GRalpha (submitted)
- 226. Olivier, S., P. Robe & V. Bours: Can NF-kappaB be a target for novel and efficient anti-cancer agents? *Biochem Pharmacol*, 72, 1054-68 (2006)
- 227. Calzado, M. A., S. Bacher & M. L. Schmitz: NF-kappaB inhibitors for the treatment of inflammatory diseases and cancer. *Curr Med Chem*, 14, 367-76 (2007)
- 228. Auboeuf, D., D. H. Dowhan, M. Dutertre, N. Martin, S. M. Berget & B. W. O'Malley: A subset of nuclear receptor coregulators act as coupling proteins during

synthesis and maturation of RNA transcripts. *Mol Cell Biol*, 25, 5307-16 (2005)

- 229. Auboeuf, D., E. Batsche, M. Dutertre, C. Muchardt & B. W. O'Malley: Coregulators: transducing signal from transcription to alternative splicing. *Trends Endocrinol Metab*, 18, 122-9 (2007)
- 230. Michalides, R., A. Griekspoor, A. Balkenende, D. Verwoerd, L. Janssen, K. Jalink, A. Floore, A. Velds, L. van't Veer & J. Neefjes: Tamoxifen resistance by a conformational arrest of the estrogen receptor alpha after PKA activation in breast cancer. *Cancer Cell*, 5, 597-605 (2004)
- 231. Irusen, E., J. G. Matthews, A. Takahashi, P. J. Barnes, K. F. Chung & I. M. Adcock: p38 Mitogen-activated protein kinase-induced glucocorticoid receptor phosphorylation reduces its activity: role in steroid-insensitive asthma. *J Allergy Clin Immunol*, 109, 649-57 (2002)
- 232. Vanden Berghe, W., M. N. Ndlovu, R. Hoya-Arias, N. Dijsselbloem, S. Gerlo & G. Haegeman: Keeping up NF-kappaB appearances: epigenetic control of immunity or inflammation-triggered epigenetics. *Biochem Pharmacol*, 72, 1114-31 (2006)
- 233. Tsuchiya, Y., M. Nakajima, S. Takagi, T. Taniya & T. Yokoi: MicroRNA regulates the expression of human cytochrome P450 1B1. *Cancer Res*, 66, 9090-8 (2006)
- 234. Hossain, A., M. T. Kuo & G. F. Saunders: Mir-17-5p regulates breast cancer cell proliferation by inhibiting translation of AIB1 mRNA. *Mol Cell Biol*, 26, 8191-201 (2006)
- 235. Adams, B. D., H. Furneaux & B. A. White: The micro-ribonucleic acid (miRNA) miR-206 targets the human estrogen receptor-alpha (ERalpha) and represses ERalpha messenger RNA and protein expression in breast cancer cell lines. *Mol Endocrinol*, 21, 1132-47 (2007)

Abbreviations: NR: nuclear receptor, TF: transcription factor, GR: glucocorticoid receptor, ER: estrogen receptor, AR: androgen receptor, PR: progesterone receptor, MR: mineralocorticoid receptor, PPAR: peroxisome proliferatoractivated receptor, GC: glucocorticoid, RXR: retinoid X receptor, AF: activation function, LBD: ligand binding domain, DBD: DNA binding domain, PGC: PPARgamma coactivator, SRC: steroid receptor coactivator, TIF: transcription intermediary factor, CBP: cAMP responsive element binding protein, HAT: histone acetyltransferase, HDAC: histone deacetylase, SMRT: silencing mediator of retinoic acid and thyroid hormone receptors, NF-kappaB: nuclear factor-kappaB, AP-1: activator protein 1, STAT: signal transducer and activator of transcription, IRF: interferon regulatory factor, IkappaB: inhibitor of NFkappaB, IKK: IkappaB kinase complex, MMP: matrix metalloproteinase, iNOS: inducible NO synthase, COX: cyclooxygenase, IL: interleukin, TNF: tumor necrosis factor, GRE: glucocorticoid response element, E2: estradiol, MKP: MAP kinase phosphatase, ERK: extracellular regulated kinase, JNK: c-Jun N-terminal kinase, PKA: cAMP-dependent protein kinase, PKAc: the catalytical subunit of PKA, BTM: basal transcription machinery, pol II: RNA polymerase II, CTD: carboxy-terminal domain

**Key Words:** NF-kappaB, nuclear receptors, glucocorticoid receptor, estrogen receptor, peroxisome proliferatoractivated receptor, cancer, inflammation, therapeutic implications

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