

Advances in malignant melanoma: genetic insights from mouse and man

Omar Kabbarah¹ and Lynda Chin^{1,2}

¹ Department of Medical Oncology, Dana-Farber Cancer Institute, ² Department of Dermatology, Harvard Medical School, Boston, Massachusetts 02115, USA

TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Genetic predisposition to melanoma
 - 3.1. The CDKN2A locus
 - 3.1. INK4A
 - 3.3. CDK4
 - 3.4. ARF
4. Mouse models for genetic dissection of *Cdkn2a*
5. MC1R and the RHC phenotype
6. Somatic mutations in melanoma
 - 6.1. The MAPK cascade
 - 6.1.1. RAS
 - 6.1.2. BRAF
 - 6.2. HGF/SF-MET signaling
 - 6.3. PTEN and melanoma
7. Gene-environment interaction in melanoma
8. Candidate targets for melanoma therapies
9. Challenges and opportunities
10. Acknowledgements
11. References

1. ABSTRACT

Notorious for its proclivity for metastases and resistance to known therapies, malignant melanoma represents a major health concern. Genetic, epidemiological and genomic investigations are highlighting a repertoire of stereotypical mutations that are associated with human melanoma genesis. The functional significance of many of these genetic alterations is being ascertained through the use of *in vivo* mouse models. Insights from human and mouse studies, coupled with the development of novel tools for high-resolution characterization of the melanoma genome, hold promise for the identification of better diagnostic markers and potential therapeutic targets. With the rapid improvements in drug design, these recent advances are generating optimism for the development of better therapeutic options for melanoma patients.

2. INTRODUCTION

Cutaneous melanoma arises from the malignant transformation of pigment-producing melanocytes that reside at the epidermal-dermal junction of the human skin. Both genetic and environmental factors are known to contribute to disease progression. The first report on familial predisposition to melanoma in individuals with pale complexion and light hair color dates back to the 1820s (1). Two centuries later, the genetic and environmental factors that drive melanoma pathogenesis are beginning to be elucidated.

It is now clear that melanoma incidence is influenced by geographical location, such as altitude and latitude, suggesting a link between ultraviolet (UV) light exposure and melanoma risk (2). The current rise in melanoma incidence is attributable, in part, to the

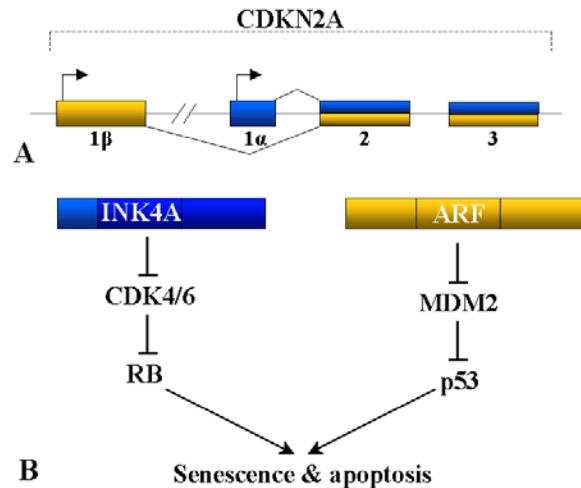


Figure 1. The CDKN2A locus at chromosome 9p21. **A.** The locus has a unique genomic organization and codes for the melanoma suppressors INK4A and ARF (9,10). Two distinct promoters drive the 1α (INK4A) and 1β (ARF) exons and result in alternatively spliced transcripts that share exons 2 and 3. Although shared, different open reading frames within exon 2 give rise to two distinct protein products. **B.** INK4A inhibits CDK4/6-cyclin D-mediated hyperphosphorylation of RB; thereby, insuring that RB is in complex with the transcription factor E2F (11). RB-E2F complexes sequester histone deacetylases that repress transcription resulting in G1 cycle arrest (12). Loss of INK4A allows CDK4/6 to phosphorylate RB, which, in turn, frees E2F to initiate transcription for entry in S phase. The melanoma suppressor ARF blocks MDM2-mediated ubiquitylation and subsequent degradation of p53 (13-16). This helps stabilize p53 and preserves its melanoma suppressive activities. Loss of ARF activity can result in uncontrolled MDM2 degradation of p53, which, in turn, can lead to loss of cell cycle control and compromised tumor suppression.

popularity of sun tanning and the increase in migration or travel of fair-skinned individuals to sun-intense locations (3). On the other hand, recent decline in melanoma incidence in Australia has been attributed to an elevated level of awareness of the dangers of excessive sun exposure (4). Genetic determinants of melanoma risk also include pigmentation patterns of the hair and skin and multiple benign or dysplastic nevi (moles). However, our current understanding of the genetic basis for these phenotypes and the mechanisms through which their underlying genetic lesions can contribute to the genesis of melanocytic tumors remains incomplete.

At present, early detection and prevention are the only effective tools for combating melanoma. With the advent of new technologies and the development of model systems, our insights into the mechanisms of melanoma formation are expanding, along with the potential for developing new and more effective therapies. Genomic investigations are highlighting lesions in melanoma-relevant genes, the functional significance of which is

being validated in refined mouse models. These advances will help identify early disease markers and rational targets for drug and biomarker discovery efforts, which should improve drastically the detection and treatment of this disease.

3. GENETICS PREDISPOSITION TO MELANOMA

Key genetic lesions that underlie familial melanoma, which represents approximately 8-12% of all cases (5), have been identified through traditional linkage mapping studies. These analyses have pointed to the two susceptibility genes, *CDKN2A* and *CDK4*, as potent regulators of melanoma genesis. More recently, polymorphic variants of the *pigmentation-associated melanocortin-1 receptor (MC1R)* have been found to be associated with predisposition to melanoma, particularly those variants encoding red hair, freckling, fair skin and sun sensitivity.

3.1. The CDKN2A locus

Identified for its frequent homozygous deletion in cancer cell lines of various types, the 9p21 locus was designated a melanoma susceptibility gene when it was found to be commonly altered in patients with familial melanoma (6-8). The *CDKN2A* locus is unique in that it encodes the two distinct cancer suppressor loci p16^{INK4A} and *ARF* (also referred to as *p14* in humans and *p19* in rodents) (9,10). Both products of the *CDKN2A* locus are potent inhibitors of melanomagenesis, and their loss predisposes to the development of the disease (Figure 1). *INK4A*, which is the founding member of the inhibitor of cyclin-dependent kinase protein family, prevents CDK4/6-mediated phosphorylation and subsequent inactivation of the *RB* tumor suppressor and results in G1 cell cycle arrest (11,12). The product of the alternative reading-frame of the 9p21 locus is *ARF*, which protects against melanoma by inhibiting MDM2-mediated ubiquitination and subsequent degradation of p53 (10,13-16).

3.2. INK4A

Inherited mutation in the *CDKN2A* locus in melanoma-prone families ranges from 25-40% (17,18). Evidence that INK4A is a melanoma suppressor comes from the presence of mutations that target exon 1α, which is the *INK4A*-specific exon within the locus in the germline of melanoma-prone individuals. Recent associations have been found between melanoma incidence and polymorphisms in the 5' and 3' untranslated regions of *CDKN2A*, suggesting that the familial predisposition alleles should be expanded to include mutations that affect the translation efficacy and those that could modulate message stability (19,20).

3.3. CDK4

Germline and acquired sporadic *CDK4* mutations that affect INK4A binding have been described in melanoma patients, reinforcing the importance of *CDKN2A* locus in suppressing melanoma. The most prevalent of these mutant alleles targets CDK4 Arg24, a residue that is conserved between CDK4 and CDK6 (Arg24Cys or Arg24His) and is known to be crucial for binding to INK4A (21-25). As expected, these *CDK4* mutations are

epistatic to *INK4A* loss, supporting the importance of *INK4A* in the regulation of CDK4/6 activity and, subsequently, inhibiting melanoma development. In accordance, the manifestations of germline *INK4A* and *CDK4* mutations are clinically indistinguishable, with a similar mean age of melanoma onset and number of melanocytic lesions (26). Finally, mice expressing the Arg24Cys mutant form of Cdk4 are susceptible to carcinogen-induced melanoma (27). Interestingly, no *Cdkn2a* loss or *Trp53* mutations were detected in melanomas arising in this model, suggesting that the Arf-p53 pathway is intact in these tumors and that defective Ink4a-Cdk4-Rb function is specifically driving melanoma genesis in these mice (27).

3.4. ARF

The finding that *CDKN2A* mutations frequently target both *INK4A* and *ARF*, that ARF plays an important regulatory role of p53, and that p53 mutation is rare in melanoma suggest the possibility that ARF may possess melanoma-suppressing properties (28). In support of this notion, somatic mutations that are specific to ARF-coding sequences in the shared exon 2 of *CDKN2A*, and insertions and deletions in *ARF* exon 1 β have been reported in two metastatic human melanomas (29-31). Inherited susceptibility to melanoma has also been documented in two patients with germline *ARF* mutations –the first was a 14-kb deletion in exon 1 β that spared both *INK4A* and *INK4B* genes, consistent with an *ARF*-specific event, and the second was a 16-bp insertion in exon 1 β that generated a frame-shifted ARF mutant defective in cell cycle arrest (32,33). However, a lack of demonstration of intact *INK4A* activity in these lesions precludes designation of *ARF* as a *bona fide* melanoma suppressor. Although partial or total deletion of exon 2 clearly result in abrogation of both p16 and ARF function, there is some evidence that point mutagenesis of exon 2 may still be preferentially targeting the p16 transcript. A recent survey of reported exon 2 point mutations (both germline and somatic) demonstrates a higher proportion of synonymous changes and a lower percentage of nonsense mutations in the ARF reading frame when compared to the p16 reading frame (34). *In silico* modeling of sequence variants that yield missense mutations in both frames further suggests that the impact of the codon change is more severe for p16 than ARF (34). Thus, although ARF clearly exhibits genetic and biochemical evidence of tumor suppressive function, unlike p16, its precise contribution to human melanoma pathogenesis remains to be clarified.

The advent of mouse models to study the tumor suppressive activities of the *Cdkn2a* locus, whose organization is evolutionarily conserved between mouse and man, provided a genetic platform upon which to assess the contributions of Ink4a and Arf to melanoma suppression (10,35-39).

4. MOUSE MODELS FOR GENETIC DISSECTION OF CDKN2A

To dissect the melanoma suppressive capabilities of the *CDKN2A* locus, exons 2 and 3 of murine *Cdkn2a*

were deleted using a conventional gene-targeting approach (40). This allele lacked both *Ink4a* and *Arf* and exhibited a high incidence of fibrosarcoma and lymphoma (40). When combined with a constitutively active *HRAS* mutation in the melanocytic lineage (*Tyr-RAS*), *Ink4a/Arf*-null animals were highly prone to cutaneous melanoma, with a short latency and high penetrance (41). Interestingly, melanomas arising in *Tyr-RAS/Cdkn2a*^{+/-} showed a loss of heterozygosity (LOH) pattern of the wild-type allele that always included exon 2, pointing to the need for elimination of both Ink4a and Arf in mouse melanoma development (41).

The role of the p53 pathway in protecting against human melanoma development has remained controversial. In fact, very few mutations or deletions of *TP53* have been reported in human melanocytic tumors (42). Early mouse modeling studies using the SV40 T antigens gave rise to a highly aggressive melanoma phenotype, pointing to a potentially important role for the p53 pathway in melanoma formation (43). Consistent with these observations, later studies on *Tyr-RAS/Trp53*^{+/-} mice also yielded a cutaneous melanoma phenotype (44). Importantly, melanomas arising in the *Tyr-RAS/Trp53*^{+/-} model were associated with LOH for the wild-type *Trp53* allele and retention of *Arf*, which provided direct evidence of the anti-melanoma capabilities of the p53 pathway (42,44). In contrast to melanomas from *Tyr-RAS/Arf*^{-/-} mice, in which Ink4a was lost in approximately 50% of the cases (discussed below), melanomas arising in *Tyr-RAS/Trp53*^{-/-} mice always retained *Ink4a* activity (44). Interestingly, the Rb pathway in *Ink4a*-competent melanomas was frequently inactivated by amplification and overexpression of *Myc*, which can act to relieve the G1 cell cycle block by acting in parallel and downstream of Ink4a (44-46). As in the case of mouse tumors, human melanomas exhibit an increase in *MYC* copy number coupled with overexpression of the oncogene (47,48). Although the basis for the discrepancy in preference for Rb-pathway inactivation between *Arf*- and *Trp53*-null mice is currently unknown, it is clear that there exists cooperation between *Myc* overexpression and *Arf* loss in some murine systems, and that inactivation of *Ink4a* can cooperate with *p53* loss (49,50). Although additional investigations are warranted to dissect the precise roles of these genetic combinations, it is evident that both the RB and p53 pathways play key roles in melanoma suppression.

To address more fully the contributions of the *Cdkn2a* locus to melanoma prevention, *Ink4a*- and *Arf*-specific knockouts have been generated. Surprisingly, mice that were deficient for *Ink4a* but carried a wild-type *Arf* allele exhibited weak spontaneous and DMBA (7,12-dimethylbenzanthracene) carcinogen-induced melanoma susceptibility phenotypes (50-52). Melanoma genesis in *Ink4a*-deficient animals was greatly enhanced in the setting of *Arf* haploinsufficiency, suggesting that *Arf* does, indeed, contribute in *Cdkn2a* melanoma suppression (53). When combined with the *Tyr-RAS* transgene, either the *Ink4a*- or *Arf*-knockout facilitated melanoma formation (53). It is noteworthy that the Rb-pathway was frequently targeted in *Tyr-RAS/Arf*^{-/-} mice, while the p53-pathway was compromised in melanomas from *Tyr-RAS/Ink4a*^{-/-}

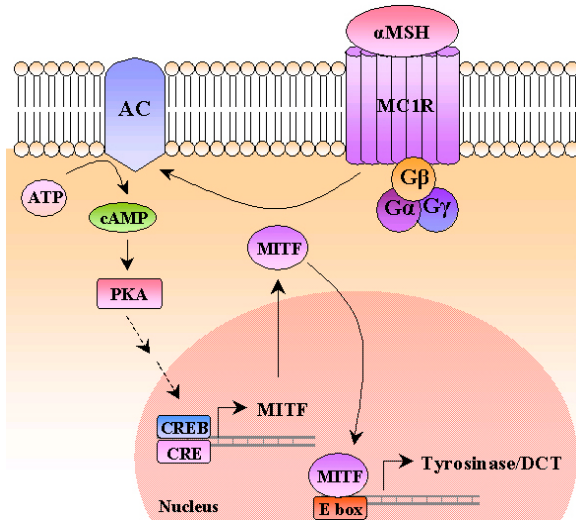


Figure 2. The melanocortin signaling pathway. Melanocytic growth is stimulated by the ligand αMSH, which binds to and activates the G protein-coupled (GPCR) melanocortin-1 receptor MC1R. The G_s family of G proteins (including Gα, Gβ and Gγ) transmits signals from MC1R to adenylyl cyclase (AC), which, in turn, catalyzes the conversion of cytoplasmic ATP to cAMP. Increased levels of cAMP act as a second messenger to activate protein-kinase A (PKA), which, upon activation, translocates to the nucleus where it phosphorylates the CREB (cAMP-responsive-element binding protein) family of transcription factors). Phosphorylated CREBs then induce the expression of genes containing CRE (cAMP-responsive elements) consensus sequences in their promoters, such as the helix-loop-helix transcription factor MITF (64). The transcription factor MITF, which is important for melanocyte differentiation and survival binds to promoters harboring E box consensus sequences (CANNTG), such as those present in the promoters of the pigimentary genes tyrosinase TRP-1 (tyrosinase-related protein-1), and DCT (dopachrome tautomerase), which is also referred to as TRP2 (64).

animals (53). These striking genetic signatures point to both *Ink4a* and *Arf* as two potent suppressors of melanoma *in vivo*, and provide a potential explanation for the need for concomitant regulation of both genes and the evolutionary conservation of the *Cdkn2a* locus.

Taken together, data from *in vivo* mouse models clearly implicate ARF and, thus, the p53 pathway, as a *bona fide* melanoma-suppressing network. Further studies are required to explain the apparent low frequency of *TP53* mutations in human melanomas, in view of the recurrent and early loss of *CDKN2A* and the close association between ARF and p53 in melanoma pathogenesis.

5. MC1R AND THE RHC PHENOTYPE

The 'red hair color' (RHC) phenotype is known to be an independent risk factor for all skin cancers, including melanoma, and is characterized by pigimentary

traits that include red hair, fair complexion, inability to tan, and freckling. The *MC1R* gene encodes a seven-transmembrane G-protein-coupled receptor that is expressed in epidermal melanocytes. Upon stimulation by its ligand, *melanocyte-stimulating factor* (MSH), the MC1R receptor acts as a key determinant of the pigimentary characteristics of the melanocytic lineage (Figure 2). Allelic polymorphisms within the *MC1R* gene underlie, in many instances, pigimentary variation and the human skin phototype (54,55). The most common three MC1R variants –Arg151Cys, Arg160Trp and Asp294His, R151C, R160W and D294H, respectively– are associated with the RHC phenotype (56-61). Carriers of one of the RHC-linked *MC1R* polymorphisms exhibit a reduced epidermal response to UV damage, and, therefore, may be at an increased risk of developing melanoma (58,59,61,61).

The association between *MC1R* polymorphisms and pigimentary traits relates to imbalances in MSH-MC1R regulation of melanin synthesis (63,64). Certain *MC1R* variants shift the pheomelanin-eumelanin balance, favoring the accumulation of the former compound in the skin. Increased pheomelanin can contribute to melanoma pathogenesis by virtue of its diminished UV protective capacity and its mutagenic and toxic properties (65). In accordance with this hypothesis, *in vitro* cell culture studies have demonstrated that *MC1R*-variant human melanocytes are hypersensitive to UV-light exposure and its associated cytotoxic effects (66). *MC1R* polymorphisms are also linked to increased melanoma risk in carriers with dark/olive complexions, as these variants are known for their ability to modify the penetrance of *CDKN2A* mutations (59,61,62,67,68). In a study conducted by Hayward and colleagues on 15 Australian families with a history of melanoma, a single *MC1R* polymorphism was associated with a substantial increase in the penetrance of *CDKN2A* mutations from 50% to 84% and reduced the mean age of melanoma onset from 58 to 37 years (61). Consistent with these observations, an independent study on Dutch families reported an increase in *CDKN2A* melanoma penetrance from 18% to 35% and 55% in patients with one or two *MC1R* variants, respectively (68). From these two studies, it is evident that common RHC MC1R variants can act as modulators of melanoma susceptibility in *CDKN2A* mutation carriers, as can additional modifiers of disease risk (69). Ongoing and future studies should help elucidate the genetic and biochemical bases for the interaction between *MC1R* and other modifiers of melanoma risk in disease pathogenesis.

6. SOMATIC MUTATIONS IN MELANOMA

Mutations of the *CDKN2A* and *CDK4* genes underlie predisposition to melanoma in only a fraction of familial and sporadic cases, pointing to the presence of additional genes that are implicated in disease pathogenesis. Molecular studies in human melanomas and derivative cell lines have uncovered a host of melanoma genes and related pathways, and the melanoma-relevance of a subset of these genetic networks has been validated on a functional level. With the advent of technologies that allow for high-resolution scanning of the melanoma

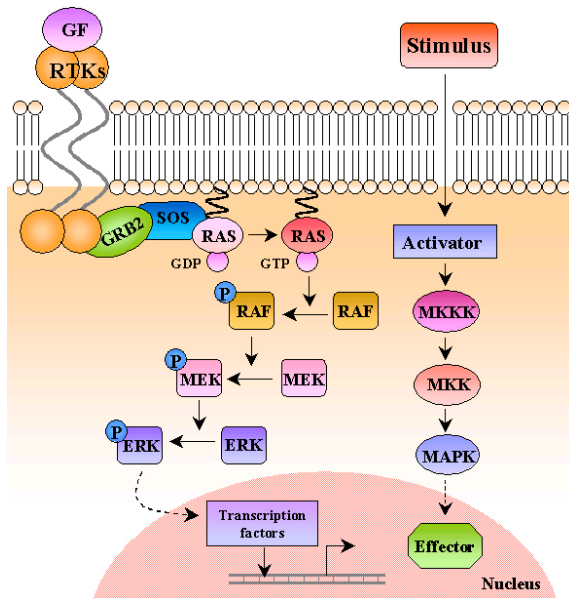


Figure 3. The MAPK-signaling pathway. Mitogen-activated protein kinases (MAPKs) relay a wide range of extracellular signals in response to a host of environmental stimuli (70). At minimum, four distinct MAPK signaling modules exist in mammalian cells. Shown here is a simplified MAPK signaling cascade with the RAS-RAF-MEK-ERK pathway. Binding of growth factors (HGF/SF, for example) to their respective receptor tyrosine kinase (RTKs), such as MET, triggers receptor dimerization and subsequent autophosphorylation of tyrosine residues present on the internal portion of the receptors. Tyrosine phosphorylation activates RTKs and allows them to bind sequence homology 2 (SH2) domains of proteins, such as GRB2. This complex then brings the cytosolic protein son of sevenless (SOS) into close proximity of RAS (HRAS, NRAS or KRAS) on the plasma membrane, and catalyzes the conversion of the inactive GDP-bound RAS into active GTP-bound RAS. Active RAS initiates the signaling cascade by phosphorylating RAF MKKK (cRAF1, ARAF or BRAF), which, in turn, phosphorylates MEK MKKs (MEK1 and MEK2). Activated MEKs phosphorylate the extracellular signal-regulated kinases ERK MAPKs (ERK1 and ERK2), which then translocate into the nucleus and phosphorylate transcription factors or other specific substrates. In addition to the classical RTK dimerization paradigm, GPCR signaling to ERK can be achieved in response to extracellular stimuli through mechanisms that are not fully understood at present (79).

genome, it is, however, increasingly clear that numerous DNA copy number alterations found in human melanomas do not harbor known melanoma targets, and, thus, may contain novel melanoma genes that have yet to be identified.

6.1. The MAPK cascade

The mitogen-activated protein kinase (MAPK) pathway regulates a broad range of cellular responses to cues from the environment (70). MAPK extracellular-

signal-regulated kinases 1 and 2 (ERK1 and ERK2) respond to diverse extracellular signals. The best-characterized signaling cascade begins with ligand-induced activation of a receptor tyrosine kinase (RTK), which in turn activates the RAS family of proto-oncogenes (NRAS, HRAS and KRAS). RAS activates the RAF serine/threonine kinases (c-RAF1, BRAF and ARAF). RAF kinases (MAPK kinase kinases (MKKK)) then phosphorylate the MAPK kinase MEK (MKK), which subsequently phosphorylates and activates ERK1 and ERK2 (Figure 3). Although many permutations of MKKK-MKK-MAPK components would appear possible, scaffolding proteins such as KSR have been suggested to confer specificity to a particular cell context (71). Additionally, this pathway is tightly regulated by several phosphatases (e.g. MKP3) whose importance in cancer biology is just becoming appreciated.

Proliferation, differentiation and survival of the melanocytic compartment is controlled in an intricate manner that requires paracrine stimulation from both GPCRs, such as endothelin-1/3 and α -melanocyte stimulating hormone (α -MSH), and RTKs, like basic fibroblast growth factor receptor (bFGFR) and hepatocyte growth factor receptor c-MET (72-76). It is also noteworthy that *Dsk5*, which represents a gain-of-function allele of the *epidermal growth factor receptor* (*Egfr*), has been found to confer an increase in epidermal melanocyte number and a concomitant hyper-pigmentation phenotype in the mouse (77). Also interesting is the fact that ERK activation has been connected to GPCR signaling through cross-talk mechanisms that are still poorly defined, placing the MAP kinase pathway at the intersection between GPCR and RTK signaling ((78,79). Given the potential cross talk between these two pathways, it is conceivable that activating mutations in *RAS*, *RAF* or other MAPK components could stimulate both GPCR and RTK signaling cascades, resulting in loss of extracellular growth-factor regulation. This might help explain the preponderance of MAPK activating mutations in many cancer types, including malignant melanoma.

6.1.1. RAS

Consistent with the frequent involvement of MAPK signaling in tumorigenesis, recent studies have reported gain-of-function *NRAS* point mutations in up to 56% of congenital nevi, 33% of primary melanomas and 26% of metastatic melanocytic tumors ((80,81). However, *NRAS* mutations have rarely been detected in dysplastic nevi, which may point to two distinct pathways of melanomagenesis; one originating from benign melanocytic lesions and the other stemming from dysplastic nevi (82-84). Interestingly, *NRAS* mutations have been reported in nodular melanomas and have been correlated with sun exposure (82,85). On the genomic level, Bastian and colleagues have reported a chromosome 11p amplification in Spitz nevi that encompasses the *HRAS* locus, and, interestingly, found that this amplified allele harbored an activating *HRAS* mutation (86). In line with the phenotype associated with activating *NRAS* mutations in humans, gain-of-function *HRAS* mutations in the melanocytic lineage of transgenic mouse leads to aberrant

proliferation and transformation, a phenotype that is exacerbated by the addition of *Cdkn2a* or *Trp53* inactivating mutations (41,44,87). Recent evidence, however, points to clear differences between *HRAS* and *NRAS* oncogenesis. Whereas transgenic *HRAS* expression results in non-metastatic melanoma, expression of dominant-active *NRAS* in Ink4a-deficient mice gives rise to metastasizing melanocytic tumors (41,88). In summary, the genetic data from both human and mouse studies highlight an important role for RAS-signaling in melanoma genesis.

6.1.2. BRAF

Activating mutations in a gene encoding another member of the MAPK signaling cascade, *BRAF*, have been identified in approximately 60% of human primary melanomas and cell lines (89). The vast majority of these *BRAF* mutations target the kinase domain, with 80% of them resulting in a single substitution (V600E) that confers constitutive *BRAF* activation (90). Significantly, *BRAF*V600E and activating *NRAS* mutations have been found to be mutually exclusive, supporting the notion that these alleles are functionally redundant in their melanoma promoting properties (89,91). In recent years, *BRAF* mutations have been detected in human nevi of both the benign and dysplastic varieties, which suggests that ERK activation may be an early event in melanoma progression (92,93). Notably, a study by Kumar *et al.* documented an inverse association in sporadic primary melanomas between activating *BRAF* and *NRAS* mutations and allelic loss on chromosome 9 (90). However, the presence of *BRAF* mutations in both benign and malignant lesions supports the need for additional cooperating genetic events to drive a full-fledged melanoma phenotype. More recently, Chudnovsky and colleagues demonstrated that Ras but not BRAF increased melanocytic neoplasia, which was attributable, in part, to the fact that RAS can activate PI3K while BRAF cannot (94). Interestingly, *BRAF* mutations were found to be significantly more common in melanomas occurring on skin subjected to intermittent sun exposure than that exposed to chronic UV (95). Furthermore, the mutated *BRAF* allele was often elevated in copy number, suggesting that *BRAF* is selected for in a subset of melanomas and pointing to the potential existence of distinct pathways of melanomagenesis (95). Ongoing studies should shed light on the genetic lesions that can cooperate with activating *BRAF* mutations in full transformation of melanocytes and on the role of BRAF in melanoma initiation and progression. The role of BRAF in melanocytic proliferation has been reviewed previously (96,97).

6.2. HGF/SF-MET signaling

The HGF/SF ligand binds to and activates tyrosine-kinase receptor c-MET, which is present on the surface of epithelial and melanocytic cells (98). Although HGF/SF-MET signaling normally acts in a paracrine manner, autocrine activation of this signaling pathway has been noted in various tumor types as well as in melanoma (99,100). Not only does HGF/SF stimulate the proliferation and motility of human melanocytes in culture, but it also downregulates E-cadherin and desmoglein-1, resulting in a disruption of normal contact between

melanocytes and keratinocytes and a cell scattering phenotype (100,101). Indeed, HGF/SF-MET activation has been linked to melanoma progression, as increased c-MET expression is characteristic of metastatic lesions and gain of the 7q33-qter region that harbors c-MET is detected in late stages of melanoma development (102-104). The melanomagenic effects of HGF/SF-MET signaling appear to be conserved between humans and mice, since studies of murine melanoma cells in explant models have demonstrated Met tyrosine-kinase activity yields a metastatic phenotype (105).

The most convincing evidence to support a role for abnormal HGF/SF-MET signaling in cancer progression comes from the observation that mice ubiquitously expressing a constitutively active HGF/SF transgene develop a variety of tumors, including cutaneous melanoma (106). Despite the relatively low incidence and long latency of melanoma in the transgenic HGF/SF model, it was found that 20% of mice presenting with melanoma develop metastatic lesions, presumably by acquiring autocrine c-Met activation (106). On the other hand, mice that carry a tissue-specific *HRAS* transgene on a *Cdkn2a*^{-/-} background exhibit a high penetrance of melanoma with a very short latency but do not show any evidence of metastases, pointing to a role for MET signaling in melanoma progression (41).

6.3. PTEN and melanoma

LOH and chromosomal studies have revealed deletions and rearrangements that target 10q24-26 in many types of tumors, including melanoma. Approximately 30-50% of human melanomas exhibit 10q LOH that encompasses both the MYC antagonist MXI1 and the tumor suppressor PTEN coding regions (107). PTEN is a well-studied tumor suppressor that negatively regulates the phosphatidylinositol 3-kinase (PI3K)-AKT pathway that relays cell proliferation and survival signals (108). Originally identified as a tumor suppressor by virtue of its homozygous deletion in breast cancers and gliomas, *PTEN* LOH or mutation have also been detected in 5-15% of primary and metastatic melanomas and in 30-40% of melanoma cell lines (109-113). Functionally, ectopic expression of PTEN was found to suppress the growth, tumorigenesis and metastasis of *PTEN*-deficient melanoma cell lines, establishing PTEN as a potent inhibitor of melanoma progression (114).

Inherited PTEN mutations in humans have been linked to three related cancer predisposition syndromes: Cowden Disease, Lhermitte-Duclos Disease and Bannayan-Zonana Syndrome (116-118). In mice, *Pten*-nullizygosity confers embryonic lethality in the mouse, while heterozygous animals exhibit an adult-onset susceptibility phenotype to a wide range of tumor types that is *Pten*-allele- and genetic background-dependent (119-122). Although not observed in *Pten*-heterozygous mice, cutaneous melanomas have been reported, albeit at low incidence, in *Pten*^{+/-}/*Cdkn2a*^{+/-} animals, which is suggestive of collaborative tumor suppression in melanoma (122).

Given that only 5-15% of human melanomas display *PTEN* mutations or allelic loss while 10q24 loss is evident in 30-50% of melanocytic tumors, and that reintroduction of *PTEN* into these tumor lines does not suppress cell growth, it is likely that other melanoma suppressors exist within the chromosome 10 locus (114). The prime candidate for the role of melanoma suppressor in the 10q24 deletion region is the *MYC* antagonist *MXI1*. Supporting evidence for a tumor suppressive role for *MXI1* in melanoma development comes from the observations that *Myc* is amplified and overexpressed in RAS-driven melanomas from *Trp53*-null mice (44), and that *Mxi1*-deficient mice display a cancer-predisposition phenotype (123). However, the role of *Mxi1* in protecting against melanoma in mice remains largely enigmatic and requires a more detailed examination. It is worth noting that a recent study has revealed high levels of phospho-AKT in 17% of normal nevi, 43% of dysplastic melanocytic lesions, 49% of primary melanomas and 77% of metastatic tumors (124). Therefore, it appears that loss of *PTEN* tumor suppression may be a more common phenomenon in melanoma than previously thought.

7. GENE-ENVIRONMENT INTERACTION IN MELANOMA

A clear link has long been established between sun and, consequently, UV exposure and skin cancer risk, including that of developing melanoma. Largely based on migration studies, it has been postulated that sun-induced damage during childhood confers a high risk of developing melanoma later in life (2,125-127). The link between sun exposure and melanoma risk is bolstered further by the fact that UV light can induce melanocytic lesions and melanoma in human skin grafts (128,129) and in mice (discussed below).

A significant number of investigations have focused on defining the molecular underpinnings of UV cancer promotion, and several mechanisms have been proposed, including a role of UV as a mutagen and mitogen, a promoter of oncogenic paracrine components and an inhibitor of immune surveillance (3, 130,131). In search of molecular targets of UV mutagenesis, studies have focused on identifying UV signature mutations in melanoma-relevant genes, such as C→T or CC→TT substitutions that result from the repair of UVB-induced damage of cyclobutane pyrimidine dimers (CPDs) and 6-4 photoproducts. *INK4A* emerged as a possible target of UV-induced mutations after the detection of C→T substitutions in the *CDKN2A* locus in human melanomas (132-134). However, the fact that similar mutations have also been observed in human gliomas, which are not linked to UV exposure, undermines the UV-*CDKN2A* connection (135). Perhaps other manifestations of UV radiation could contribute to the genesis of melanoma, such as its ability to induce crosslinks between proteins and DNA, oxidative stress and DNA single-strand breaks, chromosomal deletions associated with double-stranded breaks and altered methylation patterning that can result in epigenetic changes in gene regulation (136-138). Although our knowledge of the mechanisms of UV-mediated

carcinogenesis is rudimentary at present, studies on genetically engineered mice is providing valuable insights into the melanomagenic potential of ultraviolet light.

Using *Hgf/Sf* transgenic mice, Merlino and colleagues found that a single mild episode of sunburn in neonatal animals increased dramatically the risk of developing melanoma (139). Interestingly, chronic exposure of adult *Hgf/Sf* transgenic mice to UV light resulted in other skin cancers but not melanoma (140). Furthermore, UV exposure dramatically accelerated the onset and increased the penetrance of melanoma in *Hgf/Sf*/*Cdkn2a*^{-/-} mice (141). To examine the possibility that one or both of the *Cdkn2a* locus products might be targets of UV, *Tyr-RAS* mice neonates that were deficient for either *Ink4a* or *Arf* were treated with a single erythrogenic dose of ultraviolet radiation (142). Only the *Tyr-RAS/Arf*^{-/-} animals developed melanoma, and, surprisingly, melanomas arising in the UV-exposed mice acquired Rb-pathway lesions that were different from those observed in unexposed *Tyr-RAS/Arf*^{-/-} littermates (142). Although *Ink4a* mutations were observed in both spontaneous and UV-promoted melanomas at incidences of 50% and 25%, respectively, *Cdk6* amplification was observed only in UV-induced tumors at a frequency of 50% (142). Strikingly, loss of *Ink4a* and gain of *Cdk6* were mutually exclusive events in these UV-induced melanomas (142). In a recent study, *CDK6* amplification was found to be functionally important in driving melanomagenesis in human cells, as siRNA knockdown of the *CDK6* transcript lead to a reduction in melanoma cell growth (143). These findings suggest that Rb pathway components are critical targets of melanomagenesis in the UV-induced *Tyr-RAS/Arf*^{-/-} mouse model (142).

Much in line with findings from the *Tyr-RAS/Arf*^{-/-} model, recent studies have revealed a cooperative effect between UV exposure and *INK4A* mutation in human populations (144). In a study that included 402 melanoma patients and 713 unaffected close relatives from 80 families carrying known *INK4A* mutations, the authors assessed the penetrance of heterozygous *INK4A* mutations as well as potential environmental contributions to melanomagenesis in these individuals (144). The overall penetrance of melanoma by age 80 was found to be 67%, and, whereas only 58% of individuals of European origin developed the disease, 76% from the United States and an astounding 91% from Australia presented with melanoma (144). These observations clearly indicate that the penetrance of *INK4A* mutation is greatly influenced by geographical location, with the highest incidence of melanoma being in mutation carriers from regions of more intense UV exposure. Given the fact that UV light targets the Rb pathway in mice through inactivation of *Ink4a* or amplification of *Cdk6*, it is not surprising that *INK4A* mutation carriers living in locations of high UV exposure (Australia) would be more susceptible to melanoma than those from low UV exposure regions (Europe). Since almost all *CDKN2a* mutant humans are heterozygous, UV exposure may facilitate loss of the WT allele. It is also possible that the propensity to amplify *Cdk6* in mice and its counterpart in humans may be different.

Factors other than geographical location and *INK4A* status can also contribute to melanoma risk. For example, gene-environment interaction between *MC1R* and UV that have yet to be fully elucidated, as well as sun protection and tanning practices that are likely to contribute to melanoma risk in *INK4A* mutation carriers and non-carriers alike.

8. CANDIDATE TARGETS FOR MELANOMA THERAPIES

Traditionally, systemic chemotherapeutic or radiotherapeutic regimens have served, in many cases, as the only treatment options for cancer patients. The recent successes of the tyrosine-kinase inhibitor imatinib (Gleevec) in the treatment of gastrointestinal stromal tumors and chronic myelogenous leukaemia (CML) and the tyrosine kinase inhibitor gefitinib (Iressa), which targets the epidermal growth factor receptor in NSCLC harboring EGFR mutation (145-148) have demonstrated the power of targeted therapies. Genomic technologies and emerging proteomic tools are identifying many potential candidate targets for therapeutic development. In this respect, one of the biggest challenges will be to select those targets that are most essential for tumor maintenance. Another lesson learned from imatinib and gefitinib treatment of patients is the issue of acquired tumor drug resistance, pointing to needs for understanding drug mechanism of action as the basis for designing rational combination therapeutic regimens (149,150).

Inducible transgenic mouse models have served as an invaluable resource for studying the effects of loss of oncogene expression in established tumors, highlighting the requirement for persistent expression of certain oncogenes for tumor maintenance (151). Using such a model, a constitutively active *RAS* allele was found to be required not only for melanoma initiation, but also for tumor preservation (152). Persistent oncogenic *RAS* expression was found to play an essential role in melanoma maintenance by regulating tumor angiogenesis through vascular endothelial growth factor-dependent and -independent mechanisms (152). Given its position immediately downstream of *RAS*, it would not be surprising if *BRAF* activity were to also be required for melanoma maintenance. It is noteworthy, however, that *RAS* signals through several signaling cascades, only one of which is the *RAF-ERK* pathway. In addition, functional evaluation of the role of *BRAF* in tumor initiation and maintenance in melanoma models and in a clinical context has yet to be accomplished.

As in the case of many cancers, melanoma arises as a consequence of aberrant communications between tumor cells and the microenvironment milieu. Melanoma cells typically downregulate E-cadherin expression, which disrupts normal melanocytes-keratinocyte adhesion and allows tumors to escape into the dermis and to evade negative growth signals from keratinocytes (153). In the context of a three-dimensional skin reconstruction system, ectopic expression of E-cadherin in melanoma cells restored adhesion with keratinocytes and suppressed

melanomagenesis (154). Not only can melanoma cells escape negative growth signal from the surrounding microenvironment, but they can also elicit a paracrine response that stimulates stromal cells to produce growth factors. For example, melanoma cells often expression high levels of c-Met, which drives the production of HGF in dermal fibroblasts and provides the tumor cell with the necessary proliferative signals (100). Furthermore, numerous studies have shown that melanomagenesis is also accompanied by recruitment of endothelial cells in support of angiogenesis (155). The interdependency of melanoma cells on positive growth signals from the microenvironment and tissue vasculature provides a unique opportunity for the development of therapeutic compounds that target these tumor-supporting networks. Stromal and endothelial melanoma-supporting cells may provide better therapeutic targets, since their relative genomic stability would impede their acquisition of drug resistance.

Chromosomal instability is a major driving force behind genomic alteration that can endow a tumor with important growth advantages. Consistent with this hypothesis, melanomagenesis is accompanied by the accumulation of recurrent genomic gains and losses that are believed to contribute to disease progression (156). In fact, successful classification of melanoma has been accomplished through the use of array-based comparative genomic hybridization (aCGH), highlighting a genotype-phenotype correlation between DNA copy number and melanoma subtype (157). Interestingly, spontaneous and carcinogen-induced melanomas that arise in mouse models and derivative cell lines share some of the genomic patterns of gain and loss that are characteristic of human melanocytic tumors (158,159). The evolutionary conservation of these lesions is suggestive of their potential functional significance in driving melanomagenesis in both mice and humans. Using a high-resolution aCGH platform, we have initiated a large-scale analysis of benign nevi, primary and metastatic melanomas (160). We have observed a dramatic increase in the number of recurrent genomic gains and losses during the transition from primary to distal metastatic melanoma (unpublished observations). Although our analyses have rediscovered amplicons and deletions that target genes previously implicated in melanomagenesis, a large number of focal and recurrent events do not harbor known cancer genes (unpublished observations). The targets of a subset of these novel DNA copy number alterations will likely prove to be melanomagenic and will serve as potential targets for future drug discovery efforts. The current standing and future direction in the melanoma therapeutics field has been eloquently reviewed in several recent articles (97,161,162).

9. CHALLENGES AND OPPORTUNITIES

There is no doubt that substantial progress has been made in dissecting genetic networks and key gene-environment interactions that drive melanomagenesis. One of the major challenges will be to translate the enormous body of knowledge that has been acquired into effective therapeutic options for melanoma patients. A good example of how this is being accomplished is a *RAF*

inhibitor that is presently in clinical trials. Although activated BRAF represents a logical target for drug discovery efforts, its role in melanomagenesis and, possibly, in tumor maintenance have yet to be fully elucidated. The development of a regulatable BRAF model, analogous to the inducible HRAS mouse system, would provide the scientific community with an invaluable tool for defining the role of RAF in melanoma development and tumor maintenance. Furthermore, such an inducible BRAF model would serve as an important tool in understanding mechanisms of tumor escape that would be anticipated if RAF inhibitors were to make their way to the clinic. Finally, the use of pre-clinical models will facilitate the identification of cooperating events in melanoma development, which should help guide combination therapy initiatives, and will serve as a platform for testing the efficacy and safety of new melanoma drugs. In addition to activating *BRAF* mutations, many genetic lesions that are likely to contribute to melanoma progression have yet to be discovered. The rapidly evolving genomic technologies will likely facilitate the identification of melanoma-relevant genes, and, therefore, it will be necessary to assess their functional contributions in tumor progression and maintenance as a first step towards selecting those that would be the most suitable drug targets.

10. ACKNOWLEDGEMENTS

The authors wish to thank Dr. Norman Sharpless for critical review of the manuscript. O.K. is supported, in part, by NIH training grant T32 AR07098. L.C. is supported by NIH grants R01 CA93947 and U01 CA84313.

11. REFERENCES

- 1 Norris W: A case of fungoid disease. *Edinburgh Medicine and Surgery* 16, 562-565 (1820)
- 2 Armstrong B.K. & A. Kricke: The epidemiology of UV induced skin cancer. *J Photochem Photobiol B* 63, 8-18 (2001)
- 3 Gilchrest B.A., M.S. Eller, A.C. Geller & M. Yaar: The pathogenesis of melanoma induced by ultraviolet radiation. *N Engl J Med* 340, 1341-1348 (1999)
- 4 Marrett L.D., H.L. Nguyen & B.K. Armstrong: Trends in the incidence of cutaneous malignant melanoma in New South Wales, 1983-1996. *Int J Cancer* 92, 457-462 (2001)
- 5 Fountain J.W., S.J. Bale, D.E. Housman & N.C. Dracopoli: Genetics of melanoma. *Cancer Surv* 9, 645-671 (1990)
- 6 Nobori T., K. Miura, D.J. Wu, A. Lois, K. Takabayashi & D.A. Carson: Deletions of the cyclin-dependent kinase-4 inhibitor gene in multiple human cancers. *Nature* 368, 753-756 (1994)
- 7 Kamb A., D. Shattuck-Eidens, R. Eeles, Q. Liu, N.A. Gruis, W. Ding, C. Hussey, T. Tran, Y. Miki, J. Weaver-Feldhaus, M. McClure, J. Aitken, D. Anderson, W. Bergman, R. Frants, D. Goldgar, A. Green, R. MacLennan, N.G. Martin, L.J. Meyer, P. Youl, J.J. Zone, M.H. Skolnick & L.A. Cannon-Albright: Analysis of the p16 gene (CDKN2) as a candidate for the chromosome 9p melanoma susceptibility locus. *Nat Genet* 8, 23-26 (1994)

- 8 Hussussian C.J., J.P. Struwing, A.M. Goldstein, P.A. Higgins, D.S. Ally, M.D. Sheahan, W.H. Clark, Jr., M.A. Tucker & N.C. Dracopoli: Germline p16 mutations in familial melanoma. *Nat Genet* 8, 15-21 (1994)
- 9 Kamb A: Cell-cycle regulators and cancer. *Trends Genet* 11, 136-140 (1995)
- 10 Quelle D.E., F. Zindy, R.A. Ashmun & C.J. Sherr: Alternative reading frames of the INK4a tumor suppressor gene encode two unrelated proteins capable of inducing cell cycle arrest. *Cell* 83, 993-1000 (1995)
- 11 Serrano M., G.J. Hannon & D. Beach: A new regulatory motif in cell-cycle control causing specific inhibition of cyclin D/CDK4. *Nature* 366, 704-707 (1993)
- 12 Sherr C.J. & J.M. Roberts: CDK inhibitors: positive and negative regulators of G1-phase progression. *Genes Dev* 13, 1501-1512 (1999)
- 13 Kamijo T., J.D. Weber, G. Zambetti, F. Zindy, M.F. Roussel & C.J. Sherr: Functional and physical interactions of the ARF tumor suppressor with p53 and Mdm2. *Proc Natl Acad Sci U S A* 95, 8292-8297 (1998)
- 14 Pomerantz J., N. Schreiber-Agus, N.J. Liegeois, A. Silverman, L. Alland, L. Chin, J. Potes, K. Chen, I. Orlow, H.W. Lee, C. Cordon-Cardo & R.A. DePinho: The Ink4a tumor suppressor gene product, p19Arf, interacts with MDM2 and neutralizes MDM2's inhibition of p53. *Cell* 92, 713-723 (1998)
- 15 Stott F.J., S. Bates, M.C. James, B.B. McConnell, M. Starborg, S. Brookes, I. Palmero, K. Ryan, E. Hara, K.H. Vousden & G. Peters: The alternative product from the human CDKN2A locus, p14(ARF), participates in a regulatory feedback loop with p53 and MDM2. *EMBO J* 17, 5001-5014 (1998)
- 16 Zhang Y., Y. Xiong & W.G. Yarbrough: ARF promotes MDM2 degradation and stabilizes p53: ARF-INK4a locus deletion impairs both the Rb and p53 tumor suppression pathways. *Cell* 92, 725-734 (1998)
- 17 Aitken J., J. Welch, D. Duffy, A. Milligan, A. Green, N. Martin & N.K. Hayward: CDKN2A variants in a population-based sample of Queensland families with melanoma. *J Natl Cancer Inst* 91, 446-452 (1999)
- 18 Tsao H., X. Zhang, K. Kwitkiwski, D.M. Finkelstein, A.J. Sober & F.G. Haluska: Low prevalence of germline CDKN2A and CDK4 mutations in patients with early-onset melanoma. *Arch Dermatol* 136, 1118-1122 (2000)
- 19 Liu L., D. Dilworth, L. Gao, J. Monzon, A. Summers, N. Lassam & D. Hogg: Mutation of the CDKN2A 5' UTR creates an aberrant initiation codon and predisposes to melanoma. *Nat Genet* 21, 128-132 (1999)
- 20 Kumar R., J. Smeds, P. Berggren, O. Straume, B.L. Rozell, L.A. Akslen & K. Hemminki: A single nucleotide polymorphism in the 3'untranslated region of the CDKN2A gene is common in sporadic primary melanomas but mutations in the CDKN2B, CDKN2C, CDK4 and p53 genes are rare. *Int J Cancer* 95, 388-393 (2001)
- 21 Wolfel T., M. Hauer, J. Schneider, M. Serrano, C. Wolfel, E. Klehmann-Hieb, E. De Plaen, T. Hankeln, K.H. Meyer zum Buschenfelde & D. Beach: A p16INK4a-insensitive CDK4 mutant targeted by cytolytic T lymphocytes in a human melanoma. *Science* 269, 1281-1284 (1995)
- 22 Zuo L., J. Weger, Q. Yang, A.M. Goldstein, M.A. Tucker, G.J. Walker, N.K. Hayward & N.C. Dracopoli:

- Germline mutations in the p16INK4a binding domain of CDK4 in familial melanoma. *Nat Genet* 12, 97-99 (1996)
- 23 Soufir N, M.F. Avril, A. Chompret, F. Demenais, J. Bombléd, A. Spatz, D. Stoppa-Lyonnet, J. Benard & B. Bressac-de Paillerets: Prevalence of p16 and CDK4 germline mutations in 48 melanoma-prone families in France. The French Familial Melanoma Study Group. *Hum Mol Genet* 7, 209-216 (1998)
- 24 Tsao H, E. Benoit, A.J. Sober, C. Thiele & F.G. Haluska: Novel mutations in the p16/CDKN2A binding region of the cyclin-dependent kinase-4 gene. *Cancer Res* 58, 109-113 (1998)
- 25 Russo A.A, L. Tong, J.O. Lee, P.D. Jeffrey & N.P. Pavletich: Structural basis for inhibition of the cyclin-dependent kinase Cdk6 by the tumour suppressor p16INK4a. *Nature* 395, 237-243 (1998)
- 26 Goldstein A.M, J.P. Struwing, A. Chidambaram, M.C. Fraser & M.A. Tucker: Genotype-phenotype relationships in U.S. melanoma-prone families with CDKN2A and CDK4 mutations. *J Natl Cancer Inst* 92, 1006-1010 (2000)
- 27 Sotillo R, J.F. Garcia, S. Ortega, J. Martin, P. Dubus, M. Barbacid & M. Malumbres: Invasive melanoma in Cdk4-targeted mice. *Proc Natl Acad Sci U S A* 98, 13312-13317 (2001)
- 28 Ruas M. & G. Peters: The p16INK4a/CDKN2A tumor suppressor and its relatives. *Biochim Biophys Acta* 1378, F115-177 (1998)
- 29 Piccinin S, C. Doglioni, R. Maestro, T. Vukosavljevic, D. Gasparotto, C. D'Orazi & M. Boiocchi: p16/CDKN2 and CDK4 gene mutations in sporadic melanoma development and progression. *Int J Cancer* 74, 26-30 (1997)
- 30 Straume O, J. Smets, R. Kumar, K. Hemminki & L.A. Akslen: Significant impact of promoter hypermethylation and the 540 C>T polymorphism of CDKN2A in cutaneous melanoma of the vertical growth phase. *Am J Pathol* 161, 229-237 (2002)
- 31 Kumar R, I. Sauroja, K. Punnonen, C. Jansen & K. Hemminki: Selective deletion of exon 1 beta of the p19ARF gene in metastatic melanoma cell lines. *Genes Chromosomes Cancer* 23, 273-277 (1998)
- 32 Randerson-Moor J.A, M. Harland, S. Williams, D. Cuthbert-Heavens, E. Sheridan, J. Aveyard, K. Sibley, L. Whitaker, M. Knowles, J.N. Bishop & D.T. Bishop: A germline deletion of p14(ARF) but not CDKN2A in a melanoma-neural system tumour syndrome family. *Hum Mol Genet* 10, 55-62. (2001)
- 33 Rizos H, S. Puig, C. Badenas, J. Malvehy, A.P. Darmanian, L. Jimenez, M. Mila & R.F. Kefford: A melanoma-associated germline mutation in exon 1beta inactivates p14ARF. *Oncogene* 20, 5543-5547 (2001)
- 34 Yang R.A & H. Tsao: Recurrent Patterns of Dual RB and p53 Pathway Inactivation in Melanoma. *J Invest Dermatol* (In Press)
- 35 Duro D, O. Bernard, V. Della Valle, R. Berger & C.J. Larsen: A new type of p16INK4a/MTS1 gene transcript expressed in B-cell malignancies. *Oncogene* 11, 21-29 (1995)
- 36 Mao L, A. Merlo, G. Bedi, G.I. Shapiro, C.D. Edwards, B.J. Rollins & D. Sidransky: A novel p16^{INK4a} transcript. *Cancer Res* 55, 2995-2997 (1995)
- 37 Stone S, P. Jiang, P. Dayananth, S.V. Tavtigian, H. Katcher, D. Parry, G. Peters & A. Kamb: Complex structure and regulation of the p16(MTS1) locus. *Cancer Res* 55, 2988-2994 (1995)
- 38 Swafford D.S, S.K. Middleton, W.A. Palmisano, K.J. Nikula, J. Tesfagzi, S.B. Baylin, J.G. Herman & S.A. Belinsky: Frequent aberrant methylation of p16INK4a in primary rat lung tumors. *Mol Cell Biol* 17, 1366-1374 (1997)
- 39 Sherburn T.E, J.M. Gale & R.D. Ley: Cloning and characterization of the CDKN2A and p19ARF genes from *Monodelphis domestica*. *DNA Cell Biology* 17, 975-981 (1998)
- 40 Serrano M, H. Lee, L. Chin, C. Cordon-Cardo, D. Beach & R.A. DePinho: Role of the INK4a locus in tumor suppression and cell mortality. *Cell* 85, 27-37 (1996)
- 41 Chin L, J. Pomerantz, D. Polsky, M. Jacobson, C. Cohen, C. Cordon-Cardo, J.W. Horner, 2nd & R.A. DePinho: Cooperative effects of INK4a and ras in melanoma susceptibility in vivo. *Genes Dev* 11, 2822-2834 (1997)
- 42 Yang F.C, G. Merlino & L. Chin: Genetic dissection of melanoma pathways in the mouse. *Semin Cancer Biol* 11, 261-268 (2001)
- 43 Bradl M, A. Klein-Szanto, S. Porter & B. Mintz: Malignant melanoma in transgenic mice. *Proc Natl Acad Sci U S A* 88, 164-168 (1991)
- 44 Bardeesy N, B.C. Bastian, A. Hezel, D. Pinkel, R.A. DePinho & L. Chin: Dual inactivation of RB and p53 pathways in RAS-induced melanomas. *Mol Cell Biol* 21, 2144-2153 (2001)
- 45 Alevizopoulos K, J. Vlach, S. Hennecke & B. Amati: Cyclin E and c-Myc promote cell proliferation in the presence of p16INK4a and of hypophosphorylated retinoblastoma family proteins. *EMBO J* 16, 5322-5333 (1997)
- 46 Santoni-Rugiu E, J. Falck, N. Mailand, J. Bartek & J. Lukas: Involvement of Myc activity in a G(1)/S-promoting mechanism parallel to the pRb/E2F pathway. *Mol Cell Biol* 20, 3497-3509 (2000)
- 47 Kraehn G.M, J. Utikal, M. Udart, K.M. Greulich, G. Bezold, P. Kaskel, U. Leiter & R.U. Peter: Extra c-myc oncogene copies in high risk cutaneous malignant melanoma and melanoma metastases. *Br J Cancer* 84, 72-79 (2001)
- 48 Ross D.A. & G.D. Wilson: Expression of c-myc oncoprotein represents a new prognostic marker in cutaneous melanoma. *Br J Surg* 85, 46-51 (1998)
- 49 Schmitt C.A, M.E. McCurrach, E. de Stanchina, R.R. Wallace-Brodeur & S.W. Lowe: INK4a/ARF mutations accelerate lymphomagenesis and promote chemoresistance by disabling p53. *Genes Dev* 13, 2670-2677 (1999)
- 50 Sharpless N.E, S. Alson, S. Chan, D.P. Silver, D.H. Castrillon & R.A. DePinho: p16INK4a and p53 Deficiency Cooperate in Tumorigenesis. *Cancer Res* 62, 2761-2765 (2002)
- 51 Krimpenfort P, K.C. Quon, W.J. Mooi, A. Loonstra & A. Berns: Loss of p16INK4a confers susceptibility to metastatic melanoma in mice. *Nature* 413, 83-86 (2001)
- 52 Sharpless N.E, N. Bardeesy, K.H. Lee, D. Carrasco, D.H. Castrillon, A.J. Aguirre, E.A. Wu, J.W. Horner & R.A. DePinho: Loss of p16INK4a with retention of p19Arf

- predisposes mice to tumorigenesis. *Nature* 413, 86-91 (2001)
- 53 Sharpless N.E., K. Kannan, J. Xu, M.W. Bosenberg & L. Chin: Both products of the mouse *Ink4a/Arf* locus suppress melanoma formation in vivo. *Oncogene* 22, 5055-5059 (2003)
- 54 Valverde P., E. Healy, I. Jackson, J.L. Rees & A.J. Thody: Variants of the melanocyte-stimulating hormone receptor gene are associated with red hair and fair skin in humans. *Nat Genet* 11, 328-330 (1995)
- 55 Sturm R.A.: Skin colour and skin cancer - MC1R, the genetic link. *Melanoma Res* 12, 405-416 (2002)
- 56 Box N.F., J.R. Wyeth, L.E. O'Gorman, N.G. Martin & R.A. Sturm: Characterization of melanocyte stimulating hormone receptor variant alleles in twins with red hair. *Hum Mol Genet* 6, 1891-1897 (1997)
- 57 Smith R., E. Healy, S. Siddiqui, N. Flanagan, P.M. Steijlen, I. Rosdahl, J.P. Jacques, S. Rogers, R. Turner, I.J. Jackson, M.A. Birch-Machin & J.L. Rees: Melanocortin 1 receptor variants in an Irish population. *J Invest Dermatol* 111, 119-122 (1998)
- 58 Flanagan N., E. Healy, A. Ray, S. Philips, C. Todd, I.J. Jackson, M.A. Birch-Machin & J.L. Rees: Pleiotropic effects of the melanocortin 1 receptor (MC1R) gene on human pigmentation. *Hum Mol Genet* 9, 2531-2537 (2000)
- 59 Palmer J.S., D.L. Duffy, N.F. Box, J.F. Aitken, L.E. O'Gorman, A.C. Green, N.K. Hayward, N.G. Martin & R.A. Sturm: Melanocortin-1 receptor polymorphisms and risk of melanoma: is the association explained solely by pigmentation phenotype? *Am J Hum Genet* 66, 176-186 (2000)
- 60 Bastiaens M., J. ter Huurne, N. Gruis, W. Bergman, R. Westendorp, B.J. Vermeer & J.N. Bouwes Bavinck: The melanocortin-1-receptor gene is the major freckle gene. *Hum Mol Genet* 10, 1701-1708 (2001)
- 61 Box N.F., D.L. Duffy, W. Chen, M. Stark, N.G. Martin, R.A. Sturm & N.K. Hayward: MC1R genotype modifies risk of melanoma in families segregating *CDKN2A* mutations. *Am J Hum Genet* 69, 765-773 (2001)
- 62 Healy E., N. Flannagan, A. Ray, C. Todd, I.J. Jackson, J.N. Matthews, M.A. Birch-Machin & J.L. Rees: Melanocortin-1-receptor gene and sun sensitivity in individuals without red hair. *Lancet* 355, 1072-1073 (2000)
- 63 Busca R. & R. Ballotti: Cyclic AMP a key messenger in the regulation of skin pigmentation. *Pigment Cell Res* 13, 60-69 (2000)
- 64 Widlund H.R. & D.E. Fisher: Microphthalmia-associated transcription factor: a critical regulator of pigment cell development and survival. *Oncogene* 22, 3035-3041 (2003)
- 65 Harsanyi Z.P., P.W. Post, J.P. Brinkmann, M.R. Chedekel & R.M. Deibel: Mutagenicity of melanin from human red hair. *Experientia* 36, 291-292 (1980)
- 66 Scott M.C., K. Wakamatsu, S. Ito, A.L. Kadakaro, N. Kobayashi, J. Groden, R. Kavanagh, T. Takakuwa, V. Virador, V.J. Hearing & Z.A. Abdel-Malek: Human melanocortin 1 receptor variants, receptor function and melanocyte response to UV radiation. *J Cell Sci* 115, 2349-2355 (2002)
- 67 Kennedy C., J. ter Huurne, M. Berkhout, N. Gruis, M. Bastiaens, W. Bergman, R. Willemze & J.N. Bavinck: Melanocortin 1 receptor (MC1R) gene variants are associated with an increased risk for cutaneous melanoma which is largely independent of skin type and hair color. *J Invest Dermatol* 117, 294-300 (2001)
- 68 van der Velden P.A., L.A. Sandkuijl, W. Bergman, S. Pavel, L. van Mourik, R.R. Frants & N.A. Gruis: Melanocortin-1 receptor variant R151C modifies melanoma risk in Dutch families with melanoma. *Am J Hum Genet* 69, 774-779 (2001)
- 69 Hayward N.K.: Genetics of melanoma predisposition. *Oncogene* 22, 3053-3062 (2003)
- 70 Johnson G.L. & R. Lapadat: Mitogen-activated protein kinase pathways mediated by ERK, JNK, and p38 protein kinases. *Science* 298, 1911-1912 (2002)
- 71 Johnson G.L., H.G. Dohlman & L.M. Graves: MAPK kinase kinases (MKKKs) as a target class for small-molecule inhibition to modulate signaling networks and gene expression. *Curr Opin Chem Biol* 9, 325-331 (2005)
- 72 Imokawa G., Y. Yada & M. Miyagishi: Endothelins secreted from human keratinocytes are intrinsic mitogens for human melanocytes. *J Biochem* 267, 24675-24680 (1992)
- 73 Gilchrist B.A., H.Y. Park, M.S. Eller & M. Yaar: Mechanisms of ultraviolet light-induced pigmentation. *Photochem Photobiol* 63, 1-10 (1996)
- 74 Tada A., I. Suzuki, S. Im, M.B. Davis, J. Cornelius, G. Babcock, J.J. Nordlund & Z.A. Abdel-Malek: Endothelin-1 is a paracrine growth factor that modulates melanogenesis of human melanocytes and participates in their responses to ultraviolet radiation. *Cell Growth Differ* 9, 575-584 (1998)
- 75 Nesbit M., H.K. Nesbit, J. Bennett, T. Andl, M.Y. Hsu, E. Dejesus, M. McBrien, A.R. Gupta, S.L. Eck & M. Herlyn: Basic fibroblast growth factor induces a transformed phenotype in normal human melanocytes. *Oncogene* 18, 6469-6476 (1999)
- 76 Dupin E. & N.M. Le Douarin: Development of melanocyte precursors from the vertebrate neural crest. *Oncogene* 22, 3016-3023 (2003)
- 77 Fitch K.R., K.A. McGowan, C.D. van Raamsdonk, H. Fuchs, D. Lee, A. Puech, Y. Herault, D.W. Threadgill, M. Hrabe de Angelis & G.S. Barsh: Genetics of dark skin in mice. *Genes Dev* 17, 214-228 (2003)
- 78 Busca R., P. Abbe, F. Mantoux, E. Aberdam, C. Peyssonnaud, A. Eychene, J.P. Ortonne & R. Ballotti: Ras mediates the cAMP-dependent activation of extracellular signal-regulated kinases (ERKs) in melanocytes. *Embo J* 19, 2900-2910 (2000)
- 79 Lowes V.L., N.Y. Ip & Y.H. Wong: Integration of signals from receptor tyrosine kinases and G protein-coupled receptors. *Neurosignals* 11, 5-19 (2002)
- 80 Papp T., H. Pemsel, R. Zimmermann, R. Bastrop, D.G. Weiss & D. Schiffmann: Mutational analysis of the N-ras, p53, p16INK4a, CDK4, and MC1R genes in human congenital melanocytic naevi. *J Med Genet* 36, 610-614 (1999)
- 81 Demunter A., M. Stas, H. Degreef, C. De Wolf-Peeters & J.J. van den Oord: Analysis of N- and K-ras mutations in the distinctive tumor progression phases of melanoma. *J Invest Dermatol* 117, 1483-1489 (2001)
- 82 Jafari M., T. Papp, S. Kirchner, U. Diener, D. Henschler, G. Burg & D. Schiffmann: Analysis of ras mutations in human melanocytic lesions: activation of the

- ras gene seems to be associated with the nodular type of human malignant melanoma. *J Cancer Res Clin Oncol* 121, 23-30 (1995)
- 83 Albino A.P., D.M. Nanus, I.R. Mentle, C. Cordon-Cardo, N.S. McNutt, Bressler, J & M. Andreeff: Analysis of ras oncogenes in malignant melanoma and precursor lesions: correlation of point mutations with differentiation phenotype. *Oncogene* 4, 1363-1374 (1989)
- 84 Papp T, H. Pemsel, I. Rollwitz, H. Schipper, D.G. Weiss, D. Schiffmann & R. Zimmermann: Mutational analysis of N-ras, p53, CDKN2A (p16(INK4a)), p14(ARF), CDK4, and MC1R genes in human dysplastic melanocytic naevi. *J Med Genet* 40, E14 (2003)
- 85 van Elsas A, S.F. Zerp, S. van der Flier, K.M. Kruse, C. Aarnoudse, N.K. Hayward, D.J. Ruiter & P.I. Schrier: Relevance of ultraviolet-induced N-ras oncogene point mutations in development of primary human cutaneous melanoma. *Am J Pathol* 149, 883-893 (1996)
- 86 Bastian B.C., M. Kashani-Sabet, H. Hamm, T. Godfrey, D.H. Moore, 2nd, E.B. Brocker, P.E. LeBoit & D. Pinkel: Gene amplifications characterize acral melanoma and permit the detection of occult tumor cells in the surrounding skin. *Cancer Res* 60, 1968-1973 (2000)
- 87 Powell M.B., P. Hyman, O.D. Bell, A. Balmain, K. Brown, D. Alberts & G. Bowden: Hyperpigmentation and melanocytic hyperplasia in transgenic mice expressing the human T24 Ha-ras gene regulated by a mouse tyrosinase promoter. *Mol Carcin* 12, 82-90 (1995)
- 88 Ackermann J, M. Fruttschi, K. Kaloulis, T. McKee, A. Trumpp & F. Beermann: Metastasizing melanoma formation caused by expression of activated N-RasQ61K on an INK4a-deficient background. *Cancer Res* 65, 4005-4011 (2005)
- 89 Davies H, G.R. Bignell, C. Cox, P. Stephens, S. Edkins, S. Clegg, J. Teague, H. Woffendin, M.J. Garnett, W. Bottomley, N. Davis, E. Dicks, R. Ewing, Y. Floyd, K. Gray, S. Hall, R. Hawes, J. Hughes, V. Kosmidou, A. Menzies, C. Mould, A. Parker, C. Stevens, S. Watt, S. Hooper, R. Wilson, H. Jayatilake, B.A. Gusterson, C. Cooper, J. Shipley, D. Hargrave, K. Pritchard-Jones, N. Maitland, G. Chenevix-Trench, G.J. Riggins, D.D. Bigner, G. Palmieri, A. Cossu, A. Flanagan, A. Nicholson, J.W. Ho, S.Y. Leung, S.T. Yuen, B.L. Weber, H.F. Seigler, T.L. Darrow, H. Paterson, R. Marais, C.J. Marshall, R. Wooster, M.R. Stratton & P.A. Futreal: Mutations of the BRAF gene in human cancer. *Nature* 417, 949-954 (2002)
- 90 Kumar R, Hemminki K: Activating BRAF and N-Ras mutations in sporadic primary melanomas: an inverse association with allelic loss on chromosome 9. *Oncogene* 22, 9217-9224 (2003)
- 91 Rajagopalan H, A. Bardelli, C. Lengauer, K.W. Kinzler, B. Vogelstein & V.E. Velculescu: Tumorigenesis: RAF/RAS oncogenes and mismatch-repair status. *Nature* 418, 934 (2002)
- 92 Pollock P.M., U.L. Harper, K.S. Hansen, L.M. Yudit, M. Stark, C.M. Robbins, T.Y. Moses, G. Hostetter, U. Wagner, J. Kakareka, G. Salem, T. Pohida, P. Heenan, P. Duray, O. Kallioniemi, N.K. Hayward, J.M. Trent & P.S. Meltzer: High frequency of BRAF mutations in nevi. *Nat Genet* 33, 19-20 (2003)
- 93 Cohen C, A. Zavala-Pompa, J.H. Sequeira, M. Shoji, D.G. Sexton, G. Cotsonis, F. Cerimele, B. Govindarajan, N. Macaron & J.L. Arbiser: Mitogen-activated protein kinase activation is an early event in melanoma progression. *Clin Cancer Res* 8, 3728-3733 (2002)
- 94 Chudnovsky Y, A.E. Adams, P.B. Robbins, Q. Lin & P.A. Khavari: Use of human tissue to assess the oncogenic activity of melanoma-associated mutations. *Nat Genet* 37, 745-749 (2005)
- 95 Maldonado J.L., J. Fridlyand, H. Patel, A.N. Jain, K. Busam, T. Kageshita, T. Ono, D.G. Albertson, D. Pinkel & B.C. Bastian: Determinants of BRAF mutations in primary melanomas. *J Natl Cancer Inst* 95, 1878-1890 (2003)
- 96 Smalley K.S. & M. Herlyn: Loitering with intent: new evidence for the role of BRAF mutations in the proliferation of melanocytic lesions. *J Invest Dermatol* 123, xvi-xvii (2004)
- 97 Tuveson D.A., B.L. Weber & M. Herlyn: BRAF as a potential therapeutic target in melanoma and other malignancies. *Cancer Cell* 4, 95-98 (2003)
- 98 Bottaro D.P., J.S. Rubin, D.L. Faletto, A.M. Chan, T.E. Kmiecik, G.F. Vande Woude & S.A. Aaronson: Identification of the hepatocyte growth factor receptor as the c- met proto-oncogene product. *Science* 251, 802-804 (1991)
- 99 Vande Woude G.F., M. Jeffers, J. Cortner, G. Alvord, I. Tsarfaty & J. Resau: Met-HGF/SF: tumorigenesis, invasion and metastasis. *Ciba Found Symp* 212, 119-130 (1997)
- 100 Li G, H. Schaidt, K. Satyamoorthy, Y. Hanakawa, K. Hashimoto & M. Herlyn: Downregulation of E-cadherin and Desmoglein 1 by autocrine hepatocyte growth factor during melanoma development. *Oncogene* 20, 8125-8135 (2001)
- 101 Halaban R, J.S. Rubin, Y. Funasaka, M. Cobb, T. Boulton, D. Faletto, E. Rosen, A. Chan, K. Yoko & W. White: Met and hepatocyte growth factor/scatter factor signal transduction in normal melanocytes and melanoma cells. *Oncogene* 7, 2195-2206 (1992)
- 102 Natali P.G., M.R. Nicotra, M.F. Di Renzo, M. Prat, A. Bigotti, R. Cavaliere & P.M. Comoglio: Expression of the c-Met/HGF receptor in human melanocytic neoplasms: demonstration of the relationship to malignant melanoma tumour progression. *Br J Cancer* 68, 746-750 (1993)
- 103 Wiltshire R.N., P. Duray, M.L. Bittner, T. Visakorpi, P.S. Meltzer, R.J. Tuthill, L.A. Liotta & J.M. Trent: Direct visualization of the clonal progression of primary cutaneous melanoma: applicatio of tissue microdissection and comparative genomic hybridization. *Cancer Res* 55, 3954-3957 (1995)
- 104 Bastian B.C., P.E. LeBoit, H. Hamm, E.B. Brocker & D. Pinkel: Chromosomal gains and losses in primary cutaneous melanomas detected by comparative genomic hybridization. *Cancer Res* 58, 2170-2175 (1998)
- 105 Rusciano D, P. Lorenzoni & M.M. Burger: Expression of constitutively activated hepatocyte growth factor/scatter factor receptor (c-met) in B16 melanoma cells selected for enhanced liver colonization. *Oncogene* 11, 1979-1987 (1995)
- 106 Otsuka T, H. Takayama, R. Sharp, G. Celli, W.J. LaRochelle, D.P. Bottaro, N. Ellmore, W. Vieira, J.W. Owens, M. Anver & G. Merlino: c-Met autocrine activation induces development of malignant melanoma

- and acquisition of the metastatic phenotype. *Cancer Res* 58, 5157-5167 (1998)
- 107 Wu H, V. Goel & F.G. Haluska: PTEN signaling pathways in melanoma. *Oncogene* 22, 3113-3122 (2003)
- 108 Stambolic V, A. Suzuki, J.L. de la Pompa, G.M. Brothers, C. Mirtsos, T. Sasaki, J. Ruland, J.M. Penninger, D.P. Siderovski & T.W. Mak: Negative regulation of PKB/Akt-dependent cell survival by the tumor suppressor PTEN. *Cell* 95, 29-39 (1998)
- 109 Li J, C. Yen, D. Liaw, K. Podsypanina, S. Bose, S.I. Wang, J. Puc, C. Miliarensis, L. Rodgers, R. McCombie, S.H. Bigner, B.C. Giovanella, M. Ittmann, B. Tycko, H. Hibshoosh, M.H. Wigler & R. Parsons: PTEN, a putative protein tyrosine phosphatase gene mutated in human brain, breast, and prostate cancer. *Science* 275, 1943-1947 (1997)
- 110 Steck P, M.A. Perhouse, S.A. Jasser, W.K.A. Yung, H. Lin, A.H. Ligon, L.A. Langford, M.L. Baumgard, J. Hattier, D. T., C. Frye, R. Hu, B. Swedlund, D.H.F. Teng & S.V. Tavtigian: Identification of a candidate tumour suppressor gene, MMAC 1, at chromosome 10q23.3 that is mutated in multiple advanced cancers. *Nature Genet* 15, 356-362 (1997)
- 111 Li D.M. & H. Sun: TEP1, encoded by a candidate tumor suppressor locus, is a novel protein tyrosine phosphatase regulated by transforming growth factor beta. *Cancer Res* 57, 2124-2129 (1997)
- 112 Guldberg P, P. thor Straten, A. Birck, V. Ahrenkiel, A.F. Kirkin & J. Zeuthen: Disruption of the MMAC1/PTEN gene by deletion or mutation is a frequent event in malignant melanoma. *Cancer Res* 57, 3660-3663 (1997)
- 113 Teng D.H, R. Hu, H. Lin, T. Davis, D. Iliev, C. Frye, B. Swedlund, K.L. Hansen, V.L. Vinson, K.L. Gumpfer, L. Ellis, A. El-Naggar, M. Frazier, S. Jasser, L.A. Langford, J. Lee, G.B. Mills, M.A. Pershouse, R.E. Pollack, C. Tornos, P. Troncoso, W.K. Yung, G. Fujii, A. Berson, P.A. Steck & et al.: MMAC1/PTEN mutations in primary tumor specimens and tumor cell lines. *Cancer Res* 57, 5221-5225 (1997)
- 114 Robertson G.P, F.B. Furnari, M.E. Miele, M.J. Glendening, D.R. Welch, J.W. Fountain, T.G. Lugo, H.J. Huang & W.K. Cavenee: In vitro loss of heterozygosity targets the PTEN/MMAC1 gene in melanoma. *Proc Natl Acad Sci U S A* 95, 9418-9423 (1998)
- 115 Hwang P.H, H.K. Yi, D.S. Kim, S.Y. Nam, J.S. Kim & D.Y. Lee: Suppression of tumorigenicity and metastasis in B16F10 cells by PTEN/MMAC1/TEP1 gene. *Cancer Lett* 172, 83-91 (2001)
- 116 Liaw D, D.J. Marsh, J. Li, P.L. Dahia, S.I. Wang, Z. Zheng, S. Bose, K.M. Call, H.C. Tsou, M. Peacocke, C. Eng & R. Parsons: Germline mutations of the PTEN gene in Cowden disease, an inherited breast and thyroid cancer syndrome. *Nat Genet* 16, 64-67 (1997)
- 117 Marsh D.J, P.L. Dahia, Z. Zheng, D. Liaw, R. Parsons, R.J. Gorlin & C. Eng: Germline mutations in PTEN are present in Bannayan-Zonana syndrome. *Nat Genet* 16, 333-334 (1997)
- 118 Nelen M.R, W.C. van Staveren, E.A. Peeters, M.B. Hassel, R.J. Gorlin, H. Hamm, C.F. Lindboe, J.P. Fryns, R.H. Sijmons, D.G. Woods, E.C. Mariman, G.W. Padberg & H. Kremer: Germline mutations in the PTEN/MMAC1 gene in patients with Cowden disease. *Hum Mol Genet* 6, 1383-1387 (1997)
- 119 Di Cristofano A, B. Pesce, C. Cordon-Cardo & P.P. Pandolfi: Pten is essential for embryonic development and tumour suppression. *Nat Genet* 19, 348-355 (1998)
- 120 Podsypanina K, L.H. Ellenson, A. Nemes, J. Gu, M. Tamura, K.M. Yamada, C. Cordon-Cardo, G. Catoretti, P.E. Fisher & R. Parsons: Mutation of Pten/Mmac1 in mice causes neoplasia in multiple organ systems. *Proc Natl Acad Sci U S A* 96, 1563-1568 (1999)
- 121 Suzuki A, J.L. de la Pompa, V. Stambolic, A.J. Elia, T. Sasaki, I. del Barco Barrantes, A. Ho, A. Wakeham, A. Itie, W. Khoo, M. Fukumoto & T.W. Mak: High cancer susceptibility and embryonic lethality associated with mutation of the PTEN tumor suppressor gene in mice. *Curr Biol* 8, 1169-1178 (1998)
- 122 You M.J, D.H. Castrillon, B.C. Bastian, R.C. O'Hagan, M.W. Bosenberg, R. Parsons, L. Chin & R.A. DePinho: Genetic analysis of Pten and Ink4a/Arf interactions in the suppression of tumorigenesis in mice. *Proc Natl Acad Sci U S A* 99, 1455-1460 (2002)
- 123 Schreiber-Agus N, Y. Meng, T. Hoang, H. Hou, Jr., K. Chen, R. Greenberg, C. Cordon-Cardo, H.W. Lee & R.A. DePinho: Role of Mx1 in ageing organ systems and the regulation of normal and neoplastic growth. *Nature* 393, 483-487 (1998)
- 124 Dai D.L, M. Martinka & G. Li: Prognostic significance of activated Akt expression in melanoma: a clinicopathologic study of 292 cases. *J Clin Oncol* 23, 1473-1482 (2005)
- 125 Holman C.D & B.K. Armstrong: Cutaneous malignant melanoma and indicators of total accumulated exposure to the sun: an analysis separating histogenetic types. *J Natl Cancer Inst* 73, 75-82 (1984)
- 126 Autier P. & J.F. Dore: Influence of sun exposures during childhood and during adulthood on melanoma risk. EPIMEL and EORTC Melanoma Cooperative Group. European Organisation for Research and Treatment of Cancer. *Int J Cancer* 77, 533-537 (1998)
- 127 Whiteman D.C, C.A. Whiteman & A.C. Green: Childhood sun exposure as a risk factor for melanoma: a systematic review of epidemiologic studies. *Cancer Causes Control* 12, 69-82 (2001)
- 128 Atilasoy E.S, J.T. Seykora, P.W. Soballe, R. Elenitsas, M. Nesbit, D.E. Elder, K.T. Montone, E. Sauter & M. Herlyn: UVB induces atypical melanocytic lesions and melanoma in human skin. *Am J Pathol* 152, 1179-1186 (1998)
- 129 Berking C, R. Takemoto, R.L. Binder, S.M. Hartman, D.J. Ruiter, P.M. Gallagher, S.R. Lessin & M. Herlyn: Photocarcinogenesis in human adult skin grafts. *Carcinogenesis* 23, 181-187 (2002)
- 130 Jamal S. & R.J. Schneider: UV-induction of keratinocyte endothelin-1 downregulates E-cadherin in melanocytes and melanoma cells. *J Clin Invest* 110, 443-452 (2002)
- 131 Donawho C.K. & M.L. Kripke: Evidence that the local effect of ultraviolet radiation on the growth of murine melanomas is immunologically mediated. *Cancer Res* 51, 4176-4181 (1991)
- 132 Kamb A, N.A. Gruis, J. Weaver-Feldhaus, Q. Liu, K. Harshman, S.V. Tavtigian, E. Stockert, R.S. Day, 3rd, B.E. Johnson & M.H. Skolnick: A cell cycle regulator

potentially involved in genesis of many tumor types. *Science* 264, 436-440 (1994)

133 Pollock P.M., J.V. Pearson & N.K. Hayward: Compilation of somatic mutations of the CDKN2 gene in human cancers: non-random distribution of base substitutions. *Genes Chromosomes Cancer* 15, 77-88 (1996)

134 Peris K, S. Chimenti, M.C. Fargnoli, P. Valeri, H. Kerl & P. Wolf: UV fingerprint CDKN2a but no p14ARF mutations in sporadic melanomas. *J Invest Dermatol* 112, 825-826 (1999)

135 Kyritsis A.P., B. Zhang, W. Zhang, M. Xiao, H. Takeshima, M.L. Bondy, J.E. Cunningham, V.A. Levin & J. Bruner: Mutations of the p16 gene in gliomas. *Oncogene* 12, 63-67 (1996)

136 de Grujil F.R., H.J. van Kranen & L.H. Mullenders: UV-induced DNA damage, repair, mutations and oncogenic pathways in skin cancer. *J Photochem Photobiol B* 63, 19-27 (2001)

137 Horiguchi M, K.I. Masumura, H. Ikehata, T. Ono, Y. Kanke & T. Nohmi: Molecular nature of ultraviolet B light-induced deletions in the murine epidermis. *Cancer Res* 61, 3913-3918 (2001)

138 Jones P.A. & S.B. Baylin: The fundamental role of epigenetic events in cancer. *Nat Rev Genet* 3, 415-428 (2002)

139 Noonan F.P., J.A. Recio, H. Takayama, P. Duray, M.R. Anver, W.L. Rush, E.C. De Fabo & G. Merlino: Neonatal sunburn and melanoma in mice. *Nature* 413, 271-272 (2001)

140 Noonan F.P., T. Otsuka, S. Bang, M.R. Anver & G. Merlino: Accelerated ultraviolet radiation-induced carcinogenesis in hepatocyte growth factor/scatter factor transgenic mice. *Cancer Res* 60, 3738-3743 (2000)

141 Recio J.A., F.P. Noonan, H. Takayama, M.R. Anver, P. Duray, W.L. Rush, G. Lindner, E.C. De Fabo, R.A. DePinho & G. Merlino: Ink4a/arf deficiency promotes ultraviolet radiation-induced melanomagenesis. *Cancer Res* 62, 6724-6730 (2002)

142 Kannan K, N.E. Sharpless, J. Xu, R.C. O'Hagan, M. Bosenberg & L. Chin: Components of the Rb pathway are critical targets of UV mutagenesis in a murine melanoma model. *Proc Natl Acad Sci U S A* 100, 1221-1225 (2003)

143 Okamoto I, C. Pirker, M. Bilban, W. Berger, D. Losert, C. Marosi, O.A. Haas, K. Wolff & H. Pehamberger: Seven novel and stable translocations associated with oncogenic gene expression in malignant melanoma. *Neoplasia* 7, 303-311 (2005)

144 Bishop D.T., F. Demenais, A.M. Goldstein, W. Bergman, J.N. Bishop, B. Bressac-de Paillerets, A. Chompret, P. Ghiorzo, N. Gruis, J. Hansson, M. Harland, N.K. Hayward, E.A. Holland, G.J. Mann, M. Mantelli, D. Nancarrow, A. Platz & M.A. Tucker: Geographical variation in the penetrance of CDKN2A mutations for melanoma. *J Natl Cancer Inst* 94, 894-903 (2002)

145 Druker B.J., M. Talpaz, D.J. Resta, B. Peng, E. Buchdunger, J.M. Ford, N.B. Lydon, H. Kantarjian, R. Capdeville, S. Ohno-Jones & C.L. Sawyers: Efficacy and safety of a specific inhibitor of the BCR-ABL tyrosine kinase in chronic myeloid leukemia. *N Engl J Med* 344, 1031-1037 (2001)

146 Demetri G.D., M. von Mehren, C.D. Blanke, A.D. Van den Abbeele, B. Eisenberg, P.J. Roberts, M.C. Heinrich, D.A. Tuveson, S. Singer, M. Janicek, J.A.

Fletcher, S.G. Silverman, S.L. Silberman, R. Capdeville, B. Kiese, B. Peng, S. Dimitrijevic, B.J. Druker, C. Corless, C.D. Fletcher & H. Joensuu: Efficacy and safety of imatinib mesylate in advanced gastrointestinal stromal tumors. *N Engl J Med* 347, 472-480 (2002)

147 Paez J.G., P.A. Janne, J.C. Lee, S. Tracy, H. Greulich, S. Gabriel, P. Herman, F.J. Kaye, N. Lindeman, T.J. Boggon, K. Naoki, H. Sasaki, Y. Fujii, M.J. Eck, W.R. Sellers, B.E. Johnson & M. Meyerson: EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science* 304, 1497-1500 (2004)

148 Lynch T.J., D.W. Bell, R. Sordella, S. Gurubhagavatula, R.A. Okimoto, B.W. Brannigan, P.L. Harris, S.M. Haserlat, J.G. Supko, F.G. Haluska, D.N. Louis, D.C. Christiani, J. Settleman & D.A. Haber: Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med* 350, 2129-2139 (2004)

149 Gorre M.E., M. Mohammed, K. Ellwood, N. Hsu, R. Paquette, P.N. Rao & C.L. Sawyers: Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. *Science* 293, 876-880 (2001)

150 Haber D.A. & J. Settleman: Overcoming Acquired Resistance to Iressa/Tarceva with Inhibitors of a Different Class. *Cell Cycle* (In Press)

151 Felsher D.W.: Cancer revoked: oncogenes as therapeutic targets. *Nat Rev Cancer* 3, 375-380 (2003)

152 Chin L, A. Tam, J. Pomerantz, M. Wong, J. Holash, N. Bardeesy, Q. Shen, R. O'Hagan, J. Pantginis, H. Zhou, J.W. Horner, 2nd, C. Cordon-Cardo, G.D. Yancopoulos & R.A. DePinho: Essential role for oncogenic Ras in tumour maintenance. *Nature* 400, 468-472 (1999)

153 Hsu M.Y., M.J. Wheelock, K.R. Johnson & M. Herlyn: Shifts in cadherin profiles between human normal melanocytes and melanomas. *J Invest Dermatol Symp Proc* 1, 188-194 (1996)

154 Hsu M.Y., F.E. Meier, M. Nesbit, J.Y. Hsu, P. Van Belle, D.E. Elder & M. Herlyn: E-cadherin expression in melanoma cells restores keratinocyte-mediated growth control and down-regulates expression of invasion-related adhesion receptors. *Am J Pathol* 156, 1515-1525 (2000)

155 Streit M. & M. Detmar: Angiogenesis, lymphangiogenesis and melanoma metastasis. *Oncogene* 22, 3172-3179 (2003)

156 Bastian B.C.: Understanding the progression of melanocytic neoplasia using genomic analysis: from fields to cancer. *Oncogene* 22, 3081-3086 (2003)

157 Bastian B.C., A.B. Olshen, P.E. LeBoit & D. Pinkel: Classifying melanocytic tumors based on DNA copy number changes. *Am J Pathol* 163, 1765-1770 (2003)

158 O'Hagan R.C., C.W. Brennan, A. Strahs, X. Zhang, K. Kannan, M. Donovan, C. Cauwels, N.E. Sharpless, W.H. Wong & L. Chin: Array CGH for tumor classification and gene discovery in mouse models of malignant melanoma. *Cancer Res* 63, 5352-5356 (2003)

159 Melnikova V.O., S.V. Bolshakov, C. Walker & H.N. Ananthaswamy: Genomic alterations in spontaneous and carcinogen-induced murine melanoma cell lines. *Oncogene* 23, 2347-2356 (2004)

160 Aguirre A.J., C. Brennan, G. Bailey, R. Sinha, B. Feng, C. Leo, Y. Zhang, J. Zhang, J.D. Gans, N.

Melanoma genetics

Bardeesy, C. Cauwels, C. Cordon-Cardo, M.S. Redston, R.A. DePinho & L. Chin: High-resolution characterization of the pancreatic adenocarcinoma genome. *Proc Natl Acad Sci USA* 101, 9067-9072 (2004)

161 Thompson J, S. Menzies, H. Shaw, R. Scolyer & R. Kefford: Cutaneous melanoma. *Lancet* 365, 9476 (2005)

162 Chudnovsky Y, P.A. Khavari & A.E. Adams: Melanoma genetics and the development of rational therapeutics. *J Clin Invest* 115, 813-824 (2005)

Key Words: Tumor, Cancer, Neoplasia, Melanoma, Melanocyte, Genetics, Mouse Model, Review

Send correspondence to: Dr Lynda Chin, Department of Medical Oncology, Dana-Farber Cancer Institute, Department of Dermatology, Harvard Medical School, Boston, Massachusetts 02115, USA, Tel: 617-632-6091, Fax: 617-632-6069, E-mail: Lynda_Chin@dfci.harvard.edu

<http://www.bioscience.org/current/vol11.htm>