

## Atopic eczema: a disease modulated by gene and environment

Weian Mao<sup>1</sup>, Jingyi Mao<sup>2</sup>, Jian Zhang<sup>1</sup>, Li Wang<sup>1</sup>, Dilian Cao<sup>1</sup>, Yi Qu<sup>1</sup>

<sup>1</sup>Department of Dermatology, Shanghai Seventh People's Hospital, Shanghai, China; <sup>2</sup>Shanghai University of Traditional Chinese Medicine, Shanghai, China

### TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Epidemiology of AE
4. Molecular basis of AE
  - 4.1. Genetic factors
  - 4.2. Skin barrier dysfunction
  - 4.3. Immune system abnormalities
    - 4.3.1. Innate immunity
    - 4.3.2. Adaptive immunity
5. Microbes and food allergy in AE
6. Conclusion
7. Acknowledgements
8. References

## 1. ABSTRACT

Atopic eczema (AE) is a chronic inflammatory skin disease that is mainly characterized by pruritus and epidermal barrier dysfunction. Between 15% and 20% of children and 1%–3% of adults are affected worldwide. AE is a complex disease triggered by multiple triggers, including gene and environmental factors. Impaired skin barrier function, modifications of the immune system, and hyper-reactivity to environmental stimulation directly cause and aggravate AE. In this review, we provide an overview of the recent developments and future directions in the pathogenesis of AE.

## 2. INTRODUCTION

Atopic eczema (AE), also known as atopic dermatitis (AD), is one of the most common chronic inflammatory skin diseases with a strong family predisposition occurring worldwide (1,2). Most of the patients with AE are infants; the onset is as early as 2–7 months of age (3). Many people outgrow AE by early adulthood, however, according to the theory of Atopic March, young children with AE may suffer from airway allergy, such as asthma or allergic rhinitis, later in life (4–7).

AE is a multifactorial skin disorder characterized by pruritus, epidermal barrier dysfunction, skin lesions, high susceptibility to allergens and microbes, and an ongoing course of relapse and remission (8–10). According to the nomenclature for AE by the Nomenclature Review Committee of the World Allergy Organization, the term AE should only be used for eczema patients with elevated total serum immunoglobulin E (IgE) levels >150kU/l and IgE-specific sensitization to allergens through an IgE-antibody determination or skin test (11). Other authorities use the term non-atopic eczema with chronic inflammation and skin disorder, but with less of a change in IgE antibody (12,13).

Complex factors contribute to the development of AE. With respect to the genetic aspect, a loss-of-function mutation of filaggrin, which is known as an important gene in the regulation of the epidermal barrier, may increase the risk of AE (14–17). People with AE often exhibit elevated interleukin (IL)-4, IL-5, IL-9, IL-13, and IL-17 levels and high-affinity IgE receptors (18–22). In contrast, most AE patients have hyper-reactivity to food or aero allergens, and are always associated with allergic asthma, rhinitis, conjunctivitis, and allergic contact urticaria, and are more vulnerable to microbes and viruses (23,24). However, the exact pathogenesis and the link between other diseases are

far from understood. In this review we discuss the recent developments in the mechanism AE and future directions in this field.

### 3. EPIDEMIOLOGY OF AE

Different diagnostic guidelines have been proposed for the diagnosis of AE (25). The criteria established by Hanifin and Rajka are the most well-accepted and accurate (26,27); however, the criteria are infrequently used in large epidemiologic studies due to time constraints. Questionnaire studies, such as the International Study of Asthma and Allergies in Childhood (ISAAC), provide a worldwide method for estimation of the prevalence of AE. Although there may be slight deviation as the results vary based on the answers of the individual, the ISAAC questionnaire still has a strong correlation with the findings on clinical examination (28–31).

Currently, 15%–20% of children and 1%–3% of adults are affected with AE worldwide (32,33). According to the Scoring of Atopic Dermatitis (SCORAD), which was developed by the European Task Force of Atopic Dermatitis (ETFAD) to determine the severity of AE, the majority of patients are classified as ‘mild,’ whereas 10%–20% of patients are ‘severe,’ and this percentage seems to be higher in the adult AE population. Nearly 60% of patients experience remission (34).

The prevalence of AE has increased significantly in the last decade, reaching >10% of the population in developing countries, with a plateau at approximately 20% in Western countries (30,35). Nigeria, the United Kingdom, and New Zealand have the highest prevalence. Recent research has also shown an increase in AE in Korea, Japan, and India.

### 4. MOLECULAR BASIS OF AE

#### 4.1 Genetic factors

Recent studies have shown that AE has a strong family predisposition with a phenotype concordance of 0.72–0.77 and 0.15–0.23 in monozygotic and dizygotic twin pairs, respectively, and is highly heritable (2,36). A number of single nucleotide polymorphisms (SNPs) have been described in genes associated with AE, thus highlighting the importance of the genetic component in the pathogenesis of AE. Moreover, genes involved in epidermal barrier differentiation and immune responses have been implicated in AE development.

Filaggrin, derived from filament aggregation protein, is a major protein that facilitates terminal differentiation of the epidermis and formation of the skin barrier (16). A loss-of-function mutation in the filaggrin gene (FLG) has been demonstrated as the most significant genetic factor for the development of AE.

FLG is located in the epidermal differentiation complex on chromosome 1q21 (37), with other genes encoding loricrin and S100 calcium-binding proteins. The product of FLG is profilaggrin, a large, insoluble

polyprotein which is expressed in terminally differentiating keratinocytes in the outermost layers of the human epidermis. Profilaggrin is the major constituent of keratohyalin granules in the stratum granulosum. Profilaggrin can be dephosphorylated and cleaved by several endoproteases to produce the functional monomeric filaggrin (38–41).

In mice, loss-of-function mutations of FLG result in the absence or reduction of the FLG protein and lead to a compromised skin barrier that allows the entry of allergens, then triggers immunologic responses. Previous studies have shown that nearly 25%–50% of AE patients have FLG loss-of-function mutations (42,43). The down regulation of FLG leads to a defective skin barrier, which allows external antigens to penetrate the epidermis and initiate immune responses. Since Smith *et al.* (44) first discovered the two loss-of-function mutations, R501X and 2282del4, in 2006, 3321delA, E2422X, S3247X, and R2447 mutations have been identified. The mutations vary within the population and geographic regions and can also occur in patients with ichthyosis vulgaris, psoriasis, asthma, and allergic rhinitis (45,46).

In addition, many AE-associated genes important for inflammation and atopy have been identified. Chromosome 5q31–33 harbors genes, including IL-3, IL-4, IL-5, and IL-13, and granulocyte-macrophage colony-stimulating factor (GM-CSF), which encode Th2 cytokines and regulate IgE production. The 590C/T mutation of the IL-4 gene promoter region increases the transcriptional activity of IL-4, resulting in upregulated pro-inflammatory factors, such as IL-19, IL-20, IL-1 $\alpha$ , and IL-25, and downregulated antimicrobial factors, such as interferon (IFN)- $\gamma$ , S100s, and Toll-like receptors. Polymorphic variants of IL-13 (R130Q and R110Q) have been shown to be associated with atopy. IL-4 and IL-13 can also downregulate FLG expression in patients with AE (47,48).

Signal transducer and activator of transcription 6 (STAT6), encoded on chromosome 12q13–24, is a downstream factor of IL-4 and IL-13. IL-4 and IL-13 can activate STAT6 by a phosphorylation process, making STAT6 able to translocate to the nucleus and bind to target genes to regulate its expression (49–51). Activation of STAT6 in AE patients may contribute to the elevated serum IgE level and impaired epidermal barrier. Vladich *et al.* (52) demonstrated that an IL-13 (R130Q) mutation can induce the phosphorylation and activation of STAT6. Moreover, a very recent study showed SNPs in IL-13 (rs20541) and STAT6 (rs1059513) have a combined effect on the risk of eczema, which revealed the gene–gene interaction in AE.

Many other genes, such as SPINK5, NOD1, NOD2, CCL17, IL-18, CTLA4, and PHF11, also have strong relationships with AE development. There is still much work to be done to better understand the consequences of those mutations and the pathophysiology of AE.

### 4.2. Skin barrier dysfunction

The epidermis not only functions as a physical barrier, but also an active immunologic organ. It is the first barrier to protect individuals from microbes, allergens, viruses, and toxins from the outer environment. Dry skin and skin lesions, the most significant phenotypes of patients with AE, mainly result from lack of function of the epidermal barrier.

The stratum corneum (SC), the uppermost layer of the skin, consists of flattened keratinocytes, and lipids play a key role in the protective function of the skin. During epidermal differentiation, keratinocytes move from a proliferative cell type in the basal cell layer to flat, dead cell remnants (corneocytes) in the SC. Lipids, such as free fatty acids and cholesterol encompass the corneocytes and protect the skin from water loss (53).

Ceramides, the dominant lipids, make up approximately 50% of the human SC. Lipids play an important role in determination of the permeability barrier and water reservoir of the epidermis. The balance of ceramides is regulated by three sphingolipid hydrolysis enzymes, in which  $\beta$ -glucocerebrosidase (GlcCDase) and sphingomyelinase (SMase) contribute to the synthesis, whereas ceramidase is for the degradation of ceramides (54,55). Thus, the expression and activity of these enzymes could be factors involved with AE. Researchers have discovered that AE patients exhibit reduced levels of ceramides, especially ceramide-1, although one study showed no change in ceramide levels in uninvolved atopic skin. Hara *et al.* (56) have shown that a deficiency in ceramides is linked to the high expression of sphingomyelin deacylase, which can compete with SMase or GlcCDase for a common substrate, sphingomyelin or glucosylceramide. In contrast, the skin of AE patients is frequently colonized by bacteria, especially *Staphylococcus aureus*. The bacteria are likely to secrete significantly more of the ceramidase, but less of the sphingomyelinase in all skin types of AE than healthy patients (57). Faster degradation results, but less synthesis of ceramides and thus a decreased amount of ceramides.

In addition, several proteases located in the SC are associated with AE patients. Proteases not only act as enzymes that conduct hydrolysis of peptide bonds, but also signaling molecules that contribute to increased desquamation and skin barrier dysfunction. Serine protease (SP) can mediate pro-inflammatory effects through protease-activated receptor-2 (PAR-2), induce the secretion of pro-inflammatory cytokines, and result in skin barrier disruption. Three SPs (stratum corneum chymotryptic enzyme [SCCE], stratum corneum tryptic enzyme [SCTE], and stratum corneum cathepsin-L-like enzyme [SCCL]) have been identified in SC and are important for desquamation (58,59). The mutation of an AACC insertion in the 3'UTR of the SCCE gene has been described in some patients with AE (60). This mutation may result in a change in SCCE activity and people with this mutation are more than two times as likely to develop AE as individuals with the normal allele. Moreover, soaps and other detergents can increase skin pH, leading to

increased activity of both endogenous and exogenous proteases, which is suspected to induce abnormalities in SC integrity and permeability barrier homeostasis (61).

In contrast, the serine protease inhibitor, Karzal type 5 (SPINK5), is expressed in epidermis and the product of SPINK5, lymphoepithelial Kazal-type-related inhibitor (LEKTI), is a protease inhibitor which inhibits SCTE and SCCE. Mutation of SPINK5 has been implicated in Netherton syndrome, a rare skin disease characterized by greatly elevated IgE levels with atopic manifestations. Several studies have shown that SNPs of SPINK5 are associated with AE patients and the subsequent severe inflammation (62–64).

### 4.3. Immune system abnormalities

The deficient skin barrier of AE patients facilitates the entry of infectious microbes and allergens into the skin where they encounter immunocompetent cells and initiate rapid innate and adaptive immune responses. Alterations in both innate and adaptive immunity have been described in AE.

#### 4.3.1. Innate immunity

Innate immunity is the first host barrier and antigen-non-specific defense mechanisms can be activated immediately or within several hours after exposure to virtually any foreign agent. The components of innate immune system always focus on pattern-recognition receptors (PRRs), pathogen-associated molecular patterns (PAMPs), and antimicrobial peptides (AMPs). PRRs play a pivotal role in the induction of the innate immune system, and can respond to highly conserved PAMPs shared by many classes of pathogens, including bacterial cell-wall products (such as LPS), peptidoglycan (PGN), and lipoteichoic acid (LTA), the fungal cell wall product zymosan, and viral double-stranded RNA. To discriminate these PAMPs, numerous PRRs have been identified and characterized, such as toll-like receptor (TLRs), C-type lectin receptors (CLRs), CD14, double-stranded RNA binding kinase, and nucleotide-binding oligomerization domain (NOD; 65–67). Stimulation of PRRs by PAMPs will initiate a signal transduction cascade that leads to the release of AMPs, cytokines, and chemokines, which are important for the recruitment of effector leukocytes or have direct antimicrobial effects to limit the infection.

Among these PRRs, TLRs are the most extensively studied, with 11 identified members (TLR1–11). TLRs are expressed on various cells of the innate immune system, including macrophages, dendritic cells (DCs), neutrophils, and mucosal epithelial and endothelial cells. TLRs not only bind to PAMPs, but also recognize newly discovered self-molecules released in response to tissue damage, which are collectively referred to as damage-associated molecular patterns (DAMPs). Ligand recognition induces signal transduction through a myeloid differentiation primary response gene-88 (MyD88)-dependent pathway, activating nuclear factor  $\kappa$ B (NF- $\kappa$ B) and resulting in the production of pro-inflammatory cytokines. TLR3 and TLR4 use a MyD88-independent pathway and activate interferon regulatory factor 3, resulting in IFN- $\beta$  gene expression.

Patients with AE have been shown to have reduced TLR function. TLR2, compared to most other TLRs, can recognize a remarkably broad range of PAMPs and is essential for the response to several pathogens. As TLR2 can recognize components of *S. aureus*, such as LTA and PGN, studies have indicated that TLR2 deficiency may contribute to susceptibility to *S. aureus* and severity of AE. A missense mutation (R753Q) of TLR2 occurs at a frequency of 12% in adult AE patients and is associated with a more severe phenotype, higher serum total IgE levels, and greater susceptibility to *S. aureus* colonization, although there is one study that showed opposite results (68–70). Patients with AE have a significantly lower expression of TLR2, and reduced IL-6, IL-8, and IL-1 $\beta$  pro-inflammatory cytokines released by macrophages. However, one study showed that monocytes/macrophages from AE patients with the TLR2 R753Q SNP produced significantly more IL-6 and IL-12, which may also act as important factors to stimulate T cells and thus initiate the adaptive immune response (71).

Furthermore, an A-16934T mutation in the TLR2 promoter region, which inhibits TLR2 transcription, was identified with an increased secretion of IL-6 and high total serum IgE levels (72). This mutation was significantly overrepresented in individuals with severe AE (SCORAD>50) and was associated with allergic asthma, hay fever symptoms, and recurrent bacterial infections. TLR9, which is found within the endosome, can recognize unmethylated CpG DNA and intracellular viral antigens. Still, no SNPs were found in TLR1, TLR3, and TLR6 with AE (71,73).

NOD, also known as caspase activation and recruitment domain (CARD) are intracellular receptors which can recognize PAMPs, particularly PGN through the C-terminal leucine-rich repeat (LRR) region and trigger the downstream signaling pathway via activation of NF- $\kappa$ B. The NOD family includes five members, among which NOD1 and NOD2 are the most prominent. Keratinocytes express NOD1 and NOD2 can produce IL-6 after stimulation with PGN and AMP, and human  $\beta$ -defensin (HBD) 2 after stimulation with NOD2-specific ligand muramyl dipeptide. Furthermore, NOD1 is located on chromosome 7p14–p15, a region linked to atopy, whereas NOD2 is located on chromosome 16q12, a locus associated with several autoimmune diseases. SNPs of NOD1 (rs2907748, rs2907749, and rs2075822) as well as a NOD2 variant (R702W) are significantly associated with AE and asthma. Moreover, NOD2-deficient mice have impaired clearance of *S. aureus* after subcutaneous or intraperitoneal infection (74–76). Whether or not NOD2 SNPs are correlated with increased susceptibility to epicutaneous *S. aureus* infections in patients with AE needs to be addressed.

In addition, CD14 and mannan-binding lectin (MBL) can respond to PAMPs and have also been shown to be associated with AE development (77). These factors not only contribute to the innate immune

system in patients with AE, but also adaptive immunity, which highlights the importance of further study.

### 4.3.2. Adaptive immunity

The adaptive immune system, also known as the acquired immune system, which consists of highly specialized, systemic cells and processes, eliminates or prevents pathogen growth (78). Acquired immunity is triggered when a pathogen evades the innate immune system in vertebrates (79,80). Acquired immunity creates immunologic memory after an initial response to a specific pathogen, leading to an enhanced response to subsequent encounters with that same pathogen. The cells functioning in the acquired immune system are T and B lymphocytes (81). T cells are intimately involved in cell-mediated immune responses, whereas B cells play a large role in the humoral immune response. Immediately after recognizing foreign antigen in the cellular context, the acquired immune response is activated. The foreign antigen presented by DCs or macrophages induces a complex response, in which CD8<sup>+</sup> T lymphocytes directly kill the pathogens, the helper T cells secrete numerous cytokines, such as IFN- $\gamma$ , to clear the pathogens (82), and B cells produce antibodies to recognize and neutralize specific pathogens.

Many inflammatory skin diseases are thought to be mediated by T cell activation and proliferation in the skin (83). T cells are one of the major elements of adaptive immunity and have a critical role in the pathogenesis of AE (84). The migration of memory and effector T cells to the inflamed skin plays an essential role in the development of atopic skin inflammation. The initial phase of AE is predominated by T helper type 2 (Th2) cytokines, then switches to a more chronic Th1-dominated eczematous phase (85). As such, AE is a biphasic disease. It has been shown that AE patients exhibit characteristic features of dramatic Th2 polarization with high levels of IL-4, IL-5, and IL-13 in the acute phase in both lesional and non-lesional skin in combination with a predominance of Th2 cytokines in the blood (86,87). Increased mRNA expression of IFN- $\gamma$ , IL-5, IL-12, and GM-CSF is observed in patients with chronic AE, whereas mRNA expression for Th1 cytokines, such as IFN- $\gamma$  and IL-12, is not detectable in acute AE skin lesions. Based on the different cytokines elaborated during the chronic phase of disease, biphasic AE suggests the initiation of acute skin inflammation by Th2 cytokines and maintenance of chronic inflammation by Th1 cytokines. DCs contribute to allergic sensitization and maintenance of inflammation with the Th2-to-Th1 switch.

An increase number of peripheral blood CD4<sup>+</sup>CD25<sup>+</sup>regulatory T (Treg) cells have been demonstrated in AE patients compared to healthy controls (88,89). Treg cells control the activation of autoreactive and T effector cells and are crucial for the maintenance of peripheral tolerance to self-antigens. The balance between Th2 cells and allergen-specific Treg cells appears to be decisive in the development of

allergy. It has been shown that Treg cells from AE patients markedly inhibit the activation of IL-4-secreting Th2 cells and IFN- $\gamma$ -secreting Th1 cells stimulated with antigen *in vitro* (90,91).

In addition to T cells, B cells are also involved in the process. Enhanced IgE production by B cells occurs in AE patients. The effective production of IgE in atopic disease by B cells depends on support by Th2 cells. B cells play a critical role in antigen-specific CD4<sup>+</sup> T-cell proliferation and Th2 and IL-17 responses in a murine model of AE (86,87). CD19 expression of B cells has been found to play an important role in AE. Compared to T cells, however, the role of B cells in AE needs further exploration. Taken together, the important role of the adaptive immune system in AE has been clarified, which provides targeted therapies for the treatment of AE (86,87).

### 5. MICROBES AND FOOD ALLERGY IN AE

AE is frequently complicated by recurrent skin infections with bacterial, viral, and mycotic pathogens. The role of microbial superinfections has not been fully elucidated, but there is a general consensus that bacterial superinfections, in part due to impaired innate immunity, play a critical role in the clinical course of skin lesions (92). *S. aureus* colonization of both lesional and clinically uninvolved skin in AE has been demonstrated to increase significantly and exacerbate the disease (57). Greater than 90% and 76% *S. aureus* colonization has been demonstrated in lesional and non-lesional skin of AE patients, respectively, whereas <10% *S. aureus* colonization is associated with healthy skin (93–95). A previous study concluded that *S. aureus* colonization is both a cause and a consequence of allergic skin inflammation. Patients developing AE exhibit impaired skin barriers, increased synthesis of extracellular matrix adhesions for *S. aureus*, reduced skin lipid content, increased skin surface pH, and defective innate immune responses, which lead to a significant increase in *S. aureus* colonization (96). In contrast, the exotoxins secreted by *S. aureus* are superantigens which could be recognized by large numbers of different T cells via interaction with the major histocompatibility complex (MHC) II and  $\beta$ -chain of the T cell receptor. Skin immune response activating and cytokines releasing (tumor necrosis factor (TNF)- $\alpha$ , IFN- $\gamma$ , IL-1, IL-4, and IL-12) can in turn cause severe inflammation (97). Moreover, anti-inflammatory agents can reduce skin *S. aureus* colonization and is recommended for AE control.

*Malassezia* is a monophyletic genus of fungi that belongs to the normal cutaneous flora. Fourteen species are currently recognized, among which *M. sympodialis* has been reported to be associated with AE and can also cause systemic infections (98). Of adult AE patients, 30%–80% are reactive to *M. sympodialis* in terms of specific IgE and T cell reactivity. Products, such as zymosan, can be recognized by TLR2 and activate mast cells, leading to the release of potent

inflammatory mediators, such as histamine, proteases, chemotactic factors, cytokines, and arachidonic acid metabolites (99). In addition, *M. sympodialis* can activate mast cells to release cysteinyl leukotrienes, enhance the mast cell IgE response, modulate MAPK activation, and alter IL-6 production by signaling through the TLR2/MyD88 pathway. Thus, it may have effects on inflammation and itching in AE (100,101).

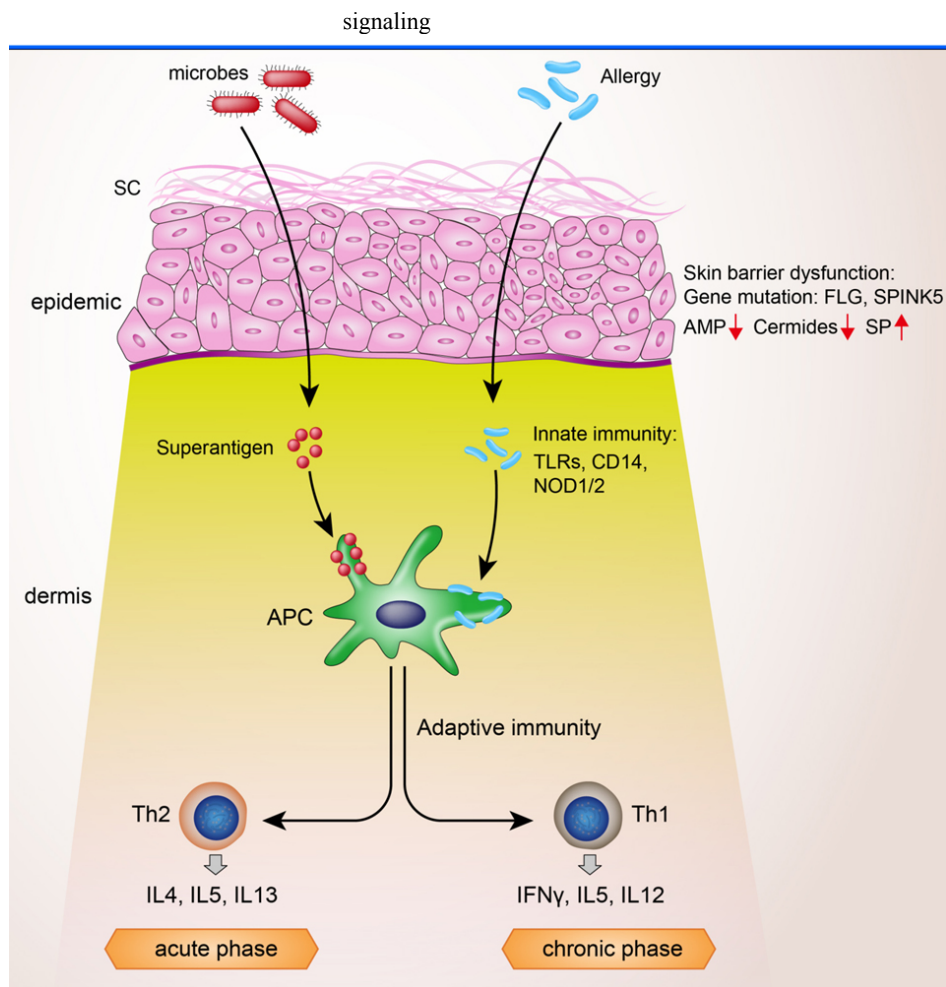
In contrast, extracellular vesicles secreted by *M. sympodialis* containing antigens and allergens from the fungi can induce a significantly higher IL-4 response in AE patients. All of this evidence indicates *M. sympodialis* may have a role in pathogenesis and severity of AE (102).

In addition, clinical studies have revealed that >50% of all children with AE can experience exacerbations triggered by certain foods (103–105). Food allergy and AE often occur in the same patient. While different foods affect people differently, it has been shown that foods, such as cow's milk and hen's eggs, can directly provoke flares of AE, particularly in sensitized infants, whereas inhaled allergens and pollen-related foods are of greater importance in older children, adolescents, and adults. Three patterns of cutaneous reactions to food may occur in patients with AE upon oral challenge. The first pattern commonly occurs a few minutes after ingestion of food, without exacerbation of AE, with the onset of gastrointestinal, respiratory, and cardiovascular symptoms. In the second pattern, pruritus occurs soon after ingestion of food, with subsequent scratching leading to an exacerbation of AE. In the third pattern, exacerbations of AE occur after 6–48 h; these exacerbations are termed late reactions (106–109). Reliable markers for the identification of patients with food-responsive eczema are still lacking. Based on a straightforward history, diagnosis of immediate symptoms provoked by a food may be evident, which is further confirmed by diagnostic tests to detect food-specific IgE antibody. Determination of the role played by food allergy in patients with AE is more difficult and may require additional diagnostic maneuvers, including elimination diets and oral food challenges (107,110). Further investigations and clinical studies need to be conducted to clarify the relationships between foods and AE.

### 6. CONCLUSION

AE is a common skin inflammatory disease with complex genetic and environmental factors that affects an increasing number of people worldwide. Fortunately, efforts from scientists have provided us much evidence in understanding this disease. Gene mutations, skin barrier abnormalities, dysfunction of the innate and adaptive immune systems, microbes, and allergens are important factors for the development and exacerbation of AE (Figure 1). These findings not only allow us to develop a precise definition of AE, but also have a great impact on clinical therapy. However, more studies are needed to discover interactions between those factors and the

subsequent



**Figure 1.** Important factors for the development and exacerbation of AE: Gene mutations, skin barrier abnormalities, dysfunction of the innate and adaptive immune systems, microbes, and allergens.

transduction pathways. It is hoped that specific biomarkers can be identified to reflect the detailed pathogenesis for AE, which is important in providing an early diagnostic strategy and targeted therapy for affected individuals.

## 7. ACKNOWLEDGEMENTS

This work was supported by the TCM-Integrated Key Disease Building Project (Eczema) of Shanghai Health Bureau (zxbz2012-04).

## 8. REFERENCES

1. Leung DY, Boguniewicz M, Howell MD, Nomura I, Hamid QA. New insights into atopic dermatitis. *J Clin Invest* 113, 651-657. (2004 )
2. Schultz Larsen F. Atopic dermatitis: a genetic-epidemiologic study in a population-based twin sample. *J Am Acad Dermatol* 28, 719-723. (1993 )

3. Lau S. Oral application of bacterial lysate in infancy diminishes the prevalence of atopic dermatitis in children at risk for atopy. *Benef Microbes* 1-3. (2013)
4. Hon KL, Wang SS, Leung TF. The atopic march: from skin to the airways. *Iran J Allergy Asthma Immunol* 11, 73-77. (2012)
5. Spergel JM. From atopic dermatitis to asthma: the atopic march. *Ann Allergy Asthma Immunol* 105:99-106; quiz 107-109, 117. (2010)
6. Spergel JM. Epidemiology of atopic dermatitis and atopic march in children. *Immunol Allergy Clin North Am* 30, 269-280. (2010)
7. Spergel JM, Paller AS. Atopic dermatitis and the atopic march. *J Allergy Clin Immunol* 112, S118-127. (2003)
8. Caffarelli C, Dondi A, Povesi Dascola C, Ricci G. Skin prick test to foods in childhood atopic eczema: pros and cons. *Ital J Pediatr* 39, 48. (2013)

9. Penders J, Gerhold K, Stobberingh EE, Thijs C, Zimmermann K, Lau S, Hamelmann E. Establishment of the intestinal microbiota and its role for atopic dermatitis in early childhood. *J Allergy Clin Immunol* 132(3), 601-607.e8 (2013)
10. Williams HC, Burney PG, Pembroke AC, Hay RJ. The U.K. Working Party's Diagnostic Criteria for Atopic Dermatitis. III. Independent hospital validation. *Br J Dermatol* 131, 406-416. (1994)
11. Johansson SG, Bieber T, Dahl R, Friedmann PS, Lanier BQ, Lockey RF, Motala C, Ortega Martell JA, Platts-Mills TA, Ring J, Thien F, Van Cauwenberge P, Williams HC. Revised nomenclature for allergy for global use: Report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. *J Allergy Clin Immunol* 113, 832-836. (2004)
12. Roguedas-Contios AM, Misery L. What is intrinsic atopic dermatitis? *Clin Rev Allergy Immunol* 41, 233-236. (2011)
13. Williams HC, Johansson SG. Two types of eczema--or are there? *J Allergy Clin Immunol* 116, 1064-1066. (2005)
14. O'Regan GM, Irvine AD. The role of filaggrin loss-of-function mutations in atopic dermatitis. *Curr Opin Allergy Clin Immunol* 8, 406-410. (2008)
15. O'Regan GM, Sandilands A, McLean WH, Irvine AD. Filaggrin in atopic dermatitis. *J Allergy Clin Immunol* 122, 689-693. (2008)
16. Osawa R, Akiyama M, Shimizu H. Filaggrin gene defects and the risk of developing allergic disorders. *Allergol Int* 60, 1-9. (2011)
17. van den Oord RA, Sheikh A. Filaggrin gene defects and risk of developing allergic sensitisation and allergic disorders, systematic review and meta-analysis. *BMJ* 339, b2433. (2009)
18. Auriemma M, Vianale G, Amerio P, Reale M. Cytokines and T cells in atopic dermatitis. *Eur Cytokine Netw* 24, 37-44. (2013)
19. Bao L, Shi VY, Chan LS. IL-4 up-regulates epidermal chemotactic, angiogenic, and pro-inflammatory genes and down-regulates antimicrobial genes in vivo and in vitro: relevant in the pathogenesis of atopic dermatitis. *Cytokine* 61, 419-425. (2013)
20. Kaminishi K, Soma Y, Kawa Y, Mizoguchi M. Flow cytometric analysis of IL-4, IL-13 and IFN-gamma expression in peripheral blood mononuclear cells and detection of circulating IL-13 in patients with atopic dermatitis provide evidence for the involvement of type 2 cytokines in the disease. *J Dermatol Sci* 29, 19-25. (2002)
21. Liu FT, Goodarzi H, Chen HY. IgE, mast cells, and eosinophils in atopic dermatitis. *Clin Rev Allergy Immunol* 41, 298-310. (2011)
22. Oppel T, Schuller E, Gunther S, Moderer M, Haberstok J, Bieber T, Wollenberg A. Phenotyping of epidermal dendritic cells allows the differentiation between extrinsic and intrinsic forms of atopic dermatitis. *Br J Dermatol* 143, 1193-1198. (2000)
23. Callesen M, Beko G, Weschler CJ, Sigsgaard T, Jensen TK, Clausen G, Toftum J, Norberg LA, Host A. Associations between selected allergens, phthalates, nicotine, PAHs and bedroom ventilation and clinically confirmed asthma, rhinoconjunctivitis and atopic dermatitis in preschool children. *Indoor Air*.. (2013)
24. Wadonda-Kabondo N, Sterne JA, Golding J, Kennedy CT, Archer CB, Dunnill MG. Association of parental eczema, hayfever, and asthma with atopic dermatitis in infancy: birth cohort study. *Arch Dis Child* 89, 917-921. (2004)
25. Samochocki Z, Dejewski J. A comparison of criteria for diagnosis of atopic dermatitis in children. *World J Pediatr* 8, 355-358. (2012)
26. Hilete M. Evaluation of Hanifin and Rajka atopic eczema diagnostic guidelines for reduced minor criteria. *Ethiop Med J* 47, 39-47. (2009)
27. Rudzki E, Samochocki Z, Rebandel P, Saciuk E, Galecki W, Raczka A, Szmurlo A. Frequency and significance of the major and minor features of Hanifin and Rajka among patients with atopic dermatitis. *Dermatology* 189, 41-46. (1994)
28. Asher MI, Keil U, Anderson HR, Beasley R, Crane J, Martinez F, Mitchell EA, Pearce N, Sibbald B, Stewart AW, et al. International Study of Asthma and Allergies in Childhood (ISAAC): rationale and methods. *Eur Respir J* 8, 483-491. (1995)
29. Asher MI, Weiland SK. The International Study of Asthma and Allergies in Childhood (ISAAC). ISAAC Steering Committee. *Clin Exp Allergy* 28 Suppl 5, 52-66; discussion 90-51. (1998)
30. Mallol J, Crane J, von Mutius E, Odhiambo J, Keil U, Stewart A. The International Study of Asthma and Allergies in Childhood (ISAAC) Phase Three: a global synthesis. *Allergol Immunopathol (Madr)* 41, 73-85. (2013)
31. Sole D, Vanna AT, Yamada E, Rizzo MC, Nasipitz CK. International Study of Asthma and Allergies in Childhood (ISAAC) written questionnaire: validation of the asthma component among Brazilian children. *J Invest Allergol Clin Immunol* 8, 376-382. (1998)
32. Levy RM, Gelfand JM, Yan AC. The epidemiology of atopic dermatitis. *Clin Dermatol* 21, 109-115. (2003)

33. Williams H, Robertson C, Stewart A, Ait-Khaled N, Anabwani G, Anderson R, Asher I, Beasley R, Bjorksten B, Burr M, Clayton T, Crane J, Ellwood P, Keil U, Lai C, Mallol J, Martinez F, Mitchell E, Montefort S, Pearce N, Shah J, Sibbald B, Strachan D, von Mutius E, Weiland SK. Worldwide variations in the prevalence of symptoms of atopic eczema in the International Study of Asthma and Allergies in Childhood. *J Allergy Clin Immunol* 103, 125-138. (1999)
34. Oranje AP, Stalder JF, Taieb A, Tasset C, de Longueville M. Scoring of atopic dermatitis by SCORAD using a training atlas by investigators from different disciplines. ETAC Study Group. Early Treatment of the Atopic Child. *Pediatr Allergy Immunol* 8, 28-34. (1997)
35. Oh JW, Pyun BY, Choung JT, Ahn KM, Kim CH, Song SW, Son JA, Lee SY, Lee SI. Epidemiological change of atopic dermatitis and food allergy in school-aged children in Korea between 1995 and 2000. *J Korean Med Sci* 19, 716-723. (2004)
36. Larsen FS, Holm NV, Henningsen K. Atopic dermatitis. A genetic-epidemiologic study in a population-based twin sample. *J Am Acad Dermatol* 15, 487-494. (1986)
37. Compton JG, DiGiovanna JJ, Johnston KA, Fleckman P, Bale SJ. Mapping of the associated phenotype of an absent granular layer in ichthyosis vulgaris to the epidermal differentiation complex on chromosome 1. *Exp Dermatol* 11, 518-526. (2002)
38. Gan SQ, McBride OW, Idler WW, Markova N, Steinert PM. Organization, structure, and polymorphisms of the human profilaggrin gene. *Biochemistry* 30, 5814. (1991)
39. Rawlings AV, Harding CR. Moisturization and skin barrier function. *Dermatol Ther* 17 Suppl 1, 43-48. (2004)
40. Sandilands A, Sutherland C, Irvine AD, McLean WH. Filaggrin in the frontline, role in skin barrier function and disease. *J Cell Sci* 122, 1285-1294. (2009)
41. Sandilands A, Terron-Kwiatkowski A, Hull PR, O'Regan GM, Clayton TH, Watson RM, Carrick T, Evans AT, Liao H, Zhao Y, Campbell LE, Schmuth M, Gruber R, Janecke AR, Elias PM, van Steensel MA, Nagtzaam I, van Geel M, Steijlen PM, Munro CS, Bradley DG, Palmer CN, Smith FJ, McLean WH, Irvine AD. Comprehensive analysis of the gene encoding filaggrin uncovers prevalent and rare mutations in ichthyosis vulgaris and atopic eczema. *Nat Genet* 39, 650-654. (2007)
42. Brown SJ, McLean WH. One remarkable molecule: filaggrin. *J Invest Dermatol* 132, 751-762. (2012)
43. Brown SJ, Relton CL, Liao H, Zhao Y, Sandilands A, McLean WH, Cordell HJ, Reynolds NJ. Filaggrin haploinsufficiency is highly penetrant and is associated with increased severity of eczema: further delineation of the skin phenotype in a prospective epidemiological study of 792 school children. *Br J Dermatol* 161, 884-889. (2009)
44. Smith FJ, Irvine AD, Terron-Kwiatkowski A, Sandilands A, Campbell LE, Zhao Y, Liao H, Evans AT, Goudie DR, Lewis-Jones S, Arsecularatne G, Munro CS, Sergeant A, O'Regan G, Bale SJ, Compton JG, DiGiovanna JJ, Presland RB, Fleckman P, McLean WH. Loss-of-function mutations in the gene encoding filaggrin cause ichthyosis vulgaris. *Nat Genet* 38, 337-342. (2006)
45. Stemmler S, Parwez Q, Petrasch-Parwez E, Epplen JT, Hoffjan S. Two common loss-of-function mutations within the filaggrin gene predispose for early onset of atopic dermatitis. *J Invest Dermatol* 127, 722-724. (2007)
46. Weidinger S, Illig T, Baurecht H, Irvine AD, Rodriguez E, Diaz-Lacava A, Klopp N, Wagenpfeil S, Zhao Y, Liao H, Lee SP, Palmer CN, Jenneck C, Maintz L, Hagemann T, Behrendt H, Ring J, Nothen MM, McLean WH, Novak N. Loss-of-function variations within the filaggrin gene predispose for atopic dermatitis with allergic sensitizations. *J Allergy Clin Immunol* 118, 214-219. (2006)
47. Micheal S, Minhas K, Ishaque M, Ahmed F, Ahmed A. IL-4 gene polymorphisms and their association with atopic asthma and allergic rhinitis in Pakistani patients. *J Invest Allergol Clin Immunol* 23, 107-111. (2013)
48. Namkung JH, Lee JE, Kim E, Kim HJ, Seo EY, Jang HY, Shin ES, Cho EY, Yang JM. Association of polymorphisms in genes encoding IL-4, IL-13 and their receptors with atopic dermatitis in a Korean population. *Exp Dermatol* 20, 915-919. (2011)
49. Goenka S, Kaplan MH. Transcriptional regulation by STAT6. *Immunol Res* 50, 87-96. (2011)
50. Hebenstreit D, Wirsberger G, Horejs-Hoeck J, Duschl A. Signaling mechanisms, interaction partners, and target genes of STAT6. *Cytokine Growth Factor Rev* 17, 173-188. (2006)
51. Kelly-Welch AE, Hanson EM, Boothby MR, Keegan AD. Interleukin-4 and interleukin-13 signaling connections maps. *Science* 300, 1527-1528. (2003)
52. Vladich FD, Brazille SM, Stern D, Peck ML, Ghittoni R, Vercelli D. IL-13 R130Q, a common variant associated with allergy and asthma, enhances effector mechanisms essential for human allergic inflammation. *J Clin Invest* 115, 747-754. (2005)
53. Imokawa G, Yada Y, Higuchi K, Okuda M, Ohashi Y, Kawamata A. Pseudo-acylceramide with linoleic acid produces selective recovery of diminished cutaneous barrier function in essential fatty acid-deficient rats and has an inhibitory effect on epidermal hyperplasia. *J Clin Invest* 94, 89-96. (1994)
54. Joo KM, Nam GW, Park SY, Han JY, Jeong HJ, Lee SY, Kim HK, Lim KM. Relationship between cutaneous



barrier function and ceramide species in human stratum corneum. *J Dermatol Sci* 60, 47-50. (2010)

55. Masukawa Y, Narita H, Sato H, Naoe A, Kondo N, Sugai Y, Oba T, Homma R, Ishikawa J, Takagi Y, Kitahara T. Comprehensive quantification of ceramide species in human stratum corneum. *J Lipid Res* 50, 1708-1719. (2009)

56. Hara J, Higuchi K, Okamoto R, Kawashima M, Imokawa G. High-expression of sphingomyelin deacylase is an important determinant of ceramide deficiency leading to barrier disruption in atopic dermatitis. *J Invest Dermatol* 115, 406-413. (2000)

57. Williams JV, Vowels B, Honig P, Leyden JJ. Staphylococcus aureus isolation from the lesions, the hands, and anterior nares of patients with atopic dermatitis. *J Emerg Med* 17, 207-211. (1999)

58. Candi E, Schmidt R, Melino G. The cornified envelope: a model of cell death in the skin. *Nat Rev Mol Cell Biol* 6, 328-340. (2005)

59. Hachem JP, Man MQ, Crumrine D, Uchida Y, Brown BE, Rogiers V, Roseeuw D, Feingold KR, Elias PM. Sustained serine proteases activity by prolonged increase in pH leads to degradation of lipid processing enzymes and profound alterations of barrier function and stratum corneum integrity. *J Invest Dermatol* 125, 510-520. (2005)

60. Vasilopoulos Y, Cork MJ, Murphy R, Williams HC, Robinson DA, Duff GW, Ward SJ, Tazi-Ahnini R. Genetic association between an AACC insertion in the 3'UTR of the stratum corneum chymotryptic enzyme gene and atopic dermatitis. *J Invest Dermatol* 123, 62-66. (2004)

61. Rance F, Boguniewicz M, Lau S. New visions for atopic eczema: an iPAC summary and future trends. *Pediatr Allergy Immunol* 19 Suppl 19, 17-25. (2008)

62. Chavanas S, Bodemer C, Rochat A, Hamel-Teillac D, Ali M, Irvine AD, Bonafe JL, Wilkinson J, Taieb A, Barrandon Y, Harper JJ, de Prost Y, Hovnanian A. Mutations in SPINK5, encoding a serine protease inhibitor, cause Netherton syndrome. *Nat Genet* 25, 141-142. (2000)

63. Lan CC, Tu HP, Wu CS, Ko YC, Yu HS, Lu YW, Li WC, Chen YC, Chen GS. Distinct SPINK5 and IL-31 polymorphisms are associated with atopic eczema and non-atopic hand dermatitis in Taiwanese nursing population. *Exp Dermatol*. 20, 975-979. (2011;)

64. Namkung JH, Lee JE, Kim E, Byun JY, Kim S, Shin ES, Cho EY, Yang JM. Hint for association of single nucleotide polymorphisms and haplotype in SPINK5 gene with atopic dermatitis in Koreans. *Exp Dermatol* 19, 1048-1053. (2010)

65. Elias PM. Stratum corneum defensive functions: an integrated view. *J Invest Dermatol* 125, 183-200. (2005)

66. Esche C, Stellato C, Beck LA. Chemokines: key players in innate and adaptive immunity. *J Invest Dermatol* 125, 615-628. (2005)

67. McGirt LY, Beck LA. Innate immune defects in atopic dermatitis. *J Allergy Clin Immunol* 118, 202-208. (2006)

68. Ahmad-Nejad P, Mrabet-Dahbi S, Breuer K, Klotz M, Werfel T, Herz U, Heeg K, Neumaier M, Renz H. The toll-like receptor 2 R753Q polymorphism defines a subgroup of patients with atopic dermatitis having severe phenotype. *J Allergy Clin Immunol* 113, 565-567. (2004)

69. Gergely P, Jr., Blazsek A, Weiszhar Z, Pazar B, Poor G. Lack of genetic association of the Toll-like receptor 4 (TLR4) Asp299Gly and Thr399Ile polymorphisms with spondylarthropathies in a Hungarian population. *Rheumatology (Oxford)* 45, 1194-1196. (2006)

70. Xiong Y, Song C, Snyder GA, Sundberg EJ, Medvedev AE. R753Q polymorphism inhibits Toll-like receptor (TLR) 2 tyrosine phosphorylation, dimerization with TLR6, and recruitment of myeloid differentiation primary response protein 88. *J Biol Chem* 287, 38327-38337. (2012)

71. Niebuhr M, Langnickel J, Draing C, Renz H, Kapp A, Werfel T. Dysregulation of toll-like receptor-2 (TLR-2)-induced effects in monocytes from patients with atopic dermatitis: impact of the TLR-2 R753Q polymorphism. *Allergy* 63, 728-734. (2008)

72. Oh DY, Schumann RR, Hamann L, Neumann K, Worm M, Heine G. Association of the toll-like receptor 2 A-16934T promoter polymorphism with severe atopic dermatitis. *Allergy* 64, 1608-1615. (2009)

73. Niebuhr M, Lutat C, Sigel S, Werfel T. Impaired TLR-2 expression and TLR-2-mediated cytokine secretion in macrophages from patients with atopic dermatitis. *Allergy* 64, 1580-1587. (2009)

74. Nolte T, Zadeh-Khorasani M, Safarov O, Rueff F, Varga R, Herbach N, Wanke R, Wollenberg A, Mueller T, Gropp R, Wolf E, Siebeck M. Induction of oxazolone-mediated features of atopic dermatitis in NOD-scid IL2Rgamma(null) mice engrafted with human peripheral blood mononuclear cells. *Dis Model Mech* 6, 125-134. (2013)

75. Voron'ko OE, Dmitrieva-Zdorova EV, Latysheva EA, Aksenova MG, Storozhakov GI, Bodoev NV, Archakov AI. [CARD15 and TLR4 genes polymorphisms in atopic bronchial asthma]. *Mol Biol (Mosk)* 45, 831-839. (2011)

76. Weidinger S, Klopp N, Rummeler L, Wagenpfeil S, Novak N, Baurecht HJ, Groer W, Darsow U, Heinrich J, Gauger A, Schafer T, Jakob T, Behrendt H, Wichmann HE, Ring J, Illig T. Association of NOD1 polymorphisms with atopic eczema and related phenotypes. *J Allergy Clin Immunol* 116, 177-184. (2005)

77. Carrera MC, Moura P, Crovella S, de Souza PR, de Alencar LC, Sarinho E. High polymorphism of the MBL2 gene in patients with atopic dermatitis. *Ann Allergy Asthma Immunol* 105, 39-42. (2010)
78. Sinclair C, Bains I, Yates AJ, Seddon B. Asymmetric thymocyte death underlies the CD4:CD8 T-cell ratio in the adaptive immune system. *Proc Natl Acad Sci U S A* 110, E2905-2914. (2013)
79. Schenten D, Medzhitov R. The control of adaptive immune responses by the innate immune system. *Adv Immunol* 109, 87-124. (2011)
80. de Brito CA, Goldoni AL, Sato MN. Immune adjuvants in early life: targeting the innate immune system to overcome impaired adaptive response. *Immunotherapy* 1, 883-895. (2009)
81. Namkoong H, Song MY, Seo YB, Choi DH, Kim SW, Im SJ, Sung YC, Park Y. Enhancement of antigen-specific CD8 T cell responses by co-delivery of Fc-fused CXCL11. Vaccine pii: S0264-410X(13)01052-9 (2013)
82. van Essen D. Long-range control of T-cell development. *Blood* 122, 854-856. (2013)
83. Kondo H, Ichikawa Y, Imokawa G. Percutaneous sensitization with allergens through barrier-disrupted skin elicits a Th2-dominant cytokine response. *Eur J Immunol* 28, 769-779. (1998)
84. Matsui K, Nishikawa A. Lipoteichoic acid from *Staphylococcus aureus* induces Th2-prone dermatitis in mice sensitized percutaneously with an allergen. *Clin Exp Allergy* 32, 783-788. (2002)
85. Maintz L, Novak N. Getting more and more complex: the pathophysiology of atopic eczema. *Eur J Dermatol* 17, 267-283. (2007)
86. Kimura M, Tsuruta S, Yoshida T. Unique profile of IL-4 and IFN-gamma production by peripheral blood mononuclear cells in infants with atopic dermatitis. *J Allergy Clin Immunol* 102, 238-244. (1998)
87. Kimura M, Tsuruta S, Yoshida T. Correlation of house dust mite-specific lymphocyte proliferation with IL-5 production, eosinophilia, and the severity of symptoms in infants with atopic dermatitis. *J Allergy Clin Immunol* 101, 84-89. (1998)
88. Groux H, O'Garra A, Bigler M, Rouleau M, Antonenko S, de Vries JE, Roncarolo MG. A CD4+ T-cell subset inhibits antigen-specific T-cell responses and prevents colitis. *Nature* 389, 737-742. (1997)
89. Akdis M, Verhagen J, Taylor A, Karamloo F, Karagiannidis C, Cramer R, Thunberg S, Deniz G, Valenta R, Fiebig H, Kegel C, Disch R, Schmidt-Weber CB, Blaser K, Akdis CA. Immune responses in healthy and allergic individuals are characterized by a fine balance between allergen-specific T regulatory 1 and T helper 2 cells. *J Exp Med* 199, 1567-1575. (2004)
90. Wahl SM, Vazquez N, Chen W. Regulatory T cells and transcription factors: gatekeepers in allergic inflammation. *Curr Opin Immunol* 16, 768-774. (2004)
91. Taylor A, Verhagen J, Akdis CA, Akdis M. T regulatory cells in allergy and health: a question of allergen specificity and balance. *Int Arch Allergy Immunol* 135, 73-82. (2004)
92. Baker BS. The role of microorganisms in atopic dermatitis. *Clin Exp Immunol* 144, 1-9. (2006)
93. Voorhees T, Chang J, Yao Y, Kaplan MH, Chang CH, Travers JB. Dendritic cells produce inflammatory cytokines in response to bacterial products from *Staphylococcus aureus*-infected atopic dermatitis lesions. *Cell Immunol* 267, 17-22. (2011)
94. Kozman A, Yao Y, Bina P, Saha C, Yao W, Kaplan MH, Travers JB. Encoding a superantigen by *Staphylococcus aureus* does not affect clinical characteristics of infected atopic dermatitis lesions. *Br J Dermatol* 163, 1308-1311. (2010)
95. Leyden JJ, Marples RR, Kligman AM. *Staphylococcus aureus* in the lesions of atopic dermatitis. *Br J Dermatol* 90, 525-530. (1974)
96. Lin YT, Wang CT, Chiang BL. Role of bacterial pathogens in atopic dermatitis. *Clin Rev Allergy Immunol* 33, 167-177. (2007)
97. Nada HA, Gomaa NI, Elakhras A, Wasfy R, Baker RA. Skin colonization by superantigen-producing *Staphylococcus aureus* in Egyptian patients with atopic dermatitis and its relation to disease severity and serum interleukin-4 level. *Int J Infect Dis* 16, e29-33. (2012)
98. Saunders CW, Scheynius A, Heitman J. *Malassezia* fungi are specialized to live on skin and associated with dandruff, eczema, and other skin diseases. *PLoS Pathog* 8, e1002701. (2012)
99. Yim SM, Kim JY, Ko JH, Lee YW, Choe YB, Ahn KJ. Molecular analysis of *malassezia* microflora on the skin of the patients with atopic dermatitis. *Ann Dermatol* 22, 41-47. (2010)
100. Aleyas AG, Han YW, Patil AM, Kim SB, Kim K, Eo SK. Impaired cross-presentation of CD8alpha+ CD11c+ dendritic cells by Japanese encephalitis virus in a TLR2/MyD88 signal pathway-dependent manner. *Eur J Immunol* 42, 2655-2666. (2012)
101. Selander C, Engblom C, Nilsson G, Scheynius A, Andersson CL. TLR2/MyD88-dependent and -independent activation of mast cell IgE responses by the skin

## Atopic eczema

commensal yeast *Malassezia sympodialis*. *J Immunol* 182, 4208-4216. (2009)

102. Gehrmann U, Qazi KR, Johansson C, Hultenby K, Karlsson M, Lundeberg L, Gabrielsson S, Scheynius A. Nanovesicles from *Malassezia sympodialis* and host exosomes induce cytokine responses--novel mechanisms for host-microbe interactions in atopic eczema. *PLoS One* 6, e21480. (2011)

103. Bohle B. The impact of pollen-related food allergens on pollen allergy. *Allergy* 62, 3-10. (2007)

104. Bollinger ME, Dahlquist LM, Mudd K, Sonntag C, Dillinger L, McKenna K. The impact of food allergy on the daily activities of children and their families. *Ann Allergy Asthma Immunol* 96, 415-421. (2006)

105. Pajno GB, Morabito L, Barberio G. Allergy to house dust mite and snails: a model of cross-reaction between food and inhalant allergens with a clinical impact. *Pediatr Pulmonol Suppl* 18, 163-164. (1999)

106. Kijima A, Murota H, Takahashi A, Arase N, Yang L, Nishioka M, Yamaoka T, Kitaba S, Yamauchi-Takahara K, Katayama I. Prevalence and impact of past history of food allergy in atopic dermatitis. *Allergol Int* 62, 105-112. (2013)

107. Oppenheimer J, Bender B. The impact of food allergy and bullying. *Ann Allergy Asthma Immunol* 105, 410-411. (2010)

108. Herman EM, Burks AW. The impact of plant biotechnology on food allergy. *Curr Opin Biotechnol* 22, 224-230. (2011)

109. Jones SM, Scurlock AM. The impact of food allergy: the real "fear factor". *Ann Allergy Asthma Immunol* 96, 385-386. (2006)

110. Nguyen M, Wainstein BK, Hu W, Ziegler JB. Parental satisfaction with oral peanut food challenges; perception of outcomes and impact on management of peanut allergy. *Pediatr Allergy Immunol* 21, 1119-1126. (2010)

**Abbreviations:** AE, atopic eczema; AD, atopic dermatitis; IgE, immunoglobulin E; IL, interleukin; ISAAC, International Study of Asthma and Allergies in Childhood; SCORAD, Scoring of Atopic Dermatitis; SNPs, single nucleotide polymorphisms; FLG, filaggrin gene; GM-CSF, granulocyte-macrophage colony-stimulating factor; IFN, interferon; PAR-2, protease-activated receptor-2; SPINK5, Karzai type 5; SCCE, stratum corneum chymotryptic enzyme; SCTE, stratum corneum tryptic enzyme; SCCL, stratum corneum cathepsin-L-like enzyme; PRRs, pattern-recognition receptors; PAMPs, pathogen-associated molecular patterns; AMPs, antimicrobial peptides; PGN, peptidoglycan; LTA, lipoteichoic acid; TLRs, toll-like receptors; CLRs, C-type lectin receptors; DCs, dendritic cells; DAMPs, damage-associated molecular patterns; NF- $\kappa$ B, nuclear factor  $\kappa$ B; MyD88, myeloid differentiation

primary response gene-88; CARD, caspase activation and recruitment domain; HBD, human  $\beta$ -defensin; MBL, mannan-binding lectin; MHC, major histocompatibility complex; TNF, tumor necrosis factor

**Key Words:** Atopic eczema, Skin Diseases, Genetic Factors, Immune System Abnormalities, Microbes, Review

**Send correspondence to:** Weian Mao, Department of Dermatology, Shanghai Seventh People's Hospital, 358 Datong Road, Pudong New Area, Shanghai 200137, China, Tel: 86-18930837777, Fax: 86-21-51323092, E-mail: Weian\_Mao@yeah.net